

Synthesis, Conformational Analysis, and Complexation Study of an Iminosugar-Aza-Crown, a Sweet Chiral Cyclam Analog

Alexandra Bordes, Ana Poveda, Thibault Troadec, Antonio Franconetti, Ana Ardá, Flavie Perrin, Mickaël Ménand, Matthieu Sollogoub, Jérôme Guillard, Jérôme Désiré, Raphaël Tripiet, Jesús Jiménez-Barbero,* and Yves Blériot*



Cite This: <https://dx.doi.org/10.1021/acs.orglett.0c00503>



Read Online

ACCESS |



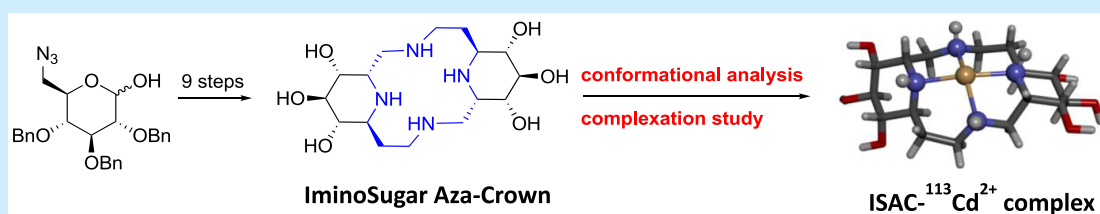
Metrics & More



Article Recommendations



Supporting Information



ABSTRACT: A new family of chiral C_2 symmetric tetraazamacrocycles, coined ISAC for IminoSugar Aza-Crown, incorporating two iminosugars adopting a 4C_1 conformation is disclosed. Multinuclear NMR experiments on the corresponding Cd^{2+} complex show that the ISAC is a strong chelator in water and its tetramine cavity adopts a conformation similar to that of the parent Cd -cyclam complex. Similar behavior is observed with Cu^{2+} in solution, with enhanced stability compared to the Cu -cyclam complex.

Oxygen- and nitrogen-based macrocycles, often referred to as “crowns”, have been long known for their coordinating abilities toward anions and cations. In this context, sugar-based macrocyclic compounds are of high interest and demonstrated biological, environmental, and medical properties.¹ Several advantages are associated with their glycosidic nature including their availability from natural sources and their polyhydroxylated pattern allowing the modulation of their physicochemical properties. Furthermore, their chirality combined with the geometric constraints brought by the five- or six-membered sugar rings is useful for the design of conformationally restricted chiral cavities. Thus, the design of sugar-based macrocycles bearing a combination of O- and N-donors is appealing and a vast array has been reported,² incorporating furanoses, pyranoses, and disaccharides. Depending on the size of the macrocycle and/or the number of oxygen and nitrogen donor atoms, different guests can be accommodated. Large 15- to 19-membered rings bearing up to three nitrogen atoms (1–3, Figure 1) have been used as excellent ligands for large and/or hard (according to HSAB theory) guests, such as amino acids³ or ammonium^{4–7} and alkali metal cations.⁸ Smaller 14-membered rings with two oxygen and two nitrogen donors (4–5, Figure 1) have been designed to accommodate softer or smaller transition metal cations, such as cadmium(II), mercury(II), and copper(II).^{9–11}

Tetra-azamacrocycles, such as cyclam (1,4,8,11-tetraazacyclotetradecane), constitute another specific class of ligands that have been extensively studied for their complexation properties.^{12,13} They have been exploited for the selective

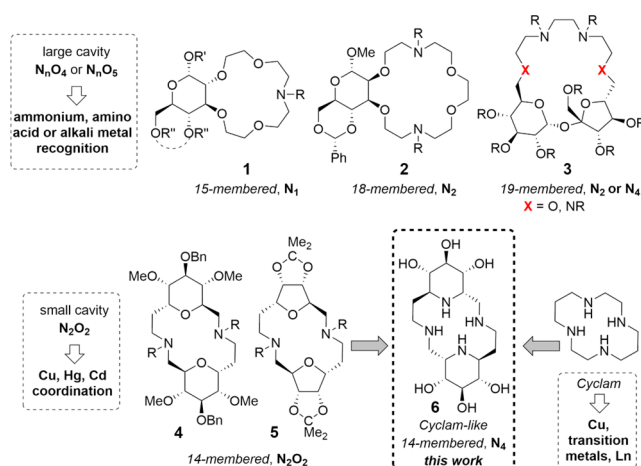


Figure 1. Structures and targeted guests of known and new polyaza-sugar-based macrocycles 1–6.

