

Design and synthesis of a *trans*-fused polycyclic ether skeleton as an α -helix mimetic scaffold

Hiroki Oguri,^{*,†} Akifumi Oomura, Shintaro Tanabe and Masahiro Hirama^{*}

Department of Chemistry, Graduate School of Science, Tohoku University, and SORST,
Japan Science and Technology Corporation (JST), Sendai 980-8578, Japan

Received 12 January 2005; revised 31 January 2005; accepted 8 February 2005

Abstract—Inspired by the common skeletal feature of potent marine toxins, a ladder-like polycyclic ether scaffold was designed as the basis for the synthesis of structurally defined α -helix mimics. A synthetic route to *trans*-fused 6/6/6/6/6 pentacyclic ether skeletons is presented, involving the iterative assembly of tetrahydropyran rings via the SmI₂-mediated coupling reaction of α -sulfonyl ketones with aldehydes.

© 2005 Elsevier Ltd. All rights reserved.

The potent ladder-like polycyclic ether marine toxins are based on a common *trans*/*syn*-fused cyclic ether array, with ring sizes ranging from five to nine members.¹ Among these potent toxins, brevetoxins^{2,3} and ciguatoxins^{4,5} (Fig. 1) are believed to bind to a common site (site

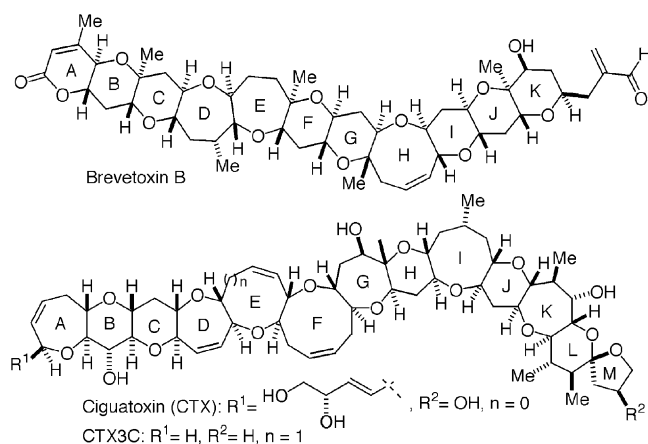


Figure 1. Structures of brevetoxin B, ciguatoxin and CTX3C.

Keywords: Polycyclic ether; α -Helix; Scaffold; Samarium iodide.

^{*} Corresponding authors. Tel.: +81 22 217 6563; fax: +81 22 217 6566; e-mail addresses: oguri@sci.hokudai.ac.jp; hirama@ykbcs.chem.tohoku.ac.jp

[†] Present address: Division of Chemistry, Graduate School of Science, Hokkaido University.

5) on the α subunit of voltage-sensitive sodium channels (VSSCs),⁶ where the α subunit consists of 24 transmembrane helices. Although the three-dimensional structure of the receptor site in the VSSC remains unknown, and the extremely limited availability of the toxins has hampered further biological investigation, recent studies have suggested a relationship between the size of the polycyclic ether skeleton and the inhibitory activity against the binding of labeled brevetoxins with the VSSC.⁷ It appears that both the length and the relatively straight conformation of the cyclic ether skeletons are critical factors in this relationship, and are required for the binding and modulation of the VSSC function.⁸ The interactions between the polycyclic ethers and the receptor site are surmised to be primarily hydrophobic and to be supplemented by hydrogen bonding between skeletal oxygen atoms and the OH/NH groups of the α -helical peptides.⁸ In addition to this hydrogen bonding, axially oriented CH groups adjacent to the skeletal oxygen atoms are also expected to play an important role in molecular recognition of the aromatic amino acid residues through CH/ π interactions, as exemplified by the crystal structures of complexes between the carbohydrate-binding proteins and specific substrates.^{9,10}

On analysis of the mode of action of these ladder-like toxins, it was noticed that the *trans*-fused cyclic ethers may be topologically analogous to the α -helical peptides. In the 6/6/6 tricyclic system (Fig. 2a), the distance between skeletal oxygen atoms on the same side (4.8 Å) in the cyclic ethers is almost identical to the interval

