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Synthesis of Alkenyldiarylmethane (ADAM) Non-Nucleoside HIV-1 Reverse Transcriptase Inhibitors with Non-Identical Aromatic Rings

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Abstract—The existing methods for the synthesis of alkenyldiarylmethane (ADAM) non-nucleoside reverse transcriptase inhibitors proceed from symmetrical benzophenones and therefore result in products with identical aromatic rings. New methods have therefore been devised for the preparation of stereochemically defined ADAMs with non-identical aromatic rings. The new routes rely on palladium-catalyzed reactions, including Sonogashira, Suzuki, Stille, and hydroarylation methodology. Several of the new ADAMs inhibited the cytopathic effect of HIV-1 in cell culture and HIV-1 reverse transcriptase at submicromolar concentrations. © 2001 Elsevier Science Ltd. All rights reserved.

The non-nucleoside reverse transcriptase inhibitors (NNRTIs) are a structurally diverse set of compounds that act by an allosteric mechanism.¹ A number of NNRTIs, including nevirapine, delavirdine, and efavirenz, are used in combination chemotherapy for the treatment of AIDS.^{2–4} However, drug incompatibilities, adverse effects, the emergence of resistant viral strains, and cross-resistance continue to limit the clinical usefulness of the NNRTIS.^{5–9} Additional NNRTIS are therefore needed that might have improved pharmacokinetics, limited toxicities, and more favorable resistance mutation profiles. The alkenyldiarylmethanes (ADAMs) have recently been identified as a new class of NNRTIs.^{10–13} Several of the ADAMs, including compounds 1-3, are potent inhibitors of the cytopathic effect of HIV-1 in cell culture, with EC₅₀ values in the low nanomolar range. A number of HIV-1 strains containing AZT resistance mutations have shown increased sensitivity to some of the ADAMs, indicating a possible therapeutic role for the ADAMs in combination with AZT.¹²

The existing ADAM syntheses include the attachment of the alkenyl chain to symmetrical benzophenone deriva-tives by Wittig,^{10–12,14} McMurry,^{13,15} or Horner– Emmons¹² reactions. These methods are not ideal for the synthesis of ADAMs having differently substituted aromatic rings because they would lead to mixtures of cis and *trans* isomers when employed with benzophenones having non-identical aromatic rings. In addition, benzophenones with different substituents on each aromatic ring are not as readily available as those with identical substituents. Since the two aromatic rings of the ADAMs are not in identical environments when bound in the NNRTI binding pocket of HIV-1 reverse transcriptase,^{12,13} it is logical to conclude that ADAMs with identical aromatic substituents are not the ideal choice to 'fit' the binding site. Consequently, a need does exist for ADAMs with non-identical aromatic rings, and we have therefore recently investigated ADAM syntheses that do not proceed from benzophenones, including the use of palladium-catalyzed reactions in the solid phase synthesis of ADAMs 4 and 5.¹⁶

The present communication details the use of the Sonogashira, Suzuki, Stille, and hydroarylation reactions in stereochemically defined, solution phase syntheses of new ADAMs 6–8 with differently substituted

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aromatic rings. These compounds were chosen to explore the importance of the aromatic methyl ester groups and halogen atoms for biological activity. In addition, the truncated compounds 10 and 11, in which one of the aromatic rings of 3 has been eliminated, were prepared. The corresponding reduction product 12, as well as the oxidized compound 13, were also synthesized. The investigation of the biological activities of these compounds should be helpful in the identification of the ADAM 'pharmacophore'.



Results and Discussion

3-Methylsalicylic acid (14) was quantitatively converted into its methyl ester using $TMSCHN_2$ in a mixture of



Scheme 1. Reagents and conditions: (a) (i) TMSCHN₂, MeOH, benzene, 23 °C (2 h); (ii) NaI, NaOH, MeOH, then NaOCl (5.25%), 0– 3 °C (3 h); (b) Me₂SO₄, K₂CO₃, acetone, reflux (24 h); (c) methyl 5hexynoate, Pd(OAc)₂, PPh₃, CuI, triethylamine, EtOAc, 23 °C (12 h); (d) Bu₃SnH, Pd(PPh₃)₄, THF, 23 °C (1 h); (e) I₂, CH₂Cl₂, 23 °C (50 min); (f) 3,4-dimethoxyphenylboronic acid, Na₂CO₃, Pd(PPh₃)₄, CH₃CN/H₂O, 90 °C (19 h).

methanol and benzene.¹⁷ The resulting methyl ester was iodinated using sodium iodide in the presence of sodium hypochlorite as an oxidant to afford intermediate 15 (Scheme 1).¹⁸ Treatment of the iodinated ester 15 with dimethyl sulfate and K₂CO₃ produced methyl 5-iodo-3methyl-2-methoxybenzoate 16 in 90% overall yield. Palladium-catalyzed Sonogashira coupling of methyl 5hexynoate with iodinated ester 16 afforded the alkyne 17,^{19,20} which underwent hydrostannation in the presence of Pd(PPh₃)₄ to give the regiochemically and stereochemically defined vinylstannane 18 in 39% overall yield.²¹ Subsequent treatment of 18 with I₂ in CH₂Cl₂ produced vinyl iodide 19.22 The Suzuki coupling of 19 with 3,4-dimethoxyphenylboronic acid in the presence of Pd(PPh₃)₄ and Na₂CO₃ afforded the desired ADAM 6 in 53% yield.^{23,24}

Palladium-catalyzed Sonogashira coupling of 2-iodopyrazine (**20**) with methyl 5-hexynoate afforded alkyne **21** in 81% yield (Scheme 2). Subsequent hydrostannation and I₂ displacement of the tributyltin group produced vinyl iodide **23** in 76% yield. Coupling of **23** with 3,4-dimethoxyphenylboronic acid under Suzuki conditions afforded ADAM **7** in 52% yield.

As shown in Scheme 3, treatment of 5-iodosalicylic acid 24 with dimethyl sulfate and K_2CO_3 produced 25, which coupled with methyl 5-hexynoate under Sonogashira reaction conditions to afford alkyne 26. Palladium-catalyzed hydroarylation of alkyne 26 served as a key step to construct the second aromatic ring in compound 8.²⁵

Since the addition can happen at both positions of the carbon–carbon triple bond, two structural isomers (8 and 27) are formed in almost equal yield. Careful preparative TLC isolation afforded pure ADAM 8 in 31% yield, and well as pure 27 in 30% yield.

