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Facile fabrication of AIE/AIEE-active fluorescent nanoparticles based on barbituric for cell imaging applications†

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Four barbituric derivatives (1–4), were synthesized with obvious aggregation induced emission (AIE) or aggregation induced emission enhancement (AIEE) behaviors. The optical properties in different states and the mechanisms of the enhanced emission of 1–4 were investigated. We found that there were a large number of globular or blocky nanoparticles with average diameters from 229 to 394 nm in tetrahydrofuran (THF)/water solutions when the water fraction (f_w) was 90%. The single crystal data of 2 and 4 reveal that multiple intermolecular interactions restrict the intramolecular motions, and promote the formation of a herringbone arrangement (4), thus permitting intense emission in the aggregate state. In addition, the frontier molecular orbital energy and band gap calculated by density functional theory (DFT) are consistent with the experimental results. Compound 4 is cell-permeable: upon entering the live cells, the dye molecules can accumulate in the lysosomes and turn on the fluorescence.

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1. Introduction

Most luminescent materials suffer from a severe effect called aggregation-caused quenching (ACQ), and their luminescence is quenched in high molecular concentrations or in the aggregation state.^{1,2} These organic materials exhibit relatively strong fluorescence in dilute solution, while only weak or non-emissive fluorescence is detected in the aggregated states or solid states, because strong intermolecular interactions facilitate the non-radiative decay pathway. In contrast, AIE is an opposite phenomenon to ACQ, and luminogens with AIE characters can light up in the aggregated states which conventional fluorophores can never achieve.

Due to the unique fluorescence properties, luminescent materials with AIE/AIEE characteristics have attracted considerable attention since the discovery of the AIE phenomenon by Tang in 2001.^{3,4} AIE/AIEE luminophores have invaluable potential for sensing and bioimaging applications,^{5–7} such as optoelectronic devices,⁸ bio-sensing,^{9–11} explosive sensors,¹² and stimuli responsive materials.^{13,14} Up to now, the range of emission wavelengths of AIE-active molecules that have been developed for

cell imaging covers the entire visible light range from blue to red.^{15–18} However, AIE-luminophores with red and near-infrared emission are desired for avoiding possible autofluorescence from biological samples and reducing photocytotoxicity.^{19–21} To address these issues, a few examples have been reported to extend the electronic conjugation of the fluorophores, thus decreasing the energy gap between the electronic ground state and excited state and consequently tuning the absorption and emission characters bathochromically.²² Nonetheless, such a strategy has the disadvantages of tedious synthesis, high synthetic workload, interchromophoric induced fluorescence quenching, poor photostability and solubility.²³ Alternatively, electron-donating (D) and accepting (A) moieties can be incorporated into the design of molecular skeleton. D- π -A molecules typically form charge-transfer (CT) state with a smaller band gap in polar solvents, thus shifting the emission to longer wavelengths.²⁴ Barbituric acid derivatives display diverse biological activities, *e.g.*, anesthetic, hypotensive, antibacterial and anti-tumor activities, and sedative effects.²⁵ And they can be readily linked with chromophore substrates through extensively-studied, highly-efficient Knoevenagel reactions, under mild conditions and with no requirement for a special catalyst.²⁶ Triphenylamine and barbituric acid derived push- π -pull systems have been reported by Kim *et al.* to act as dye molecules.²⁷ Shi *et al.* reported a type of optical probe for Hg²⁺ via the formation of a mercury(II)-barbituric acid coordination complex.²⁸ Zhou *et al.* synthesized and evaluated near-infrared probes with barbituric acid acceptors for *in vivo* detection of amyloid plaques.²⁹

The AIE properties of these compounds have not been investigated except compound 3, and in particular in organism.

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So far, a series of AIE active chromophores have been reported and various mechanistic pathways such as conformational planarization,³⁰ J-aggregate formation,^{31,32} twisted intramolecular charge transfer (TICT),³³ excited state intramolecular proton transfer (ESIPT),³⁴ and restriction of intramolecular motion (RIM) have been explored to operate AIE principles by suppressing the intermolecular π - π stacking interactions.³⁵

Herein, based on the RIM and intramolecular charge transfer (ICT),³⁶ we present a series of AIE/AIEE-active and structurally simple luminogens bearing D- π -A systems. Barbituric acid is incorporated as the electron-withdrawing group, while triphenylamine and carbazole moieties are installed as the donor groups because of their electron-rich nature. For these compounds, they exhibit obvious AIE/AIEE behaviors and red emission. Their mechanisms of enhanced emission behaviors can be well explained by intermolecular interaction in their crystal structures. With the D- π -A structures, compound **4** emits at the red visible region of the electromagnetic spectrum in its aggregate state. Finally, *in vivo* cell imaging experiments of compound **4** have been performed in *Drosophila* S2 cells, and intense intracellular fluorescence signal can be detected with minimal background noise.

2. Experimental section

2.1. Materials

4-Fluorobenzaldehyde (Jiu Ding Chemistry, 97%), triphenylamine (Aladdin, 99%), carbazole (energy, 98%), *N,N*-dimethylamino benzaldehyde (Jiu Ding Chemistry, 99%), barbituric acid (Aladdin, 98%), 1,3-dimethyl-barbituric acid (Aladdin, 99%), were used without further purification. DMF was generally dried by molecular sieves (4 Å), sodium sands and CaH₂ successively, and distilled prior to use. Schneider's *Drosophila* line 2 [*D. mel.* (2), SL2] (ATCC® CRL-1963™) cell line (*Drosophila* S2 cell) was purchased from ATCC.

2.2. Characterizations

Absorption spectra were recorded using a UV-2500 spectrometer. The fluorescence spectra were measured with an F-4600 fluorescence spectrophotometer. The luminescence quantum yields were determined relative to Coumarin 307 in ethanol solution as a quantum yield standard ($\Phi = 0.56$). Confocal Laser Scanning Microscope (CLSM) experiments were carried out with Leica SP5.

2.3. Computation

The geometric and electronic structures of **4** compounds were optimized by density functional theory (DFT) at the B3LYP level. The HOMO and LUMO energy levels are predicated by 6-31G(d) basis set. The compositions of molecular orbits were analyzed using the Gauss View 5.0 program.

2.4. Synthesis of compounds

The target luminogens were synthesized by the Knoevenagel condensation of *N,N*-dimethylamino benzaldehyde, *N,N*-

diphenylamino benzaldehyde and *N*-carbazolyl benzaldehyde with (1,3-dimethyl) barbituric acid, respectively (Scheme 1). Then, these luminogens were characterized by ¹H NMR (nuclear magnetic resonance spectrometry) and the FTIR (Fourier transform infrared spectrometry). Single crystals of compounds **2** and **4** were obtained by slow evaporation of these solutions in the *n*-hexane/CHCl₃ solution at room temperature.

