

SALVIANDULINE C, A 5,6-SECOCLERODANE DITERPENOID FROM *SALVIA LAVANDULOIDES**

EMMA MALDONADO,† JORGE CÁRDENAS, BEATRIZ SALAZAR, RUBÉN A. TOSCANO, ALFREDO ORTEGA, CHRISTOPHE K. JANKOWSKI, ANDRÉ AUMELAS‡ and MARIE R. VAN CALSTEREN‡§

Instituto de Química, Universidad Nacional Autónoma de México, Circuito Exterior, Ciudad Universitaria, Coyoacán 04510 México, D. F.; ‡Faculté des Etudes Supérieures et de la Recherche, Université de Moncton, Moncton, N. B., Canada E1A 3E9

(Received 15 April 1991)

Key Word Index—*Salvia lavanduloides*; Labiatae; diterpene; 5,6-secoclerodane.

Abstract—The structure and configuration of salvianduline C, a novel 5,6-secoclerodane diterpenoid isolated from *Salvia lavanduloides*, have been established by high resolution NMR spectroscopy and X-ray crystallographic analysis.

INTRODUCTION

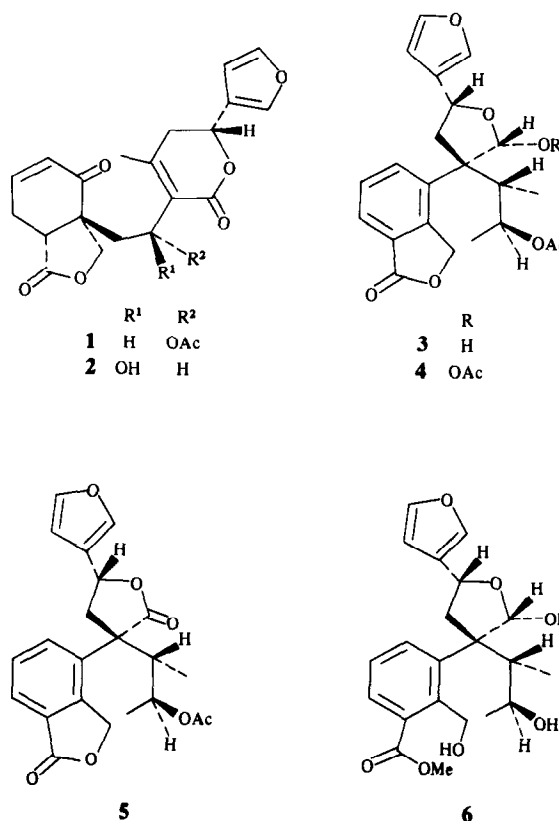
Previously [1], we described the characterization of salviandulines A (1) and B (2), two 9,10-secoclerodane diterpenoids from the aerial parts of *Salvia lavanduloides* L. We now describe the isolation and structural elucidation of a further secoclerodane, salvianduline C (3), from the same plant.

RESULTS AND DISCUSSION

The new compound (3) had the molecular formula $C_{22}H_{24}O_7$ by mass spectrometry ($[M]^+$ at m/z 400). Its IR spectrum showed absorptions for a hydroxyl group, an α - β unsaturated- γ -lactone, an ester and a β -substituted furan ring. The 1H -H and 1H - ^{13}C COSY and COLOC spectra allowed us to propose the secoclerodanic structure 3 for salvianduline C.

The ^{13}C NMR spectrum (Table 1) revealed the presence of a trisubstituted benzene. Thus it contained six aromatic carbons signals, three as singlets at δ 126.0 (C-4), δ 145.2 (C-5) and δ 136.5 (C-10) and three as doublets at δ 135.4 (C-1), δ 128.7 (C-2) and δ 124.2 (C-3). The last signal correlates in the 1H - ^{13}C COSY spectrum with a double doublet signal at δ 7.82 ($J=6$ and 3 Hz, H-3) which is *ortho* to H-2 (δ 7.46 t, $J=6$ Hz) and *meta* to H-1 (δ 7.4–7.5), so establishing the substitution pattern of the aromatic ring as vicinal trisubstituted. This aromatic ring should correspond to the ring A of a clerodane diterpene in which the aromatization of ring A proceeds with cleavage of the C-5/C-6 linkage, but retention of the C-18 C-19- γ -lactone ring system (IR 1762 cm^{-1} ; ^{13}C : δ 170.2 (C-18), δ 71.3 (C-19); 1H NMR δ 5.57 d, $J=15$ Hz (H-19), δ 5.46 d, $J=15$ Hz (H-19'), the presence of which is frequently present in clerodanes from Mexican *Salvia* species [2]. Furthermore, the above assumptions also

account for the presence of a primary and secondary methyl groups in salvianduline C. The primary methyl group was responsible for the doublet at δ 1.23 ($J=6.5$ Hz, H-6) and the signal at δ 20.1 (C-6) in the 1H and ^{13}C NMR spectra. It was located at C-7 and geminal to an oxygenated function, because of the chemical shift of the H-7 signal, which appeared as a quartet at δ 5.28 ($J=6.5$ Hz). The secondary methyl group was located at C-8, as the H-8 signal was also a quartet (δ 1.93, $J=7$ Hz).