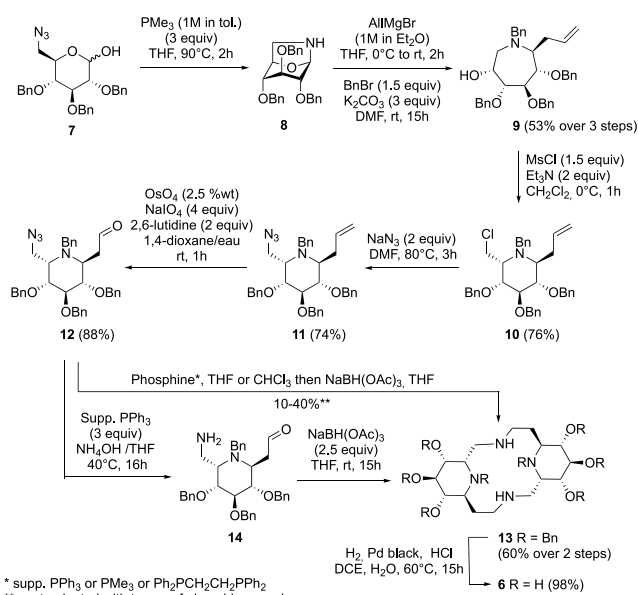
binding of transition metals, heavy metals, and lanthanides, especially after their mono- or poly-N-functionalization. In particular, cyclam is one of the azamacrocycles that has been

Received: February 11, 2020

advantageously employed during these past decades in extraction,¹⁴ catalysis,^{15–17} supramolecular,¹⁸ sensing,¹⁹ bio-inorganic,²⁰ and medicinal chemistry.²¹ Although they have been decorated with appended carbohydrates on side chains, for formulation or medical imaging purposes,^{22–24} sugar motifs have never been, to the best of our knowledge, directly included within the skeleton of such polyazamacrocycles. In order to take the best of both worlds, we postulated that incorporation of iminosugars, sugar analogues in which the endocyclic oxygen has been replaced by nitrogen, could be of interest to generate new “sweet” constrained cyclam derivatives, which physicochemical properties could be tuned thanks to their polyhydroxylated nature. Iminosugars have thus far been almost exclusively developed as therapeutic candidates^{25,26} targeting carbohydrate processing enzymes. Herein, we report the synthesis, conformational analysis, and preliminary ion-binding study of a new nitrogen-containing sugar-based macrocycle coined iminosugar azacrown (ISAC) **6** (Figure 1).

To build the tetraazamacrocyclic framework, we explored an intermolecular macrocyclization, relying either on a reductive amination or on a Staudinger-aza-Wittig (SAW) reaction, a strategy successfully applied for the construction of sugar-azacrown (SAC) ethers.^{11,27} The required iminosugar-based azidoaldehyde **7**²⁸ was submitted to a SAW reaction using PMe_3 ²⁹ to provide the corresponding bicyclic hemiaminal **8**³⁰ that was directly treated with AlMgBr and *N*-benzylated to afford the hydroxyazepane **9**. Mesylation of the free OH in **9** provided the ring-contracted chlorinated piperidine **10**. A subsequent azide displacement yielded the azido-piperidine **11** in 74% yield. Finally, oxidative cleavage of the allylic moiety with $\text{OsO}_4/\text{NaIO}_4$ in the presence of 2,6-lutidine³¹ furnished the target azidoaldehyde **12** (Scheme 1), the structure of which is guided by the necessity to have relatively flexible arms provided by the two carbon and three carbon appendages to favor dimerization and disposed in a *trans*-relationship to avoid intramolecular coupling. The dimerization of **12** was examined next, the crude expected di-imine being further reduced with

Scheme 1. Synthesis of ISAC 6



$\text{NaBH}(\text{OAc})_3$. Two solvents (THF, CHCl_3) and three sources of phosphine (supported PPh_3 , PMe_3 , and $\text{Ph}_2\text{PCH}_2\text{CH}_2\text{PPh}_2$ ³²) were screened, all leading to the expected ISAC **13** albeit in poor yield (10–40%) that was contaminated with traces of phosphine species in all cases (Scheme 1). To gain insight into this unsatisfactory dimerization, the reaction was monitored by ^1H NMR in CDCl_3 at room temperature over a 4.5 h period using PPh_3 as the phosphine source (Figure 2). We observed the progressive

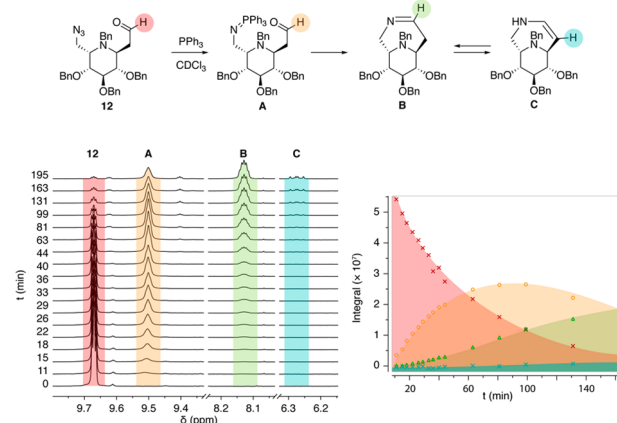


Figure 2. Monitoring of the Staudinger-aza-Wittig reaction of azidoaldehyde **12**. Top: Proposed reaction pathway for the Staudinger/Aza-Wittig reaction of azidoaldehyde **12**. Bottom: selected regions of the ^1H NMR monitoring (CDCl_3 , 400 MHz, 300 K) of the reaction of **12** with PPh_3 and graph of the integrals intensity variation during the reaction