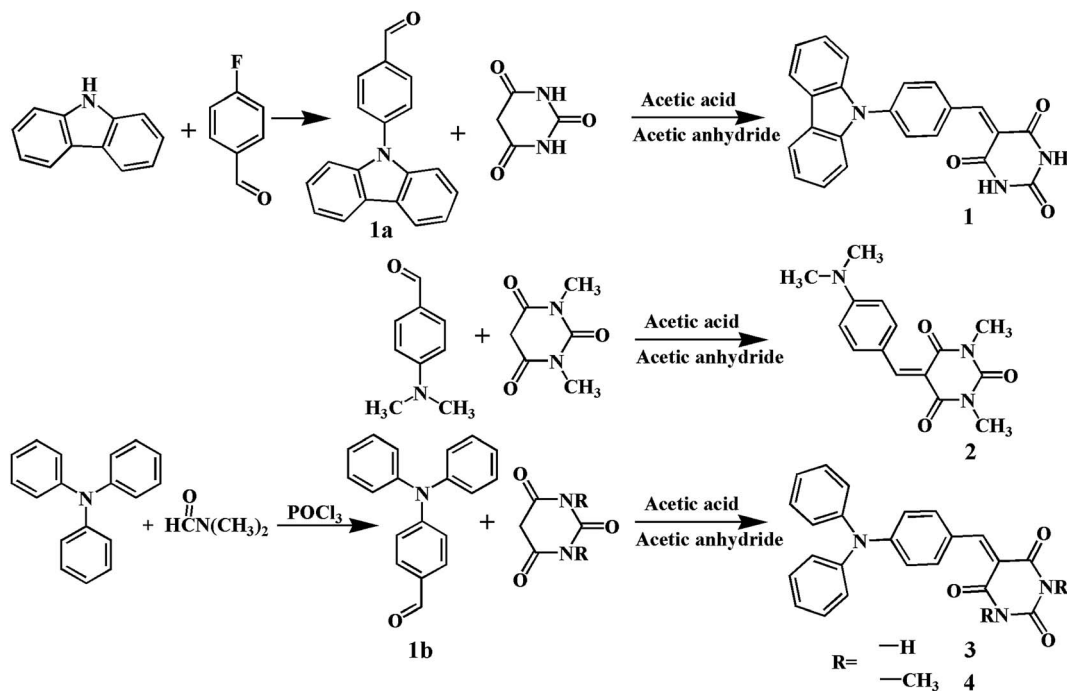
2.4.1. Synthesis of 4-(9H-carbazolyl) benzaldehyde (1a). A mixture of 9H-carbazole (1.7 g, 0.01 mol), and sodium hydroxide (1.2 g, 0.03 mol) in dry dimethylformamide (DMF) was heated with stirring. Then 4-fluorobenzaldehyde (1.5 g, 0.012 mol) was added, and the reaction mixture was stirred at 90 °C for 24 h. After cooling to room temperature, the mixture was poured into distilled water and extracted with ethyl acetate. The organic layer was dried with anhydrous magnesium sulfate (MgSO₄) and concentrated by vacuum evaporation. The crude product was purified by column chromatography using the mixture of petroleum and dichloromethane (v/v = 15 : 1) as an eluent to get the desired compound as a yellow solid (1.19 g) with a yield of 70.1%. ¹H NMR (CDCl₃) δ (ppm): 10.13 (s, 1H), 8.16 (m, 4H), 7.81 (d, 2H), 7.52 (d, 2H), 7.45 (t, 2H), 7.34 (t, 2H).

2.4.2. Synthesis of 4-(diphenylamino)-benzaldehyde (1b). 2 ml of dry DMF (25.7 mmol) was added dropwise to 5 ml of phosphorus oxychloride (54 mmol) maintained at 0 °C. After the solution was stirred for 15 min, triphenylamine (1 g, 4 mmol) was added to the above Vilsmeier reagent, and the mixture was heated to 45 °C for an additional 2.5 h. After cooling, the clear red solution was added dropwise to ice water with stirring, and the resulting mixture was allowed to stand for 2 h. The crude product was collected by filtration, washed with water, and dissolved in dichloromethane. The organic solution was washed with H₂O (150 ml) and dried over MgSO₄. After the solvent was evaporated off, the residue was recrystallized from ethyl acetate to afford aldehyde as a pale yellow crystal with a yield of 0.78 g (80%). ¹H NMR (CDCl₃) δ (ppm): 9.82 (s, 1H), 7.69 (d, 2H), 7.35 (t, 4H), 7.19 (m, 6H), 7.02 (m, 2H).

2.4.3. Synthesis of 5-(*N*-carbazole styryl)-barbituric acid (1). A mixture of barbituric acid (0.128 g, 1 mmol) and *N*-carbazolyl benzaldehyde (0.274 g, 0.7 mmol) in acetic (15 ml), acetic anhydride (15 ml) and benzene (5 ml) was added to dissolve the undissolved *N*-carbazolyl benzaldehyde, then the mixture was refluxed for 2 h. The reaction was cooled to room temperature and the solid particles were filtered. The product was then purified by recrystallization with acetic acid to give 0.30 g with 75% yield. ¹H NMR (DMSO) δ (ppm): 11.43 (s, 1H), 11.30 (s, 1H), 8.43 (d, 2H), 8.39 (s, 1H), 8.26 (d, 2H) 7.78 (d, 2H), 7.46 (m, 2H), 7.34 (m, 4H).

2.4.4. Synthesis of 5-(4-dimethylamino styryl)-1,3-dimethyl-barbituric acid (2). A mixture of 1,3-dimethyl-barbituric acid (0.78 g, 5 mmol) and *N,N*-dimethylamino benzaldehyde (0.745 g, 5 mmol) in acetic (5 ml) and acetic anhydride (5 ml) was refluxed for 2 h. The reaction was cooled to room temperature and the solid particles were filtered. The product was then purified by recrystallization with acetic acid to give 0.97 g with 68% yield. ¹H NMR (CDCl₃) δ (ppm): 8.45 (s, 1H), 8.40 (d, 2H), 6.76 (d, 2H), 3.41 (d, 6H), 3.17 (s, 6H).





Scheme 1 Synthetic routes to 1–4.

2.4.5. Synthesis of 5-(4-diphenylamino styrene)-barbituric acid (3). A mixture of barbituric acid (0.384 g, 3 mmol) and 4-(diphenylamino) benzaldehyde (0.447 g, 3 mmol) in acetic (5 ml) and acetic anhydride (5 ml) was refluxed for 2 h. The reaction was cooled to room temperature and the solid particles were filtered. The product was then purified by recrystallization with acetic acid to give 0.87 g with 78% yield. ^1H NMR (DMSO) δ (ppm): 11.21 (s, 1H), 11.07 (s, 1H), 8.28 (d, 2H), 8.15 (s, 1H), 7.44 (t, 4H), 7.25 (m, 6H), 6.76 (d, 2H).

