

4-Acylamino-6-arylfuro[2,3-*d*]pyrimidines: potent and selective glycogen synthase kinase-3 inhibitors

Yutaka Maeda,^a Masato Nakano,^{a,*} Hideyuki Sato,^a Yasushi Miyazaki,^a
Stephanie L. Schweiker,^b Jeffery L. Smith^b and Anne T. Truesdale^b

^aChemistry Department, Tsukuba Research Laboratories, GlaxoSmithKline K.K., Wadai 43, Tsukuba, Ibaraki 300-4247, Japan

^bGlaxoSmithKline Inc., Five Moore Drive, Research Triangle Park, NC 27709, USA

Received 16 April 2004; revised 24 May 2004; accepted 24 May 2004

Available online 17 June 2004

Abstract—Modeling studies of a furo[2,3-*d*]pyrimidine GSK-3 hit compound **1** superimposed onto the X-ray crystal structure of a legacy pyrazolo[3,4-*c*]pyridazine GSK-3 inhibitor **2** led to the identification of 4-acylamino-6-arylfuro[2,3-*d*]pyrimidine template **3**. Synthesis of analogues based on template **3** has resulted in a number of potent and selective GSK-3 β inhibitors. The most potent and selective compound was the *m*-pyridyl analogue **24**.

© 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Glycogen synthase kinase-3 (GSK-3) is a serine/threonine kinase involved in the regulations of many cell functions.¹ GSK-3 is implicated in type-2 diabetes because of its ability to phosphorylate glycogen synthase, which effectively inhibits the enzyme in catalyzing the synthesis of glycogen. Recent publications have also suggested that the β -isomer (GSK-3 β) of its two isoforms (α and β) plays a pivotal role in the genesis of Alzheimer's disease (AD) and other dementia.^{2,3} A selective inhibitor of GSK-3 potentially can be an effective agent for the treatment of type-2 diabetes and/or AD, as well as other neurodegenerative disorders. A number of GSK-3 inhibitors having different structural classes have been reported, and they showed varying degrees of efficacy and specificity.⁴

In the course of our exploratory research targeting the VEGFR2 and TIE2 tyrosine kinases,⁵ furo[2,3-*d*]pyrimidine **1** was identified as a GSK-3 inhibitor having an IC₅₀ value of 0.081 μ M versus GSK-3 β . Modeling studies in which **1** was superimposed onto the X-ray crystal structure of the complex of the potent

pyrazolo[3,4-*c*]pyridazine GSK-3 inhibitor **2**⁶ with GSK-3 β suggested that both compounds bind in the enzyme with similar space requirements. The studies also suggested the existence of a unique binding interaction between **1** and the hinge region in the ATP binding site (Figs. 1 and 2).⁷

The outcome of the modeling studies prompted us to modify **1** with a view to increasing its affinity for GSK-3. This included replacement of the *p*-methoxy group by other groups, the acylation of the 4-amino group, and removal of the pyridyl moiety, which resulted in the creation of a new template **3** (Fig. 2).

We first investigated the structure–activity relationship (SAR) on the 4-*N*-acyl portion. Thus a set of analogues of **3** with varying *N*-acyl groups and with a constant 6-*p*-methoxyphenyl group was synthesized. Their kinase inhibitory activity is summarized in Table 1. The hexanoyl analogue **4** exhibited submicromolar inhibitory activity versus GSK-3 β . The closely related methylthio analogue **5** was equipotent to **4**. On the other hand, the isobutyryl analogue **6** was approximately 4-fold more potent than **5**. This interesting result provided the impetus for the synthesis of cycloalkyl analogues **7–10**. Unfortunately, **7** and **9** exhibited only slight improvements in the potency (IC₅₀ = ca. 0.3 μ M) over that of **4** or **5**, and the cyclohexyl analogue **10** was much less potent. Surprisingly, cyclopentyl analogue **8** was 10-fold

Keywords: Glycogen synthase kinase-3 inhibitors.

