

## D-2-Deoxy-2-fluoro-*chiro*-inositol—a new member of the deoxy fluoro inositol family

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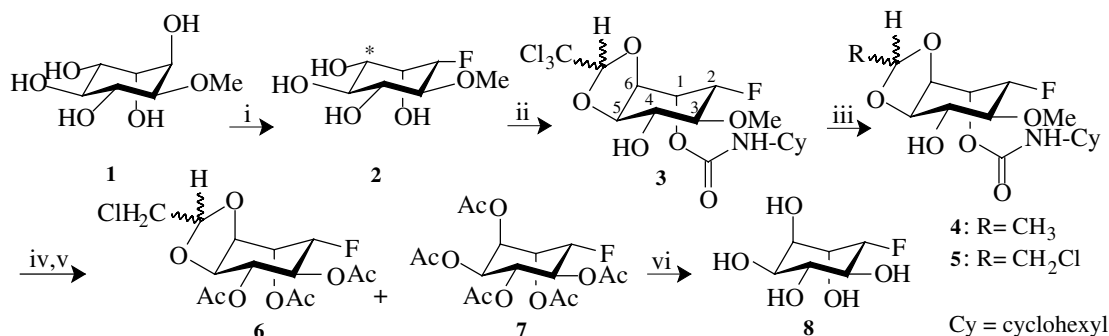
Dedicated to Professor Klaus Peseke on the occasion of his 65th birthday

**Abstract**—An efficient synthetic route to D-2-deoxy-2-fluoro-*chiro*-inositol has been developed with inversions of the C-1 and C-5 configuration of L-quebrachitol. The key steps of the route are two consecutive one pot epimerization procedures which do not require time-consuming protecting groups chemistry.

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Biological activity and/or transport properties of organic compounds are influenced by fluorine and fluorinated moieties, respectively.<sup>1,2</sup> A number of diastereomeric deoxyfluoro inositols have been synthesized and evaluated as probes of the phosphatidylinositol pathway, most notably 5-deoxy-5-fluoro-*myo*-inositol, which is incorporated into the pathway at about 25% the level of *myo*-inositol itself.<sup>3</sup> The set of the six regioisomeric monodeoxy fluoro analogues of *myo*-inositol<sup>3–14</sup> as well as L-3-deoxy-3-fluoro-*chiro*-inositol,<sup>15</sup> 1-deoxy-1-fluoro-*scyllo*-,<sup>9,16,17</sup> 2-deoxy-2-fluoro-*scyllo*<sup>14</sup> and 2-deoxy-2-

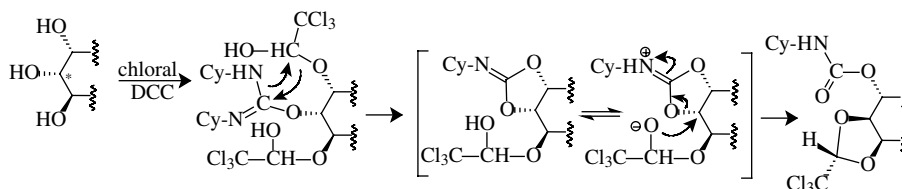
fluoro-*neo*-inositol<sup>3</sup> were synthesized so far. In this letter we report about an access to D-2-deoxy-2-fluoro-*chiro*-inositol (**8**) being a new member of the family of monofluoroinositols. The compound was synthesized from L-quebrachitol (**1**) by combination of efficient reaction steps as shown in Scheme 1. The essential points of the synthetic strategy are two epimerization procedures which evade a time-consuming protecting groups chemistry. The first regioselective epimerization occurs at fluorination of L-quebrachitol with diethylaminosulfur trifluoride (DAST) in dichloromethane.<sup>12,18</sup> This



**Scheme 1.** Synthesis of D-2-deoxy-2-fluoro-*chiro*-inositol (**8**) via non-classical epimerization of L-1-deoxy-1-fluoro-6-O-methyl-*myo*-inositol (**2**).<sup>7,12,18</sup> Reagents and conditions: (i) DAST, CH<sub>2</sub>Cl<sub>2</sub>, rt, 4 h;<sup>18</sup> (ii) chloral, DCC, (CH<sub>2</sub>Cl)<sub>2</sub>, reflux, 6 h; (iii) Raney-Nickel/H<sub>2</sub>, EtOH, rt, 16 h; (iv) 57% hydriodic acid, reflux, 2 h; (v) Ac<sub>2</sub>O, pyridine, rt, 12 h; (vi) CsF/Al<sub>2</sub>O<sub>3</sub>, MeOH, rt, 10–12 h.<sup>23</sup>