2.4.6. Synthesis of 5-(4-diphenylamino styrene)-1,3-dimethyl-barbituric acid (4). A mixture of 1,3-dimethyl-barbituric acid (0.163 g, 1 mmol) and 4-(diphenylamino) benzaldehyde (0.276 g, 1 mmol) in acetic (5 ml) and acetic anhydride (5 ml) was refluxed for 2 h. The reaction was cooled to room temperature and the solid particles were filtered. The product was then purified by recrystallization with acetic acid to give 0.27 g with 70% yield. ^1H NMR (CDCl_3) δ (ppm): 8.44 (s, 1H), 8.24 (d, 2H), 7.38 (t, 4H), 7.23 (m, 6H), 6.95 (d, 2H), 3.40 (d, 6H).

2.5. Cell culture and imaging

Drosophila melanogaster S2 cells were cultured in Shields and Sang M3 Insect Medium (39.4 g L^{-1}) with fetal bovine serum (15%, v/v), penicillin–streptomycin solution (1% v/v at a final concentration of 100k units per L penicillin and 100 mg L^{-1} streptomycin) and KHCO_3 (0.5 g L^{-1}) and incubated at 22°C . The cells were seeded in glass-bottom plates at a density of 5×10^4 cells per mL and incubated for 30 min at 22°C . A DMSO solution of 4 (10 mM) was diluted with phosphate buffered saline (PBS) to a final concentration of 0.1 mM. The cultured cells were incubated with a PBS solution (50%, v/v) of compound 4 (0.1 mM) for 2 hour, washed three times with PBS

(100 μL) and imaged. All images were recorded with a Leica SP5 confocal laser scanning microscope (CLSM).³⁷

For the colocalization experiments, a PBS solution (50%, v/v) of 4 (0.1 mM) was added to the cells for 2 hour, and washed three times with PBS (100 μL). Then PBS solutions (50%, v/v) of 4',6-diamidino-2-phenylindole (DAPI) or LysoTracker Blue (10 μM) were added to the cells for 30 min and washed three times with PBS (100 μL). The fluorescence of DAPI or LysoTracker Blue was recorded in blue channels ($\lambda_{\text{Ex}} = 405 \text{ nm}$, $\lambda_{\text{Em}} = 420\text{--}470 \text{ nm}$), the fluorescence of compound 4 was recorded in red channel ($\lambda_{\text{Ex}} = 488 \text{ nm}$, $\lambda_{\text{Em}} = 500\text{--}600 \text{ nm}$).

3. Results and discussion

3.1. Optical properties

Optical properties of the compounds 1–4 were investigated by UV-vis and fluorescence spectroscopy. All the compounds demonstrate good solubility in typical organic solvents such as THF, DMF, chloroform, and DMSO, but they are hardly soluble in water. The UV-vis absorption spectra of compounds 1–4 are presented in Fig. 1.

As shown in Fig. 1, they exhibit quite similar absorption spectra in THF. The absorption bands ranging between 270 and 350 nm are assigned to $\pi\text{--}\pi^*$ electronic transition of the respective benzene ring, carbazole and diphenylamine units. The molecules exhibit strong absorption peaks at 412, 454, 454, and 458 nm for 1–4, respectively, which can be attributed to intramolecular charge transfer (ICT) from the carbazole, dimethylamino and diphenylamine units to the (1,3-dimethyl) barbiturate moiety.³⁸ Density functional theory (DFT) calculations also confirmed the ICT possibilities of these compounds (Fig. 9).



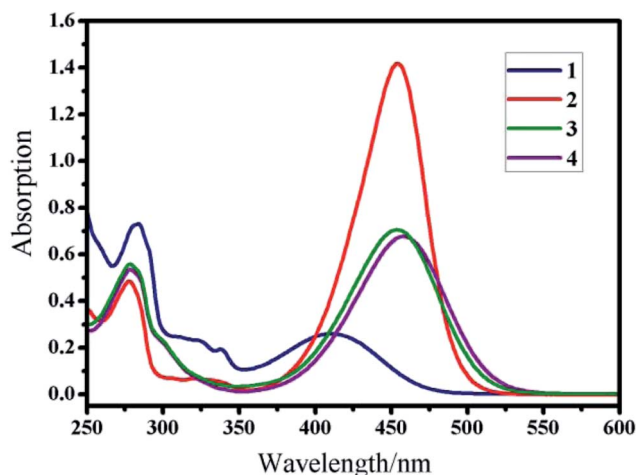


Fig. 1 UV-visible absorption spectra of 1–4 (24 μM) in THF solution.

We investigated the emission of the four compounds in different states. As shown in Table 1, in pure THF with low concentration, these compounds are almost molecularly isolated without any interactions between the adjacent molecules. All the molecules exhibit distorted conformation due to the intramolecular steric hindrance.³⁹ The optimized geometric structure of **1** show twisted conformation with a dihedral angle of 56.63° (Fig. S17[†]). Thus the molecules are not in good conjugation state, which contribute to the shortest emission wavelength (Fig. 2).

The molecules of **2** in the crystal state are arranged so regularly that the rotatable single bonds are locked due to the multiple intermolecular interactions (Fig. 6). These interactions help to further solidify the molecular conformation in the aggregated state, and they block the nonradiative pathways.⁴⁰

3.2. AIE/AIEE properties

Emission spectra of 1–4 in THF and THF/water media were recorded to facilitate the aggregation due to their water insoluble characters (Fig. 3). Therefore, the luminogens are able to aggregate in the aqueous mixtures with high water fractions (f_w). Mixture of THF and water was used for preparation of nanoaggregates by increasing the percentage of water in the THF solution. Their fluorescence (FL) spectra with different f_w

are shown in Fig. 3, and the changes of FL peak depict the variation tendency of fluorescence intensity with different f_w .

We can see from the photographs that in pure THF with low concentration, compounds **2** and **4** are nonemissive, while **1** and **3** show weak emission in dilute pure THF (Fig. 4). In Fig. 3, the data indicate that all chromophores exhibit different fluorescent behaviors in mixed solvents with different f_w . The FL intensities of **2** and **4** in pure THF are so low that only weak signals are recorded with maxima at 542.8 and 633.4 nm, respectively (Fig. 3b and d). Then they start to increase when the f_w is increased to 70% and 60%, respectively. The solutions emit orange and red light with λ_{Em} at 603.4 and 627.2 nm at $f_w = 90\%$, respectively. The FL intensity of **2** enhances 23.6-fold and the fluorescence quantum yield (Φ_F) increases from 0.4% to 8.2% when f_w increases from 0% to 90%. While the FL intensity of **4** enhances 42.2-fold (Φ_F : 0.1% to 3.2%). These data demonstrate **2** and **4** are AIE-active molecules.