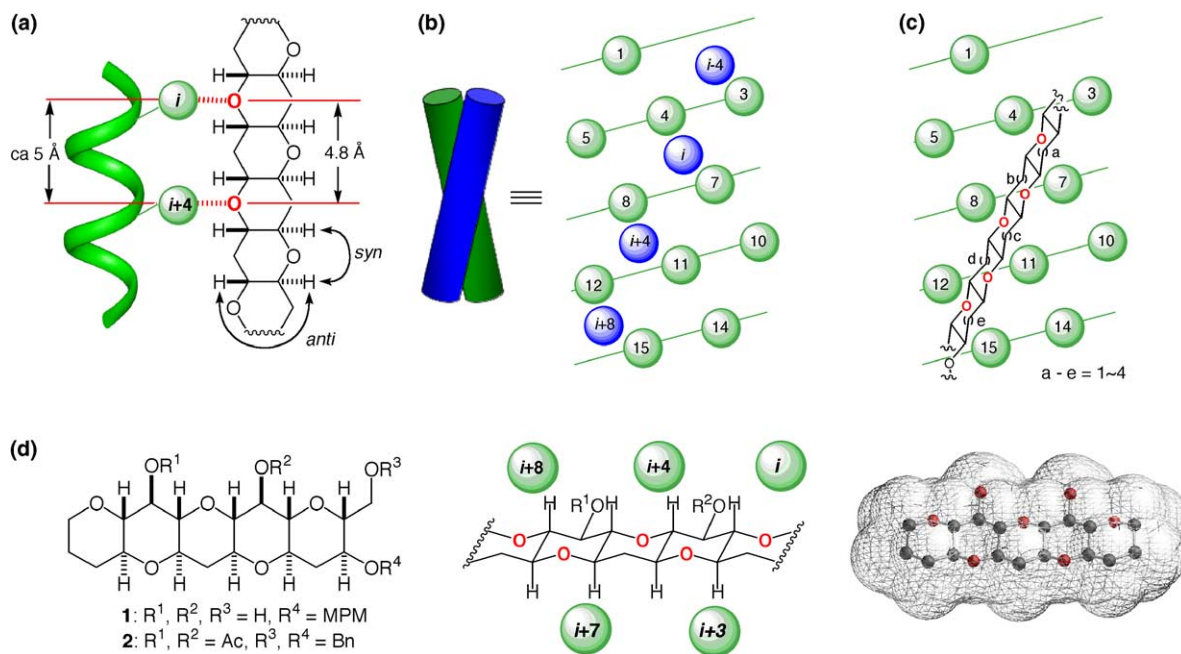


Figure 2. (a) Structural features of a cyclic ether skeleton and an α -helix; (b) Typical helix–helix interaction, showing projection of the surface defined by the ridges and grooves of two helices (green and blue); (c) Hypothetical binding model for the cyclic ether with an α -helix; (d) Design of cyclic ether **1** as an α -helix mimetic scaffold. The water-accessible surface (Connolly surface) for the skeletal cyclic ether part of **1** was generated by Chem 3D Ver. 5.0 using a probe radius of 1.4 Å.

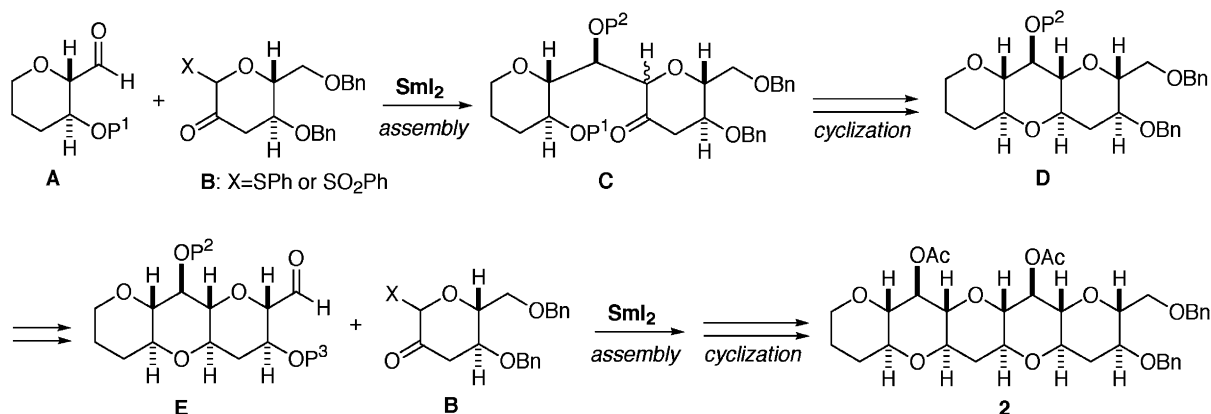
between side-chains in the α -helical peptides in the canonical *i*, *i*+4 relationship (ca. 5 Å). As illustrated in Figure 2b, a typical mode of packing between α -helices (green and blue) involves insertion of the ridges of one helix (blue) into the grooves of an adjacent helix (green).¹¹ Inspired by this potentially analogous topology, the hypothetical binding model shown in Figure 2c was derived. In this model, the ladder-like polycyclic ether fits into the grooves of the helix.

