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Original article

Synthesis and antitumor activity of α -aminophosphonate derivatives containing thieno[2,3-d]pyrimidines

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ABSTRACT

Two series of thieno[2,3-d]pyrimidine derivatives were designed and synthesized, in which bioactive α aminophosphonate subunits were introduced at the N3 position through an N–N bond connection. The *in vitro* cytotoxic activity of the novel compounds was tested against human esophageal carcinoma cells (EC109), human hepatocarcinoma cells (HepG2), human gastric carcinoma cells (MGC-803), respectively, by the MTT method. The evaluation results revealed that compounds **6mb**, **6mf**, **6mg**, **6nd** and **6nh** exerted the most potent inhibition against HepG2, MGC-803 and EC109 cells, respectively. In particular, compound **6mg** presented excellent inhibitory effect against HepG2 (91.2%) and MGC-803 (94.4%) cells.

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1. Introduction

It is well known that fused pyridines and thienopyrimidines have many valuable pharmacological properties, such as antioxidative [1,2], antimalarial [3,4], antibacterial [5], and antiviral effects [6]. The pyrimidine-based derivatives, *e.g.* thieno[2,3d]pyrimidines, were proved to have pharmacological and therapeutic properties in medicinal chemistry, such as anti-inflammatory [7], antibacterial [8,9], antifungal [10,11], antitumor [12], antidepressant [13], antiplatelet [14], antihypertensive [15], herbicidal [16] and plant growth regulatory [17] properties.

As phosphorus analogs of α -amino acids and their esters, α aminophosphonates have received much attention owing to their potential biological activities such as antibacterial, anticancer, as well as insecticidal and herbicidal properties, [18–22] and were investigated as starting materials for the syntheses of phosphonopeptides. Moreover, if the aminophosphonate subunit was suitably embedded into potential antitumor agents, it could effectively improve their bioactivity [23], e.g. benzothiazole moiety

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or fluorine moiety containing α -aminophosphonates [24–27] were proved to have high antitumor activity.

Based on the concept of bioisosterism and inspired by the above observations, we were led to the proposal to incorporate thieno[2,3-*d*]pyrimidine ring into the α -aminophosphonate compounds, to construct novel classes of thieno[2,3-d]pyrimidine moiety containing α -aminophosphonate derivatives, and to evaluate their antitumor activity. Therefore, two series of such compounds, incorporating the α -aminophosphonate and the thieno[2,3-d]pyrimidines, were designed and synthesized and their antitumor properties were investigated.

Twenty-two new diethyl or diisopropyl [(3-substituted or 4-substituted or 2,4-disubstituted) (2-methyl-6-ethyl-4-oxo-4-*H*-thieno[2,3-d]pyrimidin-3-yl-amino) methyl] phosphonates were synthesized and their *in vitro* antitumor activity under cell membrane conditions were evaluated by the MTT method. Compounds **6ma-6mk** and **6na-6nk**, as well as the intermediates **4a-4k**, were tested for their anti-proliferation activity against three human cancer cell lines (EC109, HepG2 and MGC-803) at different concentrations. This represents the first report about the synthesis and *in vitro* antitumor activity evaluation of thieno[2,3-d]pyrimidines containing α -aminophosphonate derivatives.

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2. Experimental

All starting materials were commercially available and analytically pure. ¹H NMR spectra were recorded on a BRUKER DPX-400 spectrometer (Bruker Company, Germany), using TMS as an internal standard and CDCl₃ as a solvent. Chemical shifts (δ values) and coupling constants (*J* values) are given in ppm and Hz, respectively. TLC analysis was carried out on silica gel plates GF254 (Qindao Haiyang Chemical, China). High-resolution mass spectra were performed on an ESI Q-TOF MS spectrometer (Micromass, England). Melting points were determined on an XT4A apparatus (Beijing Keyi Electro-optic Instrument Plant, China) and were uncorrected. The intensity data were collected on an Agilent xcalibur Eos-II- CCD-diffractometer (Agilent Technologies).