*Contribution No. 1069 of Instituto de Química, UNAM.

†Author to whom correspondence should be addressed.

§Permanent address: CRASH, 3600 Bv Casavant, St. Hyacinthe, QC, Canada.

and the ^{13}C signal for this methyl (C-17) was observed at $\delta 10.0$. It is interesting that there was no coupling between H-7 and H-8, indicating a rigid conformation of the chain (C-6/C-9) bonded to C-10. This can be explained if the acetoxy group (IR 1728 cm^{-1} ; ^{13}C $\delta 170.2$ and 20.7) is located at C-7. At this position, the ester group can settle over the aromatic ring, which induces a strong diamagnetic shift upon the acetyl methyl group ($\delta 1.38$) [3, 4]. The NMR spectra also showed signals attributable to a β -substituted furan at $\delta 6.44$ (H-14), 7.38 (H-15) and 7.44 (H-16), and $\delta 126.9$, 109.0 , 143.9 and 140.2 (C-13 to C-16, respectively).

The remaining signals correspond to a γ -lactol closed to C-12 ($\delta 72.2$). The H-12 signal appeared as a doublet at $\delta 5.18$ coupled with H-11 β ($\delta 2.92$) and H-11 α ($\delta 2.61$) in an axial-equatorial and axial-axial relationship. The hemiacetalic proton, H-20, appeared as a doublet at $\delta 5.77$ by interaction with the hydroxylic proton ($\delta 2.88$). From the above data structure **3** was proposed for *salvianduline C*.

Acetylation of **3** confirmed the presence of the C-20 hydroxyl group, because in the ^1H NMR spectrum of the acetoxy derivative **4**, the H-20 signal was shifted downfield, appearing at $\delta 6.82$ and the signals for the methyl acetates at C-7 and C-20 were observed at $\delta 1.35$ and 1.77 , respectively. In this spectrum, the signals for the aromatic protons H-1, H-2 and H-3 were clearly observed at $\delta 7.3$ (dd , $J = 7$ and 2 Hz), $\delta 7.48$ (t , $J = 7$ Hz) and $\delta 7.82$ (dd , $J = 7$ and 2 Hz), thus confirming the existence of a vicinal trisubstituted benzene ring.

The formation of the dilactone **5** on oxidation of **3**, gave further support to the proposed structure. In the ^1H NMR spectrum of **5**, the H-20 signal was not observed and the H-12 signal was shifted downfield ($\delta 5.54$), while the ^{13}C NMR spectrum (Table 1) showed an additional

carbonyl signal at $\delta 175.6$ (C-20). In this compound, the signal for the ester methyl group appeared at lower field ($\delta 1.92$) probably due to a conformational change in the molecule.

Basic hydrolysis of **3** followed by methylation of the resulting product gave **6**. Its ^1H NMR spectrum showed the H-7 signal at $\delta 3.54$ as a quartet of doublets as result of interaction with H-6 and H-8. In the IR spectrum, γ -lactone and the acetyl absorptions were replaced by an aromatic carbomethoxy band at 1688 cm^{-1} .

In order to establish the relative stereochemistry of **3**, NOE differential spectra were determined. These allowed us to postulate that H-12, H-11 β , H-20 and the chain on C-9 were at the same side of the lactol ring (Fig. 1). However, this information was not enough to establish the orientation of the substituent at C-7 and C-8. Therefore, X-ray analysis of **3** was performed. It showed that the isobenzofuran-18(19H)-one and the furan ring are essentially planar (maximum deviation of C-19 is 0.055 \AA) and almost perpendicular (84.7° between least-squares planes). The γ -lactol ring displays an β -envelop conformation, with C-9 as the flap, in which the hydroxyl at C-20 in α -orientation holds an *anti* relation to the side chain. In this conformation the acetoxy group at C-7 lies then almost coplanar to the isobenzofuranone in order to minimize steric repulsion, confirming the upfield signal for this group observed in the ^1H NMR spectrum. Application of the Karplus equation to the solid state conformation ($\theta[\text{H-7/C-7/C-8/H-8}] = 70.3 (1^\circ)$ in order to estimate the vicinal coupling constants expected for H-7 and H-8 and comparison with the observed value, confirms the extreme rigidity of the side chain.