The same strategy was employed to construct ADAM 9 as outlined in Scheme 4. 2-Chlorophenol was iodinated using sodium iodide in the presence of sodium hypochlorite as an oxidant. Treatment of the iodinated phenol 29 with dimethyl sulfate and K_2CO_3 produced 30. Sonogashira coupling of iodo compound 30 with methyl 5-hexynoate gave alkyne 31. Palladium-catalyzed hydroarylation of alkyne 31 gave a mixture of compounds 9 and 32. Purification of both 9 and 32 was achieved by preparative TLC.

The syntheses of compounds **10–13** are illustrated in Scheme 5. Hydrogenation of **17** using Lindlar's catalyst gave the *cis*-alkene **10**. Palladium-catalyzed Heck coupling of methyl 5-hexenoate with **16** produced the *trans*-alkene **11**. Additional analogues (**12–13**) were also made



Scheme 2. Reagents and conditions: (a) methyl 5-hexynoate, Pd(OAc)₂, PPh₃, CuI, triethylamine/EtOAc, $23 \degree C$ (12 h); (b) Bu₃SnH, Pd(PPh₃)₄/THF, $23 \degree C$ (1 h); (c) I₂/CH₂Cl₂, $23 \degree C$ (30 min); (d) 3,4-dimethoxyphenylboronic acid, Na₂CO₃, Pd(PPh₃)₄, CH₃CN/H₂O, 90 °C (14 h).



Scheme 3. Reagents and conditions: (a) $(CH_3)_2SO_4$, K_2CO_3 , acetone, reflux (14 h); (b) methyl 5-hexynoate, Pd(OAc)₂, PPh₃, CuI, TEA, EtOAc, 23 °C (12 h); (c) Compound 25, Pd₂(dba)₃, Et₂NH, HCOOH, EtOAc, reflux (12 h).

through complete hydrogenation of 17 or by heating 17 in refluxing formic acid.²⁶

The new ADAMs synthesized in this study were evaluated for inhibition of the cytopathic effect of HIV- 1_{RF} in CEM-SS cells, and the resulting EC₅₀ values are listed in Table 1. The cytotoxic concentrations in uninfected CEM-SS cells (CC₅₀ values) were determined as well. These compounds were also tested for inhibition of HIV-1 RT, and the resulting IC₅₀ values are also included in Table 1. Two of the new ADAMs (6 and 8) inhibited the cytopathic effect of HIV- 1_{RF} with EC₅₀ values of 0.21 and 0.47 μ M, respectively. These analogues were also found to inhibit HIV-1 RT with $poly(rC) \cdot oligo(dG)$ as the template primer with IC₅₀ values of 0.074 and 0.504 µM. Compound 7, with a heterocyclic ring, was inactive. The esters on the aromatic ring seem to be essential for the anti-HIV activity (ADAM 9 was inactive, vs ADAM 2 with EC_{50} of 13 nM). However, the halogen atoms on the aromatic ring



Scheme 4. Reagents and conditions: (a) NaI, NaOH, NaOCl/MeOH, $23 \degree C (2 h)$; (b) (CH₃)₂SO₄, K₂CO₃, acetone, reflux (14 h); (c) methyl 5hexynoate, Pd(OAc)₂, PPh₃, CuI, TEA, EtOAc, $23 \degree C (12 h)$; (d) Compound 30, Pd₂(dba)₃, Et₂NH, HCOOH, EtOAc, reflux (12 h).



Scheme 5. Reagents and conditions: (a) Methyl 5-hexenoate, $Pd(OAc)_2$, TEA, EtOAc, 70 °C (18 h); (b) Methyl 5-hexynoate, $Pd(OAc)_2$, PPh₃, CuI, TEA, EtOAc, 23 °C (12 h); (c) H₂, Pd (5%) on BaSO₄, quinoline, 23 °C (18 h); (d) H₂, Pd (5%) on activated carbon (14 h); (e) HCOOH, reflux (2–4 h).

significantly increase the activity (ADAM 2, $EC_{50} = 13$ nM; ADAM 8, $EC_{50} = 470$ nM). The large difference in lipophilicity between ADAM 2 and ADAM 8 due to the incorporation of halogen atoms on the aromatic ring may account for the lower activity of ADAM 8. Both *cis*- and *trans*-alkenes 10 and 11 were inactive against HIV-1 viruses and HIV-1 RT. This suggests that both aromatic rings are required for targeting the hydrophobic pocket of the RT enzyme, and is also consistent with the 'butterfly model' proposed for the binding of NNRTIs to HIV-1 RT.^{10,12,13,27}

It is apparent from the data in Table 1 that there is not a perfect correlation between the anti-HIV-1 EC₅₀ values and the reverse transcriptase IC₅₀ values. For example, compound 6 is one of the most potent reverse transcriptase inhibitors ever synthesized in the ADAM series (IC₅₀ 0.074 μ M), but it is about three fold less potent as an inhibitor of the cytopathic effect of HIV-1 in cell culture (EC₅₀ 0.21 μ M). In contrast, ADAM 1 is less potent than 6 versus RT in the cell-free assay system (IC₅₀ 0.3 μ M), but it is considerably more potent as an antiviral agent (EC₅₀ 0.0013 µM). However, this difference is not unusual for the non-nucleoside reverse transcriptase inhibitors.²⁸⁻³¹ The reverse transcriptase inhibition studies were performed in a cell-free system with a synthetic template/primer, poly(rC).oligo(dG). As discussed elsewhere, the discrepancy in EC₅₀ values for inhibition of the cytopathic effect of the virus and the IC₅₀ values for reverse transcriptase inhibition may simply reflect the differences between the in vitro assay, in which synthetic template/primer has been added, and the cellular system.³⁰ In spite of the lack of close correlation of reverse transcriptase inhibitory activity with antiviral activity in the ADAM series, prior work with ADAM-resistant HIV-1 strains having mutations in reverse transcriptase leaves little doubt that in general,

Table 1. Anti-HIV activities of the ADAMs

Compd	RT (IC50, µM) ^a	$EC_{50}~(\mu M)^b$	CC ₅₀ (µM) ^c	TI ^d
1 ¹³	0.3	0.0013	13	10,000
2 ¹²	0.3	0.013	31.6	2431
3 ³³	1.0	0.25	6.0	24
4 ¹⁶	0.516	0.020	1.46	73
5 ¹⁶	0.587	0.080	2.17	27
6	0.074	0.21	1.14	5.48
7	100	NA ^e	19.40	
8	0.504	0.47	2.0	4.27
9	100	NA	≥ 6.25	
10	88.2	NA	157.0	
11	100	NA	1.70	
12	1.75	NA	108	
13	100	NA	200	
17	100	NA	62.50	
32	100	NA	≥ 6.25	

^aInhibitory activity versus HIV-1 reverse transcriptase with rCdG as the template primer.

