Synthesis and Characterization of Disulfide-Schiff Base Derivatives and *in vitro* Investigation of Their Antibacterial Activity Against Multidrug-Resistant *Acinetobacter baumannii* Isolates: A New Study¹

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Abstract—In this study two different methods without a catalyst and with a CeO_2 nano catalyst were used for the synthesis of dimeric disulfide-Schiff bases. The dimeric disulfide-Schiff base derivatives were characterized by FT-IR, NMR, and MS spectra, and elemental analysis. The disulfide-Schiff bases and their derivatives **2–5c** were screened for *in vitro* antibacterial activity against 40 multidrug-resistant strains of *Acinetobacter baumannii*, and their minimum inhibitory concentrations were determined. Most of products exhibited high antibacterial activity against *Acinetobacter baumannii*.

Keywords: dimeric disulfide-Schiff bases, antibacterial activity, *Acinetobacter baumannii*, minimum inhibitory concentration (MIC), nanoceria, multidrug-resistant **DOI:** 10.1134/S1070363218020184

INTRODUCTION

Metal nanoparticles have attracted considerable attention in synthetic organic chemistry due to their high catalytic activity and reusability. However, nano CeO₂ catalyzed reactions are not common in organic synthesis [1]. Condensation of 2,2'-diaminodiphenyl 2,2'-disulfide with different aromatic aldehydes gives rise to a series of interesting Schiff bases of SNNS, $N_2S_2O_2$ donors, with potential applications in medicine. Steric and electronic properties of these compounds can be finely adjusted, thus making them attractive in coordination and inorganic chemistry [2, 3].

Over recent years, there have been many reports on applications of Schiff base compounds as antibacterial, antifungal, anticancer, antioxidant, anti-inflammatory, antimalarial, and antiviral agents [4, 5].

The *Acinetobacter* species are gram negative, nonmotile, non-fermentative, opportunistic pathogens that are widely spread in soil and water and can colonize in hospitals [6]. Antimicrobial treatment of multidrugresistant *Acinetobacter baumannii* (MDRAB) poses a significant problem at present time [7].

RESULTS AND DISCUSSION

In the current study, the title compounds were synthesized for the first time using a CeO_2 nanocatalyst, thus adding a new approach to the existing methods. Synthesis of the dimeric disulfide-Schiff bases **3–5c** was carried out by reacting compound **2** with several aldehydes including 2-hydroxy benzaldehyde, 2,3-dihydroxy benzaldehyde, 2,4-dihydroxy benzaldehyde, 2,5-dihydroxy benzaldehyde, 2-hydroxy-3-methoxy benzaldehyde, 2-hydroxy-5-methoxy benzaldehyde. Synthesis of these dimeric disulfide-Schiff bases was carried out in two different ways without catalysts and using CeO₂ nanocatalyst (Schemes 1 and 2). Both methods were determined to be efficient, however, the yield of the process catalyzed by CeO₂ nanocatalyst

¹ The text was submitted by the authors in English.

Scheme 1. Synthesis of 2,2'-disulfanediyldianiline (2).



was considerably higher. The reaction time of the catalyzed reaction was reduced from hours to minutes (Table 1). The structures of all synthesized compounds were characterized by FT-IR, ¹H, and ¹³C NMR spectroscopic methods [8].

In IR spectra, except compounds 4a-4c, the characteristic bands of the OCH₃ and OH groups were not clearly observed probably due to the effect of tautomerism [9–12]. The characteristic for Schiff bases C=N group was recorded by sharp stretching vibra-

tions bands in the range of 1632–1606 cm⁻¹ [13, 14]. Specific peaks of $v(C-C_{Ar})$, $v(C-O_{Ar})$, v(C-S), and v(S-S) were observed in the range of 1485–1439, 1278–1225, 758–729, and 577–553 cm⁻¹, respectively. Intensity of vibrations depended on the presence of different functional groups such as CH₃, OH, and NH₂.

¹H and ¹³C NMR spectra of the synthesized compounds were measured in DMSO- d_6 and CDCl₃. The azomethine (H–C=N) proton was recorded as a singlet in the range of 8.54–9.04 ppm. Purity of compounds **2–5c** was determined by evaluating the melting point together with TLC and NMR spectra.