The 5,6-secoclerodane diterpene structure for *salvianduline C* (**3**) was thus confirmed to be as shown in Fig. 1 [stereochemistry relative to C-12 (*R*)]. To our knowledge, the diterpenoid **3** is only the second one of this type to be found so far in nature [5].

Table 1. ^{13}C NMR spectral data of compounds **3** and **5** (CDCl_3 , 90 MHz)

C	3	5
1 <i>d</i>	135.4	133.3
2 <i>d</i>	128.7	129.5
3 <i>d</i>	124.2	125.6
4 <i>s</i>	126.0 ^a	127.3
5 <i>s</i>	145.2	144.0
6 <i>q</i>	20.1	20.3
7 <i>d</i>	68.7	69.2
8 <i>d</i>	42.3	45.4
9 <i>s</i>	60.9	55.4
10 <i>s</i>	136.5	136.0
11 <i>t</i>	39.8	40.4
12 <i>d</i>	72.2	71.7
13 <i>s</i>	126.9 ^a	124.1
14 <i>d</i>	109.0	107.8
15 <i>d</i>	143.9	144.3
16 <i>d</i>	140.2	139.8
17 <i>q</i>	10.0	9.9
18 <i>s</i>	170.2	170.5
19 <i>t</i>	71.3	70.4
20	99.4	175.6
OAc <i>s</i>	170.2	170.2
<i>q</i>	20.7	21.1

^a Values may be interchangeable.

EXPERIMENTAL

General. ^{13}C NMR: 90 MHz; ^1H NMR: 80 and 360 MHz 1D spectra: 16 K time domain addresses and processed with 32 K addresses (accuracy 0.24 Hz/point). The CDCl_3 proton residual signal was taken as reference (7.24 ppm). Connectivities between protons were established by 2D correlated spectroscopy (COSY) using pulse sequences $\text{D}_1\text{-}90^\circ\text{-D}_0\text{-}45^\circ\text{-FID}$. COSY was obtained through $512\text{ }t_1$ values and, prior to Fourier transformation, the t_2 and t_1 domain FID_s were multiplied with a non-shifted sine-bell window function. All spectra are available on request.

Isolation of compound 3. Ground, dried aerial parts of *Salvia lavanduloides* (2460 g) collected in Morelos, México (voucher deposited in the Herbarium of the Instituto de Biología, U.N.A.M. AOH 214) were extracted with Me_2CO and MeOH to give 296.1 and 70.5 g of extracts, respectively. The Me_2CO extract was partitioned between MeOH- H_2O (4:1) and hexane- C_6H_6 (1:1) to give 185.2 and 88.3 g of residues of the polar and non-polar phases, respectively.

The polar phase was percolated through activated charcoal. The decoloured material was fractioned by CC (silica gel Merck G; hexane-EtOAc gradient elution). Frs eluted with hexane-EtOAc (7:3) contained 3.61 g of compound **3**, which was purified by recrystallization from EtOAc-hexane, mp $96\text{--}98^\circ$ (occludes EtOAc in the crystalline lattice) mp $177\text{--}179^\circ$ from Et₂O-hexane, $[\alpha]_D^{25} + 31.87$ (CHCl_3 ; c 0.251); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 227 (6599), 274 (1537), 280 (1510); IR $\nu_{\text{max}}^{\text{CHCl}_3}\text{ cm}^{-1}$: 3589, 3420,

Table 2. ^1H NMR spectral data of compounds 3–6, (80 MHz, CDCl_3 , TMS as int. standard)