As a first step in the investigation of this model, a 6/6/6/6 pentacyclic ether skeleton (**1**) was designed as a structurally defined α -helix mimetic scaffold¹² having two equatorial hydroxyl groups separated by a distance of 4.8 Å (Fig. 2d). Installation of a variety of substituents (R¹ and R²) is possible via the hydroxyl groups, and the consequent undulation of the molecular surface

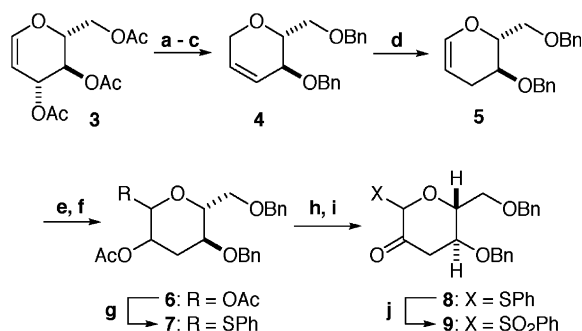
is expected to be analogous to the ridges formed by side-chains of α -helices in the *i*, *i*+4 relationship.

The strategy for the iterative synthesis of the cyclic ether scaffolds is outlined in Scheme 1. A SmI₂-mediated Reformatsky-type reaction of α -ketosulfide **B** with aldehyde **A** was employed for the assembly of the two hydroxy-pyrans.¹³ Subsequent hydroxy-ketone cyclization of the compound **C** would afford tricyclic **D**.¹⁴ After conversion of **D** into aldehyde **E**, the second assembly of two cyclic ethers **E** and **B** followed by the hydroxy-ketone cyclization would yield the 6/6/6/6/6 cyclic ether skeleton **2** with two equatorial hydroxy groups.

The key coupling component, α -ketosulfide **8**, was synthesized from tri-*O*-acetyl-D-glucal **3** (Scheme 2). Reduction of **3** with triethylsilane and BF₃·Et₂O followed by



Scheme 1. Strategy for iterative synthesis of cyclic ether skeleton **3**.

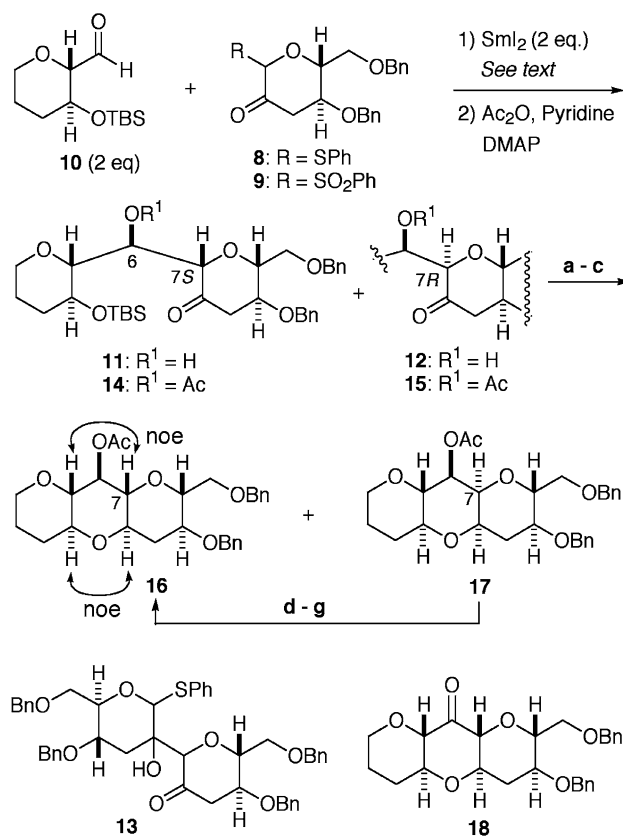


Scheme 2. Reagents and conditions: (a) Et₃SiH, BF₃·Et₂O, CH₃CN, –20 °C, 78%; (b) K₂CO₃, MeOH; (c) NaH, BnBr, THF/DMF(1/1), 90% (two steps); (d) RhCl(PPh₃)₃ (15 mol %), DBU, EtOH, 60 °C, 83%; (e) cat. OsO₄, NMO, acetone; (f) Ac₂O, pyridine, DMAP, 93% (two steps); (g) PhSH, SnCl₄ (10 mol %), CH₂Cl₂, –20 to 0 °C, 90%; (h) K₂CO₃, MeOH; (i) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, –78 to 0 °C, 92% (two steps); (j) *m*-CPBA, CH₂Cl₂, –40 to 0 °C, 88%.

methanolysis and benzylation gave **4**. The allylic ether **4** was then converted into enol ether **5** in the presence of Wilkinson's catalyst and DBU in 83% yield.¹⁵ Dihydroxylation of **5** and subsequent acetylation yielded **6**, which was then subjected to SnCl₄-catalyzed thioacetalization to afford **7**.¹⁶ Conversion of **7** to the α -ketosulfide **8** was achieved by methanolysis and subsequent Swern oxidation in 92% yield (two steps).