 $^b The \ EC_{50}$ is the 50% inhibitory concentration for cytopathicity of $HIV\text{-}1_{RF}$ in CEM-SS cells.

 $^{\circ}\text{The}$ CC $_{50}$ is the 50% cytotoxic concentration for mock-infected CEM-SS cells.

 $^d The TI$ is the therapeutical index, which is the CC_{50} divided by the $EC_{50}.$

^eNA (not active): no observed inhibition of HIV-1 cytopathicity up to the cytotoxic concentration in uninfected cells.

the ADAMs are in fact acting as non-nucleoside reverse transcriptase inhibitors.^{10,12,13}

The methyl ester groups present in the most potent ADAMs are likely to be substrates for a variety of esterases present in the body. Future work in the ADAM series should therefore focus on the exchange of these ester groups with metabolically stable replacements that will retain the anti-HIV activity.

Experimental

Melting points were obtained in capillary tubes and are uncorrected. ¹H NMR spectra were determined at 300 MHz. Chemical ionization mass spectra (CIMS) were run using isobutane as the reagent gas. Microanalyses were performed at the Purdue University Microanalysis Laboratory. Silica gel flash chromatography was accomplished using 230–400 mesh silica gel. Unless otherwise stated, chemicals and solvents were reagent grade and used as obtained from commercial sources without further purification. Compounds **15** and **16** were synthesized as described elsewhere.¹⁶

Methyl 6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyllhex-5-ynoate (17). Methyl 5-hexynoate (0.593 g, methyl 5-iodo-3-methyl-2-methoxy-4.71 mmol), benzoate 16 (1.2 g, 3.92 mmol), Pd(OAc)₂ (62.4 mg, 0.278 mmol), Ph₃P (145.8 mg, 0.556 mmol), copper(I) iodide (0.15 g, 0.788 mmol), and triethylamine (1.14 g, 11.25 mmol) were stirred in ethyl acetate (30 mL) at room temperature overnight under argon. The mixture was diluted with water (15 mL), and the layers were separated. The organic layer was washed with water and brine, dried over sodium sulfate, and concentrated to give a black residue, which was purified by flash chromatography on silica gel (120 g, column: 5×30 cm, hexanes-EtOAc 3:1) to afford 17 as a brown liquid (488 mg, 41%): ¹H NMR (CDCl₃) δ 7.67 (d, J=2.08 Hz, 1H), 7.36 (d, J = 1.69 Hz, 1H), 3.90 (s, 3H), 3.81 (s, 3H), 3.68 (s, 3H), 2.50 (t, J = 6.20 Hz, 2H), 2.46 (t, J = 5.65Hz, 2H), 2.27 (s, 3H), 1.91 (m, 2H); ¹³C NMR (CDCl₃) δ 173.51, 166.11, 157.81, 137.71, 132.82, 132.26, 124.38, 118.98, 88.64, 80.07, 61.46, 52.12, 51.49, 32.74, 23.72, 18.68, 15.78; EIMS m/z 304 (M⁺); HRMS calcd for C₁₇H₂₀O₅ 304.1311, found 304.1318. Anal. calcd for C₁₇H₂₀O₅: C, 67.07; H, 6.58. Found: C, 66.95; H, 6.47.

Methyl (5E)-6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyl]-6-(tributylstannyl)hex-5-enoate (18). To a stirred solution of alkyne 17 (67 mg, 0.22 mmol) in dry THF (20 mL) at room temperature was added (Ph₃P)₄Pd (7.6 mg, 0.007 mmol) and, dropwise, Bu₃SnH (96.2 mg, 0.33 mmol). The reaction mixture was kept stirring for 1 h under argon. The solvent was evaporated and the brown residue was purified by flash chromatography on silica gel (30 g, hexanes–EtOAc 3:1, v/v) to afford a colorless oil (116 mg, 94%): ¹H NMR (CDCl₃) δ 7.16 (d, J=2.25 Hz, 1H), 6.86 (d, J=2.15 Hz, 1H), 5.70 (t, J=6.92 Hz, J_{SnH} =31.24 Hz, 1H), 3.88 (s, 3H), 3.80 (s, 3H), 3.60 (s, 3H), 2.27 (s, 3H), 2.23 (t, J=7.66 Hz, 2H), 2.01 (dt, J=7.31 and 6.90 Hz, 2H), 1.65 (m, 2H), 1.18–1.44 (m, 18H), 0.83 (t, J=7.25 Hz, 9H); ESIMS m/z 595 (MH⁺). Anal. calcd for C₂₉H₄₈SnO₅: C, 58.59; H, 8.08. Found: C, 58.47; H, 7.96.

Methyl (5E)-6-iodo-6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyllhex-5-enoate (19). The vinyl tributylstannane 18 (80 mg, 0.13 mmol) was dissolved in dry CH_2Cl_2 (5 mL). Finely divided I_2 (49.3 mg, 0.19 mmol) was added and the mixture was stirred vigorously at room temperature for 50 min. Saturated aqueous $Na_2S_2O_3$ (5 mL) was added, the phases were separated, and the aqueous phase was extracted with CH₂Cl₂ $(3 \times 10 \text{ mL})$. The combined organic extracts were dried (Na_2SO_4) and concentrated. The residue was purified by flash chromatography on silica gel (30 g, hexanes-EtOAc 3:1, v/v) to afford a colorless oil (50.5 mg, 90%): ¹H NMR (CDCl₃) δ 7.51 (d, J=2.01 Hz, 1H), 7.24 (d, J = 2.0 Hz, 1H), 6.44 (t, J = 7.41 Hz, 1H), 3.91 (s, 3H), 3.83 (s, 3H), 3.62 (s, 3H), 2.30 (s, 3H), 2.26 (t, J = 7.43Hz, 2H), 2.01 (dt, J=7.41 and 7.52 Hz, 2H), 1.70 (m, 2H); CIMS m/z 433 (MH⁺). Anal. calcd for C₁₇H₂₁IO₅: C, 47.22; H, 4.86. Found: C, 47.21; H, 4.70.

