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A highly selective fluorescent turn-on probe for Al³⁺ via Al³⁺-promoted hydrolysis of ester

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ABSTRACT

A new reactive and highly selective fluorescent chemosensor (1) based on thiazole was synthesized for the quantification of aluminum ions in ethanol. The mechanism of fluorescence was based on the aluminum-promoted hydrolysis of the ester moiety and subsequent complexation.

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1. Introduction

The developments of turn-on fluorescent probes that operate through chemical reactions triggered by the target analytes have attracted attention.^{1–3} A range of chemical reactions have been implemented in the development of many reactive probes. This approach involves the use of highly selective and normally irreversible chemical reactions coupled with signal transduction induced by analytes. These reactive probes provide higher selectivity with larger spectroscopic changes than chemical probes. Previously, the synthesis and Zn^{2+} or Cu^{2+} sensing properties of several thiazole based chemosensors were reported.^{4–7} Among them, the selectivity for cations can be controlled by changing the moiety at position-4 of the thiazole ring. Schiff base fluorescent sensors have attracted special attention owing to their versatile organic blockers possessing comparable accessibility and structure variable. On the other hand, few studies have examined Schiff base chemosensors for the detection of Al^{3+} ion due to its weak coordination ability compared to transition metal ions^{8–16} Therefore, considering the Schiff base and functional carboxylic moiety¹⁷ affinity to Al^{3+} , probe **1** (Fig. 1) was prepared by introducing an ester and phenol group at positions-4 and -2 of the thiazole ring, respectively. This provided harder donors, such as oxygen and nitrogen atoms, in the molecular skeleton for the hard acid, Al³⁺.



Fig. 1. Fluorescent thiazole probes, 1-3 synthesized in this study and aluminum complex of 1.

Moreover, the hydroxyl group and ternary N atom as coordinating sites act as a bidentate ligand and show the same coordinating motif as the family of 8-hydroxyquinoline ligands that are used extensively in analytical chemistry for the detection of Al^{3+,18} Probes 2 and 3 were also prepared to understand the critical role of phenol at the 2-position of the thiazole rings and the mode of complexation with Al³⁺.

2. Results and discussion

Probes 1 and 3 were obtained in good yield by a Hantzsch condensation reaction of 2-hydroxyphenvlthiobenzamide and thiobenzamide with ethyl bromopyruvate in refluxing ethanol. and probe **2** was prepared in good yield by the hydrolysis of probe **1**.





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The structures of probes **1**, **2**, and **3** were confirmed by ¹H and ¹³C NMR spectroscopy, and elemental analyses.

UV–vis study was carried out in ethanol at a concentration of 20 μ M. The host solution was heated to 50 °C for 1 h after adding the guest to ensure complete complexation. Probe **1** displayed an obvious absorption band at 325 nm. This was attributed to a π – π transition, which is favored by the planar orientation enforced by intramolecular hydrogen bonding.¹⁹ On the other hand, upon the addition of aluminum ions to a solution of probe **1**, the absorption band at 325 nm, as shown in Fig. 2 with an isosbestic point at 339 nm. These findings are characteristic of phenol–thiazole derivatives when a complexation process is accompanied by the deprotonation of a hydroxyl group.^{6,7} Moreover, there are three clear isosbestic points at 339, 304, and 286 nm, which clearly suggests the conversion of free chemosensor **1** to an Al³⁺ complex of only one stoichiometry.^{20,21}



Fig. 2. UV–vis titration spectra of probe 1 (20 $\mu M)$ upon the addition of 10 equiv of Al(ClO₄)₃ in EtOH on heating at 50 °C for 1 h.

The emission studies were carried in a similar manner at a concentration of 2 μ M in ethanol. As a cation sensor, one of the important criteria for potential applications is the selectivity. To solutions of probe **1**, 10 equiv of a range of biologically and non-biologically relevant metal cations were added, and their complexation abilities were examined by fluorescence emission. None of the other cations except for Al³⁺, which produced a prominent fluorescence peak at 434 nm, induced a distinct emission shift with probe **1** (Figs. 3 and 4). Even the addition of Ga³⁺, which exhibits similar coordination behavior to Al³⁺, and Fe³⁺ did not produce any changes in the emission spectra. Fig. 5 shows the fluorescence spectrum of the free probe **1** and those in the presence of an incremental amount of Al³⁺.