The tetrahydropyran rings were assembled by Reformatsky-type coupling of the α -ketosulfide **8** with the aldehyde **10**¹⁷ using Mastuda's protocol (Scheme 3).¹³ A solution of **8** in THF was added dropwise to a solution of SmI₂ (2 equiv) in THF at –78 °C to generate the samarium enolate regioselectively, and the resulting solution was treated with **10** (2 equiv). Unfortunately, homocoupled **13** was the major product (34% yield), and the desired coupling products **11** were obtained in only low yield (18%).¹⁸ Thus, the samarium enolate was trapped immediately after generation by adding a mixture of **8** and **10** (2 equiv) to a solution of SmI₂ (2 equiv) at –78 °C. Although the yield of the desired **11** was improved (up to ~40%), significant amounts of **13** (~20%) were still produced.¹⁸ To suppress the formation of **13**, sulfone **9**¹⁹ was used in place of sulfide **8** to accelerate the generation of the samarium enolate. Treatment of a mixture of **9** and **10** (2 equiv) with SmI₂ (2 equiv) promoted the Reformatsky-type reaction smoothly at –78 °C, affording the desired coupling products without formation of **13**. Subsequent in situ acetylation gave a 3:2 diastereomeric mixture of **14** and **15** in 79% yield (two steps). A highly stereo-controlled addition at C6 occurred in this synthesis, presumably due to 1,2-chelation of **10** with the samarium species. The mixture of **14** and **15** was subsequently subjected to hydroxy-ketone cyclization followed by reductive etherification to afford separable tricyclic ethers **16**²⁰ and **17**. The *trans*, *cis*-fused **17** was converted to *trans*, *trans*-fused **16** in four steps involving the epimerization of ketone **18** under Swern oxidation conditions.^{14b}

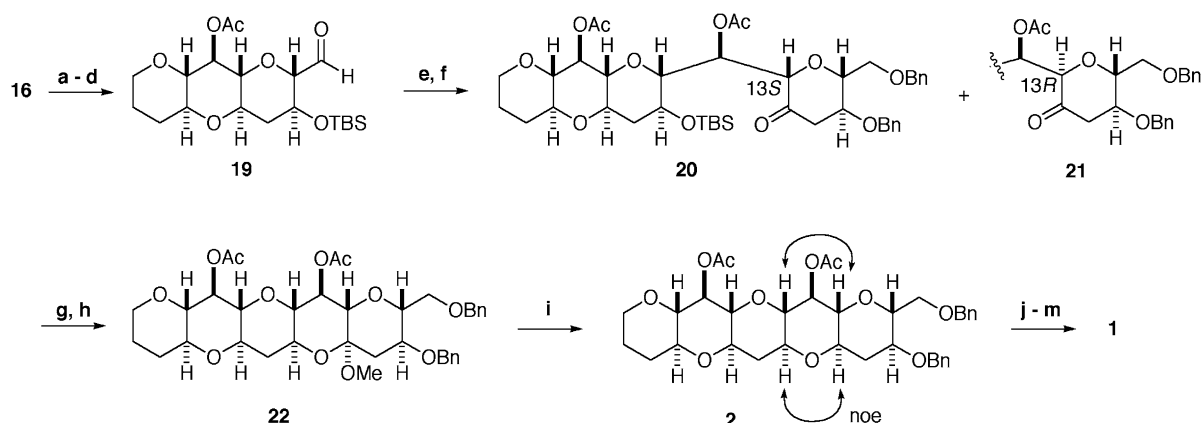
With tricyclic **16** in hand, iterative assembly with **9** was performed, leading to **1** (Scheme 4). Functional group



Scheme 3. Reagents and conditions: (a) CSA, MeOH/CH₂Cl₂ (1:7); (b) Ac₂O, pyridine, DMAP; (c) Et₃SiH, TMSOTf, CH₂Cl₂, –20 °C, **16** 34% (three steps), **17** 9% (three steps); (d) K₂CO₃, MeOH; (e) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, –78 to 0 °C, 57% (two steps); (f) NaBH₄, CeCl₃, MeOH/THF (4:1), –20 °C; (g) Ac₂O, pyridine, DMAP, 50% (two steps).

manipulation of **16** gave aldehyde **19** via a four-step procedure involving (i) removal of benzyl groups, (ii) protection of the hydroxyl groups as TBS ethers, (iii) selective removal of the primary silyl group, and (iv) Swern oxidation. The SmI₂-mediated coupling reaction of **19** (1.2 equiv) with sulfone **9** and subsequent acetylation furnished a 1:1 diastereomeric mixture of **20** and **21** in 72% yield (two steps). Exposure of the mixture to CSA in MeOH/CH₂Cl₂ (1:4) effected the removal of the TBS and in situ methyl ketal formation, as well as partial deacetylation. The desired methyl ketal **22** was isolated after acetylation in 52% yield. The products possessing 13*R* stereochemistry could not be isolated. Reduction of **22** with Et₃SiH and TMSOTf gave the *trans*-fused 6/6/6/6/6 pentacyclic ether **2** in 91% yield.²¹ Finally, protecting-group manipulation under standard conditions led to the scaffold **1**.