Microbiological assessment. Antibacterial activity of the some newly synthesized compounds was tested (Table 2). Meropenem was used as the standard antibiotic. Compounds **2**, **4a** and **5b** exhibited antibacterial effects against *A. baumannii* higher than the

Scheme 2. Synthesis of dimeric disulfide-Schiff base compounds 3–5c.



		Yield, %		Synthesis time	
Comp. no.	Structure	without CeO ₂ nanocatalyst	CeO ₂ nanocatalyst	without CeO ₂ nanocatalyst, h	CeO ₂ nanocatalyst, min
2	NH ₂	75	_	9	30
3	S ^{-S} H ₂ N	75	95	6	15
	S N N C H OH				
4 a		80	95	6	15
	S S N C H OH OH				
4b	S S N N C H OH OH	76	98	29	15

Table 1. Structures of synthesized dimeric disulfide-Schiff base compounds and reaction parameters

Table 1. (Contd.)

Comm	Structure	Yield, %		Synthesis time	
no.		without CeO ₂ nanocatalyst	CeO ₂ nanocatalyst	without CeO ₂ nanocatalyst, h	CeO ₂ nanocatalyst, min
4c	HO HO HO	74	92	6	15
5a	OH S N C H OH OH OH	70	96	5	15
5b	H_3CO	86	97	6	15
5c	$H_{3}CO \qquad \bigcup_{OH} OCH_{3} \qquad OCH_{3}$	90	96	5	15

other compounds and meropenem. This was particularly true for the compound 2.

Probably difference in the MIC results was due to the presence of one or two OH groups in the structures of the compounds. The compounds **4a**–**4c** containing two OH groups on each aromatic cycle were more effective than the compound **3**, that contained one OH group. The presence of two hydroxy groups in the ortho positions in the structure of the compound **4a** made its efficiency higher than compounds **4b** and **4c** that contained the OH groups in meta and para positions. Among compounds **5a**–**5c**, the compound **5a** demonstrated the lowest activity probably due to ortho position of the hydroxy and methoxy groups.

EXPERIMENTAL

All reagents were commercially available and purchased from Merck and Acros. The CeO_2 nanoparticles were prepared in accordance with the earlier developed method [15–17].

2,2'-Diaminodiphenyl disulfide was synthesized by oxidation of 2-aminothiophenol. The ligands were synthesized according to the earlier developed method using the CeO₂ nanocatalyst and without a catalyst [15].

Fourier transform infrared-attenuated total reflection spectroscopy (FTIR-ATR) data were recorded on a Perkin Elmer spectrophotometer. ¹H and ¹³C NMR spectra were measured in CDCl₃ and DMSO-*d*₆ on a Bruker-400 spectrometer using TMS as an internal standard. Elemental analyses were carried out using a Thermo Scientific Flash 2000. Mass spectra were measured on an AB SCIEX 4000 Q-TRAP LC-MS/ MS instrument.

2,2'-Disulfanediyldianiline (2). Shiny yellow solid, yield 75%, mp 90–92°C. IR spectrum, v, cm⁻¹: 3375 (NH₂), 1471 (C=C), 744 (C–S). ¹H NMR spectrum, δ , ppm: 4.31 s (2H, NH₂), 6.56 t.d (1H, *J* = 7.5, 1.2 Hz, Ar-H), 6.56 t.d (1H, *J* = 7.5, 1.2 Hz, Ar-H), 6.56 t.d (1H, *J* = 8.5, 1.3 Hz, Ar-H), 7.07–7.19 m (2H, Ar-H). ¹³C NMR spectrum, δ_{C} , ppm: 115.3, 118.2, 118.8, 131.7, 136.9, 148.7. Found, %: C 58.01; H 4.69; N 11.14; S 26.16. C₁₂H₁₂N₂S₂. Calculated %: C 58.03; H 4.87; N 11.28; S 25.82. *M* 249 [*M* + H]⁺.