Probe **1** showed no emission peak when excited at 364 nm and the slow addition of Al^{3+} elicited a large fluorescence peak at 434 nm. Therefore, in probe **1**, the fluorescence is turned on by binding with Al^{3+} . The inset in Fig. 5 shows the dependence of the normalized emission intensity ratio at 434 nm on the Al^{3+} concentration. The linear enhancement of fluorescence was observed with increasing Al^{3+} ion concentration when the Al^{3+} to the probe **1** concentration ratio was \leq 1:1. On the other hand, higher Al^{3+} concentrations did not lead to any further emission enhancement. To determine the stoichiometry of probe **1** and Al^{3+} in the complex, Job's method was employed using the emission changes at 434 nm as a function of the molar fraction of Al^{3+} . As shown in Fig. 6 maximum emission was observed when the molar fraction of



Fig. 3. Fluorescence spectra of probe 1 (2 μ M) in EtOH upon heating at 50 °C for 1 h on the addition of 10 equiv of various cations. λ_{ex} =364 nm.

Al³⁺ reached 0.5, which is indicative of 1:1 complexation between probe **1** and Al³⁺. The binding constant for the complex between probe **1** and Al³⁺ was determined by a non-linear fitting of the corresponding fluorescence titration data (Fig. 5) as 2.1×10^6 M⁻¹ with a satisfactory correlation coefficient value (*R*=0.9983) (Error estimated to be $\leq 10\%$).^{22,23}

A competitive binding experiment with different cations and Al^{3+} also shows that probe **1** is highly selective to Al^{3+} (Fig. S-1). It was also found that probe 1 shows 0.39 μ M of detection limit to sufficiently sense Al^{3+} (Fig. S-2).²⁴ This unique selectivity of probe **1** toward Al³⁺ could be interpreted in terms of the mechanism in Scheme 1. Aluminum is a well known Lewis acid²⁵ that is capable of hydrolyzing probe **1** and generating probe **2** that binds with Al^{3+} resulting in the liberation of ethanol and emission peak at 434 nm due to a chelation effect.^{26–28} At the initial stages, deprotonation of the phenolic proton occurs by the addition of $Al(ClO_4)_3$, which subsequently acts as a Lewis acid hydrolyzing the ester to generate acid **2**. This acid provides a better additional binding site for the chelation of Al³⁺ and increases the fluorescence dramatically through a highly efficient chelation-enhanced fluorescence (CHEF) effect. Although there are some examples of Cu²⁺-facilitated hydrolysis of an ester bond, there have been no reports of Al³⁺-promoted ester hydrolysis, which is utilized here to improve the sensitivity and broaden the methodologies for designing a range of fluorescent probes.^{29–31} This mechanism also helps explain why a solution of probe **1** (2 μ M) in the presence of 20 μ M of Al³⁺ when allowed to stand at room temperature (25 °C), shows blue fluorescence that gradually grows more intense before reaching a maximum within 72 h, whereas an elevated temperature (50 $^{\circ}$ C) could promote the generation of fluorescence within 1 h.

To understand the mechanism by which probe **1** was hydrolyzed in the presence of $Al^{3+} a {}^{1}H$ NMR titration of probe **1** with $Al(ClO_4)_3$ was checked in CD₃OD as shown in Fig. 7. The ${}^{1}H$ NMR titration spectra showed the concomitant development of a CH_3CH_2 peak for ethanol and the disappearance of a CH_3CH_2 signal of the ester within 24 h. This supports the assumption that the fluorescence of complex **1**- Al^{3+} is generated by ester hydrolysis **1** with Al^{3+} as a promoter.

The above mechanistic proposal is further supported by the fluorescent and non-fluorescent behaviors of the control probes **2** and **3** in the presence of Al^{3+} (Fig. S-4). Probe **2** produce identical changes in the photophysical properties with Al^{3+} as probe **1** except that it showed emission selectivity toward Ga^{3+} , Zn^{2+} and Cd^{2+} along with Al^{3+} (Fig. 8). On the other hand, probe **3** did not produce



Fig. 4. Fluorogenic changes of a 2 μ M solution of probe 1 in EtOH in the presence of 10 equiv of different cations upon illumination at 365 nm.



Fig. 5. Fluorescence titration spectra of probe **1** (2 μ M) with Al(ClO₄)₃ in EtOH upon heating at 50 °C. λ_{ex} =364 nm. Inset: mol ratio plots of emission at 434 nm.