In conclusion, a polycyclic ether scaffold was designed as an α -helix mimic, inspired by the topological similarity between ladder-like cyclic ether marine toxins and α -helical peptides. The synthesis presented here provides an efficient strategy for the stereo-controlled construction of *trans*-fused polycyclic ether skeletons via SmI₂-mediated Reformatsky-type coupling of α -sulfonyl ketones with aldehydes. Further investigation of the synthesis



Scheme 4. Reagents and conditions: (a) H_2 , $\text{Pd}(\text{OH})_2/\text{C}$, MeOH/AcOEt (1/4); (b) TBSCl, imidazole, DMF, 99% (two steps); (c) CSA, MeOH , -5°C , 75%; (d) $(\text{COCl})_2$, DMSO, CH_2Cl_2 , -78°C , then Et_3N , 0°C ; (e) **19** (1.2 equiv), **9**, Sml_2 (2 equiv), THF, -100°C ; (f) Ac_2O , pyridine, DMAP, **20:21** = 1:1, 72% (two steps); (g) CSA, $\text{MeOH}/\text{CH}_2\text{Cl}_2$ (1:4); (h) Ac_2O , pyridine, DMAP, **22** 52% (two steps); (i) Et_3SiH , TMSOTf, CH_2Cl_2 , -20°C , 91%; (j) H_2 , $\text{Pd}(\text{OH})_2/\text{C}$, MeOH/AcOEt (1/4); (k) $p\text{-MeOC}_6\text{H}_4\text{CH}(\text{OMe})_2$, CSA, DMF, 71% (two steps); (l) DIBAL, CH_2Cl_2 , -78 to -15°C , then Ac_2O , pyridine, DMAP, 80%; (m) K_2CO_3 , MeOH/THF (4/1), 99%.

of a variety of α -helix mimics based on cyclic ether scaffolds and evaluation of the interaction with α -helical peptides will be presented in future reports.

Acknowledgements

The authors thank Professors K. Tsumoto and M. Umetsu for valuable discussions. This work was supported by the Core Research for Evolutional Science and Technology (CREST) programs of the Japan Science and Technology Agency (JST), and by a Grant-in-Aid for Scientific Research (S) from the Japanese Society for the Promotion of Science (JSPS).