2,2'-{(1Z,1'Z)-[(Disulfanediylbis(2,1-phenylene)]bis(azanylylidene)bis(methanylylidene)}diphenol (3). Straw yellow solid, yield 95%, mp 167–169°C. IR spectrum, v, cm⁻¹: 1610 (C=N), 1462 (C=C), 1278 (C–O),

Table 2. In vitro antibacterial activity of compounds against multi-drug resistant A. baumannii

Compound	Minimum inhibitory concentrations, mg/L			
Compound	MIC_{50}^{a}	MIC ₉₀ ^b		
2	8	16		
3	>64	>64		
4 a	16	16		
4b	32	32		
4c	32	32		
5a	>64	>64		
5b	16	32		
5c	32	32		
MEM ^c	>16	>16		

^a MIC₅₀: 50% minimum inhibitory dose. ^b MIC₉₀: 90% minimum inhibitory dose. ^c Meropenem (standard antibiotic).

747 (C–S), 556 (S–S). ¹H NMR spectrum, δ , ppm: 6.94 t (1H, J = 7.1 Hz, Ar-H), 7.04 d (1H, J = 8.4 Hz, Ar-H), 7.09–7.14 m (1H, Ar-H), 7.15–7.26 m (2H, Ar-H), 7.39 d.d (2H, J = 5.7, 1.7 Hz, Ar-H), 7.62–7.67 m (1H, Ar-H), 8.59 s (1H, H–C=N), 12.87 s (1H, Ar-OH). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 117.5, 117.7, 119.2, 119.3, 127.2, 127.3, 127.7, 127.8, 131.6, 132.7, 133.7, 161.2, 162.8. Found, %: C 68.45; H 4.35; N 6.09; S 14.08. C₂₆H₂₀N₂O₂S₂. Calculated %: C 68.40; H 4.42; N 6.14; S 14.05. M 457 $[M + H]^+$.

3,3'-{(1Z,1'Z)-[(Disulfanediylbis(2,1-phenylene)]bis(azanylylidene)bis(methanylylidene){bis(benzene-1,2-diol) (4a). Shiny scarlet solid, yield 95%, mp 205-210°C. IR spectrum, v, cm⁻¹: 3306 (O-H), 1611 (C=N), 1456 (C=C), 1244 (C-O), 749 (C-S), 574 (S–S). ¹H NMR spectrum, δ , ppm: 6.87 t (1H, J = 7.8 Hz, Ar-H), 7.04 d (1H, J = 7.8 Hz, Ar-H), 7.20 d (1H, J = 7.8 Hz, Ar-H), 7.32 t (1H, J = 7.6 Hz, Ar-H),7.39 t (1H, J = 7.5 Hz, Ar-H), 7.56 d (1H, J = 7.8 Hz, Ar-H), 7.60 d (1H, J = 7.8 Hz, Ar-H), 9.04 s (1H, H–C=N), 9.39 s (1H, O–H), 12.78 s (1H, O–H). ¹³C NMR spectrum, δ_C, ppm: 118.40, 119.16, 119.47, 119.64, 123.08, 125.97, 127.96, 128.11, 130.32, 145.66, 148.97, 151.17, 164.14. Found, %: C 63.81; H 4.18; N 5.79; S 13.06. C₂₆H₂₀N₂O₄S₂ Calculated %: C 63.92; H 4.13; N 5.73; S 13.13. M 489 $[M + H]^+$.

4,4'-{(1Z,1'Z)-[(Disulfanediylbis(2,1-phenylene)]bis(azanylylidene)bis(methanylylidene)}bis(benzene-1,3-diol) (4b). Carmine solid, yield 98%, mp 237°C. IR spectrum, v, cm⁻¹: 3100 (O–H), 1606 (C=N), 1456 (C=C), 1225 (C-O), 746 (C-S), 559 (S-S). ¹H NMR spectrum, δ , ppm: 6.87 t (1H, J = 7.8 Hz, Ar-H). 6.99 d (1H, J = 7.8 Hz, Ar-H), 7.19 d (1H, J = 7.8 Hz, Ar-H), 7.29 t (1H, J = 7.6 Hz, Ar-H), 7.38 t (1H, J = 7.5 Hz, Ar-H), 7.54 d (1H, J = 7.8 Hz, Ar-H), 7.48 d (1H, J = 7.8 Hz, Ar-H), 9.01 s (1H, H-C=N), 9.28 s (1H, O-H), 12.81 s (1H, O-H). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 101.57, 108.47, 112.25, 114.23, 120.98, 126.64, 127.55, 127.82, 132.27, 145.37, 148.67, 156.85, 166.81. Found, %: C 63.99; H 4.10; N 5.69; S 13.04. C₂₆H₂₀N₂O₄S₂ Calculated %: C 63.92; H 4.13; N 5.73; S 13.13. M 489 $[M + H]^+$.