H	3 (360 MHz)	4	5 (360 MHz)	6
1	7.4–7.5	7.30 <i>dd</i> 7, 2	7.89 <i>d</i> 7.6	
2	7.46 <i>t</i> 6	7.48 <i>t</i> 7	7.53 <i>t</i> 7.6	
3	7.82 <i>dd</i> 6, 3	7.82 <i>dd</i> 7, 2	7.89 <i>d</i> 7.6	7.9 <i>dd</i> 6, 3
6	1.23 <i>d</i> 6.5	1.23 <i>d</i> 7	1.20 <i>d</i> 6.5	1.04 <i>d</i> 7
7	5.28 <i>q</i> 6.5	5.17 <i>q</i> 7	5.03 <i>qd</i> 6.5, 1.2	3.54 <i>qd</i> 7, 1
8	1.93 <i>q</i> 7	1.98 <i>q</i> 7	2.25 <i>qd</i> 7.1, 1.2	2.11 <i>qd</i> 7, 1
11	2.61 <i>dd</i> 13, 10	2.55 <i>dd</i> 12, 10	3.05 <i>dd</i> 13.5, 7.5	2.49 <i>dd</i> 13, 10
11	2.92 <i>dd</i> 13, 7	2.90 <i>dd</i> 12, 6	2.66 <i>dd</i> 13.5, 7	2.19 <i>dd</i> 13, 5
12	5.18 <i>dd</i> 10, 7	5.29 <i>dd</i> 10, 6	5.54 <i>ddd</i> 7.5, 7, 0.8	4.56 <i>dd</i> 10, 5
14	6.44 <i>dd</i> 2, 1	6.34 <i>m</i> $W/2=4$	6.20 <i>dd</i> 1.8, 0.8	6.34 <i>m</i> $W/2=4$
15	7.38 <i>t</i> 2	7.37 <i>m</i>	7.35 <i>dd</i> 1.8, 1.7	
16	7.44 <i>m</i>	7.37 <i>m</i>	7.28 <i>dt</i> 1.7, 0.8	
17	1.23 <i>d</i> 7	1.23 <i>d</i> 7	1.17 <i>d</i> 7.1	1.20 <i>d</i> 7
19	5.57 <i>d</i> 15	5.47 <i>d</i> 14	5.39 <i>s</i> (2 H)	5.35 <i>d</i> 16
19'	5.46 <i>d</i> 15	5.25 <i>d</i> 14		5.12 <i>d</i> 16
20	5.77 <i>d</i> 1.1	6.82 <i>s</i>		5.84 <i>s</i>
MeCO ₂	1.38 <i>s</i>	1.35 <i>s</i> 1.77 <i>s</i>		
OH or OMe	2.88 <i>d</i> (OH) 1.1			3.37 <i>s</i> (OMe)

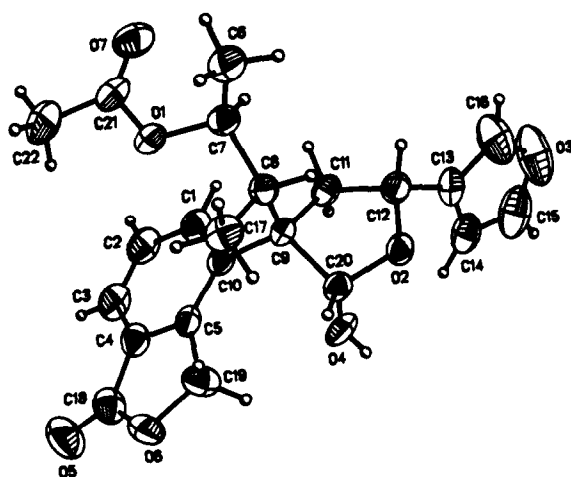


Fig. 1. The molecular conformation of salvianduline C (3), showing atom numbering.

1762, 1729, 1529, 1592, 872; EIMS m/z (rel. int.): 400 $[\text{M}]^+$ (23), 340 (35), 294 (15), 279 (19), 265 (24), 239 (28), 195 (18), 115 (29), 95 (37), 81 (36), 43 (100); HR MS 400.1853 (calc. for $\text{C}_{22}\text{H}_{24}\text{O}_7$: 400.1864).

Acetylation of compound 3. A soln of 3 (100 mg) in pyridine (1 ml) and Ac_2O (1 ml) was left to stand by 4 hr and worked-up as usual to give 83.1 mg of 4 as an oil: IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1765, 1735, 942, 871. EIMS m/z : 442 $[\text{M}]^+$ (2.5); 400 (1), 382 (0.2); 358 (0.2); 340 (1); 322 (3); 294 (3); 279 (4); 267 (7); 239 (4); 221 (3); 115 (8); 95 (15); 81 (19); 43 (100). HR MS 442.1596 (calc. for $\text{C}_{24}\text{H}_{26}\text{O}_8$: 442.1620).

Oxidation of compound 3. Jones reagent was added dropwise to a soln of 3 (101.3 mg) in Me_2CO (10 ml) at 0° . After the usual work-up 96 mg of 5 were obtained: mp $191\text{--}194^\circ$ (EtOAc–hexane), IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1768, 1732, 928, 873; EIMS m/z (rel. int.): 398 $[\text{M}]^+$ (10); 356 (6); 338 (3); 282 (39); 245 (14); 236 (12); 221 (9); 115 (22); 95 (79); 81 (28); 43 (100). HR MS 398.1700 (calc. for $\text{C}_{22}\text{H}_{22}\text{O}_7$: 398.1716).