(5Z)-6-(3,4-dimethoxyphenyl)-6-[4-methoxy-3-Methyl (methoxycarbonyl)-5-methylphenyl]hex-5-enoate (6). The vinyl iodide 19 (44.7 mg, 0.103 mmol) and 3,4-dimethoxyphenylboronic acid (18.8 mg, 0.103 mmol) were added to a solution of sodium carbonate (21.8 mg, 0.21 mmol) in water (1 mL) and CH₃CN (2 mL). To the above degassed solution was added Pd(PPh₃)₄ (6.0 mg, 0.005 mmol). The resulting mixture was heated at 90 °C (oil bath) under argon for 14 h. The reaction mixture was concentrated and subjected to flash column chromatography on silica gel (30 g, hexanes–EtOAc 3:1, v/v) to afford a light yellow oil (24 mg, 52.7%): ¹H NMR $(CDCl_3) \delta 7.43 (d, J = 1.60 Hz, 1H), 7.12 (d, J = 1.60 Hz, 1H)$ 1H), 6.79 (d, J = 1.60 Hz, 1H), 6.75 (d, J = 6.57 Hz, 1H), 6.63 (dd, J=6.57 and 1.60 Hz, 1H), 5.94 (t, J=7.46 Hz, 1H), 3.89 (s, 3H), 3.87 (s, 3H), 3.86 (s, 3H), 3.83 (s, 3H), 3.63 (s, 3H), 2.31 (s, 3H), 2.30 (t, J = 7.30 Hz, 2H), 2.10 (dt, J = 7.52 and 7.34 Hz, 2H), 1.77 (m, 2H); EIMS m/z442 (M⁺). Anal. calcd for $C_{25}H_{30}O_7$: C, 67.87; H, 6.79. Found: C, 67.85; H, 6.70.

Methyl 6-pyrazin-2-ylhex-5-ynoate (21). Methyl 5-hexynoate (800 mg, 6.35 mmol), 2-iodopyrazine (872 mg, 4.23 mmol), Pd(OAc)₂ (76 mg, 0.34 mmol), PPh₃ (178 mg, 0.68 mmol), copper(I) iodide (161 mg, 0.85 mmol), and triethylamine (930 mg, 9.2 mmol) were added to ethyl acetate (40 mL). The reaction mixture was stirred at room temperature overnight under argon and then concentrated. The residue was purified by flash chromatography on silica gel (80 g, EtOAc–hexanes 2:3, v/v) to afford **21** as a brown liquid (700 mg, 81.1%): ¹H NMR (CDCl₃) δ 8.61 (br, 1H), 8.50 (d, *J*=2.37 Hz, 1H), 8.43 (d, *J*=2.37 Hz, 1H), 3.68 (s, 3H), 2.56 (t, *J*=6.98 Hz, 2H), 2.52 (t, *J*=7.35 Hz, 2H), 1.96 (m, 2H); CIMS *m*/*z* 204 (M⁺). Anal. calcd for C₁₁H₁₂N₂O₂: C, 64.71; H, 5.88. Found: C, 64.64; H, 5.81.

Methyl (5E)-6-pyrazin-2-yl-6-(tributylstannyl)hex-5-enoate (22). To a stirred solution of alkyne 21 (350 mg, 1.72 mmol) in dry THF (20 mL) at room temperature was added (Ph₃P)₄Pd (99 mg, 0.086 mmol) and, dropwise, Bu₃SnH (749 mg, 2.57 mmol). The reaction mixture was kept stirring for 1 h under argon. The solvent was evaporated and the brown residue was purified by flash chromatography on silica gel (30 g, hexanes– EtOAc 3:1, v/v) to afford a colorless oil (750 mg, 88.3%): ¹H NMR (CDCl₃) δ 8.48 (br, 1H), 8.34 (s, 1H), 8.28 (d, *J*=1.90 Hz, 1H), 5.93 (t, *J*=7.10 Hz, *J*_{SnH}=31.56 Hz, 1H), 3.63 (s, 3H), 2.36 (t, *J*=7.47 Hz, 2H), 2.28 (t, *J*=7.34 Hz, 2H), 1.77 (m, 2H), 1.23–1.47 (m, 12H), 0.90 (t, *J*=7.48 Hz, 6H), 0.84 (t, *J*=7.27 Hz, 9H); EIMS *m*/*z* 436/438 (M⁺ – C₄H₉). Anal. calcd for C₂₃H₄₀N₂SnO₂: C, 55.87; H, 8.10; N, 5.67. Found: C, 55.87; H, 8.17; N, 5.62.

Methyl (5E)-6-iodo-6-pyrazin-2-ylhex-5-enoate (23). The vinyl tributylstannane 22 (600 mg, 1.21 mmol) was dissolved in dry CH_2Cl_2 (10 mL). Finely divided I_2 (463) mg, 1.82 mmol) was added and the mixture was stirred vigorously at room temperature for 30 min. Saturated aqueous $Na_2S_2O_3$ (10 mL) was added, the phases were separated, and the aqueous phase was extracted with CH_2Cl_2 (3×10 mL). The combined organic extracts were dried (Na₂SO₄) and concentrated. The residue was purified by flash chromatography on silica gel (35 g, hexanes–EtOAc 3:1, v/v) to afford a light yellow oil (347) mg, 86.4%): ¹H NMR (CDCl₃) δ 8.64 (s, 1H), 8.58 (br, 1H), 8.44 (br, 1H), 6.76 (t, J = 7.82 Hz, 1H), 3.63 (s, 3H), 2.29 (t, J=7.30 Hz, 2H), 2.18 (dt, J=7.29 and 7.60 Hz, 2H), 1.77 (m, 2H); EIMS *m*/*z* 332 (M⁺). Anal. calcd for C₁₁H₁₃N₂IO₂: C, 39.76; H, 3.92; N, 8.43. Found: C, 39.79; H, 4.00; N, 8.24.

(5E)-6-(3,4-dimethoxyphenyl)-6-pyrazin-2-yl-Methyl hex-5-enoate (7). The vinyl iodide 23 (214 mg, 0.64 mmol) and 3,4-dimethoxyphenylboronic acid (117 mg, 0.64 mmol) were added to sodium carbonate (150 mg, 1.42 mmol) in water (1 mL) and CH₃CN (2 mL). To the above degassed solution was added Pd(PPh₃)₄ (52.1 mg, 0.045 mmol). The resulting mixture was heated at 90 °C (oil bath) under argon for 14 h. The reaction mixture was concentrated and subjected to flash column chromatography on silica gel (80 g, hexanes–EtOAc 1:1, v/v) to afford 7 as a light yellow oil (114 mg, 51.8%): ¹H NMR (CDCl₃) δ 8.64 (s, 1H), 8.49 (br, 2H), 6.78 (d, J = 8.28 Hz, 1H), 6.77 (br, 1H), 6.65 (dd, J = 8.28 and 1.86 Hz, 1H), 6.17 (t, J = 7.25 Hz, 1H), 3.87 (s, 3H), 3.83 (s, 3H), 3.64 (s, 3H), 2.33 (t, J = 7.40 Hz, 2H), 2.22 (dt, J = 7.46 and 7.45 Hz, 2H), 1.83 (m, 2H); EIMS m/z 342 (M^+) . Anal. calcd for $C_{19}H_{22}N_2O_4$: C, 66.67; H, 6.43. Found: C, 66.91; H, 6.68.