**Keywords:** *chiro*-Inositols; L-Quebrachitol; Epimerization; Fluorination; Deoxy fluoro inositol.

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**Scheme 2.** Key steps of the non-classical epimerization of cyclic triols with *cis-trans* sequence of three vicinal OH-groups; Cy = cyclohexyl.

fluorination was originally reported by Kozikowski et al.<sup>7</sup> in 1989, first without use of any solvent and without characterization of **2**. However, the Kozikowski group modified later the protocol<sup>12</sup> and noted that *myo*-inositol derivative **2** is the sole product of the fluorination of L-quebrachitol with DAST. The structure of L-1-deoxy-1-fluoro-6-*O*-methyl-*myo*-inositol (**2**)<sup>19</sup> was meanwhile also confirmed by X-ray analysis.<sup>18</sup>

L-1-Deoxy-1-fluoro-6-*O*-methyl-*myo*-inositol (**2**) meets the requirements for a non-classical acetalization/epimerization reaction with chloral/DCC,<sup>20</sup> because it shows a vicinal triol unit with *cis-trans* sequence (Scheme 1). The course of this epimerization reaction is simplistically shown in Scheme 2; for detailed explanations see review.<sup>20</sup>

Refluxing of **2** with chloral/DCC in 1,2-dichloroethane for 6–8 h gave D-1-*O*-cyclohexylcarbamoyl-2-deoxy-2-fluoro-3-*O*-methyl-5,6-*O*-(*R/S*)-2,2,2-trichloroethylidene]-*chiro*-inositol (**3**). Consequently, the configuration at the marked C-atom of precursor **2** was inverted (Schemes 1 and 2). The yield of **3** was 61%.<sup>21</sup> Note that the numbering of the C-atoms is to change when a *myo*-inositol derivative was transformed into a D-*chiro*-inositol derivative (IUPAC rules for inositols<sup>22</sup>).

The next synthetic step was the hydrodechlorination of the acid-stable chloral acetal unit with H<sub>2</sub>/Raney nickel in ethanol. It resulted a 1.3:1 mixture of ethylidene derivative **4** and chloroethylidene derivative **5**.<sup>23</sup> After refluxing of this mixture in 57% hydriodic acid, the intensive dark-colored crude product obtained was peracetylated before it was column chromatographically prepurified. The main fraction contained both, chloroethylidene derivative **6** and pentaacetyl derivative **7**. This mixture was treated with alumina supported caesium fluoride in methanolic suspension at room temperature overnight.<sup>23,24</sup> Target compound **8** was obtained in a yield of 75% related to **3**.

The structures of the new products are supported by LC–MS data and NMR spectra; the NMR data of the compounds **4**, **5**, **7** and **8** are given in Ref. 23. Moreover, an X-ray analysis confirms configuration and conformation of target compound **8**.

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23. D-2-Deoxy-2-fluoro-*chiro*-inositol (**8**): (1): To a stirred solution of **3**<sup>18,21</sup> (304 mg, 0.67 mmol) in EtOH (10 mL) Raney-Ni (1.0 g) was added (H<sub>2</sub>-atmosphere). After the suspension was stirred for 8 h at rt, triethylamine (1 mL) was added and stirring was continued for 8 h. Then the mixture was filtered through Kieselguhr, and the filtrate was concentrated under reduced pressure. After column chromatographic purification (1:1 heptane/EtOAc), 208 mg of a crystalline 1.3:1 mixture of **4** and **5** was obtained.  
**4** (*endo*-H form): <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 5.29 (q, 1H, *J* ≈ 5.0 Hz, CHCH<sub>3</sub>), 5.29 (m, 1H, H-1), 4.80 (ddd, 1H, *J*<sub>2,F</sub> ≈ 48.2, *J*<sub>1,2</sub> ≈ 3.0, *J*<sub>2,3</sub> ≈ 6.0 Hz, H-2), 4.72