Synthesis of compound 6. Compound 3 was added to an aq. soln of NaOH (40%, 15 ml). The suspension was stirred until complete solubilization, then neutralized with aq. HCl (10%),

extracted with EtOAc and dried over Na_2SO_4 . The residue was dissolved in MeOH, cooled (0°) and stirred. Then ethereal CH_2N_2 was added dropwise until the reaction was complete. After solvent evapn, a residue obtained, which after CC afforded 34 mg of **6** as a gum, IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 3472, 1714, 1688, 1591, 991, 875; EIMS m/z (rel. int.): 390 [$\text{M}]^+$ ($\text{C}_{21}\text{H}_{26}\text{O}_7$; unobserved); 372 (10); 354 (4); 341 (5); 260 (17); 243 (23); 221 (14); 171 (65); 143 (49); 128 (45); 115 (43); 97 (90); 95 (92); 69 (63); 58 (100); 41 (72).

X-Ray crystallography of 3. A crystal of salvianduline C (**3**) was mounted on a R3M Nicolet automated diffractometer. Crystal size: $0.20 \times 0.21 \times 0.40$ mm. Unit cell dimensions: $a = 10.300$ (5), $b = 12.803$ (6), $c = 19.250$ (7) Å, (orthorhombic P_{212121}) were determined by least-squares refinement of the best angular positions for 25 independent reflections in the range $5^\circ < 2\theta < 14^\circ$ using MoK_α radiation ($\lambda = 0.71073$ Å). Data (2558 reflections) were collected at room temp., using $\theta/2\theta$ scan mode to a maximum 2θ value of 50° . The intensities of two standard reflections were measured every 50 reflections and as the intensities of these reflections showed less than 3% variation corrections for decay were deemed unnecessary. Intensities were corrected for Lorentz and polarization effects, but no absorption correction was made. A total of 1932 reflections were considered [$F > 3 \sigma(F)$]. The structure was solved by direct methods included in SHELXTL package [6] to located 28 of the 35 non-hydrogen atoms having the structure. Successive cycles of least-squares blocked-cascade refinements, followed by difference Fourier synthesis, allowed location of the remainder non-H atoms. Refinements of scale factor, positional and anisotropic thermal parameters for all-non-hydrogen atoms was carried out to convergence minimizing the function $\Sigma w (|F_o| - |F_c|)^2$. All H-atoms attached to C-atoms were located at geometric positions with constant $U = 0.06$ Å². The hydroxylic H-atom was located on a difference Fourier synthesis at advanced stage and refined isotropically. The final cycle of refinements led to a final agree-

ment factor $R = 0.06$ ($R_w = 0.06$) with maximum residual density in the final difference map of ± 0.24 .

Atomic scattering factor as for International Tables for X-ray Crystallography [7], were calculated on a NOVA 4S computer using the SHELXTL system.

Acknowledgements—We are greatly indebted to R. Gaviño, L. Velasco, M. Torres, J. Espiñeira y R. Villena for technical assistance. We thank Dr T. P. Ramamoorthy (Botany Department, Instituto de Biología, U.N.A.M.) for botanical classification of the plant material

REFERENCES

1. Ortega, A., Cárdenas, J., Toscano, A., Maldonado, E., Aumelas, A., Van Clasteren, M. R. and Jankowski, K. (1991) *Phytochemistry* (in press).
2. Rodríguez-Hahn, L., Esquivel, B., Sánchez, A. A., Sánchez, C., Cárdenas, J. and Ramamoorthy, T. P. (1987) *Rev. Latinoamer. Quím.* **18**, 104.
3. Vichniewski, W., Prasad, J. S. and Herz, W. (1984) *Phytochemistry* **23**, 1655.
4. Rodríguez-Hahn, L., Antúnez, L., Martínez, M., Sánchez, A. A., Esquivel, B., Soriano-García, M. and Toscano, A. (1986) *Phytochemistry* **25**, 1655.
5. Ortega, A., Mancera, C., Escamilla, E. and Cárdenas, J. (1988) *XV Symposium Internacional de Química de Productos Naturales*, Monterrey, N. L., México, April, 1988, p. 17.
6. Sheldrick, G. M. (1983) *An Integrated System for Solving, Refining and Displaying Crystal Structures from Diffraction Data SHELXTL*, Revision 4, University of Göttingen, Federal Republic of Germany.
7. *International Tables for X-Ray Crystallography* (1974) Vol. IV, pp. 202–207. Kynoch Press, Birmingham. (present distributor: D. Reidel, Dordrecht).