Fig. 6. Job's plot for probe 1 (20 μM) with Al(ClO₄)₃ (20 μM) in EtOH upon heating at 50 °C for 1 h. $\lambda_{ex}{=}364$ nm.

any change in absorption or emission upon the addition of 10 equiv of Al^{3+} due to the absence of phenolic–OH because it is unable to exhibit three ways coordination with Al^{3+} on a similar line of that observed with probes **1** and **2** (Fig. S-4). This suggests that Al^{3+} first binds to the –OH of the phenol and causes the hydrolysis of the ester. A complex of probe **1** with Al^{3+} in EtOH was also prepared and characterized by HR-FAB mass. The mass spectra of probe **1**- Al^{3+} showed 1:1 stoichiometry (between probe **1** and Al^{3+}) with the molecular ion peak at m/z, 263.9903, which corresponds to (probe **1**+ Al^{3+} –EtOH+H₂O–H)⁺ (Fig. S-5). This also provides evidence that the ester is hydrolyzed to an acid by Al^{3+} followed by complexation.

3. Conclusions

In summary, a thiazole based 'turn-on' fluorescence probe (1) for Al^{3+} was prepared. For the first time, such high selectivity to Al^{3+} over other cations was observed due to the Al^{3+} -promoted hydrolysis of the ester to give an acid that binds to Al^{3+} resulting in a dramatic change in fluorescence by CHEF.

4. Experimental section

4.1. General

The melting points were determined using a Thomas-Hoover capillary melting point apparatus and were uncorrected. The ¹H and ¹³C NMR spectra were recorded on a Bruker AM-400 spectrometer using Me₄Si as the internal standard. The FAB mass was determined at KBSI Daegu branch. Infrared spectra were recorded using a Shimadzu Prestige-21 spectrometer on a KBr pellet. The UV-vis absorption spectra were determined on a Shimadzu UV-1650PC spectrophotometer. The fluorescence spectra were measured on a Shimadzu RF-5301 fluorescence spectrometer equipped with a xenon discharge lamp and 1 cm quartz cells with slit 3. All the measurements were taken at 298 K. Analytical grade ethanol was purchased from Merck. All other materials for syntheses were purchased from Aldrich Chemical Co. and used as received. 2-Hydroxyphenylthiobenzamide was prepared by the procedure reported in the literature.⁶ Solutions of metal ions were prepared from their perchlorate salts of analytical grade, and subsequently diluted to prepare the working solutions. The quantum yield (ϕ) was calculated using the procedure reported in the literature.^{4–7}

4.2. Synthesis of probe

4.2.1. Ethyl 2-(2-hydroxyphenyl)thiazole-4-carboxylate (1). A solution of 2-hydroxyphenyhlthiobenzamide 4 (300 mg, 1.96 mmol) and ethyl bromopyruvate (458 mg, 2.35 mmol) in ethanol (15 mL) was heated under reflux for 2 h. The reaction progress was monitored by TLC analysis. The solvent was removed in vacuo, and the residue was washed with water and extracted with EtOAc. The organic laver was dried over anhydrous Na₂SO₄ and concentrated. The residue was purified by column chromatography (EtOAc:hexane=1:4) to give compound **1** as a colorless needle shaped solid (350 mg, 1.40 mmol) in 71% yield. Mp 109–111 °C (CH₂Cl₂-hexane); IR (KBr, cm⁻¹) 3140, 2978, 1720, 1474, 1211; ¹H NMR (400 MHz, CDCl₃) δ 8.09 (s, 1H), 7.60 (dd, J=7.8, 8.8 Hz, 1H), 7.35 (m, 1H), 7.08 (dd, J=8.4, 8.4 Hz, 1H), 6.92 (m, 1H), 4.42 (q, J=7.1 Hz, 2H), 1.41 (t, J=7.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 169.8, 160.9, 157.4, 146.5, 132.9, 127.7, 125.4, 119.9, 118.5, 116.7, 62.0, 14.7; Anal. Calcd for C₁₂H₁₁NO₂S: C, 57.82; H, 4.45; N, 5.62; S, 12.86, found: C, 57.78; H, 4.36; N, 5.66; S, 13.12. HR-mass Calcd for: C₁₂H₁₁NO₃S [M+H]⁺: 249.0460; found: *m*/*z* 249.0457.

4.2.2. 2-(2-Hydroxyphenyl)thiazole-4-carboxyic acid (2). Compound 1 (100 mg, 0.65 mmol) and 5 mL of 1 N KOH in ethanol (25 mL) was heated under reflux for 2 h. The reaction progress was monitored by TLC. The solvent was removed in vacuo, and the



Scheme 1. Al³⁺-promoted hydrolysis of probe 1.



Fig. 7. ¹H NMR spectrum of probe **1** (0.04 mM) in the presence of 1.0 equiv of AI^{3+} ion after 24 h at 25 °C in CD₃OD (detail titration in Fig. S-3).