consumption of the starting azidoaldehyde **12** (1H at 9.7 ppm) affording the transient iminophosphorane **A** (1H at 9.6 ppm). This latter converts to the bicyclic imine **B** (1H at 8.2 ppm) that slowly equilibrates with the detrimental bicyclic enamine **C** (1H at 6.3 ppm). The ESIMS analysis of the mixture after 195 min confirms the hypothesis, showing not only the different intermediates but also the expected cyclic imine dimer (see Supporting Information (SI)). These results forced us to examine the alternative reductive amination route. Azidoaldehyde **12** was reduced to the crude aminoaldehyde **14** that was directly engaged in a double reductive amination in the presence of $\text{NaBH}(\text{OAc})_3$ identified as the reagent of choice for imine reduction for such a motif.²⁷ Satisfyingly, the protected ISAC **13** was isolated in 60% yield after preparative HPLC purification, its hydrogenolysis affording the target ISAC **6** (Scheme 1).

The geometry in water of ISAC **6** was next studied through extensive NMR analysis, supported by molecular modeling protocols. The relative simplicity of its ^1H NMR spectrum associated with the presence of only one set of sugar protons confirmed its C_2 symmetry in the chemical shift NMR time scale. Both iminosugar rings adopts a chair conformation with the OH groups in the equatorial position as deduced from the observed large coupling constants ($J_{2,3} = J_{3,4} = J_{4,5} = 9.5$ Hz) (Figure 3A) and intraresidue NOEs (H1/H2, H2/H4, H3/H5) (see SI). Analysis of the coupling constants of the aminated arms connecting the iminosugar units suggested some flexibility around the C6–C7–N moiety. In fact, larger temperature coefficients were observed for the ^1H NMR signals at positions 7 and 8, while the rest of the ^1H NMR signals were barely altered (see SI). Moreover, medium size

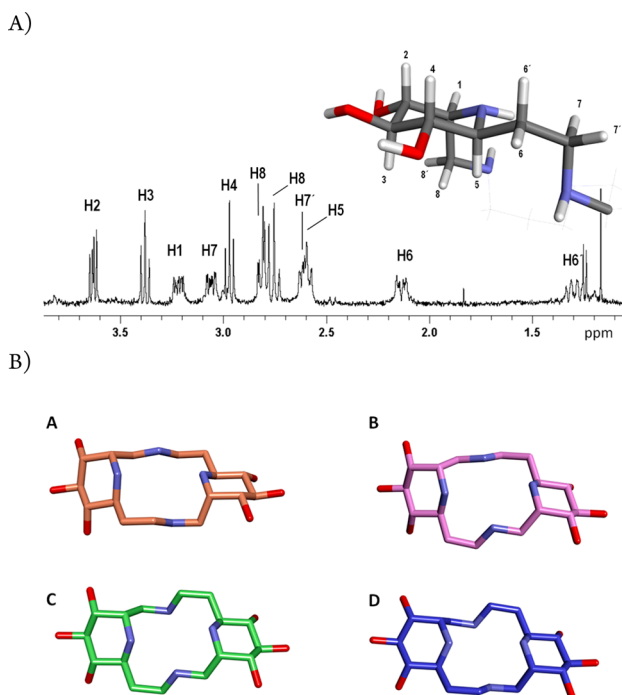


Figure 3. Conformational study of ISAC 6 in D₂O by NMR assisted by molecular mechanics. (A) ¹H NMR spectrum of ISAC 6 at 1 mM in Na₂CO₃/NaHCO₃ buffer at pH = 11 in D₂O and 298 K. (B) Possible conformations of ISAC 6 according to MM3* conformational search and NMR analysis.

coupling constants (between 6.5 and 8 Hz) were also deduced (see SI). Indeed, molecular mechanics calculations showed an ensemble of structures differing in the C6–C7 and C7–N torsion angles that could be clustered into four different families (conformers A–D), conformer A being the global energy minimum (Figure 3B).

Because of the structural similarity between ISAC 6 and cyclam, its ability to chelate divalent cations was scrutinized. To gain insight into the metal binding event, we first investigated cadmium(II) coordination as ¹¹³Cd is an NMR active nucleus with the potential of providing structural information through coupling constants and chemical shifts analysis.^{17,33} ISAC 6 was titrated with a solution of ¹¹³Cd²⁺ enriched (93.35%) CdCl₂ solution until the 1:1 complex was formed. A slow exchange process in the NMR chemical shift time scale was observed (Figure 4). Notably, the ¹H NMR signals of the NH groups were visible, but only when the ISAC 6–Cd complex was formed, strongly suggesting that the exchange process with the water solvent is slowed down, the typical nitrogen inversion is blocked, and the NH orientation becomes fixed.