(d, 1H,  $J \approx 8.0$  Hz, NH), 4.40–4.26 (m, 2H, H-5, H-6), 3.69 (t, 1H,  $J_{3,4} \approx J_{4,5} \approx 8.5$  Hz, H-4), 3.56 (s, 3H, OCH<sub>3</sub>), 3.55–3.40 (m, 2H, H-3, cyclohexyl-CH), 2.68 (br, 1H, OH), 1.96–1.11 (m, 10H, cyclohexyl-CH<sub>2</sub>), 1.36 (d, 3H,  $J \approx 5.0$  Hz, CH<sub>3</sub>CH); <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  154.1 (C(O)NH), 100.8 (CHCH<sub>3</sub>), 90.9 (d,  $J_{C,F} \approx 182.2$  Hz, C-2), 81.6 (d,  $J_{C,F} \approx 23.8$  Hz, C-3), 78.2 (C-5), 75.1 (d,  $J_{C,F} \approx 6.5$  Hz, C-6), 71.4 (d,  $J_{C,F} \approx 7.5$  Hz, C-4), 69.5 (d,  $J_{C,F} \approx 17.5$  Hz, C-1), 59.6 (OCH<sub>3</sub>), 50.3 (cyclohexyl-CH), 33.2 (2 $\times$ ), 25.4, 24.7 (2 $\times$ ) (cyclohexyl-CH<sub>2</sub>), 20.1 (CHCH<sub>3</sub>); <sup>19</sup>F{<sup>1</sup>H} NMR (235 MHz, CDCl<sub>3</sub>):  $\delta$  –199.9 (s). LC–MS (M+H):  $m/z$  = 348.

**5** (*endo*-H form): <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  5.38 (t, 1H,  $J \approx 4.0$  Hz, ClCH<sub>2</sub>CH), 5.18 (ddd, 1H,  $J_{1,F} \approx 22.5$ ,  $J_{1,2} \approx 3.0$ ,  $J_{1,6} \approx 6.3$  Hz, H-1), 4.81 (ddd, 1H,  $J_{2,F} \approx 48.5$ ,  $J_{1,2} \approx 3.0$ ,  $J_{2,3} \approx 5.0$  Hz, H-2), 4.72 (d, 1H,  $J \approx 8.0$  Hz, NH), 4.40–4.26 (m, 2H, H-5, H-6), 3.73 (t, 1H,  $J_{3,4} \approx J_{4,5} \approx 8.5$  Hz, H-4), 3.54 (s, 3H, OCH<sub>3</sub>), 3.55–3.40 (m, 4H, H-3, cyclohexyl-CH, CHCH<sub>2</sub>Cl), 2.68 (br, 1H, OH), 1.96–1.11 (m, 10H, cyclohexyl-CH<sub>2</sub>); <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  153.9 (C(O)NH), 102.1 (CHCH<sub>2</sub>Cl), 90.8 (d,  $J_{C,F} \approx 183.4$  Hz, C-2), 81.0 (d,  $J_{C,F} \approx 21.2$  Hz, C-3), 79.0 (C-5), 74.0 (d,  $J_{C,F} \approx 6.5$  Hz, C-6), 70.7 (d,  $J_{C,F} \approx 7.5$  Hz, C-4), 69.5 (d,  $J_{C,F} \approx 17.5$  Hz, C-1), 59.2 (OCH<sub>3</sub>), 50.2 (cyclohexyl-CH), 44.6 (CH<sub>2</sub>Cl), 33.2 (2 $\times$ ), 25.4, 24.7 (2 $\times$ ) (5C, cyclohexyl-CH<sub>2</sub>); <sup>19</sup>F{<sup>1</sup>H} NMR (235 MHz, CDCl<sub>3</sub>):  $\delta$  –202.4 (s). LC–MS (M+H):  $m/z$  = 382.