References and notes

- For reviews on marine polycyclic ether toxins, see: (a) Yasumoto, T.; Murata, M. *Chem. Rev.* **1993**, *93*, 1897; (b) Scheuer, P. J. *Tetrahedron* **1994**, *50*, 3; (c) Murata, M.; Yasumoto, T. *Nat. Prod. Rep.* **2000**, *17*, 293; (d) Yasumoto, T. *Chem. Rev.* **2001**, *1*, 228.
- (a) Shimizu, Y.; Chou, H. N.; Bando, H.; Van Duyne, G.; Clardy, J. *J. Am. Chem. Soc.* **1986**, *108*, 514; (b) Pawlak, J.; Tempesta, M. S.; Golik, J.; Zagorski, M. G.; Lee, M. S.; Nakanishi, K.; Iwashita, T.; Gross, M. L.; Tomer, K. B. *J. Am. Chem. Soc.* **1987**, *109*, 1144; (c) Lin, Y.-Y.; Risk, M.; Ray, S. M.; Van Engen, D.; Clardy, J.; Golik, J.; James, J. C.; Nakanishi, K. *J. Am. Chem. Soc.* **1981**, *103*, 6773; (d) Lee, M. S.; Repata, D. J.; Nakanishi, K.; Zagorski, M. G. *J. Am. Chem. Soc.* **1986**, *108*, 7855.
- Total synthesis of brevetoxins: (a) Nicolaou, K. C.; Rutjes, F. P. J. T.; Theodorakis, E. A.; Tiebes, J.; Sato, M.; Untersteller, E. J. *Am. Chem. Soc.* **1995**, *117*, 10252; (b) Nicolaou, K. C.; Gunzner, J. L.; Shi, G.-q.; Agrios, K. A.; Gärtner, P.; Yang, Z. *Chem.-Eur. J.* **1999**, *5*, 646; (c) Matsuo, G.; Kawamura, K.; Hori, N.; Matsukura, H.; Nakata, T. *J. Am. Chem. Soc.* **2004**, *126*, 14374, and references cited therein.
- (a) Murata, M.; Legrand, A.-M.; Ishibashi, Y.; Fukui, M.; Yasumoto, T. *J. Am. Chem. Soc.* **1990**, *112*, 4380; (b) Satake, M.; Murata, M.; Yasumoto, T. *Tetrahedron Lett.* **1993**, *34*, 1975; (c) Satake, M.; Morohashi, A.; Oguri, H.; Oishi, T.; Hiram, M.; Harada, N.; Yasumoto, T. *J. Am. Chem. Soc.* **1997**, *119*, 11325; (d) Satake, M.; Fukui, M.; Legrand, A.-M.; Cruchet, P.; Yasumoto, T. *Tetrahedron Lett.* **1998**, *39*, 1197; (e) Lewis, R. J.; Vernoux, J.-P.; Brereton, I. M. *J. Am. Chem. Soc.* **1998**, *120*, 5914, and references cited therein.
- Total synthesis of ciguatoxins: (a) Hiram, M.; Oishi, T.; Uehara, H.; Inoue, M.; Maruyama, M.; Oguri, H.; Satake, M. *Science* **2001**, *294*, 1904; (b) Inoue, M.; Hiram, M. *Synlett* **2004**, 577; (c) Inoue, M.; Miyazaki, K.; Uehara, H.; Maruyama, M.; Hiram, M. *Proc. Natl. Acad. Sci. U.S.A.* **2004**, *101*, 12013; (d) Inoue, M.; Hiram, M. *Acc. Chem. Res.* **2004**, *37*, 961, and references cited therein.
- (a) Bidard, J.-N.; Vijverberg, H. P. M.; Frelin, C.; Chungue, E.; Legrand, A.-M.; Bagnis, R.; Lazdunski, M. *J. Biol. Chem.* **1984**, *259*, 8353; (b) Poli, M. A.; Mende, T. J.; Baden, D. G. *Mol. Pharmacol.* **1986**, *30*, 129; (c) Lombet, A.; Bidard, J.-N.; Lazdunski, M. *FEBS Lett.* **1987**, *219*, 355; (d) Baden, D. G. *FASEB J.* **1989**, *3*, 1807; (e) Trainer, V. L.; Baden, D. G.; Catterall, W. A. *J. Biol. Chem.* **1994**, *269*, 19904; (f) Dechraoui, M.-Y.; Naar, J.; Pauillac, S.; Legrand, A.-M. *Toxicol.* **1999**, *37*, 125; (g) Yamaoka, K.; Inoue, M.; Miyahara, H.; Miyazaki, K.; Hiram, M. *Br. J. Pharmacol.* **2004**, *142*, 879; For recent reviews, see: (h) Catterall, W. A. *Neuron* **2000**, *26*, 13; (i) Anger, T.; Madge, D. J.; Mulla, M.; Riddall, D. *J. Med. Chem.* **2001**, *44*, 115.
- Inoue, M.; Hiram, M.; Satake, M.; Sugiyama, K.; Yasumoto, T. *Toxicol.* **2003**, *41*, 469.
- (a) Rein, K. S.; Baden, D. G.; Gawley, R. E. *J. Org. Chem.* **1994**, *59*, 2101; (b) Rein, K. S.; Lynn, B.; Gawley, R. E.; Baden, D. G. *J. Org. Chem.* **1994**, *59*, 2107; (c) Gawley, R. E.; Rein, K. S.; Jeglitsch, G.; Adams, D. J.; Theodorakis, E. A.; Tiebes, J.; Nicolaou, K. C.; Baden, D. G. *Chem. Biol.* **1995**, *2*, 533; (d) Purkerson-Parker, S. L.; Fieber, L. A.; Rein, K. S.; Podona, T.; Baden, D. G. *Chem. Biol.* **2000**, *7*, 385.
- (a) Quiocho, F. A. *Pure Appl. Chem.* **1989**, *61*, 1293; (b) Nishio, M.; Hirota, M.; Umezawa, Y. *CH/ π Interaction: Evidence, Nature, and Consequences*; Wiley-VCH: New York, 1998.