Fig. 8. Fluorescent spectra of probe 2 (2 μM) upon the addition of 10 equiv of various cations in EtOH. $\lambda_{ex}{=}364$ nm.

residue was washed with water, acidified with 10 mL of 1 N HCl and extracted with CH₂Cl₂. The organic layer was dried over anhydrous Na₂SO₄ and concentrated. The residue was purified by column chromatography (10% MeOH in CH₂Cl₂) to give compound **2** as a white solid (82 mg, 0.37 mmol) in 90% yield. Mp 249–251 °C (CH₂Cl₂–hexane); IR (KBr, cm⁻¹) 3450, 3125–2577, 1690, 1481, 1269; ¹H NMR (400 MHz, DMSO-*d*₆) δ 6.98 (dd, *J*=7.8, 7.3 Hz, 1H), 7.05 (d, *J*=8.1 Hz, 1H), 7.34 (dd, *J*=8.0, 7.3 Hz, 1H), 8.13 (d, *J*=7.8 Hz, 1H), 8.46 (s, 1H), 11.33 (s, 1H), 13.05 (s, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 164.0, 162.6, 155.6, 146.4, 131.9, 128.8, 128.0, 112.0, 119.1, 116.9; Anal. Calcd for C₁₀H₇NO₃S: C, 54.29; H, 3.19; N, 6.33; S, 14.49,

found: C, 54.57; H, 3.20; N, 6.45; S, 14.40. HR-mass Calcd for: $C_{10}H_7NO_3S$ [M+H]⁺: 222.0225; found: *m/z* 222.0221.

4.2.3. Ethyl 2-(2-phenyl)thiazole-4-carboxylate (**3**). This compound was obtained by the same procedure for synthesizing compound **1** using thiobenzamide (200 mg, 1.46 mmol) and ethyl bromopyruvate (341 mg, 1.75 mmol) in ethanol (15 mL). The product was purified by column chromatography (10% EtOAc in hexane) to give compound **3** as a colorless needle shaped solid in 70% yield. Mp 120–121 °C (CH₂Cl₂–hexane); ¹H NMR (400 MHz, CDCl₃) δ 7.78 (d, *J*=8.0 Hz, 2H), 7.77 (s, 1H), 7.31 (t, *J*=8.0 Hz, 3H), 4.34 (q, *J*=8.0 Hz, 2H), 1.34 (t, *J*=7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 169.5, 162.0, 148.3, 133.0, 129.4, 129.13, 128.4, 127.8, 127.7, 127.3, 62.1, 14.8; Anal. Calcd for C₁₂H₁₁NO₂S: C, 61.78; H, 4.75; N, 6.00; S, 13.74; found: C, 61.72; H, 4.69; N, 6.03; S, 13.67.

4.2.4. **1**- Al^{3+} complex. A mixture of compound **1** (100 mg, 0.45 mmol) and Al(ClO₄)₃·9H₂O (263 mg, 0.54 mmol) in ethanol (10 mL) was heated under reflux for 1 h. The mixture was cooled to room temperature, and the ethanol was evaporated and the residue was dried under vacuum to provide **1**- Al^{3+} complex as a white solid extremely hygroscopic in nature (110 mg, 92% yield). HR-FAB Mass: Calcd for (C₁₀H₇NO₄S·Al) 263.9911; found: 263.9903.

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Supplementary data

Figures of competitive binding of probe $1-Al^{3+}$, method of determination of detection limit of Al^{3+} by probe 1, ¹H NMR experiments spectra of probe 1, absorbance (UV–vis), fluorescence spectra of probes 1-3, and HR-FAB mass of probe $1-Al^{3+}$ complex are available. Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.tet.2013.05.062.

References and notes

- Kaur, R.; Saini, R.; Kumar, A.; Luxami, V.; Kaur, N.; Singh, P.; Kumar, S. Coord. Chem. Rev. 2012, 256, 1992–2028.
- 2. Jun, M. E.; Roy, B.; Ahn, K. H. Chem. Commun. 2011, 7583-7601.
- 3. Cho, D. G.; Sessler, J. L. Chem. Soc. Rev. 2009, 38, 1647-1662.
- 4. Helal, A.; Kim, S. H.; Kim, H.-S. Tetrahedron 2010, 66, 9925–9932.
- 5. Helal, A.; Lee, S. H.; Kim, S. H.; Kim, H.-S. Tetrahedron Lett. 2010, 51, 3531-3535.
- 6. Helal, A.; Kim, H.-S. Tetrahedron Lett. 2009, 50, 5510-5515.

