2-Nitro Thioglycoside Donors: Versatile Precursors of β -D-Glycosides of Aminosugars

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ABSTRACT



2-Nitro thioglycosides can be prepared by the Michael addition of thiophenol to 2-nitroglycal derivatives. NIS/TMSOTf activation of these 2-nitro thioglycosides, in the presence of alcohols, rapidly and cleanly led to the desired glycosides in good yield and β -selectivity. Reduction of the nitro group allowed generation of the corresponding 2-acetamido glycosides.

The application of 2-nitroglycals to the synthesis of mucintype glycopeptides has recently attracted our attention¹ because of the fundamental role the latter play in biological processes such as cell–cell adhesion, cell growth, fertilization, parasitic infection, and inflammation.^{2a,c} We previously demonstrated that 2-nitrogalactal concatenation is a useful tool for forming α -glycosidic bonds to L-serine or Lthreonine. Via this methodology we synthesized all the members of the mucin family,^{1,3} as well as many *O*-, *S*-, *P*-, and *C*-glycosides⁴ and nucleosides.⁵

Although the synthesis of 2-nitro thioglycosides from 2-nitroglycals has already been reported by Holzapfel et al.,⁶

the behavior of these compounds as glycosyl donors was never investigated. The synthesis of 2-nitro glycoside donors through Michael-type addition reactions and their use in glycosylation reactions is now reported here.

The 2-nitroglycals **1**, **2**,^{3a}, **3**,⁶ and **4**⁷ (Table 1), obtained by standard nitration conditions (addition of acetyl nitrate and elimination of acetic acid),^{3a-c} were converted to the 2-nitro thioglycoside by base-catalyzed glycosylation with thiophenol and 0.1 equiv of potassium *tert*-butoxide. The 2-nitro thioglycosides (**5–8**) were obtained in excellent yield

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Table 1.	Base-Catalyzed	Addition of	Thiophenol	to Glycals $1-4^a$
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Glycal		Reaction time	Product		α : β selectivity	Yield
BnO OTDS TDSO NO2	1 ^b	0.5 h	BNO OTDS TDSO O ₂ N SPh	5	0:1	88%
BnO OBn BnO NO ₂	2	1 h 0.3 h	BnO OBn BnO OBn O ₂ N	6a:6b	1:3 0:1	81% 80%
BnO OBn BnO NO ₂	3	0.3 h	BnO BnO O ₂ N SPh	7	0:1	70%
Ph TO O DO BnO	4	0.3 h	Ph TO O BnO O ₂ N SPh	8a:8b	1:2	70%

^a Reactions were conducted at room temperature in toluene in the presence of 0.1 equiv of potassium tert-butoxide. ^b TDS = thexyldimethylsilyl.

Table 2.	Coupling	Reactions	of 2-N	itrothiog	ycosides

Entry	Donors	Acceptors	Major product	Yields, α : β selectivity
1	BnO COBn BnO Co2N SPh	≻он 9	Bno - OBn	70% (0:1) ^a
2	"		Bno OBn Bno OBn Do DO	90% (1:5) ^a
3	"	HO ^{SiMe} ₃	Bno OBn Bno OSE 20°	70% (0:1) ^a
4 5	**	но 12	Bno OBn NHBz Bno Ozno COOMe 21a:21b	63% (1:4) ^a 90% (1:8) ^b
6 7	**	HO BOO BOO BOO SE 13°	Bno COBn Bno Con Bno CoAc Bno Con Bno CoAc Bno CoAc Bno CoAc Bno CoAc	70% (1:1.1) ^a 77% (1:15) ^b
8	"	BnO OH BnO O _{2N} 14 ^c	$BnO \qquad BnO \qquad C_2N \qquad BnO \qquad C_2NOSE$	74% (1:8) ^a
9	'n	HOLDER HOLDER	BnO + OBn + OBn + OF OTDS = 24	60% (0 : 1) ^b
10	BnO OTDS BnO SPh O ₂ N 5	^{Me—OH} 16	Bno COTDS TDSO O ₂ N 25	68% (0:1) ^a
11 12	**	HO	Bno OTDS NHBz TDSO O ₂ N COOMe 26a:26b	77% (1:1) ^a 68% (1:4) ^b
13	"	HO TOTBDMS BNO Me 17 d	DSO COLOR DOTEDMS TDSO COLOR DOTEDMS DSO COLOR DOTEDMS BDO OME DSO COLOR DOTEDMS DDSO COLOR DOTED DDSO COLOR DOTEDMS DDSO COLOR DOTED DDSO COLOR DOTED	50% (1:1.5) ^a



^{*a*} Reactions, unless stated otherwise, were conducted in CH₂Cl₂. Anomeric ratios were determined by isolated products. ^{*b*} Reactions were conducted in propionitrile at -15 °C. ^{*c*} SE = trimethylsilylethyl. ^{*d*} TBDMS = tert-butyldimethylsilyl.