Thus, ¹H–¹⁵N and ¹H–¹¹³Cd HSQC NMR experiments were performed (Figure 5). The ¹H–¹¹³Cd spectrum (Figure 5 red) showed only one signal for the ¹¹³Cd-containing complex ($\delta(^{113}\text{Cd}) = -387$ ppm). Indeed, the chemical shift of this signal did not change with the addition of more Cd²⁺ equivalents, strongly suggesting that only one type of complex was generated. Moreover, strong heteronuclear correlations to the ¹¹³Cd were found for the two NH's and for only one proton of each of the two CH₂ groups 7 and 8, those close to NH(b), indicating that these methylene protons display an *anti* orientation with respect to the cation. Interestingly, variable

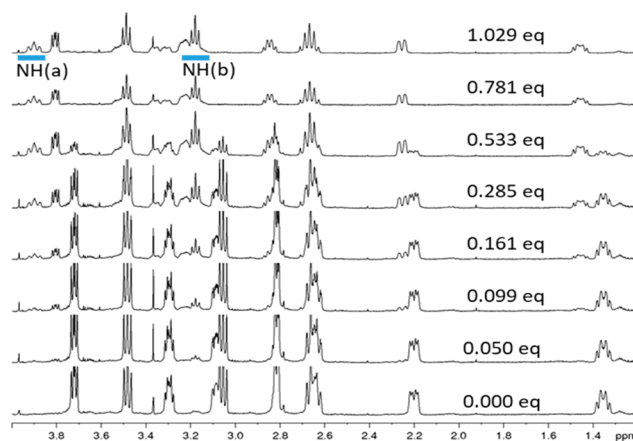


Figure 4. ¹H NMR titration of the complexation of ¹¹³Cd²⁺ by ISAC 6 at 3.5 mM in Na₂CO₃/NaHCO₃ buffer at pH = 11 in H₂O/D₂O (9:1) at 298 K, with increasing amounts of CdCl₂ (93.35% ¹¹³Cd²⁺) solution in D₂O.

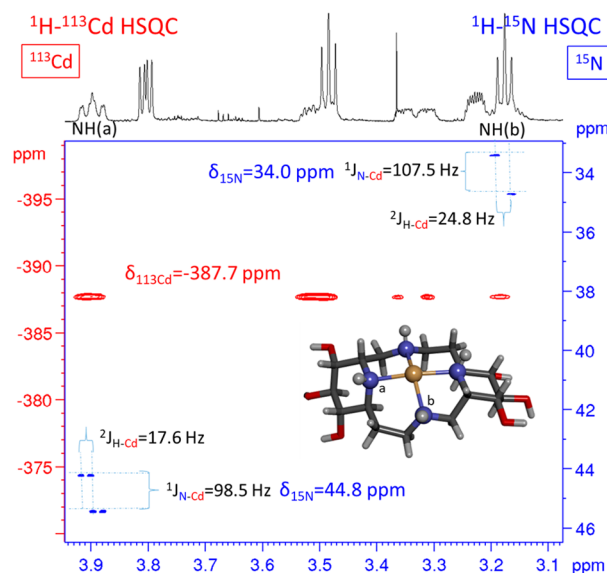


Figure 5. Superimposition of the ¹H–¹⁵N HSQC (blue) and ¹H–¹¹³Cd HSQC (red) spectra for the ISAC 6–Cd complex.

temperature experiments showed that the exposure of the NH protons to the solvent is different. The measured temperature coefficients indicated that NH(a) is more exposed to solvent than NH(b) (see SI).

Additional NOE experiments showed strong NH(a)/H6-ax and NH(a)/H4 cross peaks confirming the axial disposition of the NH(a) further supported by the ³J coupling constants with H5 (12.6 Hz, ax–ax) and H1 (3.4 Hz, ax–eq) (Figure 6). For NH(b), the ax–ax coupling pattern is evident through H5–H6ax–H7ax–NH(b), also assessing the axial orientation for this NH. Thus, the N–H bonds point toward the same face of the molecule as expected for a *trans-I* (*R,S,R,S*) type conformation, which is in full agreement with all the experimental NMR data (Figure 6 and SI).

We finally investigated copper(II) coordination by the ISAC ligand. ISAC 6 with 1 equiv of Cu(ClO₄)₂·6H₂O was placed in water at pH 6.4, and the mixture was heated for 2 h at 50 °C while adjusting the pH at 6.4. The dark blue solution suggested rapid formation of the desired complex that was isolated by precipitation in a water/acetone mixture. The targeted ISAC

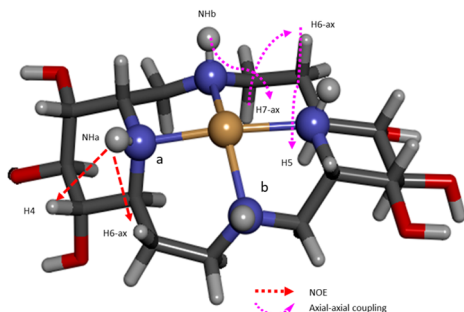


Figure 6. DFT calculated structure for the ISAC 6–Cd²⁺ complex according to the NMR data

6–Cu²⁺ complex was identified through ESI-MS spectrometry (see SI) and compared to the cyclam–Cu²⁺ complex prepared in parallel.³⁴ Its EPR analysis shows the four expected lines at low field and no superhyperfine splitting (Figure 7A) in