* Corresponding author. Tel.: +81-298-64-5541; fax: +81-298-64-5559; e-mail: masato.nakano@gsk.com

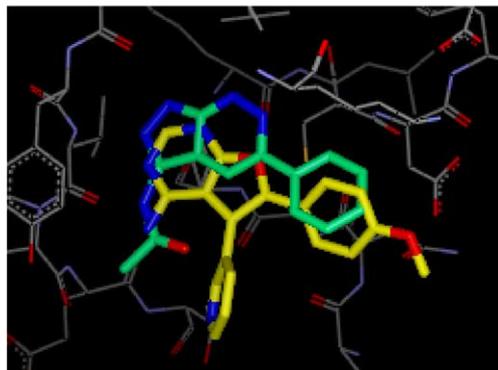
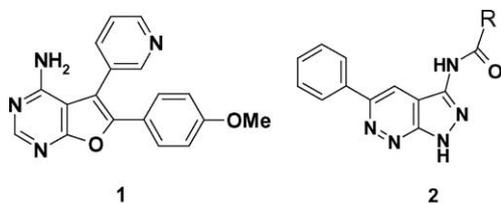


Figure 1. Docking study of furo[2,3-*d*]pyrimidine **1** (yellow) in an X-ray crystal structure of the complex of pyrazolo[3,4-*c*]pyridazine **2** (light green) and GSK-3 β .

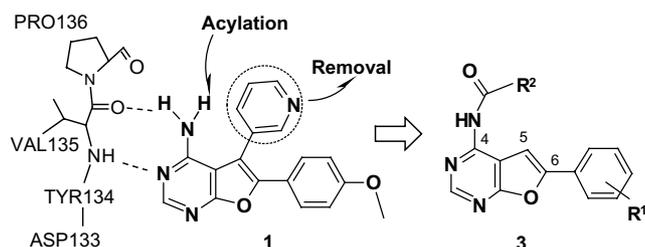


Figure 2. Proposed binding of a furo[2,3-*d*]pyrimidine hit **1** with the hinge region and design of a new series of GSK-3 inhibitor **3**.

more potent than **7** or **9** and was by far the most potent analogue versus GSK-3. According to the modeling studies of analogue **8** (see Fig. 3), the methine hydrogen and vertical orientation of the cyclopentyl group appeared to have a unique binding interaction with PRO136 in the front hydrophobic pocket. To test this hypothesis, several compounds **11–14**, which were void of the methine hydrogen, were synthesized. These compounds ranged from morpholino and pyrrolidino analogues (**11** and **12**) having in essence a urea-type functional group to other aryl groups (**13** and **14**), and they exhibited weak GSK-3 inhibitory activity, maybe due to unfavorable affinity of the 4-*N*-acyl or urea substituent with PRO136. All the analogues **4–14** showed weak inhibitory activity against CDK-2 and VEGFR2.

In the ensuing SAR studies, we utilized cyclopropyl analogue **7** as the starting point and explored the modification of the 6-position of the furo[2,3-*d*]pyrimidine scaffold. The resultant analogues are listed in Table 2. As shown in the table, the unsubstituted **15** showed improved potency against GSK-3 β . However, replace-

Table 1. Inhibition of hGSK-3 β ⁸ by 4-*N*-acyl analogues **4–14**

No.	R	GSK-3 β IC ₅₀ , μ M	CDK2 IC ₅₀ , μ M	VEGFR2 IC ₅₀ , μ M
4	-(CH ₂) ₄ CH ₃	0.437	>19	>48
5	-(CH ₂) ₂ SCH ₃	0.537	>20	>47
6	-CH(CH ₃) ₂	0.132	>21	>48
7		0.316	>20	3.55
8		0.032	>19	1.02
9		0.324	>19	>48
10		1.82	>20	>48
11		>34	>20	>48
12		1.95	>21	12.3
13		>35	>21	21.9
14		2.40	>20	>48

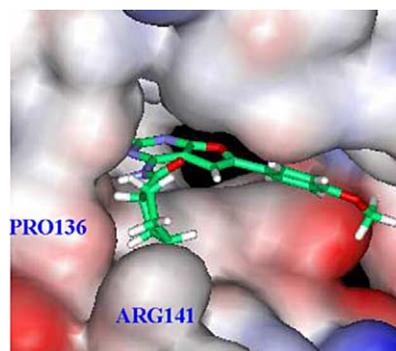
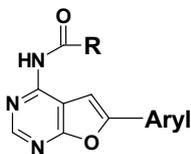


Figure 3. In silico surface model of the GSK-3 ATP-binding pocket with compound **8**.

ment with *p*-chloro, *p*-methyl, and *p*-fluoro groups (compounds **16–18**) gave analogues having reduced inhibitory activity, suggesting the potentially detrimental effects of a *p*-substituent.