10. Our recent studies indicated a high content (more than 30%) of aromatic amino acids in the complementarity-determining region of specific antibodies against the tricyclic polyether fragment of CTX3C, see: Nagumo, Y.; Oguri, H.; Tsumoto, K.; Shindo, Y.; Hiramama, M.; Tsumuraya, T.; Fujii, I.; Tomioka, Y.; Mizugaki, M.; Kumagai, I. *J. Immunol. Methods* **2004**, *289*, 137.
11. (a) Chothia, C.; Levitt, M.; Richardson, D. *Proc. Natl. Acad. Sci. U.S.A.* **1977**, *74*, 4130; (b) Branden, C.; Tooze, J. *Introduction to Protein Structure*, 2nd ed.; Garland Pub.: New York, 2000.
12. Selected examples for non-peptide based α -helix mimetics: (a) Albert, J. S.; Goodman, M. S.; Hamilton, A. D. *J. Am. Chem. Soc.* **1995**, *117*, 1143; (b) Horwell, D. C.; Howson, W.; Nolan, W. P.; Ratcliffe, G. S.; Rees, D. C.; Willems, H. M. G. *Tetrahedron* **1995**, *51*, 203; (c) Peczu, M. W.; Hamilton, A. D.; Sanchez-Quesada, J.; de Mendoza, J.; Haack, T.; Giralt, E. *J. Am. Chem. Soc.* **1997**, *119*, 9327; (d) Xuereb, H.; Maletic, M.; Gildersleeve, J.; Pelczer, I.; Kahne, D. *J. Am. Chem. Soc.* **2000**, *122*, 1883; (e) Orner, B. P.; Ernst, J. T.; Hamilton, A. D. *J. Am. Chem. Soc.* **2001**, *123*, 5382; (f) Ernst, J. T.; Kutzki, O.; Debnath, A. K.; Jiang, S.; Lu, H.; Hamilton, A. D. *Angew. Chem., Int. Ed.* **2002**, *41*, 278; (g) Jacoby, E. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 891; (h) Kutzki, O.; Park, H. S.; Ernst, J. T.; Omer, B. P.; Yin, H.; Hamilton, A. D. *J. Am. Chem. Soc.* **2002**, *124*, 11838; (i) Ernst, J. T.; Kutzki, O.; Debnath, A. K.; Jiang, S.; Lu, H.; Hamilton, A. D. *Angew. Chem., Int. Ed.* **2002**, *41*, 278; For recent reviews, see: (j) Peczu, M. W.; Hamilton, A. D. *Chem. Rev.* **2000**, *100*, 2479.
13. (a) Ichikawa, S.; Shuto, S.; Matsuda, A. *Tetrahedron Lett.* **1998**, *39*, 4525; (b) Ichikawa, S.; Shuto, S.; Matsuda, A. *J. Am. Chem. Soc.* **1999**, *121*, 10270; (c) Kodama, T.; Shuto, S.; Ichikawa, S.; Matsuda, A. *J. Org. Chem.* **2002**, *67*, 7706.
14. Selected examples for assembly of tetrahydropyran rings affording tricyclic ether systems: (a) Nicolaou, K. C.; Theodorakis, E. A.; Rutjes, F. P. J. T.; Sato, M.; Tiebes, J.; Xiao, X.-Y.; Hwang, C.-K.; Duggan, M. E.; Yang, Z. *J. Am. Chem. Soc.* **1995**, *117*, 10239; (b) Alvarez, E.; Pérez, R.; Rico, M.; Rodríguez, R. M.; Martín, J. D. *J. Org. Chem.* **1996**, *61*, 3003; (c) Oishi, T.; Nagumo, Y.; Hiramama, M. *Synlett* **1997**, 980; (d) Sasaki, M.; Fuwa, H.; Inoue, M.; Tachibana, K. *Tetrahedron Lett.* **1998**, *39*, 9027; (e) Sasaki, M.; Fuwa, H. *Tetrahedron* **2002**, *58*, 1889; (f) Takakura, H.; Sasaki, M.; Honda, S.; Tachibana, K. *Org. Lett.* **2002**, *4*, 2771; (g) Tsukano, S.; Sasaki, M. *J. Am. Chem. Soc.* **2003**, *125*, 14294; (h) Sasaki, M.; Fuwa, H. *Synlett* **2004**, 1851, and references cited therein.
15. Leeuwenburgh, M. A.; Overkleeft, H. S.; van der Marel, G. A.; van Boom, J. H. *Synlett* **1997**, 1263.