Twenty-two α -aminophosphonate derivatives (**6ma-6mk** and **6na-6nk**) were prepared according to the reaction sequences shown in Scheme 1. The starting materials, ethyl 2-amino-5ethylthiophene-3-carboxylate (1) [28] and dialkyl phosphite (5), were prepared according to the literature methods. The target analogs were easily obtained in high yields with the established reaction conditions. The acetylation of compound (1) with acetyl chloride in acetonitrile gave the ethyl ester of 2-acetylamino-5ethylthiophene-3-carboxylic acid (2) [29]. The acetylated intermediate (2) then underwent condensation-cyclization reactions with hydrazine hydrate to afford compound (3), 3-amino-6-ethyl-2-methylthieno[2,3-d]pyrimidin-4-one, in good yield [30]. The Schiff base type key intermediates (4), obtained by the reaction of (3) with various substituted benzaldehvdes, reacted with different dialkyl phosphite (5) under nitrogen atmosphere to generate the corresponding α -aminophosphonate derivatives (**6ma-6mk** and **6na-6nk**). All the new compounds were characterized using ¹H NMR and ¹³C NMR, mass spectrometry and high resolution mass spectrometry, respectively (see Supporting information for structure characterization, yields and melting points).

Fortunately, the single crystals of compound **6mi** and **6mj** were obtained. The molecular structures of the two compounds are illustrated in Figs. 1 and 2. Colorless single crystals of the compound **6mi** $(0.35 \text{ mm} \times 0.30 \text{ mm} \times 0.30 \text{ mm})$ and **6mj** $(0.30 \text{ mm} \times 0.25 \text{ mm} \times 0.22 \text{ mm})$ were selected for X-ray diffraction analysis. The data were collected on an Agilent xcalibur Eos-II-CCD-diffractometer equipped with a graphite-monochromatic Cu Kα radiation (λ = 1.5418 Å) by using a ω scan mode in the range of $4.46 < \theta < 72.28^{\circ}$ for **6mi** and $4.42 < \theta < 67.07^{\circ}$ for **6mj** at 291(2) K. The entire structure was solved by the direct methods using the SHELXS-97 program [31] and refined by the full-matrix leastsquares method on F2 with anisotropic thermal parameters for all non-hydrogen atoms using SHELXL-97. [32] For compound 6mi, a total of 8957 reflections were recorded and 4476 were unique (*R* int. = 0.0225), among which 4245 ($-13 \le h \le 13$, $-9 \le k \le 14$, $-19 \le l \le 21$) were observed. For compound **6mj**, a total of 8872 reflections were recorded and 4183 were unique ($R_{\text{int.}} = 0.0232$), among which 3930 $(-12 \le h \le 8, -14 \le k \le 13, -19 \le l \le 21)$ were observed. X-ray diffraction analysis revealed that the thieno [2,3-d]pyrimidine ring in two compounds did not exhibit good coplanarity and the molecular structure is stabilized by intermolecular N(1)-H(1)...N(3) hydrogen bonds. Thereby, the structure of 6mi and 6mj were further confirmed.

A 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay was performed to investigate *in vitro* cytotoxic activity. Human esophageal carcinoma cells (EC109), human hepatocarcinoma cells (HepG2) and human gastric carcinoma cells (MGC-803) seeded into the 96-well plate (100 μ L each well) were incubated at 37 °C under 5% CO₂ and 95% O₂ until cell adherence was observed. Cells were exposed to suspension of



Scheme 1. Synthetic routes for the targeted compounds.

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Fig. 1. X-ray structure of compound 6mi.



Fig. 2. X-ray structure of compound 6mj

compounds **6ma-6mk**, **6na-6nk**, as well as the intermediates **4a-4k** at different concentrations. After 48 h, 20 μ L of MTT with a concentration of 5 μ g/mL was added into the 96-well plate and incubated for 4 h at 37 °C. Then, the supernatant was aspirated, 150 μ L of DMSO was added to each well, and the absorbance was measured at 490 nm. The formula for calculating inhibition of cell proliferation was as follows: inhibition of cell proliferation rate (%) = (OD value of the control group - OD value of drug group)/ control OD value × 100%. This was used to plot an inhibition curve.

