

Accepted Manuscript

Title: Synthesis and antiproliferative activity of novel 4-substituted-phenoxy-benzamide derivatives

Author: Chi-Yu Sun Yang-Sheng Li Ai-Long Shi Ya-Fei Li
Rui-Fang Cao Huai-Wei Ding Qing-Qing Yin Li-Juan Zhang
Hua-Chuan Zheng Hong-Rui Song



PII: S1001-8417(15)00273-9
DOI: <http://dx.doi.org/doi:10.1016/j.cclet.2015.06.017>
Reference: CCLET 3374

To appear in: *Chinese Chemical Letters*

Received date: 24-3-2015
Revised date: 13-4-2015
Accepted date: 29-5-2015

Please cite this article as: C.-Y. Sun, Y.-S. Li, A.-L. Shi, Y.-F. Li, R.-F. Cao, H.-W. Ding, Q.-Q. Yin, L.-J. Zhang, H.-C. Zheng, H.-R. Song, Synthesis and antiproliferative activity of novel 4-substituted-phenoxy-benzamide derivatives, *Chinese Chemical Letters* (2015), <http://dx.doi.org/10.1016/j.cclet.2015.06.017>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

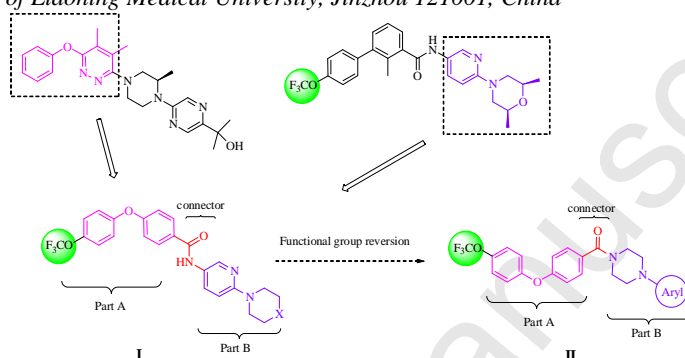
Graphical Abstract

Synthesis and antiproliferative activity of novel 4-substituted-phenoxy-benzamide derivatives

Chi-Yu Sun^a, Yang-Sheng Li^a, Ai-Long Shi^a, Ya-Fei Li^a, Rui-Fang Cao^a, Huai-Wei Ding^{a,*}, Qing-Qing Yin^a, Li-Juan Zhang^a, Hua-Chuan Zheng^b, Hong-Rui Song^{a,*}

^a Key Laboratory of Structure-Based Drug Design and Discovery, Ministry of Education, Shenyang Pharmaceutical University, Shenyang 110016, China

^b The First Affiliated Hospital of Liaoning Medical University, Jinzhou 121001, China



A series of 4-substituted-phenoxy-benzamide derivatives bearing an aryl cycloaliphatic amine moiety were synthesized and evaluated for their antiproliferative activity. Three compounds were further examined for their hedgehog pathway inhibition.

Original article

Synthesis and antiproliferative activity of novel 4-substituted-phenoxy-benzamide derivatives

Chi-Yu Sun^a, Yang-Sheng Li^a, Ai-Long Shi^a, Ya-Fei Li^a, Rui-Fang Cao^a, Huai-Wei Ding^{a,*}, Qing-Qing Yin^a, Li-Juan Zhang^a, Hua-Chuan Zheng^b, Hong-Rui Song^{a,1}

^a Key Laboratory of Structure-Based Drug Design and Discovery, Ministry of Education, Shenyang Pharmaceutical University, Shenyang 110016, China

^b The First Affiliated Hospital of Liaoning Medical University, Jinzhou 121001, China

ARTICLE INFO

ABSTRACT

Article history:

Received 24 March 2015

Received in revised form 13 April 2015

Accepted 29 May 2015

Available online

Keywords:

aryl cycloaliphatic amine

antiproliferative activity

hedgehog signaling

A series of novel 4-substituted-phenoxy-benzamide derivatives bearing an aryl cycloaliphatic amine moiety were synthesized and evaluated for antiproliferative activity against SW620, HT29 and MGC803 cancer cell lines in vitro. The pharmacological data demonstrated that the majority of target compounds exhibited moderate efficacy in HT29 and MGC803 cell lines. Compound **10c** showed promising inhibition of hedgehog (Hh) signaling pathway in an Hh-related assay. In addition, the superposition pattern of **10c** showed a good fit for a pharmacophoric model generated by Hh inhibitors and provided a basis for further structural optimization.