2,2'-{(1Z,1'Z)-[(Disulfanediylbis(2,1-phenylene)]bis(azanylylidene)bis(methanylylidene){bis(benzene-1,4-diol) (4c). Shiny brown solid, yield 92%, mp 225°C. IR spectrum, v, cm⁻¹: 3372 (O–H), 1617 (C=N), 1439 (C=C), 1252 (C-O), 758 (C-S), 575 (S-S), ¹H NMR spectrum, δ , ppm: 6.88 d (1H, J = 8.8 Hz, Ar-H), 6.96 d.d (1H, J = 8.8, 2.9 Hz, Ar-H), 7.17 d (1H, J =2.9 Hz, Ar-H), 7.28–7.33 m (1H, Ar-H), 7.37 t.d (1H, J = 7.6, 1.3 Hz, Ar-H), 7.52 d.d (1H, J = 7.8, 0.9 Hz, Ar-H), 7.58 d.d (1H, J = 7.8, 1.2 Hz, Ar-H), 8.96 s (1H, H-C=N), 9.23 s (1H, O-H), 11.83 s (1H, O-H). 13 C NMR spectrum, δ_{C} , ppm: 116.79, 117.37, 118.34, 119.36, 121.84, 125.70, 127.75, 127.92, 130.39, 146.17, 149.78, 153.06, 163.02. Found, %: C 63.80; H 4.19; N 5.79; S 13.02. C₂₆H₂₀N₂O₄S₂ Calculated %: C 63.92; H 4.13; N 5.73; S 13.13. M 489 $[M + H]^+$.

6,6'-{(1Z,1'Z)-[(Disulfanedivlbis(2,1-phenylene)]bis(azanylylidene)bis(methanylylidene){bis(2-methoxyphenol) (5a). Bright orange solid, yield 96%, mp 174°C. IR spectrum, v, cm⁻¹: 2839 (OCH₃), 1610 (C=N), 1455 (C=C), 1247 (C-O), 729 (C-S), 559 (S–S). ¹H NMR spectrum, δ , ppm: 3.96 s (3H, OCH₃), 6.98 d (3H, J = 51.5 Hz, Ar-H), 7.19 d (3H, J =10.2 Hz, Ar-H), 7.68 s (1H, Ar-H), 8.66 s (1H, H–C=N), 13.26 s (1H, O–H). ¹³C NMR spectrum, δ_C , ppm: 56.39, 116.52, 118.93, 119.38, 119.84, 124.47, 126.68, 128.41, 128.62, 130.88, 146.36, 148.44, 150.78, 164.23. Found, %: C 65.13; H 4.65; N 5.49; S 12.38. C₂₈H₂₄N₂O₄S₂ Calculated %: C 65.09; H 4.68; N 5.42; S 12.41. M 517 $[M + H]^+$.

6,6'-{(1Z,1'Z)-[(Disulfanediylbis(2,1-phenylene)]bis(azanylylidene)bis(methanylylidene){bis(3-methoxyphenol) (5b). Bright orange solid, yield 97%, mp 198°C. IR spectrum, v, cm⁻¹: 2839 (OCH₃), 1632 (C=N), 1461 (C=C), 1243 (C-O), 757 (C-S), 553 (S–S). ¹H NMR spectrum, δ , ppm: 3.86 s (3H, OCH₃),

