

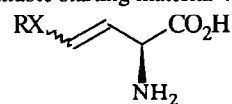
A NEW CLASS OF UNUSUAL α -AMINOACIDS : α -AMINOACID β,γ -ENOL THIOETHERS

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Summary : Two representative α -aminoacid β,γ -enol thioethers have been synthesized starting from the corresponding easily available saturated α -aminoacids by a Pummerer-like reaction.

In the course of our investigations in the field of potential enzyme inhibitors, we decided to synthesize α -aminoacids bearing unusual side chain functions from easily available starting material¹.

α -Aminoacid β,γ -enol thioethers **1** are a new class of exotic α -aminoacids to study since only one example : 2S-2-amino-4-methylthio-but-3E-enoic acid ("3,4-dehydro-L-methionine" **1**, R=CH₃, E) is described in the literature². Furthermore, none of its biological properties have been reported to date despite its potential antimetabolic activities. The oxygen-



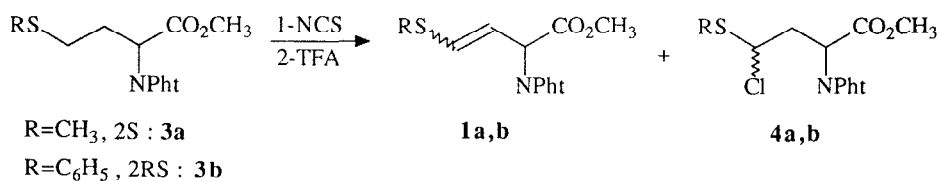
1 : X=S

2 : X=O

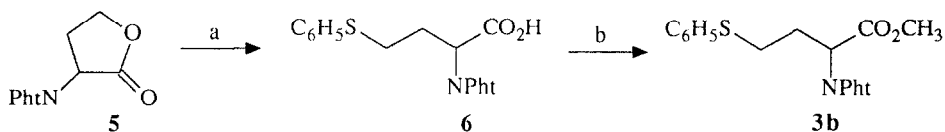
analogues, the α -aminoacid β,γ -enol ethers **2** are a well-known class of naturally occurring unusual α -aminoacids of biological interest³ : 2S-2-amino-4-methoxy-but-3E-enoic acid (L-trans-AMB) **2** (R=CH₃, E), Rhizobitoxine **2** (R=(R)-CH₂CH(CH₂OH)NH₂, E), 2S-2-amino-4-(2-aminoethoxy)-but-3E-enoic acid (AVG) **2** (R=CH₂CH₂NH₂, E) are potent inhibitors of enzymes involved in methionine metabolism. Moreover, synthetic 2S-2-amino-4-methoxy-but-3Z-enoic acid (L-cis-AMB) **2** (R=CH₃, Z) is a potent methionine analogue inhibitor of S-adenosylmethionine synthetase⁴. The thioethers **1** might have similar biological properties and could among others act as plant growth regulators by inhibition of ethylene biosynthesis in plants^{3c}. As methionine analogues, they are also potential anticancer agents⁴.

"3,4-Dehydro-L-methionine" **1** (R=CH₃, E) and L-trans-AMB **2** (R=CH₃, E) have been synthesized in a similar way starting from diethyl acetamidomalonate^{2,5}. Racemic fully protected "dehydromethionine" was thus obtained by a six step procedure in 10% overall yield, alkaline hydrolysis and enzymatic resolution leading to optically active aminoacid.

The present paper deals with a general and straightforward synthesis of β,γ -dehydrohomocysteine derivatives **1** starting from the corresponding saturated derivatives **3** based on a Pummerer-like reaction using N-chlorosuccinimide (NCS)⁶ to afford the transient chlorinated derivatives **4**, followed by an acid catalyzed dehydrochlorination⁷ (scheme 1).