The compounds **1** and **3** display a different fluorescent behavior from **2** and **4** in mixed solvents with different f_w . In pure THF, **1** and **3** show weak emission at 592.6 and 619.4 nm (Table 1). The emission of **3** is obviously red-shifted (26.8 nm) compared with **1**, because the diphenylamino group is a much stronger electron donor than the carbazole group, resulting in an increase in the HOMO energy.⁴¹ And the subsequent theoretical calculations support this speculation (Fig. 9). The FL intensity of **1** and **3** gradually decreases when $f_w < 70\%$ which may be caused by ICT.⁴² When $f_w > 70$, the FL intensity increases rapidly. The FL intensity of **1** increases 3.3-fold and the Φ_F increases from 5.4% to 11.4% when f_w increases from 0% to 90%, while the FL intensity of **3** enhances 6.5-fold (Φ_F : 1.8% to 7.4%). These data demonstrate that **1** and **3** are AIEE-active molecules.

It is also noted that the emission peak varies with f_w (Fig. 3, right). The emission peak of **2** is red-shifted from 542.8 to 603.4 nm. The large red-shift may be caused by π – π stacking when the **2** aggregate. However, compound **3** exhibits a more complicated tendency with the increase of f_w . The emission peak of **3** red-shifts from 619.4 nm to 648.4 nm when f_w increases from 0% to 70%. Further, it blue-shifts to 624.4 nm as f_w increase to 90%. When f_w is no more than 70%, molecule **3** is still completely dissolved in the mixtures, and increased f_w enhances the polarity of mixture, generating red-shift emission due to the ICT effect. As f_w reaches 70%, due to the decreased solvating power, molecule **3** starts to aggregate, resulting in less

Table 1 Optical properties of compounds 1–4

	Monomer/THF				Aggregates ^d		λ_{solid} (nm)	
	λ_{abs} ^a (nm)	λ_{Em} ^b (nm)	Stokes shift	Φ_F ^c (%)	λ_{Em} (nm)	Φ_F (%)	Powder	Crystal
1	412.0	592.6	180.6	5.4	612.0	11.4	597.0	—
2	454.0	542.8	88.8	0.4	603.4	8.2	620.0	621.0
3	454.0	619.4	162.4	1.8	624.4	7.4	637.6	—
4	458.0	633.4	175.4	0.1	627.2	3.2	608.0	582.0

^a The maxima absorption peak. ^b The maxima emission peak, in dilute THF solution (24 μM). ^c Fluorescence quantum yields were determined relative to Coumarin 307 in ethanol solution as quantum yield standard ($\Phi = 0.56$). ^d In mixed water/THF solution with $f_w = 90\%$.



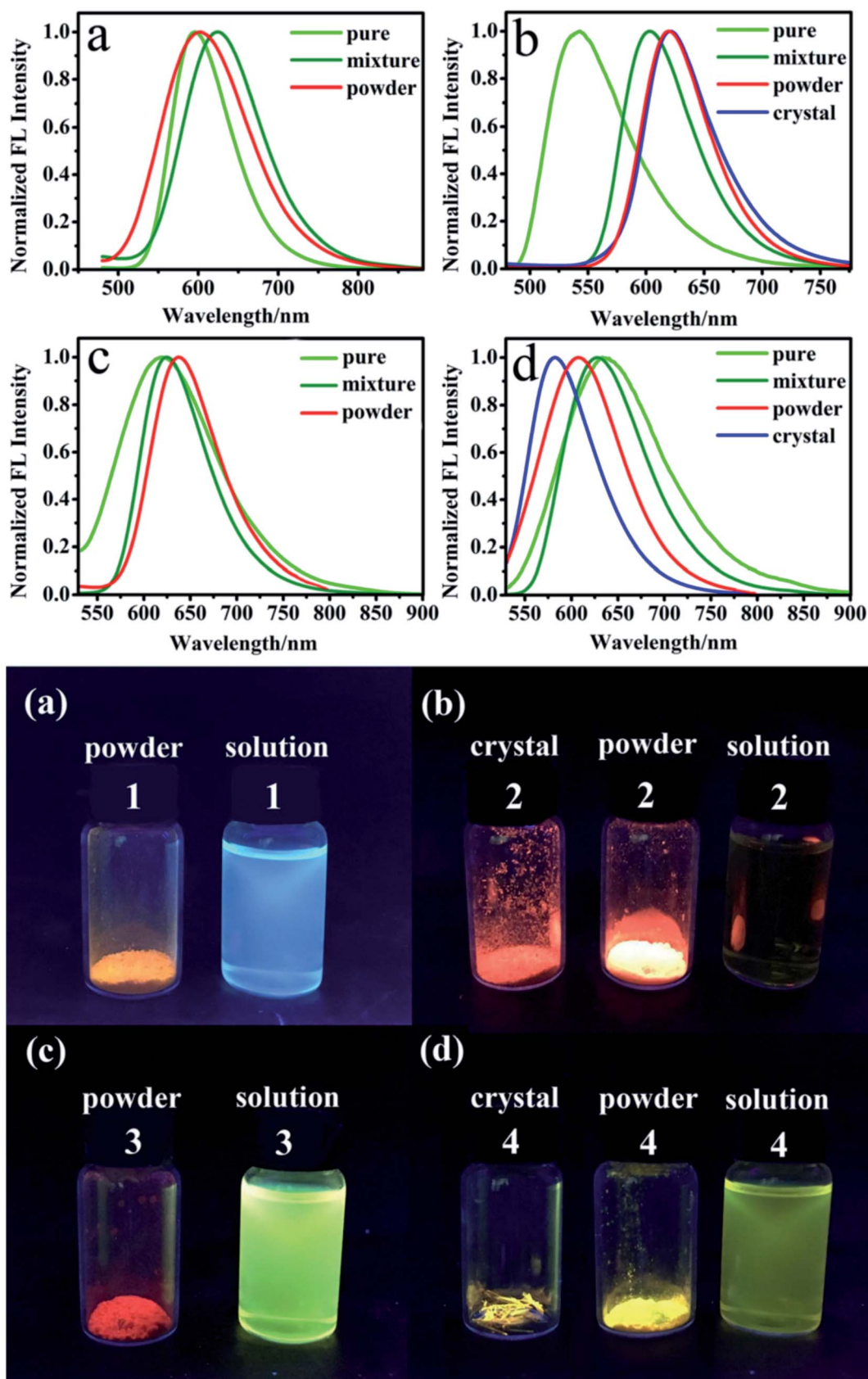


Fig. 2 Normalized FL spectra and photographs of (a) 1, (b) 2, (c) 3 and (d) 4 in different state (THF solution, solid powders, crystal).