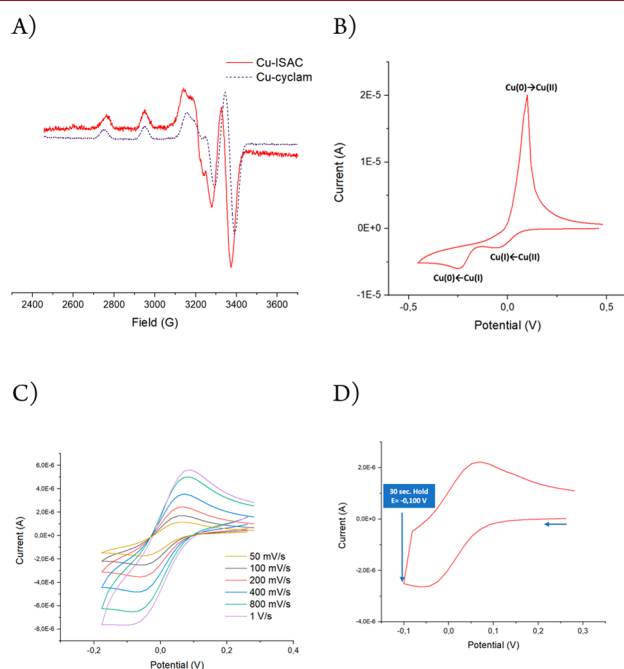


Figure 7. Study of the Cu²⁺–ISAC 6 complex: (A) By EPR (X-band): Overlap of Cu–ISAC and Cu–cyclam at 143 K in DMF/water (1:1); (B–D) by cyclic voltammetry: (B) scan from 0.5 to –0.5 V at 100 mV·s^{–1}; (C) overlap of different scan rates from 0.3 to –0.1 V; (D) pseudo-electrolysis (30 s hold at –0.1 V).

agreement with a mononuclear copper(II) complex with a square planar geometry (see SI). Comparison with the Cu–cyclam spectrum showed high similarity in the electronic parameters demonstrating a similar conformation in these conditions. One of the main pathways in copper(II) complex demetalation, and particularly in the case of *in vivo* use, is the reduction to copper(I) and subsequent decoordination.³⁵ Thus, to investigate the stability of the ISAC 6–Cu²⁺ complex against reduction, electrochemical measurements were conducted. A large scan showed the contribution of the other oxidation states (Figure 7B); thus, the redox behavior of the Cu²⁺/Cu⁺ couple was assigned upon scanning from 0.3 to –0.180, with increasing scan rates (Figure 7C). Quasi-reversible behavior of the redox couple was evidenced, with $E_{1/2} = 0.01$ V (vs Ag/AgCl) and ΔE_p ranging from 110 to 150

mV when scanning from 50 to 1000 mV·s^{–1}. This demonstrates the enhanced stability of the ISAC 6–copper(I) complex compared to its cyclam parent complex, for which an irreversible reduction wave is described in the literature.³⁶ To further gain insight into this ISAC 6–copper(I) stability, a pseudo-electrolysis was conducted by scanning from 0.3 to –0.1 V and holding this potential for 30 s to trigger local generation of the Cu⁺ complex (Figure 7D). When scanning back to 0.3 V, the current jump and the intact oxidation wave confirmed the stability of the Cu⁺ complex at the cyclic voltammetry scale, which is very promising in this ligand family. Another important factor in the characterization of coordination complexes is the kinetic inertness of the complex when placed in competitive media. As the nitrogen-based macrocycles are highly basic (cyclam pK_a's = 11.29, 10.19, 1.61, 1.91),³⁷ the kinetic inertness of their complexes can be assessed by competition with protons in acidic solutions and their degradation followed by UV–vis with the typical decrease of the copper d–d transition band over time. ISAC 6–copper(I) complex inertness was assessed at 20 °C in 1.5 M HCl solution in distilled water. It proved very stable, as its UV spectrum did not change after 15 days while a half-life of 42 min was recorded for the cyclam–Cu complex (see SI).

In conclusion, the synthesis of an iminosugar azacrown is disclosed. In the resulting C₂ symmetric macrocycle, some flexibility around the aminated arms has been observed by NMR, which disappears upon binding to Cd²⁺, suggesting an interesting interplay between preorganization and flexibility. Importantly, the ISAC 6–Cu²⁺ complex adopts a geometry comparable to that of the parent cyclam–Cu²⁺ complex, with significantly enhanced coordination ability in solution. This constitutes a first entry in the coordination chemistry of this new class of azamacrocycles.

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.orglett.0c00503>.