Table 1

3. Results and discussion

The *in vitro* cytotoxic activity of compounds **6ma-6mk**, **6na-6mk**, as well as the intermediates **4a-4k** against human esophageal carcinoma cells (EC109), human hepatocarcinoma cells (HepG2), human gastric carcinoma cells (MGC-803) was evaluated by the MTT method and the potency was expressed as inhibition rate. The results are summarized in Table 1.

Results in Table 1 show that the changes of substituents affected the antitumor activity. Among analogs **6ma-6mk** and **6na-6nk**, even intermediates Schiff bases **4a-4k**, compounds **6ma-6mk** and **6na-6nk** have a relatively higher antitumor activity than that of Schiff bases **4a-4k**. These observations indicate that the introduction of phosphoryl group improves antitumor activity to EC109, HepG2 and MGC-803 cells.

The antitumor data also indicate that the substitutes of phosphonate has no apparent influence on antitumor activity to EC109, HepG2 and MGC-803 cells. For example, when R was 4-NO₂ and R' was Et, compound **6me** exhibited moderate antitumor activity with an inhibition rate of 54.0% against EC109 cells at 50 μ g/mL; the antitumor activity of **6ne**, where the phosphonate was substituted by an *i*-Pr, was little higher than the activity of **6me** in EC109 with the inhibition rate of 58.5%. Compound **6mk** with 2-Cl (R), 4-Cl(R), and Et (R') has antitumor activity in HepG2 cells, with an inhibition rate of 77.8% at the concentration of 50 μ g/mL. Compound **6nk** with 2-Cl (R), 4-Cl(R), and *i*-Pr (R') has a lower antitumor activity (60.8%) than that of **6mk**. This same trend was observed for compounds **6md/6nd** and **6mh/6nh** against EC109, HepG2 and MGC-803 cells.

Compounds **6mg** (R:4-Cl, R':Et) and **6nh** (R:3-Br, R':*i*-Pr) showed potent inhibition against EC109 cells with the inhibition rate of 86.8% and 85.4% at 50 μ g/mL, respectively. Compounds **6mg** (R:4-Cl, R':Et) and **6nd** (R:3-NO₂, R':*i*-Pr) showed potent inhibition against HepG2 cells with the inhibition rate of 91.2% and 90.8% at 50 μ g/mL, respectively. Compounds **6mb** (R:3-CH₃, R':Et),**6mf** (R:3-Cl, R':Et),**6mg** (R:4-Cl, R':Et) and **6nh** (R:3-Br, R':*i*-Pr) showed potent inhibition against MGC-803 cells with the

Comp.	EC109			HepG2			MGC-803		
	50 µg/mL	10 µg/mL	5 μg/mL	50 µg/mL	10 µg/mL	5 μg/mL	50 µg/mL	10 µg/mL	5 μg/mL
6ma	66.3	26.1	27.5	66.0	35.4	32.4	50.3	29.0	20.6
6mb	79.8	43.7	11.5	81.6	41.9	31.4	91.3	53.1	44.7
6mc	64.4	8.0	1.0	62.3	12.8	6.6	64.6	8.2	1.1
6md	43.3	16.4	17.2	72.9	43.3	32.9	49.6	29.2	27.8
6me	54.0	10.8	4.4	61.1	15.3	9.5	59.8	-3.0	-10.4
6mf	81.4	36.1	30.3	86.7	43.7	28.2	94.4	41.4	41.5
6mg	86.8	37.6	36.7	91.2	39.7	32.8	94.4	42.2	25.8
6mh	40.4	44.8	20.9	53.8	23.7	14.9	46.3	27.0	18.2
6mi	76.5	15.3	7.6	74.3	12.3	4.6	65.4	0.6	-7.5
6mj	58.2	23.0	26.6	70.3	42.4	35.9	65.4	19.2	8.7
6mk	73.6	57.7	25.6	77.8	64.4	34.3	75.3	62.6	16.6
6nd	45.3	11.2	-2.0	90.8	40.3	26.0	63.6	40.1	27.7
6ne	58.5	5.3	-1.8	58.1	24.3	6.9	84.3	39.2	31.8
6nh	85.4	13.8	11.2	48.7	39.7	21.3	93.7	39.9	41.6
6nk	51.0	41.7	36.7	60.8	48.1	36.2	64.5	63.3	36.9
4a	51.4	2.9	-8.9	57.0	19.5	16.7	74.6	20.3	22.3
4b	49.5	14.7	-6.9	38.7	25.9	26.1	35.5	28.2	22.8
4d	29.2	17.7	16.7	39.6	29.1	26.4	33.3	10.5	9.4
4e	18.8	14.9	17.8	37.8	20.9	9.0	55.9	21.8	2.8
4f	17.0	9.3	5.6	40.7	18.9	4.6	28.2	20.5	6.0
4g	32.0	24.4	5.4	74.8	55.2	43.1	80.7	40.5	-3.0
4h	19.3	19.7	7.9	33.1	9.0	22.8	30.6	27.1	11.2
4i	43.1	8.5	3.4	64.8	39.9	17.7	63.4	57.3	33.2
4j	3.54	0.2	64.7	77.0	66.2	37.3	74.8	62.5	16.8
4k	-7.0	-4.5	-27.5	33.1	23.8	6.1	41.1	9.0	-4.6