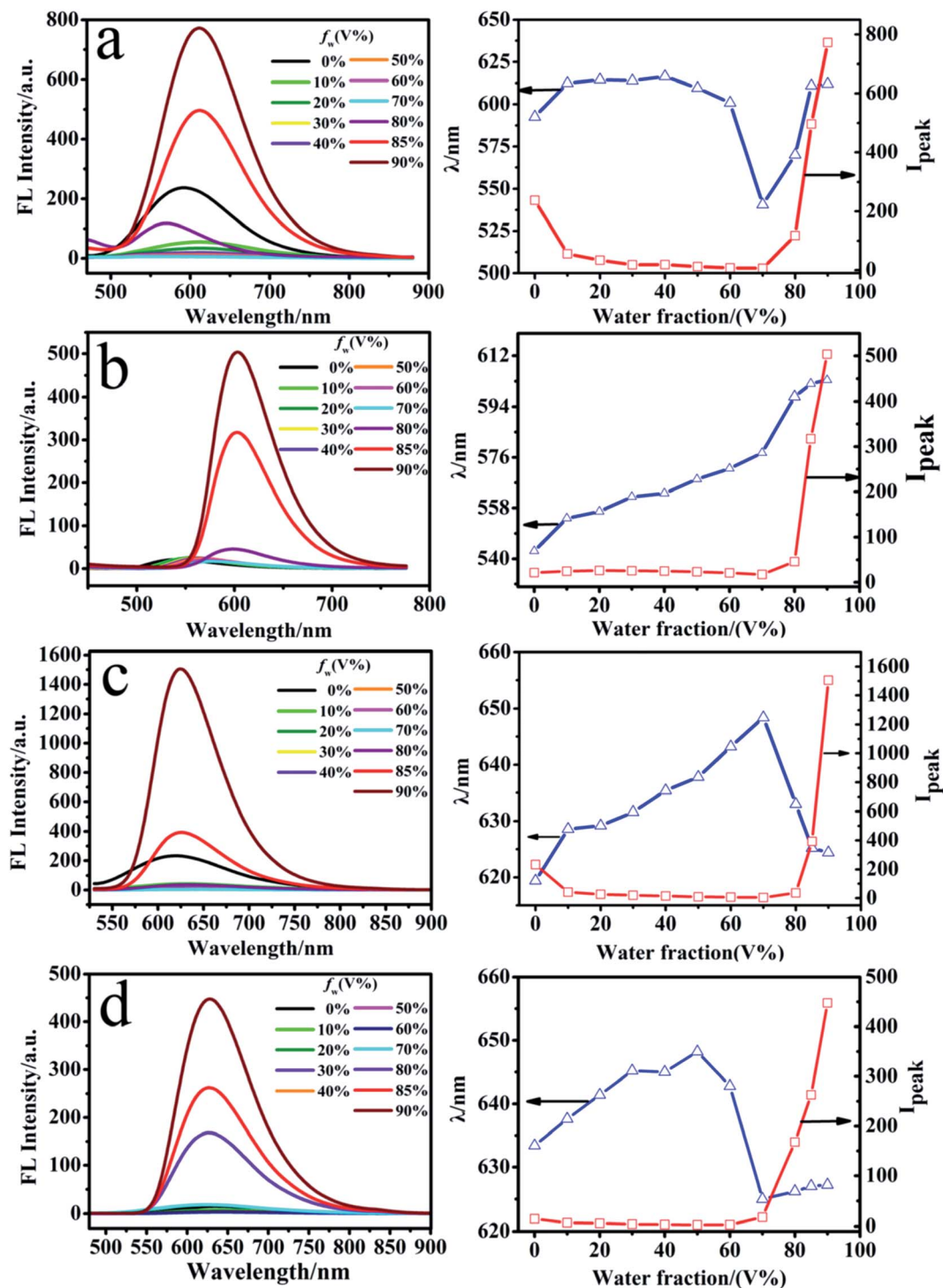


Fig. 3 FL spectra of (a) 1, (b) 2, (c) 3 and (d) 4 in THF/H₂O mixtures with different water fraction (f_w) (left); excitation wavelength = 450 nm, 398 nm, 510 nm, 460 nm. Plot of relative FL peak location (blue) and emission intensity (red) of 1–4 versus the water fraction in the aqueous mixtures (right).

polar micro-environment for the luminogens because of self-wrapping. Thus reversely giving blue-shifted emissions.^{43,44} Compounds 1 and 4 behave similarly to 3.

The morphology and size of the nanoaggregates in the THF/H₂O solvent with different f_w were detected by scanning

electron microscopy (SEM) and dynamic light scattering (DLS) (Fig. 5), which show the presence of spherical and blocky aggregates.

As shown in Fig. 5, at $f_w = 90\%$, the average diameters (d) of the nanoaggregates for 1–4 are 394.2, 229.3, 382.4 and 346 nm,



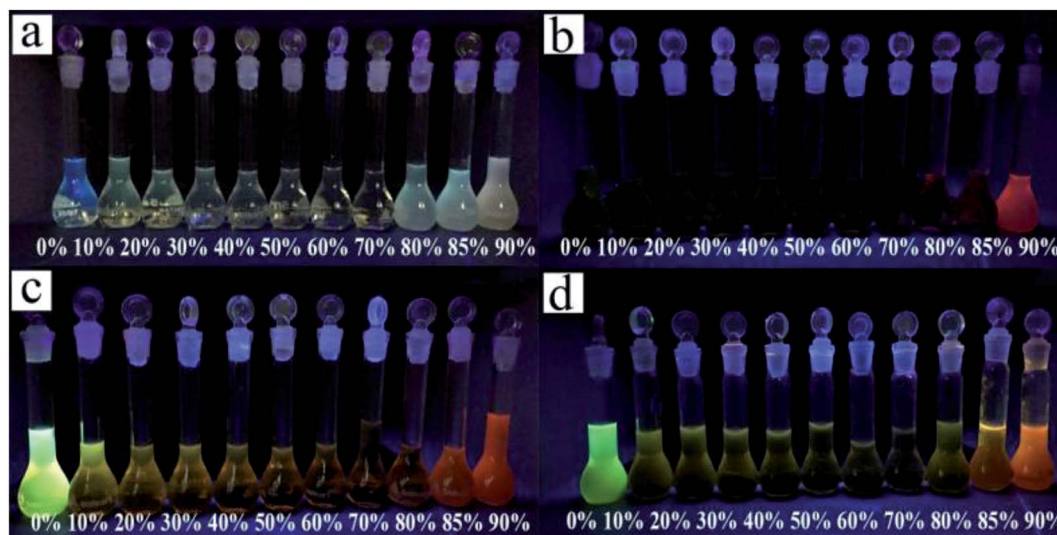


Fig. 4 Photographs of (a) 1, (b) 2, (c) 3 and (d) 4 at $f_w = 0\%$ to 90% under 365 nm UV light illumination.

respectively. The size of nanoaggregates decrease gradually with increasing f_w (Table S1†), due to the rapidly decreased solvating power of the mixed solvent (THF/ H_2O).⁴³ These data indicate that the enhanced emission of compounds is related to the formation of nanoaggregates.