Experimental procedures, characterization data for compounds 7–13, copies of ¹H and ¹³C NMR spectra for compounds 6, 12, and 13; monitoring of the Staudinger–aza-Wittig reaction with 12 by NMR and MS; study of the ISAC 6–Cu²⁺ complex (stability, analogy with cyclam–Cu²⁺ complex) by EPR, cyclic voltammetry and UV–vis spectroscopy; NMR conformational study of ISAC 6 and its complex with Cd²⁺ (PDF)

■ AUTHOR INFORMATION

Corresponding Authors

Yves Blériot – Université de Poitiers, Equipe “Synthèse Organique”, Groupe Glycochimie 4 rue Michel Brunet, 86073 Poitiers cedex 9, France; orcid.org/0000-0002-3209-9282; Email: yves.bleriot@univ-poitiers.fr

Jesús Jiménez-Barbero – CIC bioGUNE, Parque Tecnológico de Bizkaia, Derio-Bizkaia 48160, Spain; IKERBASQUE, Basque Foundation for Science, 48009 Bilbao, Spain; Dept. Organic Chemistry II, Faculty of Science and Technology, University of the Basque Country, 48940 Leioa, Spain; orcid.org/0000-0001-5421-8513; Email: jjbarbero@cicbiogune.es

Authors

Alexandra Bordes – Université de Poitiers, Equipe “Synthèse Organique”, Groupe Glycochimie 4 rue Michel Brunet, 86073 Poitiers cedex 9, France

Ana Poveda – CIC bioGUNE, Parque Tecnológico de Bizkaia, Derio-Bizkaia 48160, Spain; orcid.org/0000-0001-5060-2307

Thibault Troadec – Université de Brest, UMR-CNRS 6521 CEMCA, 29200 Brest, France

Antonio Franconetti – CIC bioGUNE, Parque Tecnológico de Bizkaia, Derio-Bizkaia 48160, Spain

Ana Ardá – CIC bioGUNE, Parque Tecnológico de Bizkaia, Derio-Bizkaia 48160, Spain; orcid.org/0000-0003-3027-7417

Flavie Perrin – Université de Poitiers, Equipe “Synthèse Organique”, Groupe Glycochimie 4 rue Michel Brunet, 86073 Poitiers cedex 9, France

Mickaël Ménand – Sorbonne Université, CNRS, Institut Parisien de Chimie Moléculaire, 75005 Paris, France; orcid.org/0000-0003-0174-9120

Matthieu Sollogoub – Sorbonne Université, CNRS, Institut Parisien de Chimie Moléculaire, 75005 Paris, France; orcid.org/0000-0003-0500-5946

Jérôme Guillard – Université de Poitiers, Equipe “Synthèse Organique”, Groupe Glycochimie 4 rue Michel Brunet, 86073 Poitiers cedex 9, France

Jérôme Désiré – Université de Poitiers, Equipe “Synthèse Organique”, Groupe Glycochimie 4 rue Michel Brunet, 86073 Poitiers cedex 9, France

Raphaël Tripiet – Université de Brest, UMR-CNRS 6521 CEMCA, 29200 Brest, France; orcid.org/0000-0001-9364-788X

Complete contact information is available at:

<https://pubs.acs.org/10.1021/acs.orglett.0c00503>

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

YB acknowledges financial support from the European Union (ERDF) and “Région Nouvelle Aquitaine”. J.J.B. thanks the European Research Council for financial support (ERC-2017-AdG, Project Number 788143-RECGLYC-ANMR). J.J.B. also thanks Agencia Estatal Investigación of Spain, AEI (Grant RTI2018-094751-B-C21, a Juan de la Cierva contract to A.F., and the Severo Ochoa Excellence Accreditation (SEV-2016-0644). T.T. and R.T. thank the “Service Commun RMN-RPE” from UBO for EPR measurements, and Dr. Noémie Lalaoui (UMR CNRS 6521, Brest) for cyclic voltammetry studies.