Very interestingly, the most potent compound in the series was the *m*-pyridyl analogue **19**. Potency of **15–19** against VEGFR2 increased somewhat but those were still in the submicromolar to low micromolar range. These findings prompted us to prepare analogues **20–24** having identical 6-aryl groups as those in **15–19** but with

Table 2. Inhibition of hGSK-3 β by 6-aryl analogues **7**, **8**, and **15–24**

No.	R	Aryl	GSK-3 β IC ₅₀ , μ M	CDK2 IC ₅₀ , μ M	VEGFR2 IC ₅₀ , μ M
7			0.316	>20	3.55
15			0.174	>21	0.832
16			1.00	>20	2.04
17			0.617	>21	0.631
18			0.575	>19	2.04
19			0.005	0.447	3.24
8			0.032	>19	1.02
20			0.023	>19	>23
21			0.550	>20	>20
22			0.240	>20	>20
23			0.219	>19	>19
24			0.005	0.457	>24

a cyclopentyl group in the 4-position. As a result, analogues **20–24** showed comparable potency versus GSK-3 β to that of compounds **15–19**. However and importantly, these compounds showed a lack of inhibitory activity against CDK-2 and VEGFR2.

Having identified a series of potent GSK-3 inhibitors, we next profiled the most potent analogue **24** against a panel of more than 20 kinases, including GSK-3 β (Table 3).⁹ The data showed that analogue **24** indeed had excellent selectivity over all the kinases in the panel, including CDK-2. Overlaying of docking structures of **24** in GSK-3 β and CDK-2/cyclin A¹⁰ also suggested that the selectivity of **24** was likely due to a steric clash between a cyclopentyl group and its interacting site in CDK-2. Based on the modeling studies, however, *m*-pyridyl effect on the high potency was not clear. Finally, representative three analogues **19**, **20**, and **24** were examined in an assay that measured glycogen accumu-

lation in L6 cells (Table 4). The 6-phenyl analogue **20** did not show potency in such an assay, but both *m*-pyridyl analogues **19** and **24** exhibited excellent potency of glycogen accumulation.¹¹

2. Chemistry¹²

The furo[2,3-*d*]pyrimidine analogues were prepared according to procedures outlined in Scheme 1. The chemistry started with alkylation of commercially available 2-bromoacetophenones **25i–v** with malononitrile to give benzoylmethylmalononitriles **26i–v**, which were treated with hydrochloric acid in acetic acid at room temperature to afford 2-amino-3-cyanofurans **27i–v**. Cyclization of furan **27i–v** with formamide resulted in 4-amino-6-phenyl-furo[2,3-*d*]pyrimidine **28i–v**, which

Table 3. Selectivity of *m*-pyridyl analogue **24** for GSK-3 β ^a

Compd no.	24
AMPK	0
Chk1	0
CKII	0
JNK	0
LCK	2
MAPK	5
RSK-2	NT
MAPKAP-K2	0
MEK1	0
MSK1	10
p70S6K	0
PDK1	0
PHOS.K	9
PKA	0
PKB α	1
PKC α	NT
PRAK	7
ROK α	0
SAPK2a	0
SAPK2b	0
SAPK3	0
SAPK4	17
SGK	8
CDK2/Cyclin A	66
GSK-3 β	91

^a Values are % I @ 10 μ M using 100 μ M ATP (see Ref. 9 for kinases used and assay details).

Table 4. Potency of **19**, **20**, and **24** in glycogen accumulation in L6 cells

No.	Potency of glycogen accumulation (EC ₅₀ , μ M)
19	0.74
20	>10
24	0.39

were reacted with acylchlorides to afford the desired products. With regard to the synthesis of a *m*-pyridyl analogues **19** and **24**, the precursor **28vi** was obtained by a different method. Obtained 2-amino-3-cyano-furan **27vi** was converted into an ethoxyimino derivative, followed by treatment with ammonia in a solution of ethanol and tetrahydrofuran. Subsequently, cyclization was

completed by addition of sodium ethoxide in the same solvent to give a precursor, 4-amino-6-(*m*-pyridyl)-furo[2,3-*d*]pyrimidine **28vi**. Cyclopentylcarbonylation of this precursor in a usual manner led to analogue **24**.