1. Introduction

The Hedgehog (Hh) protein family, which includes Sonic (Shh), Indian (Ihh), and Desert (Dhh) hedgehogs, regulated cell growth and migration during embryonic development [1-3]. Activation of the Hh signaling pathway is initiated by Shh ligands bound to its receptor Patched (Ptch), which relieves its inhibition of Smoothen (Smo) receptor. Smo activation triggers a series of intracellular events ultimately leads to specific gene expression mediated by the Gli family transcription factors [4,5]. Hh signaling pathway was normally silent in adult tissues, nevertheless aberrant activation of the Hh pathway was associated with certain cancers. Thus, the blockade of Hh pathway had been investigated as a novel strategy in cancer chemotherapy [6,7].

Inhibition of Smo activity has shown some promise in the treatment of cancers driven by activating mutations of the Hh pathway [8-10]. Furthermore, several Hh pathway antagonists have proceeded to clinical development, among which vismodegib (**1**, Fig.1) has obtained marketing authorization in the United States in 2012 [11, 12]. Sonidegib (**2**, Fig.2), a clinical stage Hh inhibitor developed by Novartis, is awaiting for the registration in the U.S. for the treatment of patients with advanced basal cell carcinoma. Sonidegib (**2**) bearing a morpholinopyridine unit suppressed the growth of Hh pathway-dependent tumors by selective inhibition of the positive regulator smoothened (Smo) [13, 14]. LEQ506 (**3a**), a second-generation Hh inhibitor, is currently being investigated in a Phase I clinical trial for patients with solid tumors. SAR studies had demonstrated the replacement of the benzylic methylene linker with an oxygen atom (**3b**) was well tolerated, whereas replacement with an NH group (**3c**) resulted in a 10-fold decrease in inhibition of the Hh pathway [15].

Inspired by the structural characteristics of Sonidegib (**2**) and LEQ506 (**3a**), we envisioned that the merging of these two bioactive components would afford a hybrid structure with the potential for antiproliferative activity. We therefore adopted the biaryl ether (Part A) active fragment from LEQ506 analog (**3b**) and morpholino pyridine (Part B) unit from Sonidegib (**2**) in target compounds, and a carbonyl group or an amide group was selected as the linker between the two parts. Additionally, the order of heteroaryl or aryl cycloaliphatic amine units in the target compounds attaching to the linker was reversed based on the functional group reversion principle. Eventually, a series of 4-substituted-phenoxy-benzamide derivatives (I, II Fig.2) were obtained. In this paper, the synthesis of these benzamide derivatives was reported, and their biological activities were evaluated.

2. Experimental

The key intermediates **7a-d** were synthesized according to the routes outlined in Scheme 1. Aryl iodides were synthesized from aryl amines by a diazotization reaction. The generated diazonium salts were iodinated with KI to give compounds **5a-d** [16]. The

¹ Corresponding author.

E-mail address: dinghuaiwei627@163.com (H.-W. Ding); hongruisonghrs@126.com (H.-R. Song);

substituted iodobenzenes **5a-d** were coupled to methylparaben to afford compounds **6a-d**, via an *N,N*-dimethyl glycine-promoted Ullmann coupling. Compounds **6a-d** were further hydrolyzed to compounds **7a-d** in excellent yields under reflux [17].

Compounds **10a-f** were synthesized according to the routes outlined in Scheme 2. 2-Chloro-5-nitropyridine was converted to compounds **8a-b** in the presence of morpholine or piperidine [18]. The resulting nitro compounds **8a-b** were reduced to the amino compounds **9a-b** using stannous chloride dihydrate and hydrochloric acid in aqueous ethanol, and then alkalinized with sodium hydroxide solution [19]. **7a-d** were treated with thionyl chloride to produce the corresponding acyl chloride. **9a-b** reacted with acyl chloride to give the target compounds **10a-f** [20].

Compounds **14a-n**, **15a-b**, **16a-j** were synthesized according to the routes outlined in Scheme 3. Compound **11** was synthesized from 6-chloronicotinic acid via an esterification reaction and then converted to compound **12** using a substitution reaction [21]. Compound **12** was deprotected to give compound **13** under acid conditions [22]. Intermediate **13** was reacted with the corresponding acyl chlorides **7a-d** to give the target compounds **14a-d**, which were further hydrolyzed to compounds **15a-b**. Similarly, compounds **16a-j** were obtained by the reaction of the acyl chloride **7c** and various aryl piperazine [23].

General procedure for preparation of target compounds was given in the Supporting Information.

3. Results and discussion

All the target compounds (**10a-f**, **14a-d**, **15a-b** and **16a-j**) were evaluated for their antiproliferative activity against human colorectal cancer cell lines (SW620 and HT29), and human gastric cancer cell line (MGC803) using the MTT assay with vismodegib as a positive control. The results expressed as half maximal inhibitory concentration (IC_{50}) values are summarized in Table 1.