- 7. Helal, A.; Rashid, M. H. O.; Choi, C. H.; Kim, H.-S. Tetrahedron 2011, 67, 2794–2802.
- 8. (a) Datta, B. K.; Kar, C.; Bau, A.; Das, G. Tetrahedron Lett. 2013, 54, 771-774; (b) Upadhyay, K. K.; Kumar, A. Org. Biomol. Chem. **2010**, 8, 4892–4897; (c) Ma, T.-H.; Dong, M.; Dong, Y.-M.; Wang, Y.-W.; Peng, Y. Chem.-Eur. J. 2010, 16,
- 10313-10318. 9. (a) Oliveira, E.; Santos, H. M.; Capelo, J. L.; Lodeiro, C. *Inorg. Chim. Acta* 2012, *381*, 203–211; (b) Lu, Y.; Huang, S.; Liu, Y.; He, S.; Zhao, L.; Zeng, X. Org. Lett. 2011, *13*, 5274–5277; (c) Kim, S. H.; Choi, H. S.; Kim, J.; Lee, S. J.; Quang, D. T.; Kim, J. S. Org. Lett. 2010, 12, 560-563.
- 10. Samanta, S.; Nath, B.; Baruah, J. B. Inorg. Chem. Commun. 2012, 22, 98-100.
- Kim, S.; Noh, J. Y.; Kim, K. Y.; Kim, H. H.; Kang, H. K.; Nam, S.-W.; Kim, S. H.; Park, 11. S.; Kim, C.; Kim, J. Inorg. Chem. 2012, 51, 3597–3602.
- Park, H. M.; Oh, B. N.; Kim, J. H.; Qiong, W.; Hwang, I. H.; Jung, K. D.; Kim, C.; Kim, J. *Tetrahedron Lett.* **201**, 52, 5581–5584.
- 13. Sahana, A.; Banerjee, A.; Das, S.; Lohar, S.; Karak, D.; Sarkar, B.; Mukhopadhyay, S. K.; Mukaherjee, A. K.; Das, D. Org. Biomol. Chem. **2011**, 9, 5523–5529.
- 14. Jiang, X.; Wang, B.; Wang, Z.; Liu, Y.; Li, T.; Liu, Z. Inorg. Chem. Commun. 2011, 14, 1224-1227. 15
- Wang, J.-Q.; Huang, L.; Gao, L.; Zhu, J. H.; Wang, Y.; Fan, X.; Zou, Z. *Inorg. Chem. Commun.* **2008**, *11*, 203–206.
- 16. Kim, S. D.; Lee, D. H.; Kim, J. S. Bull. Korean Chem. Soc. 2008, 29, 245-248.

- 17. Li, X.; Wang, J.; Sun, L.; Wang, Z. Chem. Commun. 2010, 988-990.
- 18. Zhao, Y.; Lin, Z.; Liao, H.; Duanand, C.; Meng, O. Inorg. Chem. Commun. 2006, 9,
- 966-968. 19. Keck, J.; Kramer, H. E. A.; Port, H.; Hirsch, T.; Fischer, P.; Rytz, G. J. Phys. Chem. 1996, 100, 14468-14475.
- Farruggia, G.; Savage, P. B. J. Am. Chem. Soc. 2006, 128, 344–350.
 Mameli, M.; Aragoni, M. C.; Lippolis, V. Chem. Lett. 2009, 48, 9236–9249.
- 22. Thordarson, P. Chem. Soc. Rev. **2011**, 40, 1305–1323.
- 23. Forgues, S. F.; LeBris, M. T.; Gutte, J. P.; Valuer, B. J. Phys. Chem. 1988, 92, 6233-6237.
- 24. Shortreed, M.; Kopelman, R.; Kuhn, M.; Hoyland, B. Anal. Chem. 1996, 68, 1414-1418.
- 25. Su, Z.; Lee, H. W.; Kim, C. K. Org. Biomol. Chem. 2011, 9, 6402-6409.
- Gunnlaugsson, T.; Lee, T. C.; Parkesh, R. Org. Lett. 2003, 5, 4065–4068.
 Kim, J. S.; Noh, K. H.; Lee, S. H.; Kim, S. K.; Yoon, J. J. Org. Chem. 2003, 68, 597-600.
- Kim, J. S.; Shon, O. J.; Rim, J. A.; Kim, S. K.; Yoon, J. J. Org. Chem. 2002, 67, 2348–2351. 28
- 29. Polyzos, A.; Hughes, A. B.; Christie, J. R. Langmuir 2007, 23, 1872–1879.
- 30. Fife, T. H.; Przystas, T. J. J. Am. Chem. Soc. 1985, 107, 1041-1047.
- 31. Kovacs, J.; Mokhir, A. Inorg. Chem. 2008, 47, 1880–1882.