REFERENCES

- Xie, J.; Bogliotti, N. Synthesis and Applications of Carbohydrate-Derived Macrocyclic Compounds. *Chem. Rev.* **2014**, *114* (15), 7678–7739.
- Potopnyk, M. A.; Jarosz, S. Nitrogen-Containing Macrocycles Having a Carbohydrate Scaffold. In *Advances in Carbohydrate Chemistry and Biochemistry*; Elsevier: 2014; Vol. 71, pp 227–295. DOI: 10.1016/B978-0-12-800128-8.00003-0.
- Pietraszkiewicz, M.; Kozbial, M.; Pietraszkiewicz, O. Chiral recognition of amino acids by diazacrown ethers and crowns incorporating a mannopyranoside unit, immobilized in a supported liquid membrane. *Enantiomer* **1997**, *2*, 319–326.
- Pietraszkiewicz, M. Dynamic LH Nuclear Magnetic Resonance Spectroscopic Studies of Complexes Formed between Substituted Ammonium Cations and t w o Chiral Diaza-Crown Ethers Incorporating Asymmetric Carbohydrate Units. *J. Chem. Soc., Perkin Trans. 2* **1985**, *2*, 1559–1562.
- Jarosz, S.; Lewandowski, B. Synthesis and Complexation Properties towards the Ammonium Cation of Aza-Coronand Analogues Containing Sucrose. *Carbohydr. Res.* **2008**, *343* (5), 965–969.
- Lewandowski, B.; Jarosz, S. Chiral Recognition of α -Phenylethylamine by Sucrose-Based Macrocyclic Receptors. *Chem. Commun.* **2008**, *47*, 6399.
- Jarosz, S.; Listkowski, A.; Lewandowski, B.; Ciunik, Z.; Brzuszkiewicz, A. Macrocyclic Receptors Containing Sucrose Skeleton. *Tetrahedron* **2005**, *61* (35), 8485–8492.
- Bakó, P.; Vízvárdi, K.; Toppet, S.; Van der Eycken, E.; Hoornaert, G. J.; Töke, L. Synthesis, Extraction Ability and Application in Asymmetric Synthesis of Azacrown Ethers Derived from D-Glucose. *Tetrahedron* **1998**, *54* (49), 14975–14988.
- Hsieh, Y.-C.; Chir, J.-L.; Wu, H.-H.; Chang, P.-S.; Wu, A.-T. A Sugar-Aza-Crown Ether-Based Fluorescent Sensor for Hg²⁺ and Cu²⁺. *Carbohydr. Res.* **2009**, *344* (16), 2236–2239.
- Hsieh, Y.-C.; Chir, J.-L.; Yang, S.-T.; Chen, S.-J.; Hu, C.-H.; Wu, A.-T. A Sugar-Aza-Crown Ether-Based Fluorescent Sensor for Cu²⁺ and Hg²⁺ Ions. *Carbohydr. Res.* **2011**, *346* (7), 978–981.
- Xie, J.; Ménand, M.; Maisonneuve, S.; Métivier, R. Synthesis of Bispirenyl Sugar-Aza-Crown Ethers as New Fluorescent Molecular Sensors for Cu(II). *J. Org. Chem.* **2007**, *72* (16), 5980–5985.
- Izatt, R. M.; Pawlak, K.; Bradshaw, J. S.; Bruening, R. L. Thermodynamic and Kinetic Data for Macrocyclic Interactions with Cations and Anions. *Chem. Rev.* **1991**, *91* (8), 1721–2085.
- Delgado, R.; Félix, V.; Lima, L. M. P.; Price, D. W. Metal Complexes of Cyclen and Cyclam Derivatives Useful for Medical Applications: A Discussion Based on Thermodynamic Stability Constants and Structural Data. *Dalton Transactions* **2007**, *26*, 2734–2745.
- Perceley, L.; Louvet, V.; Handel, H.; Appriou, P. Application d'un tétraazamacrocycle greffé sur polymère à l'extraction et au dosage du cuivre en eau de mer. *Anal. Chim. Acta* **1985**, *169*, 325–330.
- Alves, L. G.; Madeira, F.; Munhá, R. F.; Maulide, N.; Veiros, L. F.; Martins, A. M. Cooperative Metal–Ligand Hydroamination Catalysis Supported by C–H Activation in Cyclam Zr(IV) Complexes. *Inorg. Chem.* **2018**, *57* (20), 13034–13045.
- Kuehnel, M. F.; Sahm, C. D.; Neri, G.; Lee, J. R.; Orchard, K. L.; Cowan, A. J.; Reisner, E. ZnSe Quantum Dots Modified with a Ni(Cyclam) Catalyst for Efficient Visible-Light Driven CO₂ Reduction in Water. *Chemical Science* **2018**, *9* (9), 2501–2509.
- Liang, X.; Parkinson, J. A.; Parsons, S.; Weishäupl, M.; Sadler, P. J. Cadmium Cyclam Complexes: Interconversion of *Cis* and *Trans* Configurations and Fixation of CO₂. *Inorg. Chem.* **2002**, *41* (17), 4539–4547.
- Bartoli, F.; Bencini, A.; Conti, L.; Giorgi, C.; Valtancoli, B.; Paoli, P.; Rossi, P.; Le Bris, N.; Tripiet, R. Catching Anions with Coloured Assemblies: Binding of PH Indicators by a Giant-Size Polyammonium Macrocycle for Anion Naked-Eye Recognition. *Org. Biomol. Chem.* **2016**, *14* (35), 8309–8321.
- Su, H.; Chen, X.; Fang, W. ON–OFF Mechanism of a Fluorescent Sensor for the Detection of Zn(II), Cd(II), and Cu(II) Transition Metal Ions. *Anal. Chem.* **2014**, *86* (1), 891–899.
- Kaden, T. A. Labelling Monoclonal Antibodies with Macrocyclic Radiometal Complexes. A Challenge for Coordination Chemists. *Dalton Transactions* **2006**, *30*, 3617.
- Labrosse, B.; Brelot, A.; Heveker, N.; Sol, N.; Schols, D.; De Clercq, E.; Alizon, M. Determinants for Sensitivity of Human Immunodeficiency Virus Coreceptor CXCR4 to the Bicyclam AMD3100. *J. Virol.* **1998**, *72* (8), 6381–6388.
- Larpent, C.; Laplace, A.; Zemb, T. Macrocyclic Sugar-Based Surfactants: Block Molecules Combining Self-Aggregation and

Complexation Properties. *Angew. Chem., Int. Ed.* **2004**, *43* (24), 3163–3167.