3.3. Single crystal structure

Single crystal X-ray diffraction (SXRD) analysis can provide a direct insight into the molecular conformation and the packing structure of a compound. Compound 2 adopts planar construction, and it has good planarity (Fig. 6b). The dihedral angles of 4 between the terminal unit and the phenyl ring are 59.50° and 87.23° , respectively, which indicates that compound 4 shows a large torsion (Fig. 7b).

As shown in Fig. 6b, in crystal 2, the C–H \cdots O interaction between carbonyl group and methyl group links the adjacent molecules into one-dimensional linear chain, and the distance of C–H \cdots O interaction are 2.581 \AA . Then, the adjacent chains are further linked into two-dimensional network structures through C–H \cdots O interactions (the distances are 2.548 and 2.665 \AA , respectively) between carbonyl groups and methyl groups, C–H \cdots H–C ($d = 2.310\text{ \AA}$) and two C–H \cdots π interactions ($d = 2.896\text{ \AA}$). These interactions make the molecular conformations stable.

As depicted in Fig. 7, the molecule 4 adapts a twisted conformation due to the steric hindrance between the diphenylamino unit and the 1,3-dimethylbarbiturate moiety. And the adjacent molecules are connected *via* intermolecular C–H \cdots O interactions into one-dimensional linear bands, the distances of C–H \cdots O interactions are 2.531 \AA and 3.211 \AA , respectively. As shown in Fig. 7c, the adjacent molecules in crystal 4 are herringbone stacking by C–H \cdots O ($d = 2.564\text{ \AA}$), C–H \cdots π ($d = 2.762\text{ \AA}$), C=O \cdots C and C=O \cdots N interactions ($d = 2.973\text{ \AA}$, 3.048 \AA , and 3.155 \AA). The crystal also possesses a π – π interaction between two terminal phenyl of adjacent molecules and the distance is 3.384 \AA in Fig. 7d. These interactions make their configuration stable.⁴⁰

3.4. Mechanisms of enhanced emission

As discussed above, compounds 1–4 are practically nonemissive when molecularly dissolved in solvents due to the active intramolecular rotations and vibrations. The intermolecular interactions are mainly reflected in restriction of intramolecular motions and some specific molecular stacking modes, such as J-aggregation and herringbone stacking,^{45,46} which can restrain the radiative decay process and promotes the enhanced emission behavior. On the contrary, the H-aggregation, where molecules are arranged in face-to-face type to form excimers or exciplexes, can promote the nonradiative deactivation process.⁴⁷

In diluted solutions, the molecules 1–4 are almost isolated and adopt a twisted conformation due to the steric hindrances in the molecules which decreases the π -conjugation. The intramolecular motions such as rotations, vibrations, stretching, will consume the energy of excited state and suppress the radiative decay process. While in the aggregated state, there are numerous intermolecular interactions between the hydrogen and oxygen, carbon or nitrogen atoms; besides, some π – π stacking or C–H \cdots π interactions exist in the aggregate structures. These interactions lead to the planarity and stability of molecular conformation. Under the effect of intermolecular interactions, intramolecular vibrations or rotations are locked, and the molecular conformations are more rigid and planar. These motions are suppressed greatly due to the spatial constraint, which blocks the nonradiative decay pathway and promotes the radiative decay of the excited state.³³

The restriction of intramolecular motion (RIM) induced by the molecular aggregation can be considered as the mechanism of enhanced emission for 1–3. Furthermore, the molecule 4 has a herringbone stacking conformation in crystal. Thus the enhanced emission of 4 was attributed to the synergetic effects of the herringbone stacking formation and RIM. Therefore, the phenomenon of fluorescence enhancement can be ascribed as the formation of nanoaggregates of molecules 1–4 at high f_w according to the SEM and DLS results.