3. Conclusion

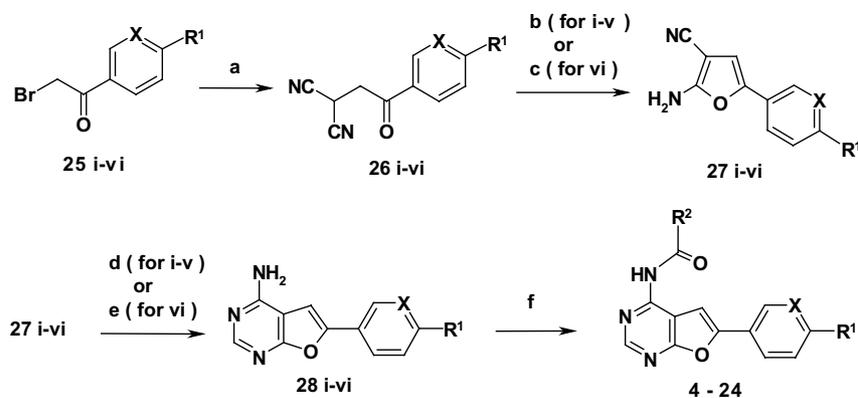
Modeling studies of a GSK-3 β hit compound **1** by superimposing onto the X-ray crystal structure of the complex of **2** with GSK-3 β led to the development of a template represented by structure **3**. Synthesis based on the template has led to the identification of a series of potent GSK-3 β specific analogues. One such compound, the *m*-pyridyl analogue **24**, was found to exhibit potent GSK-3 β inhibitory activity, possess excellent selectivity over other kinases including CDK-2 and VEGFR2, and exhibit excellent potency in cellular assays.

Acknowledgements

The authors acknowledge with gratitude the invaluable assistance of colleagues from GlaxoSmithKline RTP, USA and Harlow, UK. Additionally, we are grateful to the Division of Signal Transduction Therapy, Department of Biochemistry, The University of Dundee for selective screening.

References and notes

- (a) Cohen, P.; Frame, S. *Nat. Rev. Mol. Cell Biol.* **2001**, *2*, 769; (b) Coghlan, M. P.; Culbert, A. A.; Cross, D. A. E.; Corcoran, S. L.; Yates, J. W.; Pearce, N. J.; Rausch, O. L.; Murphy, G. J.; Carter, P. S.; Cox, L. R.; Mills, D.; Brown, M. J.; Haigh, D.; Ward, R. W.; Smith, D. G.; Murray, K. J.; Reith, A. D.; Holder, J. C. *Chem. Biol. (London)* **2000**, *7*, 793; (c) Cohen, P.; Yellowlees, D.; Aitken, A.; Donella-Deana, A.; Hemmings, B. A.; Parker, P. J. *Eur. J. Biochem.* **1982**, *124*, 21.



(i) X=CH, R¹=OCH₃; (ii) X=CH, R¹=H; (iii) X=CH, R¹=Cl; (iv) X=CH, R¹=CH₃; (v) X=CH, R¹=F; (vi) X=N, R¹=H

Scheme 1. Reagents and conditions: (a) malononitrile, NaOEt; (b) HCl–AcOH; (c) piperidine, reflux EtOH; (d) formamide, 200 °C; (e) 1. (EtO)₂CHOAc, 2. NH₃, EtOH–THF, 3. NaOEt, EtOH–THF; (f) acylchloride, NaH, DMF.