**scheme 1** (NPht=NPhthaloyl)

The phthaloyl moiety was chosen as amino protecting group to avoid any internal amine participation⁸. N-Phthaloyl-L-methionine methyl ester **3a** was readily obtained by classical amino protection⁹ of L-methionine methyl ester hydrochloride using phthalic anhydride / triethylamine in toluene. N-Phthaloyl-S-phenyl-DL-homocysteine methyl ester **3b** was synthesized as outlined in scheme 2¹⁰ : after usual protection, lactone **5** was opened using sodium thiophenolate in refluxed THF / HMPA to obtain N-Phthaloyl-S-phenyl-DL-homocysteine **6**. Esterification using anhydrous sulfuric methanol yielded **3b**.

**scheme 2**

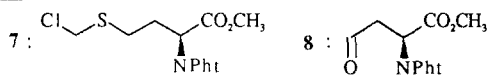
a- $\text{C}_6\text{H}_5\text{SH} : 2 \text{ eq.}, \text{NaH} : 2.2 \text{ eq.}, \text{HMPA} : 2.4 \text{ eq.}, \text{THF}, \text{reflux}, 24\text{h}, \text{yield} : 70\%$ b- $\text{H}_2\text{SO}_4, \text{CH}_3\text{OH}, \text{rt}, 18\text{h}, \text{yield} : 80\%$

α -Chlorination was chosen because of its regioselectivity towards branched carbon compared with methyl group^{11a,b}. Thus the fully protected homocysteine derivatives **3a,b** were subjected to Pummerer-like reaction using NCS in carbon tetrachloride or diethyl ether / benzene^{11c} (table 1).

sulfide	reaction conditions		% crude reaction products ^a					entry
	1	2	1 E	1 Z	4	7 ^c	8 ^c	
3a	NCS : 1.04 eq., CCl_4 0°C-reflux, 4 days	—	30	9	39	16	8	1
	NCS : 1.04eq., $\text{Et}_2\text{O}/\text{C}_6\text{H}_6$: 1/2 rt, 24h	—	14	2	66	15	2	2
		cat. TFA, CCl_4 reflux, 2h	36	9	32	17	7	3
3b	NCS : 1.04 eq.	—	0	0	100 ^b	—	0	4
	CCl_4	cat. TFA, CCl_4 rt : 17h	20	5	60	—	15	5
	rt, 16h	-20°C, 1 month	37	5	53	—	5	6