(23) Burke, H. M.; Gunnlaugsson, T.; Scanlan, E. M. Glycosylated Lanthanide Cyclen Complexes as Luminescent Probes for Monitoring Glycosidase Enzyme Activity. *Org. Biomol. Chem.* **2016**, *14* (38), 9133–9145.

(24) Moats, R. A.; Fraser, S. E.; Meade, T. J. A “Smart” Magnetic Resonance Imaging Agent That Reports on Specific Enzymatic Activity. *Angew. Chem., Int. Ed. Engl.* **1997**, *36* (7), 726–728.

(25) Horne, G.; Wilson, F. X. Therapeutic Applications of Iminosugars: Current Perspectives and Future Opportunities. In *Progress in Medicinal Chemistry*; Elsevier: 2011; Vol. 50, pp 135–176. DOI: 10.1016/B978-0-12-381290-2.00004-5.

(26) Nash, R. J.; Kato, A.; Yu, C.-Y.; Fleet, G. W. Iminosugars as Therapeutic Agents: Recent Advances and Promising Trends. *Future Med. Chem.* **2011**, *3* (12), 1513–1521.

(27) Ménand, M.; Blais, J.-C.; Valéry, J.-M.; Xie, J. De Novo Synthesis of Sugar-Aza-Crown Ethers via a Domino Staudinger Aza-Wittig Reaction. *J. Org. Chem.* **2006**, *71* (8), 3295–3298.

(28) Gradnig, G.; Legler, G.; Stütz, A. E. A Novel Approach to the 1-Deoxynojirimycin System: Synthesis from Sucrose of 2-Acetamido-1,2-Dideoxynojirimycin, as Well as Some 2-N-Modified Derivatives. *Carbohydr. Res.* **1996**, *287* (1), 49–57.

(29) Zoidl, M.; Ginzalez Santana, A.; Torvisco, A.; Tysøe, C.; Siriwardena, A.; Withers, S. G.; Wrodnigg, T. M. The Staudinger/aza-Wittig/Grignard Reaction as key step for the concise synthesis of 1-C-alkyl-iminoalditol glycomimetics. *Carbohydr. Res.* **2016**, *429*, 62–70.

(30) Mondon, M.; Fontelle, N.; Désiré, J.; Lecornuë, F.; Guillard, J.; Marrot, J.; Blériot, Y. Access to l- and d- Iminosugar C-Glycosides from a d - Gluco -Derived 6-Azidolactol Exploiting a Ring Isomerization/Alkylation Strategy. *Org. Lett.* **2012**, *14* (3), 870–873.

(31) Yu, W.; Mei, Y.; Kang, Y.; Hua, Z.; Jin, Z. Improved Procedure for the Oxidative Cleavage of Olefins by OsO₄ – NaIO₄. *Org. Lett.* **2004**, *6* (19), 3217–3219.

(32) Lucas, R.; Balbuena, P.; Errey, J. C.; Squire, M. A.; Gurucha, S. S.; McNeil, M.; Besra, G. S.; Davis, B. G. Glycomimetic Inhibitors of Mycobacterial Glycosyltransferases: Targeting the TB Cell Wall. *ChemBioChem* **2008**, *9* (14), 2197–2199.

(33) El Majzoub, A.; Cadiou, C.; Déchamps-Olivier, I.; Tinant, B.; Chuburu, F. Cyclam-Methylbenzimidazole: A Selective OFF-ON Fluorescent Sensor for Zinc. *Inorg. Chem.* **2011**, *50* (9), 4029–4038.

(34) Prevedello, A.; Bazzan, I.; Dalle Carbonare, N.; Giuliani, A.; Bhardwaj, S.; Africh, C.; Cepek, C.; Argazzi, R.; Bonchio, M.; Caramori, S.; et al. Heterogeneous and Homogeneous Routes in Water Oxidation Catalysis Starting from Cu^{II} Complexes with Tetraaza Macrocyclic Ligands. *Chem. - Asian J.* **2016**, *11* (8), 1281–1287.

(35) Woodin, K. S.; Heroux, K. J.; Boswell, C. A.; Wong, E. H.; Weisman, G. R.; Niu, W.; Tomellini, S. A.; Anderson, C. J.; Zakharov, L. N.; Rheingold, A. L. Kinetic Inertness and Electrochemical Behavior of Copper(II) Tetraazamacrocyclic Complexes: Possible Implications for in Vivo Stability. *Eur. J. Inorg. Chem.* **2005**, *2005* (23), 4829–4833.

(36) Wadas, T. J.; Wong, E. H.; Weisman, G. R.; Anderson, C. J. Coordinating Radiometals of Copper, Gallium, Indium, Yttrium, and Zirconium for PET and SPECT Imaging of Disease. *Chem. Rev.* **2010**, *110* (5), 2858–2902.

(37) Havlíčková, J.; Medová, H.; Vitha, T.; Kotek, J.; Císařová, I.; Hermann, P. Coordination Properties of Cyclam (1,4,8,11-Tetraaza-cyclotetradecane) Endowed with Two Methylphosphonic Acid Pendant Arms in the 1,4-Positions. *Dalton Transactions* **2008**, *39*, 5378.