(70–88%). As previously reported,⁸ concatenation with soft nucleophiles such as thiophenol yields mainly the β -anomer in a time-dependent manner. The β -linkage was confirmed by ¹H NMR studies, which showed a coupling constant $J_{1,2}$ = 6 Hz, which contrasted with a $J_{1,2}$ of 4 Hz for the α -anomer.

With the requisite thiophenylglycosides in hand, a series of coupling reactions with different acceptors **9**, **10**, ⁹ **11**, **12**, **13**, ¹⁰ **14**, ¹¹ **15**, ¹⁰ **16**, and **17**¹² (Table 2) were carried out. Activation simply involved cooling a 1:1.2:1.5 mixture of the thioglycoside, the acceptor, and NIS in CH₂Cl₂ at 0 °C and then adding 0.15 equiv of TMSOTf (Method A). Alternatively, the same protocol could be carried out at -15 °C in propionitrile (Method B). Workup thereafter afforded the desired glycosides.

A series of compounds, selected to illustrate the wide range of possible coupling types, was prepared. The results are presented in Table 2. In the case of highly reactive acceptor alcohols (entries 1, 3, 10, and 16) only the β anomers were detected. For less reactive primary and secondary hydroxy acceptors, β -selectivity was conserved but was not exclusive. To improve selectivity, propionitrile was used as solvent, and with the help of the nitrile effect,¹³ the β -selectivity was further increased (entries 5, 7, and 12). The presence of a nitro group at position 2 seemed to allow anchimeric assistance and consequently orientated the glycosylations in the direction of the β -anomer as the major product.

To investigate the influence of a 4,6-O-benzylidene acetal protecting group on the outcome of the glycosidation,^{14a-d} nitro thioglucoside donor **8a**, **b** was tested under the same reaction conditions. Unfortunately, in this case, activation

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of the thiophenyl group by NIS was found to be difficult, the α/β -ratios and the yields often being poor. The rigidity induced by the 4,6-*O*-benzylidene ring could explain this result.

Also the use of 1-benzenesulfinyl piperidine (BSP)/ trifluoromethanesulfonic anhydride in the presence of 2,4,6tri-*tert*-butylpyrimidine (TTBP) was investigated for thioglycoside activation.^{14a-d} Unfortunately, however, the 2-nitrothiophenyl donors were not activated under these conditions. Only low yields of the desired compounds were usually obtained; side products from elimination of the intermediate triflate^{14a-d} typically predominated (Figure 1).



Figure 1. Activation with BSP, Tf₂O, and TTBP.

Overall, our studies have demonstrated that glycosidations with 2-nitro thiophenylglycosides generate β -anomers with high yield and selectivity. These results, along with Michael-type additions of alcohols to 2-nitrogalactals,^{3a-d} thus allow access to both types of glycoside, as described in Table 3.

 Table 3.
 Comparison between Glycosidation and Michael Addition



Because in nature most amino sugars are mainly *N*-acetamido glycosides, some of our nitroglycoside compounds were submitted to reduction of the nitro group, as depicted in Table 4. A method using an excess of zinc dust¹⁵ and

Table 4.	Reduction of t	the 2-Nitro Group	
Entry	Compounds	Products	Yields
1	18	BnO COBn BnO AcNH 41	81%
2	38	Bno OBn Bno AcNH 42	75%
5	21b	Bno COBn NHBz 43	76%
6	23b	BRO COBR BRO BRO 44 AcNH BRO ACNH OSE	60%

aqueous 1 N HCl in aqueous AcOH was employed and then followed by acetylation. This method gave access to the desired acetamido derivatives 41-44.^{4c,11} A one-pot reaction, using acetic anhydride instead of acetic acid, was also explored. Unfortunately, although reduction of the nitro group occurred without hitch, the one-pot acetylation was never observed and only the free amine could ever be isolated at the end of reaction. An additional step of acetylation was necessary.

In summary, 2-nitro thioglycoside donors can be successfully activated in the presence of NIS/TMSOTf to allow the preparation of β -glycosides in good yield and selectivity. The nitro group could then be reduced to afford the corresponding acetamido glycosides. Starting from 2-nitroglycals, both α or β selectivity can now be attained using Michael-type addition or thioglycoside coupling, respectively. This demonstrates the versatility of 2-nitro sugars as glycosyl donors in glycoside synthesis. Applications of these donors are currently underway in the synthesis of core 3 and 6 mucin types; the results of these studies will be reported in due course.

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Supporting Information Available: Experimental procedures and spectral/analytical data for key intermediates and products. This material is available free of charge via the Internet at http://pubs.acs.org.

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