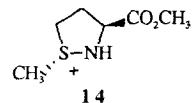
table 1

a-estimated by NMR ^1H in CDCl_3 ¹³; b-pure **4b** is obtained; c-



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- 12- HPLC conditions : Delta pak column Waters C18, 100 Å, 15 μ , flow rate : 10 ml/mn, eluent : CH₃OH/H₂O : 60/40 (**1a**) ; 70/30 (**1b**). In the case of **1b**, HPLC analysis is less efficient than for **1a**.
 The stereochemistry of the stereoisomers **10-I**, **10-II** has not been determined.
- 13- α -Aminoacid β,γ -enol thioethers :
 NMR (CDCl₃, δ ppm vs TMS). **1a-E** : ¹H, 200MHz, 2.26 (s, 3H, CH₃S) ; 3.75 (s, 3H, CO₂CH₃) ; 5.43 (d, 1H, 8.6Hz, CHN) ; 5.85 (dd, 1H, 15Hz, 8.6Hz, SCH=CH) ; 6.47 (dd, 1H, 15Hz, 0.8Hz, SCH=CH) ; 7.65-7.9 (m, 4H, NPh). ¹³C, 50.3MHz, 14.2 (CH₃S) ; 53, 53.7 (CHN, CO₂CH₃) ; 115.6 (SCH=CH) ; 132.9 (SCH=CH) ; 123.5, 131.7, 134.2 (C arom.) ; 166.8, 168.6 (CO). **1a-Z** : ¹H, 200MHz, 2.29 (s, 3H, CH₃S) ; 3.77 (s, 3H, CO₂CH₃) ; 5.77 (dd, 1H, 7.8Hz, 0.9Hz, CHN) ; 6.17 (dd, 1H, 7.8Hz, 9.4Hz, SCH=CH) ; 6.35 (dd, 1H, 9.4Hz, 0.9Hz, SCH=CH) ; 7.65-7.90 (m, 4H, NPh). ¹³C, 50.3 MHz, 17.2 (CH₃S) ; 49.7, 53 (CHN, CO₂CH₃) ; 119.4 (SCH=CH) ; 123.5, 131.7, 134.1 (C arom.) ; 134.2 (SCH=CH) ; 166.8, 168.6 (CO). **1b-E** : ¹H, 250MHz, 5.48 (dd, 1H, 8Hz, 0.8Hz, CHN) ; 6.28 (dd, 1H, 15Hz, 8Hz, SCH=CH) ; 6.55 (dd, 1H, 15Hz, 0.8Hz, SCH=CH). **1b-Z** : ¹H, 200MHz, 5.94 (d, 1H, 8Hz, CHN) ; 6.39 (dd, 1H, 8Hz, 9.3Hz, SCH=CH) ; 6.62 (dd, 1H, 9.3Hz, 0.8Hz, SCH=CH).
 MS : (CI in NH₃) : **1a-Z** 309 (M+NH₄)⁺, 292 (M+H)⁺ ; **1a-E** + **8-I** 341, 324, 309, 292, 276.
 Optical rotation : **1a-Z** [α]_D²⁵ = -32 (c=3.6, CHCl₃).



For the following compounds, only characteristic analytic data are indicated :

3a : m.p. 40-41°C (diethyl ether/petroleum ether) ; [α]_D²⁵ = -42 (c=1.49, CHCl₃). **4a** : ¹H NMR, 200MHz, δ 5.15, 4.95 (2m, SCHCl, CHN). **4b** : ¹H NMR, 250MHz, δ 5.2 (dd, 1H, 5.5Hz, 8.9Hz, CHN) ; 5.25-5.4 (m, 1H, SCHCl). **5** : m.p. 178-179°C (chloroform/petroleum ether). **6** : m.p. 153-154°C (methylene chloride/petroleum ether). **7** : ¹H NMR, 200MHz, δ 4.65 (s, ClCH₂S). **8** : ¹H NMR, 200MHz, δ 9.75 (t, 1H, 0.8Hz) ; [α]_D²⁵ = -9 (c=2.76, CCl₄). **9** : ¹H NMR, 200MHz, δ 4.60 (s, 2H, SCH₂O) ; [α]_D²⁵ = -31 (c=4.9, CHCl₃). **10-I** : ¹H NMR, 200MHz, δ 4.41 (m, 1H, OCHS) ; 5.14 (m, 1H, CHN) ; [α]_D²⁵ = +59 (c=6.3, CHCl₃). **10-II** : ¹H NMR, 250MHz, δ 4.14 (dd, 1H, 3.6Hz, 9.9Hz, OCHS) ; 5.13 (dd, 1H, 3.9Hz, 10.4Hz, CHN) ; [α]_D²⁵ = -70 (c=17.9, CHCl₃). **11** : ¹H NMR, 200MHz, δ 4.40 (t, 1H, 5.7Hz, OCHO) ; 5.00 (dd, 1H, 4.9Hz, 9.7Hz, CHN) ; [α]_D²⁵ = -20 (c=9.1, CHCl₃). **12** : ¹H NMR, 200MHz, δ 3.64 (s, 2H, SCH₂S) ; [α]_D²⁵ = -23 (c=3.2, CHCl₃). **13** : ¹H NMR, 200MHz, δ 3.68 (dd, 1H, 6.4Hz, 8.5Hz, SCHS) ; 5.37 (dd, 1H, 5.5Hz, 8.9Hz, CHN) ; [α]_D²⁵ = -29 (c=4.5, CHCl₃).

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