Methyl 5-iodo-2-methoxybenzoate (25). 5-Iodosalicylic acid (7.0 g, 26.5 mmol) was dissolved in acetone (40 mL). Anhydrous K₂CO₃ (5.5 g, 39.75 mmol) and dimethylsulfate (3.8 mL, 39.75 mmol) were added. The reaction mixture was heated at reflux for 14 h, and then the inorganic salts were filtered. The solvent was evaporated and the residue was washed with water (3×20 mL). The crude residue was crystallized from acetone to afford 25 as white solid (7.0 g, 90.4%): mp 47–48 °C; ¹H NMR (CDCl₃) δ 8.05 (d, *J*=2.15 Hz, 1H), 7.70 (dd, *J*=8.57 and 2.30 Hz, 1H), 6.73 (d, *J*=8.68 Hz, 1H), 3.88 (s, 6H).

Methyl 6-[4-methoxy-5-methoxycarbonyl)phenyl]hex-5ynoate (26). Methyl 5-hexynoate (441 mg, 3.5 mmol), iodide 25 (984 mg, 3.5 mmol), Pd(OAc)₂ (63 mg, 0.28 mmol), copper(I) iodide (133 mg, 0.70 mmol), and triethylamine (990 mg, 9.8 mmol) were mixed in ethyl acetate (40 mL) and the mixture was kept stirring at room temperature overnight under argon. Then the mixture was concentrated and the residue was purified by flash chromatography on silica gel (80 g, solvent: EtOAchexanes 1:2, v/v) to give 26 as a yellow oil (380 mg, 26%): ¹H NMR (CDCl₃) δ 7.81 (d, J=2.15 Hz, 1H), 7.45 (dd, J=8.57 and 2.30 Hz, 1H), 6.87 (d, J=8.68 Hz, 1H), 3.87 (s, 3H), 3.86 (s, 3H), 3.66 (s, 3H), 2.48 (t, J = 7.40 Hz, 2H), 2.44 (t, J = 6.87 Hz, 2H), 1.90 (m, 2H); ¹³C NMR (CDCl₃) δ 173.47, 165.80, 158.37, 136.33, 134.83, 119.91, 115.63, 111.85, 88.00, 80.01, 55.96, 51.96, 51.46, 32.76, 23.75, 18.68; CIMS m/z 291 $(MH^{+}).$

Hydroarylation of compound 26 to afford 8 and 27. Compound 26 (374 mg, 1.29 mmol), aryl iodide 25 (414 mg, 1.42 mmol), and Pd₂(dba)₃ (83 mg, 0.09 mmol) were dissolved in ethyl acetate (40 mL). Diethylamine (310 mg, 4.26 mmol) was added, followed by formic acid (131 mg, 2.84 mmol), and the solution was heated to reflux overnight. The reaction mixture was then cooled to room temperature and washed with diluted HCl and brine. The organics were dried over Na₂SO₄, and the solvent was removed under reduced pressure. The products 8 and 27 were isolated by flash chromatography (hexane/EtOAc 3:1, v/v; silica gel: 100 g) and preparative TLC (hexanes/EtOAc 5:1, v/v).

Methyl 6,6-bis[4-methoxy-3-(methoxycarbonyl)phenyl]hex-5-enoate (8). Light yellow oil. Yield: 188 mg (32%). ¹H NMR (CDCl₃) δ 7.65 (d, J=2.39 Hz, 1H), 7.56 (d, J=2.22 Hz, 1H), 7.23 (dd, J=2.03 and 2.08 Hz, 1H), 7.18 (d, J=2.49 Hz, 1H), 6.98 (d, J=8.62 Hz, 1H), 6.85 (d, J=8.76 Hz, 1H), 5.96 (t, J=7.45 Hz, 1H), 3.93 (s, 3H), 3.88 (s, 3H), 3.86 (s, 6H), 3.62 (s, 3H), 2.29 (t, J=7.50 Hz, 2H), 2.14 (dt, J=7.32 and 7.47 Hz, 2H), 1.78 (m, 2H); ¹³C NMR (CDCl₃) δ 173.83, 166.71, 166.52, 158.14, 139.77, 134.80, 134.71, 132.88, 132.25, 131.52, 129.98, 128.54, 119.85, 111.91, 111.70, 56.05, 52.02, 51.44, 33.48, 29.11, 25.03; CIMS *m*/*z* 457 (MH⁺). Anal. calcd for C₂₅H₂₈O₈: C, 65.78; H, 6.14. Found: C, 65.13; H, 6.22.

Methyl 5,6-bis[4-methoxy-3-(methoxycarbonyl)phenyl]hex-5-enoate (27). Light yellow oil. Yield: 176 mg (30%). ¹H NMR (CDCl₃) δ 7.88 (d, J=2.30 Hz, 1H), 7.74 (d, J=2.22 Hz, 1H), 7.54 (dd, J=2.30 and 2.22 Hz, 1H), 7.41 (dd, J=2.30 and 2.22 Hz, 1H), 6.98 (s, 1H), 6.96 (s, 1H), 6.62 (s, 1H), 3.91 (s, 3H), 3.90 (s, 3H), 3.89 (s, 3H), 3.59 (s, 3H), 2.68 (t, J=7.73 Hz, 2H), 2.28 (t, J=7.32 Hz, 2H), 1.71 (m, 2H); CIMS m/z 457 (MH⁺). Anal. calcd for C₂₅H₂₈O₈: C, 65.78; H, 6.14. Found: C, 65.47; H, 6.35.