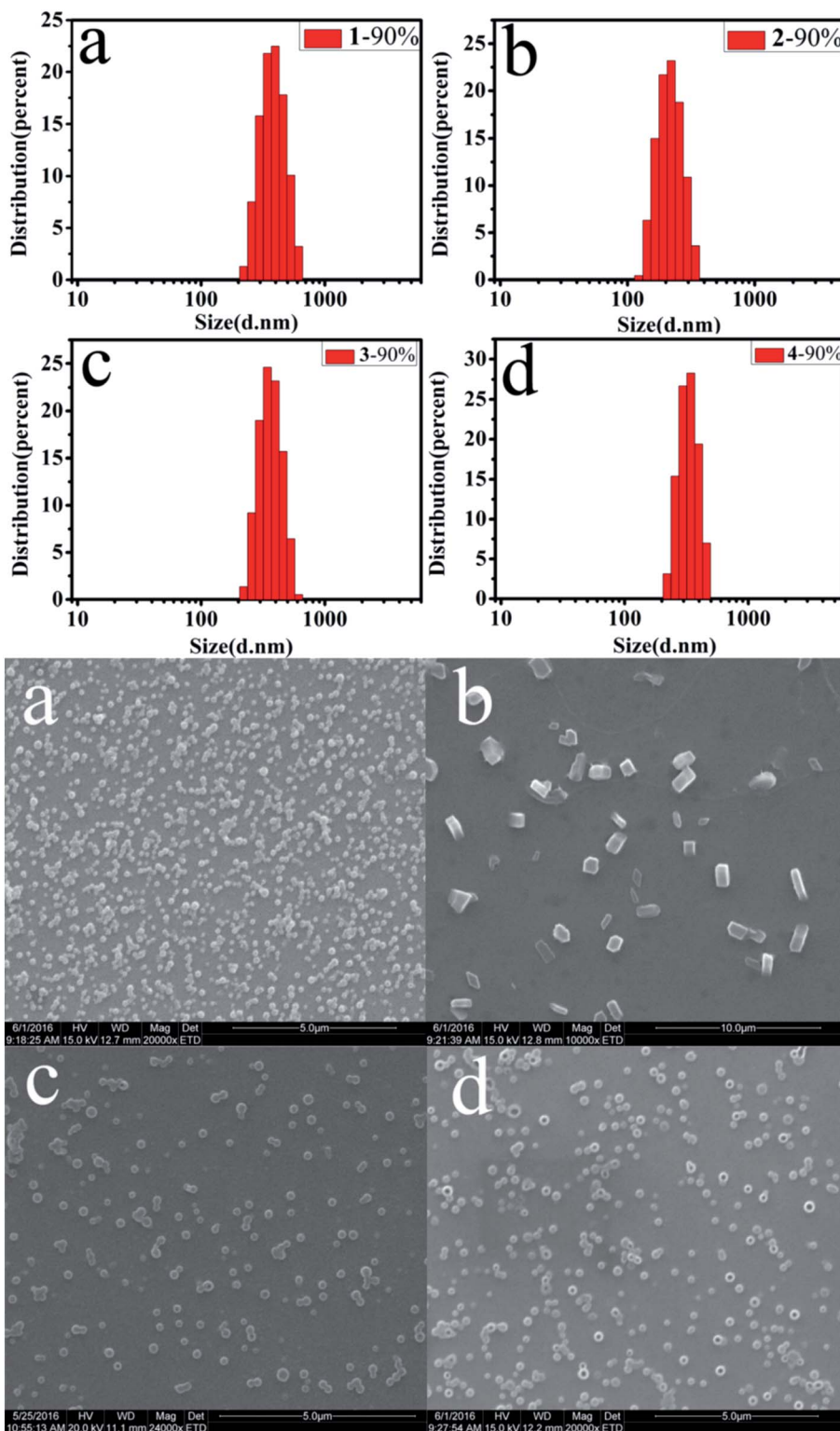


Fig. 5 SEM images and particle size distributions of (a) 1, (b) 2, (c) 3 and (d) 4 in THF/H₂O mixtures ($f_w = 90\%$) at concentrations of 24 μM .

3.5. *In vivo* imaging

Before exploring the application for live cell imaging, we examined the biocompatibility of compound 4. Comparing the morphologies of S2 cells with (Fig. 8a) and without (Fig. S19†) the presence of compound 4, they are almost identical shape

in the white-field channel, which suggest its non-cytotoxic feature. The excellent biocompatibility of compound 4 motivated us to further investigate their application in cell imaging. The time scanning spectra of compound 4 at $f_w = 0.9$ were shown in Fig. S18.† As can be seen, the fluorescence



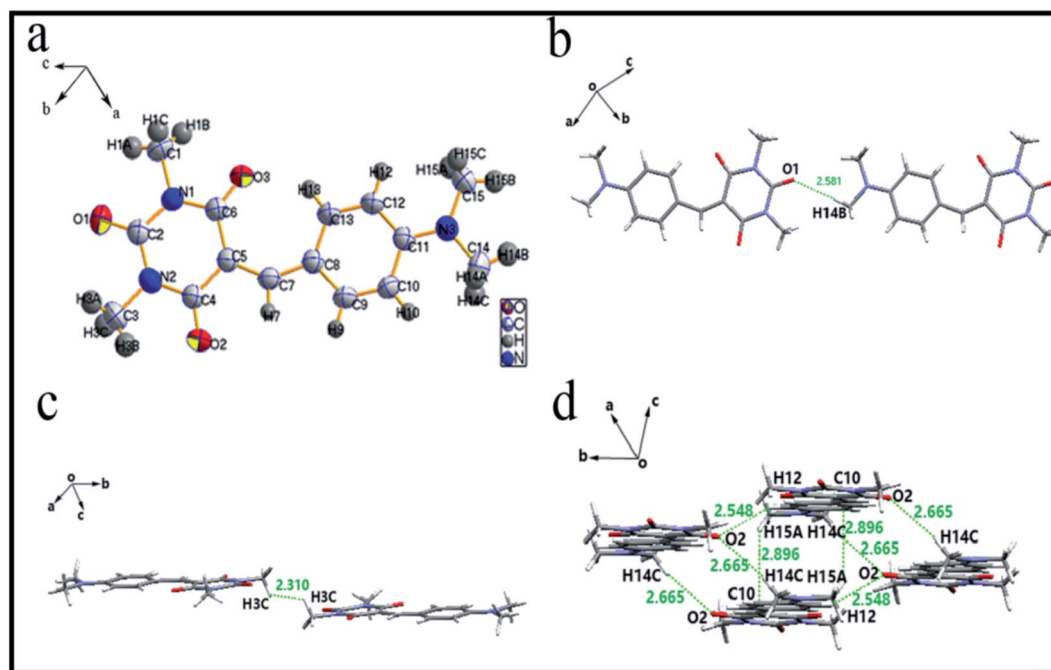


Fig. 6 ORTEP (thermal ellipsoids at the 50% probability level) diagram (a), one-dimensional structure (b), two-dimensional network structure (c) and three-dimensional structure (d) of **2**.

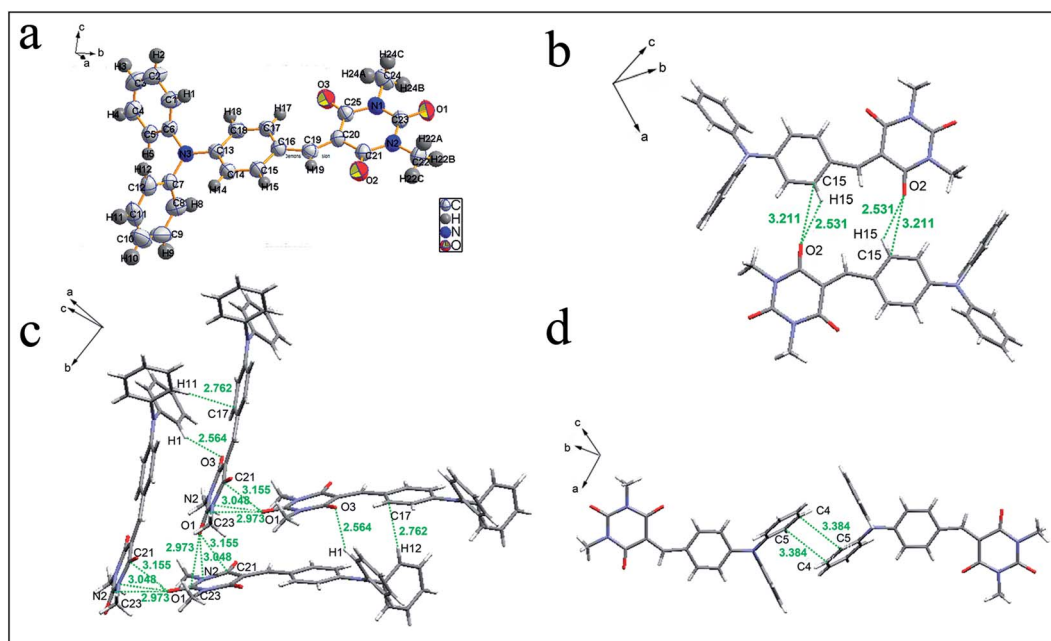


Fig. 7 ORTEP (thermal ellipsoids at the 50% probability level) diagram (a), one-dimensional structure (b), herringbone stacking structure (c) and π - π interactions (d) of **4**.

intensity hardly changed in 1800 s. These data showed that the nanoparticles exhibited excellent stability under light irradiation.