Results compared with the blank control group.

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inhibition rate of 91.3%, 94.4%, 94.4% and 93.7% at 50 μ g/mL, respectively. It is worth mentioning that compound **6mg** presented the highest inhibition against EC109 (IC₅₀ = 10.82 μ ^m mol/L), HepG2 (IC₅₀ = 10.44 μ mol/L) and MGC-803 (IC₅₀ = 10.59 μ mol/L) cells.

Analogs 6ma-6mk and 6na-6nk, with different substituents on the phenyl ring, showed various degrees of inhibitory activity against EC109, HepG2 and MGC-803 cells. Compounds 6mb/6nb and **6mc/6nc** with hydrophobic alkyl chains (-CH₃) exerted moderate inhibitory activity except for 6mb. Analogs 6md/6nd and 6me/6ne with strong electron-withdrawing groups -NO₂ as meta-substitutions or para-substitutions showed favorable antibacterial activity, but only 6nd demonstrated potent inhibition against HepG2 cells. Analogs 6mf/6nf (3-Cl), 6mg/6ng (4-Cl), 6mh/6nh (3-Br) and 6mi/6ni (4-Br) showed different inhibitory activity against EC109, HepG2 and MGC-803 cells. The antitumor inhibition of 6mf/ 6nf (3-Cl) and 6mg/6ng (4-Cl) with the substituents at meta- or para-position were better than 6mh/6nh (3-Br) and 6mi/6ni (4-Br), respectively. No activity improvement was exhibited by 6mj/6nj (4-OCH₃) and **6mk/6nk** (2,4-dichloride) compared to **6ma**(H). The results indicate that different substituents at different positions of the benzene ring significantly affect the antitumor activity of these compounds and the introduction of the electron-withdrawing groups to the benzene ring led to an increase of the antitumor activity. As to compound 6mg, it is possible that the electronic and steric effects of chlorine atoms were more complementary to the receptors. Further studies will focus on structural optimization and structure-activity relationships of these compounds.

4. Conclusion

In summary, two series of novel α -aminophosphonate derivatives containing thieno[2,3-d]pyrimidines were synthesized and their antitumor activity *in vitro* against EC109, HepG2 and MGC-803 cells has been evaluated. Biological assays revealed that the substitutions on the phenyl ring influenced the antitumor activity remarkably, while the substitutes of phosphonate had no apparent influence on the antitumor activity. Analogs **6mf/6nf** (3-Cl) **and 6mg/6ng** (4-Cl) with the substituents at *meta*- or *para*-position showed better inhibition against all of the tested tumor cells. Amongst them, compound **6mg** demonstrated excellent inhibitory effect against HepG2 (91.2%) and MGC-803(94.4%) cells. These results are useful for the design of more potent novel antitumor compounds and further study are ongoing.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.cclet.2015.03.026.

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