2-Chloro-4-iodophenol (29). 2-Chlorophenol (10.0 g, 77.82 mmol) was dissolved in MeOH (200 mL). Sodium iodide (11.70 g, 77.82 mmol) and sodium hydroxide (3.12 g, 77.82 mmol) were added, and the mixture was

cooled to 0 °C in an ice-water bath. Aqueous sodium hypochlorite (119.2 g of 5% solution) was added dropwise. The mixture was stirred at room temperature for 2 h and the excess hypochlorite was quenched with 10% aqueous sodium thiosulfate (30 mL). The mixture was adjusted to pH 4 using 1 N HCl. The mixture was extracted with ether (3×40 mL). The ether layers were combined and dried over Na₂SO₄. After the solvent was evaporated, the crude solid was crystallized from ace-tone–hexanes (1:1, v/v) to give 2-chloro-4-iodophenol as a light brown solid (19.35 g, 99%): mp 54.5–55.5 °C; ¹H NMR (CDCl₃) δ 7.62 (d, *J*=1.98 Hz, 1H), 7.45 (dd, *J*=8.47 and 1.90 Hz, 1H), 6.78 (d, *J*=8.65 Hz, 1H), 5.59 (s, 1H); ¹³C NMR (CDCl₃) δ 151.34, 137.29, 136.90, 121.08, 118.17, 81.54.

2-Chloro-4-iodoanisole (30). The above phenol **29** (8.0 g, 31.45 mmol) was dissolved in acetone (50 mL), followed by addition of anhydrous K₂CO₃ (6.52 g, 47.17 mmol) and dimethylsulfate (4.46 mL, 47.17 mmol). The reaction mixture was heated at reflux for 14 h, and then the inorganic salts were filtered. The solvent was evaporated and the residue was washed with water (3×20 mL). The crude residue was crystallized from acetone to afford **30** as light brown needles (8.0 g, 94%): mp 83–84 °C: ¹H NMR (CDCl₃) δ 7.65 (d, *J*=2.03 Hz, 1H), 7.49 (dd, *J*=8.58 and 1.96 Hz, 1H), 6.69 (d, *J*=8.63 Hz, 1H), 3.97 (s, 3H); ¹³C NMR (CDCl₃) δ 155.05, 138.15, 136.51, 123.75, 113.89, 81.81, 56.13; EIMS *m*/*z* 269 (MH⁺).

Methyl 6-(3-chloro-4-methoxyphenyl)hex-5-ynoate (31). Methyl 5-hexynoate (504 mg, 4.0 mmol), iodide compound **30** (1.07 g, 4.0 mmol), Pd(OAc)₂ (71.8 mg, 0.32 mmol), copper(I) iodide (152 mg, 0.80 mmol), and triethylamine (1.17 g, 11.56 mmol) were mixed in ethyl acetate (40 mL) and the mixture was kept stirring at room temperature overnight under argon atmosphere. The mixture was then concentrated and the residue was purified by flash chromatography on silica gel (80 g, eluent: EtOAc-hexanes 1:4, v/v) to give 31 as a brown oil (827 mg, 78%): ¹H NMR (CDCl₃) δ 7.41 (d, J=1.95 Hz, 1H), 7.25 (dd, J = 8.57 and 2.0 Hz, 1H), 6.83 (d, J = 8.57 Hz, 1H), 3.89 (s, 3H), 3.68 (s, 3H), 2.50 (t, J = 7.42 Hz, 2H), 2.46 (t, J = 7.86 Hz, 2H), 1.91 (m, 2H); ¹³C NMR (CDCl₃) δ 173.51, 154.66, 133.15, 131.08, 122.12, 116.79, 111.63, 88.28, 79.89, 56.10, 51.55, 32.82, 23.80, 18.77; CIMS m/z 267 (MH⁺).

Hydroarylation of compound 31 to afford 9 and 32. Compound 31 (508 mg, 1.90 mmol), aryl iodide 30 (563.6 mg, 2.1 mmol), and $Pd_2(dba)_3$ (122 mg, 0.133 mmol) were dissolved in ethyl acetate (45 mL). Diethylamine (458 mg, 6.27 mmol) was added, followed by formic acid (210 mg, 4.56 mmol), and the solution was heated to reflux under argon overnight. The reaction mixture was then cooled to room temperature and washed with diluted HCl and brine. The organics were dried over Na₂SO₄, and the solvent was removed under reduced pressure. The products were isolated through flash chromatography (hexane–EtOAc 3:1, v/v; silica gel: 100 g) and preparative TLC (hexane–EtOAc 5:1, v/v). **Methyl 6,6-bis(3-chloro-4-methoxyphenyl)hex-5-enoate (9).** Colorless oil. Yield: 295 mg (38%). ¹H NMR (CDCl₃) δ 7.22 (d, J=2.26 Hz, 1H), 7.14 (d, J=1.99 Hz, 1H), 7.02 (dd, J=3.92 and 2.13 Hz, 1H), 6.98 (dd, J=3.50 and 1.88 Hz, 1H), 6.93 (d, J=8.38 Hz, 1H), 6.81 (d, J=8.61 Hz, 1H), 5.92 (t, J=7.44 Hz, 1H), 3.93 (s, 3H), 3.90 (s, 3H), 3.63 (s, 3H), 2.29 (t, J=7.44 Hz, 2H), 2.13 (dt, J=7.33 and 7.37 Hz, 2H), 1.77 (m, 2H); CIMS m/z 409 (MH⁺). Anal. calcd for C₂₁H₂₂Cl₂O₄: C, 61.62; H, 5.42; Cl, 17.32. Found C, 61.50; H, 5.46; Cl, 17.30.

Methyl 5,6-bis(3-chloro-4-methoxyphenyl)hex-5-enoate (32). Colorless oil. Yield: 287 mg (37%). ¹H NMR (CDCl₃) δ 7.46 (d, J=2.12 Hz, 1H), 7.30 (m, 2H), 7.16 (dd, J=8.49 and 2.0 Hz, 1H), 6.93 (d, J=8.57 Hz, 2H), 6.56 (s, 1H), 3.92 (s, 6H), 3.63 (s, 3H), 2.66 (dt, J=7.11 and 2.41 Hz, 2H), 2.29 (t, J=7.27 Hz, 2H), 1.72 (m, 2H); ¹³C NMR (CDCl₃) δ 173.52, 154.27, 153.65, 140.22, 135.61, 131.20, 130.43, 128.27, 128.03, 126.80, 125.69, 122.40, 122.16, 111.88, 111.79, 56.14, 51.49, 33.45, 29.14, 23.63; CIMS m/z 409 (MH⁺). Anal. calcd for C₂₁H₂₂Cl₂O₄: C, 61.62; H, 5.42; Cl, 17.32. Found C, 61.65; H, 5.62; Cl, 17.32.