Compound **4** was then assessed for its capability to cell imaging by confocal laser scanning microscopy (CLSM). After incubation with *Drosophila* S2 cells with **4**, compound **4** can be

internalized into the cells, and red fluorescence can be detected intracellularly with minimal background autofluorescence signals. Presumably, compound **4** can be internalized into the endosome compartments and eventually move to the lysosome compartment of the cells within 2 hours. Indeed, the water insoluble character of compound **4** facilitates the passive cell



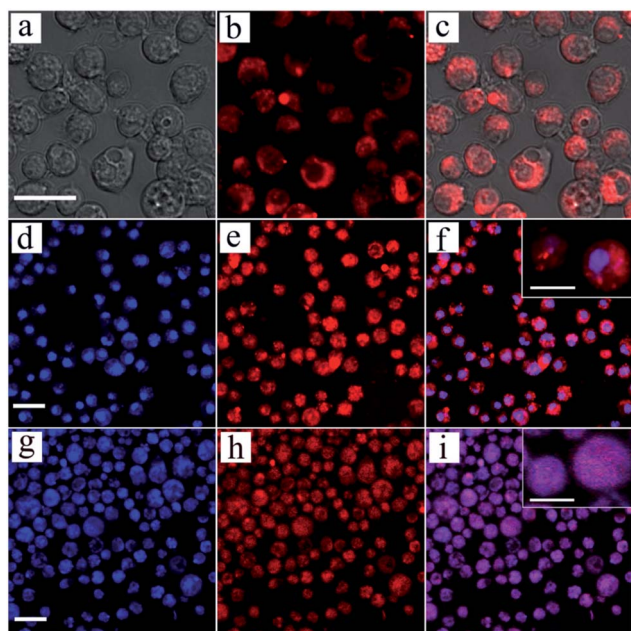


Fig. 8 (a–c) CLSM of *Drosophila* S2 cells incubated with compound **4** (50 μ M). (a) White-field channel, (b) fluorescence channel, (c) merged channel; (d–f) co-localization experiment of *Drosophila* S2 cells incubated with compound **4** (red channel) and DAPI (blue channel) and the merged images (f) (inset is an enlarged view of (f)); (g–i) co-localization experiment of *Drosophila* S2 cells incubated with compound **4** (red channel) and LysoTracker Blue (blue channel) and the merged image (i). Scale bar = 20 μ m; inset scale bar = 10 μ m.

uptake and accumulation of the molecule into the lysosome. Indeed, co-localization experiments of compound **4** with LysoTracker Blue suggested they moved into the same intracellular space – lysosomes (Fig. 8g–i).

Intracellular co-localization experiment was performed with compound **4** and 4,6-diamino-2-phenylindole (DAPI). DAPI is a nucleus-targeting fluorescence probe, and accumulates selectively in the nucleus structure of cells. In the co-localization experiment, intense blue fluorescence can be detected in nucleus of the S2 cell, while no red fluorescence can be detected in the same region, suggesting that the compound **4** is unable to pass the nuclear membrane (Fig. 8f). The images show intense fluorescence was detected exclusively in the cytoplasm part of the *Drosophila* S2 cell, which might because of the molecule aggregation in the cells. The aggregated state tends to prevent fluorescence quenching resulted from non-radiative decay caused in aqueous environment, which is actual a common phenomenon in AIE-based bioimaging probes.⁴⁸

3.6. Theoretical calculation

To have a better insight into their optical properties, we calculated frontier molecular orbitals by using the DFT at B3LYP/6-31G(d) basis set. The optimized geometries and HOMO/LUMO plots of **1–4** are illustrated in Fig. 9. The geometry optimized structures of **1–4** show highly twisted conformation, which are favorable for active intramolecular rotations of multiple phenyls and carbazole groups in solutions, thereby powerfully dissipating the excitons energy and making them nonemissive or weak emissive in solvents. The theoretical calculated HOMO levels of **1–4** are -5.61 , -5.53 , -5.47 , and -5.44 eV, respectively. The energy gaps for **1–4** are 2.89, 3.27, 2.98, and 3.01 eV, respectively. And the gaps estimated by absorption maxima of **1–4** are 3.81, 4.18, 4.12 and 4.10 eV, respectively. Therefore, the calculated values agree well with the test ones.

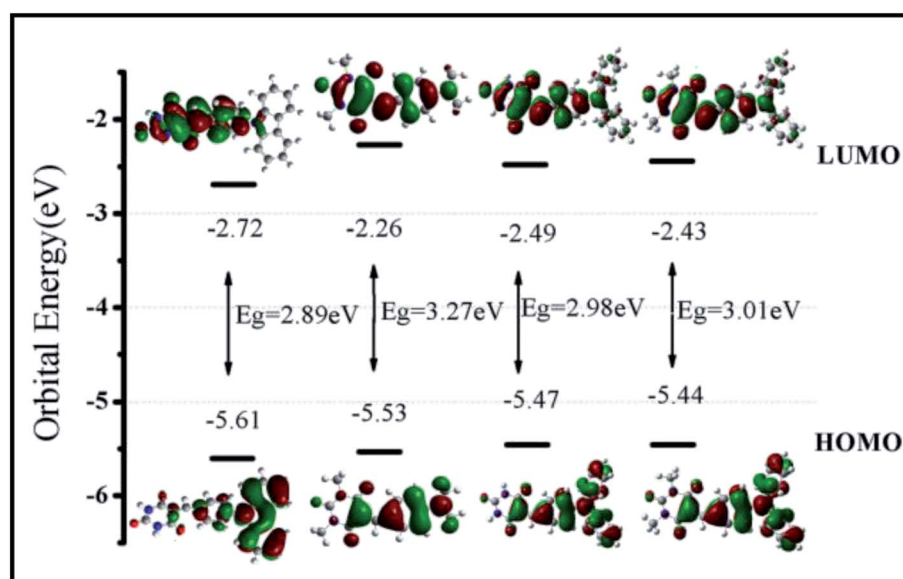


Fig. 9 Energy levels of HOMO and LUMO, energy gaps, and electron cloud distributions of molecules **1–4** calculated by the B3LYP/6-31G(d) program. (HOMO = the highest occupied molecular orbital energy; LUMO = the lowest unoccupied molecular orbital energy; E_g = energy gap between HOMO and LUMO levels.)



4. Conclusions

In summary, we report four luminogens **1–4** with typical AIE/AIEE characteristics. Their emission wavelength changed with different f_w , and their powders emit orange (597 nm and 608 nm) and red (620 nm and 637 nm) light. UV-vis absorption spectra and theoretical calculation reveal the ICT possibilities of **1–4** from the donor units to the acceptor units. And we analyze the mechanisms of enhanced emission of these compounds are RIM. Furthermore, the theoretically calculated results agree with the measured optical properties. With the advantages of excellent biocompatibility and simplicity, compound **4** represents an excellent probe for *Drosophila* S2 cells imaging. Once entering lysosomes, the intramolecular motion of compound **4** will be restricted and the emission from compound **4** will be turned on.

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