16. Nicolaou, K. C.; Li, J.; Zenke, G. *Helv. Chem. Acta* **2000**, *83*, 1977.
17. Nicolaou, K. C.; Hwang, C. K.; Marron, B. E.; DeFrees, S. A.; Couladouros, E. A.; Abe, Y.; Carroll, P. J.; Snyder, J. P. *J. Am. Chem. Soc.* **1990**, *112*, 3040.
18. Unfortunately, diastereomeric coupling products such as **12** could not be detected, presumably due to a retro-aldol reaction. In situ acetylation was thus conducted to prevent retro-reaction in the coupling of **9** with **10**.
19. SmI₂-promoted C-glycosylations using pyridyl sulfones: (a) Krintel, S. L.; Jiménez-Barbero, J.; Skrydstrup, T. *Tetrahedron Lett.* **1999**, *40*, 7565; (b) Miquel, N.; Doineau, G.; Beau, J.-M. *Angew. Chem., Int. Ed.* **2000**, *39*, 4111; (c) Mikkelsen, L. M.; Krintel, S. L.; Jimenez-Barbero, J.; Skrydstrup, T. *J. Org. Chem.* **2002**, *67*, 6297; (d) Mikkelsen, L. M.; Skrydstrup, T. *J. Org. Chem.* **2003**, *68*, 2123.
20. Data for **16**: ¹H NMR (500 MHz, CDCl₃) δ 1.46–1.58 (m, 1H), 1.49 (dt, 1H, J = 12.0, 9.0 Hz), 1.65–1.76 (m, 2H), 2.09 (s, 3H), 2.11–2.12 (m, 1H), 2.57 (dt, 1H, J = 12.0, 4.0 Hz), 3.04 (t, 1H, J = 9.0 Hz), 3.13 (t, 1H, J = 9.0 Hz), 3.23 (td, 2H, J = 9.0, 4.5 Hz), 3.30 (td, 1H, J = 11.5, 3.5 Hz), 3.40 (ddd, 1H, J = 9.0, 5.5, 2.0 Hz), 3.49 (td, 1H, J = 9.0, 4.5 Hz), 3.62 (dd, 1H, J = 11.0, 5.5 Hz), 3.80 (dd, 1H, J = 11.0, 2.0 Hz), 3.95 (ddd, 1H, 11.0, 3.5, 1.0 Hz), 4.44 (d, 1H, J = 11.0 Hz), 4.52 (d, 1H, J = 11.0 Hz), 4.58 (d, 2H, J = 11.0 Hz), 5.11 (t, 1H, J = 9.0 Hz), 7.23–7.34 (m, 10H); ¹³C NMR (125 MHz, CDCl₃) δ 21.00, 24.90, 29.06, 35.03, 67.80, 68.93, 71.05, 71.97, 73.05, 73.17, 73.93, 75.94, 79.41, 80.13, 80.92, 127.28, 127.42, 127.57, 127.71, 127.86, 128.10, 128.17, 128.30, 137.71, 138.42, 170.32; IR (neat) ν 3030, 2948, 1741, 1454, 1367, 1237, 1099 cm⁻¹; MALDI-TOFMS calcd for C₂₈H₃₄O₇Na [M+Na]⁺ 505.220; found 505.218; [α]_D²⁶ +26.4 (c 1.00, CHCl₃).
21. Data for **2**: ¹H NMR (500 MHz, CDCl₃) ¹H NMR (500 MHz, CDCl₃) δ 1.37–1.48 (m, 1H), 1.46 (dt, 1H, J = 11.5, 9.5 Hz), 1.51 (dt, 1H, J = 11.5, 9.5 Hz), 1.59–1.70 (m, 2H), 1.97 (s, 3H), 2.01 (s, 3H), 2.01–2.08 (m, 1H), 2.38 (dt, 1H, J = 11.5, 4.5 Hz), 2.51 (dd, 1H, J = 11.5, 4.5 Hz), 2.98 (t, 1H, J = 9.5 Hz), 3.02 (t, 1H, J = 9.5 Hz), 3.03 (t, 1H, J = 9.5 Hz), 3.07 (t, 1H, J = 9.5 Hz), 3.12–3.20 (m, 2H), 3.21–3.30 (m, 3H), 3.34 (ddd, 1H, J = 9.5, 5.5, 1.5 Hz), 3.44 (td, 1H, J = 9.5, 4.5 Hz), 3.57 (dd, 1H, J = 11.0, 5.0 Hz), 3.71 (dd, 1H, J = 11.0, 1.5 Hz), 3.89 (m, 1H), 4.38 (d, 1H, J = 11.5 Hz), 4.49 (d, 1H, J = 12.0 Hz), 4.52 (d, 1H, J = 11.5 Hz), 4.52 (d, 1H, J = 12.0 Hz), 5.04 (t, 1H, J = 9.5 Hz), 5.07 (t, 1H, J = 9.5 Hz), 7.17–7.28 (m, 10H); ¹³C NMR (125 MHz, CDCl₃) δ 20.66, 20.75, 24.98, 29.09, 34.91, 35.03, 67.90, 68.97, 71.22, 71.98, 72.40, 72.72, 73.29, 74.00, 74.31, 74.56, 76.33, 79.23, 79.99, 80.04, 80.08, 81.02, 127.42, 127.56, 127.82, 127.84, 128.29, 128.42, 137.72, 138.42, 169.43, 169.75; MALDI-TOFMS calcd for C₃₆H₄₄O₁₁Na [M+Na]⁺ 675.273; found 675.289.