Methyl (5Z)-6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyllhex-5-enoate (10). The alkyne 17 (112 mg, 0.37 mmol) was hydrogenated using 5% palladium on barium sulfate (20 mg) and pure quinoline (20 mg) in 10 mL of MeOH. After 18 h, the catalyst was removed by filtration and the solvent was evaporated. The resulting residue was purified by flash chromatography on silica gel (20 g, column: 1×30 cm), using hexanes–EtOAc (3:1 v/v) as eluent, to give a colorless oil (100 mg, 88.5%): ¹H NMR (CDCl₃) δ 7.51 (d, J=2.03 Hz, 1H), 7.23 (d, J = 1.58 Hz, 1H), 6.36 (d, J = 11.63 Hz, 1H), 5.62 (dt, J=11.60 and 5.84 Hz, 1H), 3.91 (s, 3H), 3.78 (s, 3H), 3.64 (s, 3H), 2.34 (t, J=5.67 Hz, 2H), 2.31 (s, 3H), 2.23 (dt, J=7.2 and 5.9 Hz), 1.78 (m, 2H); ¹³C NMR (CDCl₃) δ 173.78, 166.77, 156.79, 135.16, 132.76, 132.31, 131.76, 129.18, 128.31, 124.08, 61.35, 52.03, 51.34, 33.28, 27.61, 24.84, 15.96; EIMS m/z 306 (M⁺); HRMS calcd for C₁₇H₂₂O₅ 306.1467, found 306.1467. Anal. calcd for C₁₇H₂₂O₅: C, 66.66; H, 7.19. Found: C, 66.47; H, 7.28.

Methyl (5E)-6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyllhex-5-enoate (11). A solution of methyl 5iodo-3-methyl-2-methoxybenzoate 16 (673 mg, 2.2 mmol), methyl 5-hexenoate (256 mg, 2.0 mmol), palladium acetate (7 mg, 0.03 mmol), tri-o-tolyphosphine (37 mg, 0.12 mmol), triethylamine (7 mL), and acetonitrile (15 mL) was heated under argon at 70 °C for 18 h. The cooled reaction mixture was evaporated and diluted with water (8 mL) and methylene chloride (40 mL). The methylene chloride layer was separated, washed with water, and dried over anhydrous sodium sulfate. Evaporation of the solvent then gave a brown residue, which was further purified by flash chromatography on silica gel (80 g, hexanes/EtOAc 4:1, v/v) to afford a yellow oil (300 mg, 42.1%): ¹H NMR (CDCl₃) δ 7.60 (d, J=2.35 Hz, 1H), 7.32 (d, J=2.06 Hz, 1H), 6.32 (d, J = 15.83 Hz, 1H), 6.12 (dt, J = 15.72 and 6.96 Hz, 1H),

3.91 (s, 3H), 3.80 (s, 3H), 3.66 (s, 3H), 2.36 (t, J=7.42 Hz, 2H), 2.29 (s, 3H), 2.24 (dt, J=7.18 and 6.83 Hz, 2H), 1.80 (m, 2H); EIMS m/z 306 (M⁺); HRMS calcd for C₁₇H₂₂O₅ 306.1467, found 306.1465. Anal. calcd for C₁₇H₂₂O₅: C, 66.66; H, 7.19. Found: C, 66.54; H, 7.30.

Methyl 6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyl|hexanoate (12). The alkyne 17 (231 mg, 0.76 mmol) was hydrogenated at atmospheric pressure over palladium (5%) on activated carbon (40 mg) in ethyl acetate (15 mL). After TLC (silica gel, hexanes–EtOAc 4:1, v/v) had shown that all the starting material was consumed (14 h), the catalyst was removed by filtration and the solvent was removed at reduced pressure to give a brown oil. This crude product was flash chromatographed on silica gel (10 g) using hexanes–EtOAc (4:1 v/ v) as eluent. Evaporation of the solvent gave the product 12 (200 mg, 85.5%) as a light yellow oil: ¹H NMR $(CDCl_3) \delta 7.42 (d, J = 2.22 Hz, 1H), 7.14 (d, J = 2.11 Hz,$ 1H), 3.90 (s, 3H), 3.79 (s, 3H), 3.66 (s, 3H), 2.54 (t, J = 7.72 Hz, 2H), 2.30 (t, J = 7.0 Hz, 2H), 2.28 (s, 3H), 1.62 (m, 4H), 1.34 (m, 2H); ${}^{13}C$ NMR (CDCl₃) δ 174.11, 166.98, 156.65, 137.56, 135.10, 132.32, 128.57, 124.43, 61.34, 52.00, 51.37, 34.72, 33.84, 30.88, 28.57, 24.62, 15.91; CIMS m/z 309 (MH⁺); HRMS calcd for C₁₇H₂₅O₅ (MH⁺) 309.1702, found 309.1689. Anal. calcd for C₁₇H₂₄O₅: C, 66.23; H, 7.79. Found: C, 66.20; H, 7.81.

Methyl 6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyl]-6-oxohexanoate (13). The alkyne 17 (124 mg, 0.41 mmol) and formic acid (10 mL) were placed in a 25 mL round-bottomed flask equipped with reflux condenser that was properly connected to a 100 mL graduate cylinder inverted in a breaker of water in order to monitor the evolution of carbon monoxide gas. The reaction flask was immersed in an oil bath at 100 °C. Gas evolution ceased after 1 h, and the system was left for 1 h to reach ambient temperature. The solvent was evaporated to give pure ketone 13 as a yellow oil (120 mg 90.9%): ¹H NMR (CDCl₃) δ 8.20 (d, J=2.17 Hz, 1H), 7.94 (d, J = 1.47 Hz, 1H), 3.93 (s, 3H), 3.86 (s, 3H), 3.66 (s, 3H), 2.96 (t, J = 6.78 Hz, 2H), 2.36 (t, J = 7.09 Hz, 2H), 2.34 (s, 3H), 1.73 (m, 4H); ¹³C NMR (CDCl₃) δ 198.16, 173.77, 166.14, 162.12, 134.24, 133.22, 131.93, 129.50, 124.14, 61.51, 52.32, 51.44, 37.84, 33.75, 24.39, 23.45, 16.15; CIMS m/z 323 (MH⁺); HRMS calcd for $C_{17}H_{23}O_6$ (MH⁺) 323.1495, found 323.1489. Anal. calcd for C₁₇H₂₂O₆: C, 63.35; H, 6.83. Found: C, 63.20; H, 6.79.

In vitro anti-HIV assay. Anti-HIV screening of test compounds against various viral isolates and cell lines was performed as previously described.³² This cell-based microtiter assay quantitates the drug-induced protection from the cytopathic effect of HIV-1. Data are presented as the percent control of MTS (CellTiter[®], Promega, Madison, WI, USA) values for the uninfected, drug-free control. EC₅₀ values reflect the drug concentration that provides 50% protection from the cytopathic effect of HIV-1 in infected cultures, while the CC₅₀ reflects the concentration of drug that causes 50% cell death in the uninfected cultures. MTS-based results were confirmed by measurement of cell-free supernatant RT and p24 levels. All MTS cytoprotection data were derived from triplicate tests on each plate with the EC_{50} value representing the average of the triplicates. The error of the triplicates was less than 10% for all determinations.