2. GSK-3 β is known to phosphorylate the microtubule associated protein tau in mammalian cells. Lovestone, S.; Reynolds, C. H.; Latimer, D.; Davis, D. R.; Anderton, B. H.; Gallo, J.-M.; Hanger, D.; Mulot, S.; Marquardt, B. *Curr. Biol.* **1994**, *4*, 1077.
3. GSK-3 β binds to presenilin 1 and its binding complex mediates β -amyloid-induced neurotoxicity. Planel, E.; Sun, X.; Takashima, A. *Drug Develop. Res.* **2002**, *56*, 491.
4. (a) Leclerc, S.; Garnier, M.; Hoessel, R.; Marko, D.; Bibb, J. A.; Snyder, G. L.; Greengard, P.; Biernat, J.; Wu, Y. Z.; Mandelkow, E. M.; Eisenbrand, G.; Meijer, L. [Indirubin]. *J. Biol. Chem.* **2001**, *276*, 251; (b) Leost, M.; Schultz, C.; Link, A.; Wu, Y. Z.; Biernat, J.; Mandelkow, E. M.; Bibb, J. A.; Snyder, G. L.; Greengard, P.; Zaharevitz, D. W.; Gussio, R.; Senderowicz, A. M.; Sausville, E. A.; Kunick, C.; Meijer, L. [Paullone]. *Eur. J. Biochem.* **2000**, *267*, 5983; (c) Smith, D. G.; Buffet, M.; Fenwick, A. E.; Haigh, D.; Ife, R. J.; Saunders, M.; Slingsby, B. P.; Stacey, R.; Ward, R. W. [Maleimide, SB-415286]. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 635; (d) Meijer, L.; Thunnissen, A. M. W. H.; White, A. W.; Garnier, M.; Nikolic, M.; Tsai, L. H.; Walter, J.; Cleverley, K. E.; Salinas, P. C.; Wu, Y. Z.; Biernat, J.; Mandelkow, E. M.; Kim, S. H.; Pettit, G. R. [Hymenialdisine]. *Chem. Biol.* **2000**, *7*, 51; (e) Naerum, L.; Norskov-lauritsen, L.; Olesen, P. H. [5-(1-pyridyl)-[1,3,4]-oxadiazol]. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1525; (f) Martinez, A.; Alosa, M.; Castro, A.; Perez, C.; Moreno, F. J. [Thiadiazolidinone]. *J. Med. Chem.* **2002**, *45*, 1292; (g) Wagman, A. S.; Harrison, S. D.; Johnson, K.; Ring, D. B.; Bussiere, D. E.; Nuss, J. M.; Pfister, K. B.; Goff, D.; Boyce, R. S.; Ramurthy, S.; Brown, S. P.; Renhowe, P. A.; Subramanian, S.; Zhou, X. A. [CHIR73911]. In *225th National Meeting of the American Chemical Society, New Orleans, LA, March 23–27, 2003*; American Chemical Society: Washington, DC, 2003, MEDI22; (h) Olesen, P. H.; Sorensen, A. R.; Urso, B.; Kurtzhals, P.; Bowler, U. E.; Hansen, B. F. [Triazole derivative]. *J. Med. Chem.* **2003**, *46*, 3333; (i) Gallet, T.; Lardenois, P.; Lothead, A. W.; Nedelec, A.; Marguerie, S.; Saady, M.; Yaiche, P. [6,7,8,9-tetrahydropyrimido(1,2-a)pyrimidin-4-one]. European Patent Application EP1295885, 2003; (j) Kuo, G.-H.; Prouty, C.; DeAngelis, A.; Shen, L.; O'Neil, D. J.; Shah, C.; Connolly, P. J.; Murray, W. V.; Conway, B. R.; Cheung, P.; Westover, L.; Xu, J. Z.; Look, R. A.; Demarest, K. T.; Emanuel, S.; Middleton, S. A.; Jolliffe, L.; Beavers, M. P.; Chen, X. [Macrocyclic polyoxy-generated bis-7-aza-indolylmaleimides]. *J. Med. Chem.* **2003**, *46*, 4021.
5. Adams, J. J.; Bryan, D. D.; Feng, Y.; Matsunaga, S.; Miyazaki, Y.; Nakano, M.; Rocher, J.-P.; Sato, H.; Semones, M.; Silvia, D. J.; Tang, J. PCT International Application WO 03/022852, 2003.
6. Witherington, J.; Boardas, V.; Haigh, D.; Hickey, D. M.; Ife, R. J.; Rawlings, A. D.; Slingsby, B. P.; Smith, D. G.; Ward, R. W. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 1581.
7. Docking model of **1** was made by using Quanta/CHARMm (Accelrys Inc., San Diego, CA, USA).
8. hGSK-3 β enzyme assay was carried out by scintillation proximity assay (SPA).
9. Davis, S. P.; Reddy, H.; Caivano, M.; Cohen, P. *Biochem. J.* **2000**, *351*, 95.
10. In-house X-ray crystal structure data.
11. The method of the glycogen accumulation assay in L6 cells was described by: Peat, A. J.; Garrido, D.; Boucheron, J. A.; Schweiker, S. L.; Dickerson, S. H.; Wilson, J. R.; Wang, T. Y.; Thomson, S. A. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 2127.
12. Several experimental conditions are available in Ref. 5.