6.48–6.59 m (2H, Ar-H), 7.13 d.d (1H, J = 7.7, 1.2 Hz, Ar-H), 7.17 d.d (1H, J = 7.7, 1.4 Hz, Ar-H), 7.23 t.d (1H, J = 7.4, 1.3 Hz, Ar-H), 7.30 d (1H, J = 8.5 Hz)Ar-H), 7.65 d.d (1H, J = 7.8, 1.3 Hz, Ar-H), 8.54 s (1H, H–C=N), 13.27 s (1H, O–H). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 55.55, 101.13, 107.42, 113.19, 117.52, 126.97, 127.19, 127.47, 131.39, 133.88, 146.48, 161.79, 163.61, 164.33. Found, %: C 65.25; H 4.65; N 5.38; S 12.37. C₂₈H₂₄N₂O₄S₂ Calculated %: C 65.09; H 4.68; N 5.42; S 12.41. M 517 $[M + H]^+$.

2,2'-{(1Z,1'Z)-[(Disulfanedivlbis(2,1-phenylene)]bis(azanylylidene)bis(methanylylidene){bis(4-methoxyphenol) (5c). Bright orange solid, yield 96%, mp 170°C. IR spectrum, v, cm⁻¹: 2832 (OCH₃), 1614 (C=N), 1485 (C=C), 1271 (C-O), 748 (C-S), 577 (S–S). ¹H NMR spectrum, δ , ppm: 3.82 s (3H, OCH₃), 6.91 d (1H, J = 2.7 Hz, Ar-H), 7.02 d.t (2H, J = 14.0, 5.8 Hz, Ar-H), 7.16 t.d (1H, J = 7.3, 1.4 Hz, Ar-H), 7.20 d.d (1H, J = 7.6, 1.6 Hz, Ar-H), 7.24 d.d (1H, J =7.5, 1.6 Hz, Ar-H), 7.66 d.d (1H, J = 7.7, 1.5 Hz, Ar-H), 8.59 s (1H, H–C=N), 12.43 s (1H, O–H). ¹³C NMR spectrum, δ_{C} , ppm: 55.98, 115.45, 117.63, 118.32, 118.76, 121.15, 127.22, 127.64, 127.77, 131.67, 146.34, 152.36, 155.50, 162.51. Found, %: C 65.17; H 4.64; N 5.38; S 12.38. C₂₈H₂₄N₂O₄S₂ Calculated %: C 65.09; H 4.68; N 5.42; S 12.41. M 517 $[M + H]^+$.

Antimicrobial susceptibility patterns of Acinetobacter baumannii. Multiple-resistant A. baumannii isolates were obtained from various clinical samples Microbiology (Medical Laboratory of Duzce University Medical Faculty). Identification of A. baumannii isolates was performed by conventional microbiological methods (Gram staining, oxidase, three sugar iron broth and motion test) and an automatized-bacterial identification system (Vitek 2, bioMerieux, France).

Meropenem and other antibiotic susceptibilities of A. baumannii isolates were determined by the Vitek 2 system and evaluated according to CLSI [18]. Detection of resistance to typical antibiotics belonging to at least three antibiotic classes was defined as multiple antibiotic resistance. Antibacterial activities of all the compounds were determined by the broth microdilution method [18]. The compounds were dissolved in DMSO to a final concentration of 60 µg/mL. The isolates were adjusted to McFarland's 0.5 standard and 100 µL of the Mueller Hinton broth was placed in each 96-well plate. Afterwards, serial dilutions of the solutions were carried out. MIC₅₀ and MIC₉₀ values of

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all compounds **2–5c** against *A. baumannii* isolates were compared with standard antibiotic meropenem MIC values.

CONCLUSIONS

All compounds were synthesized with high yield. However, in the new method that involved the CeO_2 nanocatalyst, the yield was increased. The catalyzed reactions were complete within minutes. The CeO_2 nanocatalyst could be used repeatedly, making the processes less costly.

Several synthesized compounds demonstrated high antibacterial activity. The present study exhibited that disulfide-Schiff base compounds were partially responsible for antibacterial activity against multi-drugresistant *A. baumannii*, thus indicating that these can be used in advanced pharmacological studies for treatment of multi-drug-resistant bacterial strains.

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