Assay for inhibition of HIV-1 RT. The effects of the compounds on the HIV-1 RT enzyme were performed using purified p66/51 RT (a kind gift of S. Hughes, ABL-Basic Research Program, NCI-FCRDC). Inhibition of reverse transcription was measured by the level of incorporation of [³²P]GTP into the poly(rC)-oligo(dG) (rCdG) homopolymer template primer system.⁵

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References and Notes

- 1. De Clercq, E. Med. Virol. 1996, 6, 97.
- 2. Drake, S. M. J. Antimicrob. Chemother. 2000, 45, 417.
- 3. Pedersen, O. S.; Pedersen, E. B. Antiviral Chem. Chemother.
- **1999**, *1999*, 285.
- 4. De Clercq, E. Farmaco 1999, 54, 26.
- 5. Carpenter, C. C. J.; Fischl, M. A.; Hammer, S. M.; Hirsch, M. S.; Jacobsen, D. M.; Katzenstein, D. A.; Montaner, J. S. G.; Richman, D. D.; Saag, M. S.; Schooley, R. T.; Thompson, M. A.; Vella, S.; Yeni, P. G.; Volberding, P. A. J. Am. Med. Assoc 1998, 280, 78.

6. Barry, M.; Mulcahy, F.; Back, D. J. Br. J. Clin. Pharmacol. 1998, 45, 221.

- 7. Fischl, M. A. AIDS 1999, 13 (Suppl. 1), S49.
- 8. Vajpayee, M.; Dar, L. Indian J. Pharmacol. 1999, 31, 96.
- 9. Beach, J. W. Clin. Ther. 1998, 20, 2.
- 10. Cushman, M.; Golebiewski, W. M.; Graham, L.; Turpin, J. A.; Rice, W. G.; Fliakas-Boltz, V.; Buckheit, R. W., Jr. *J. Med. Chem.* **1996**, *39*, 3217.
- 11. Cushman, M.; Casimiro-Garcia, A.; Williamson, K.; Rice,
- W. G. Bioorg. Med. Chem. Lett. 1998, 8, 195.
- 12. Cushman, M.; Casimiro-Garcia, A.; Hejchman, E.; Ruell,
- J. A.; Huang, M.; Schaeffer, C. A.; Williamson, K.; Rice,
- W. G.; Buckheit, R. W., Jr. J. Med. Chem. 1998, 41, 2076.

13. Casimiro-Garcia, A.; Micklatcher, M.; Turpin, J. A.; Stup, T. L.; Watson, K.; Buckheit, R. W., Jr.; Cushman, M. *J. Med. Chem.* **1999**, *42*, 4861.

14. Cushman, M.; Golebiewski, M.; Buckheit, R. W., Jr.; Graham, L.; Rice, W. G. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 2713.

- 15. McMurry, J. E. Chem. Rev. 1989, 89, 1513.
- 16. Xu, G.; Loftus, T. L.; Wargo, H.; Turpin, J. A.; Buckheit, R. W.; Cushman, M. J. Org. Chem. **2001**, 66, 5958.
- 17. Hashimoto, N.; Aoyama, T.; Shioiri, T. Chem. Pharm. Bull. 1981, 29, 1475.
- 18. Chumpradit, S.; Kung, M. P.; Mach, R.; Kung, H. F. J. Med. Chem. 1993, 36, 221.
- 19. Fagnola, M. C.; Candiani, I.; Visentin, G.; Cabri, W.; Zarini, F.; Mongelli, N.; Bedeschi, A. *Tetrahedron Lett.* **1997**, *39*, 2307.
- 20. Sonogashira, K.; Tohda, Y.; Hagihara, N. Tetrahedron Lett. 1975, 4467.
- 21. Zhang, H. X.; Guibé, F.; Balavoine, G. J. Org. Chem. 1990, 55, 1857.
- 22. Piers, E.; Coish, P. D. Synthesis 1995, 47.
- 23. Piettre, S. R.; Baltzer, S. Tetrahedron Lett. 1997, 38, 1179.
- 24. Gong, Y.; Pauls, H. W. Synlett. 2000, 829.
- 25. Hay, L. A.; Koenig, T. M.; Ginah, F. O.; Copp, J. D.; Mitchell, D. J. Org. Chem. **1998**, 63, 5050.
- 26. Menashe, N.; Dvora, R.; Shvo, Y. J. Org. Chem. 1991, 56, 2912.
- 27. Patch, R. J.; Roberts, J. C.; Gao, H.; Shi, Z.; Gopalsamy, A.; Kongsjahju, A.; Daniels, K.; Kowalczyk, P. J.; van Schravendijk, M.-R.; Gordon, K. A.; Pallai, P. V. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 2983.
- Pauwels, R.; Andries, K.; Desmyter, J.; Schols, D.; Kukla,
 M. J.; Breslin, H. J.; Raeymaeckers, A.; Van Gelder, J.;
 Woestenborghs, R.; Heykants, J.; Schellekens, K.; Janssen,
 M. A. C.; De Clercq, E.; Janssen, P. A. J. *Nature* 1990, 343, 470.
- 29. Baba, M.; De Clercq, E.; Tanaka, H.; Ubasawa, M.; Takashima, H.; Sekiya, K.; Nitta, I.; Umezu, K.; Nakashima, H.; Mori, S.; Shigeta, S.; Walker, R. T.; Miyasaka, T. *Proc. Natl. Acad. Sci. U.S.A.* **1991**, *88*, 2356.

30. Debyser, Z.; Pauwels, R.; Andries, K.; Desmyter, J.; Kukla, M.; Janssen, P. A. J.; De Clercq, E. *Proc. Natl. Acad. Sci. U.S.A.* **1991**, *88*, 1451.

- 31. Balzarini, J.; Pérez-Pérez, M.-J.; San-Félix, A.; Camarasa, M.-J.; Bathurst, I. C.; Barr, P. J.; De Clercq, E. *J. Biol. Chem.* **1992**, *267*, 11831.
- 32. Rice, W. G.; Bader, J. P. Adv. Pharmacol. (San Diego) 1995, 33, 389.
- 33. Xu, G.; Micklatcher, M.; Silvestri, M.; Loftus, T. L.; Turpin, J. A.; Buckheit, R. W.; Cushman, M. unpublished results.