(2): The 1.3:1 mixture of **4** and **5** (208 mg) was refluxed in 2 mL of 57% aqueous HI for 2 h. After concentration of the solution under reduced pressure, the residue was peracetylated with acetic acid anhydride/pyridine at rt (8 h). Then it was again concentrated. Compounds **6** ( $R_f$  = 0.27, 2:1 heptane/EtOAc) and **7** ( $R_f$  = 0.37, 2:1 heptane/EtOAc) were column chromatographically separated as mixed fraction (200 mg); LC–MS (**6**: [M+H]:  $m/z$  393; **7**: [M]:  $m/z$  368.4). **7**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$

5.52–5.46 (m, 2H, H-3, H-1), 5.41 (‘q’,  $J_{6,F} \approx J_{1,6} \approx J_{5,6} \approx 3.5$  Hz, 1H, H-6), 5.36 (t, 1H,  $J_{4,F} \approx J_{3,4} \approx J_{4,5} \approx 10.2$  Hz, H-4), 5.21 (dd, 1H,  $J_{4,5} \approx 10.2$ ,  $J_{5,6} \approx 3.5$  Hz, H-5), 4.97 (ddd, 1H,  $J_{2,F} \approx 46.4$ ,  $J_{1,2} \approx 3.5$ ,  $J_{2,3} \approx 9.8$  Hz, H-2), 2.18, 2.15, 2.06, 2.01, 1.95 (s, 15H, 5 $\times$ CH<sub>3</sub>). <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta$  171.4, 171.4, 171.4, 170.7, 170.5 (5 C=O), 88.9 (d,  $J_{C,F} \approx 188.6$  Hz, C-2), 71.7 (d,  $J_{C,F} \approx 21.0$  Hz, C-3), 70.4 (s, C-5), 70.4 (d,  $J_{C,F} \approx 10.2$  Hz, C-4), 68.8 (d,  $J_{C,F} \approx 17.7$  Hz, C-1), 68.1 (d,  $J_{C,F} \approx 6.7$  Hz, C-6), 20.5 (CH<sub>3</sub>); <sup>19</sup>F{<sup>1</sup>H} NMR (235 MHz, CDCl<sub>3</sub>):  $\delta$  –207.5. Anal. Calcd for C<sub>16</sub>H<sub>21</sub>FO<sub>10</sub> (392.34): C, 48.98; H, 5.40. Found: C, 49.23; H, 5.57.

(3): To a stirred suspension of alumina supported caesium fluoride<sup>24</sup> (150 mg,  $c$  = 1.533 mmol CsF/g support, 1/15 mole equiv fluoride per acetyl group) in MeOH (10 mL), the mixture of **6** and **7** (200 mg) was added and stirring was continued at rt overnight. After filtration and evaporation of the solvent, the residue was column chromatographically (CHCl<sub>3</sub>/MeOH 20:1  $\rightarrow$  5:1) purified. 92 mg (75%) of **8** related to **3** were isolated. The colourless crystals were recrystallized from acetone/MeOH (10:1),  $R_f$  0.48 (CHCl<sub>3</sub>/MeOH 1:1);  $[\alpha]_D^{21}$  +46.2 ( $c$  0.66, MeOH); LC–MS [M]:  $m/z$  182. On heating compound **8** loses from 163–164 °C and up water by polycondensation; the polycondensate melts finally at 212–213 °C. <sup>1</sup>H NMR (250 MHz, CD<sub>3</sub>OD):  $\delta$  4.52 (ddd, 1H,  $J_{2,F} \approx 48.8$ ,  $J_{1,2} \approx 3.3$ ,  $J_{2,3} \approx 9.5$  Hz, H-2), 4.11 (ddd, 1H,  $J_{1,F} \approx 6.9$ ,  $J_{1,6} \approx 4.0$  Hz, H-1), 3.93 (ddd, 1H,  $J_{6,F} \approx 4.4$ ,  $J_{5,6} \approx 3.3$  Hz, H-6), 3.82 (ddd, 1H,  $J_{3,F} \approx 12.3$ ,  $J_{3,4} \approx 9.4$  Hz, H-3), 3.71 (dd, 1H,  $J_{4,5} \approx 9.7$  Hz, H-5), 3.55 (dd, 1H, H-4); <sup>13</sup>C NMR (62.9 MHz, CD<sub>3</sub>OD):  $\delta$  94.2 (d,  $J_{C,F} \approx 179.0$  Hz, C-2), 74.2 (d,  $J_{C,F} \approx 11.0$  Hz, C-4), 73.4 (d,  $J_{C,F} \approx 8.4$  Hz, C-6), 72.8 (d,  $J_{C,F} \approx 18.2$  Hz, C-3), 72.2 (s, C-5), 71.6 (d,  $J_{C,F} \approx 17.1$  Hz, C-1); <sup>19</sup>F{<sup>1</sup>H} NMR (235 MHz, CD<sub>3</sub>OD):  $\delta$  –206.5.

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