Initially, target compounds were divided into two regions: diaryl ether (Part A), aryl cycloaliphatic amine moiety (Part B). In general, most of them displayed high efficacy in HT29 and MGC803 cell lines. At the outset, our focus was on the modifications of Part A, including the substitutions on the para position of phenyl. The para-trifluoromethoxy substituent **10c** (IC_{50} = 1.15 μ mol/L [HT29], IC_{50} = 0.56 μ mol/L [MGC803]) showed decent activity, however, para-methoxy and para-chlorine led to a significant loss of activity (**10c** vs **10a**, **10d** or **14a** vs **14b**, **14d**) against MGC803 cells, confirming the beneficial impact of trifluoromethoxy in the R_1 position.

Further investigations focusing on Part B on the antiproliferative activity were performed. On comparing **10d** with **10f**, it was found that morpholino pyridine surrogates were superior to piperidyl pyridine surrogates. Through functional group reversion, pyridyl piperazine analog **16c** (IC_{50} = 4.33 μ mol/L [HT29], IC_{50} = 5.35 μ mol/L [MGC803]) was obtained and exhibited moderate potencies. Addition of a polar methoxycarbonyl (**14a**, IC_{50} = 2.36 μ mol/L [HT29]) group to pyridine increased the activity while its hydrolysate **15a** lost potency by nearly two-fold. When the pyridyl was changed to a pyrimidyl (**16b**, IC_{50} = 2.14 μ mol/L [HT29]), a two-fold increase of activity was observed. However, the trend did not hold in other analogs, as unsubstituted phenyl piperazine analogue **16j** was less potent. Attempts to increase the antiproliferative activity were made by adding various groups to the phenyl ring (Part B). It was worth noting that the para-fluorine (**16h**, IC_{50} = 2.52 μ mol/L [HT29]) and meta-methoxy (**16a**, IC_{50} = 1.95 μ mol/L [HT29]) analogues were much more active than other substituted phenyl piperazine analogues. Overall, the structure-activity relationships (SAR) study revealed that the antiproliferative activity of 3 compounds (**10a**, **10c**, **10d**) was comparable to that of the positive control vismodegib (**1**) in HT29 and MGC803 cells.

As shown in Table 2, the selected compound **10c**, **10d**, **16c** were further tested for their Hh signaling inhibition using a luciferase reporter assay [24, 25] in NIH3T3 cells carrying a stably transfected Gli-reporter construct (NIH3T3-Gli-luc reporter cell line). This assay can effectively identify hedgehog signaling pathway inhibitors. It was suggested that compound **10c** (IC_{50} = 0.082 μ mol/L) was less active as compared to the positive control vismodegib (IC_{50} = 0.013 μ mol/L). In addition, **10c** and **10d** (IC_{50} = 0.127 μ mol/L) showed higher potency than **16c** (IC_{50} = 0.364 μ mol/L) in parallel to their antiproliferative activity. Although the hedgehog pathway inhibition of **10c** was less potent than that of vismodegib, these results were encouraging and worthy of further investigation owing to the large structural changes involved.

Then we examined if compound **10c** fit the proposed pharmacophoric model for Hh antagonists. A conformational analysis was performed using the Common feature program in the Discovery studio 3.0 software. This analysis was conducted on the three compounds covering conformers with a range of 20 kcal/mol with respect to the global minimum that was used in building the best pharmacophoric model. According to this, the low energy conformers of vismodegib (**1**), sonidegib (**2**) and LEQ506 (**3a**) showed a very similar orientation and fulfilled all the pharmacophoric features of the model.

This model was built up by two hydrogen bond acceptor (HBA1-2) groups and two hydrophobic (HY1-2) regions. Fitting of vismodegib (**1**), sonidegib (**2**) and LEQ506 (**3a**) to the pharmacophoric model was shown in Fig. 3A. Analysis of the superposition pattern of **10c** showed a good fit between the molecule and the pharmacophoric model (Fig. 3B). For instance, the carbonyl oxygen in the amide moiety of compound **10c** matched HBA1, and the oxygen atom in the morpholine moiety (Part B) corresponded to the HBA2 feature of the model. Moreover, the two phenyl rings in the diaryl ether moiety (Part A) were superimposed to the hydrophobic regions HY1 and HY2 perfectly.

4. Conclusion

In summary, a series of novel and distinctive 4-substituted-phenoxy-benzamide analogues were synthesized and characterized. Furthermore, three human cancer cell lines were used to evaluate their antiproliferative activity; the majority of analogues exhibited moderate activity and high selectivity in HT29 and MGC803 cell lines. In particular, the most promising compound **10c**

(Hh pathway inhibition $IC_{50} = 0.082 \mu\text{mol/L}$) displayed desirable antiproliferative activity with IC_{50} values of $1.15 \mu\text{mol/L}$ and $0.56 \mu\text{mol/L}$ against HT29 and MGC803 cell lines, respectively. The analysis of SAR indicated that compounds with a para-trifluoromethoxyl group on Part A were more potent than those with other substituents. In addition, the morpholino pyridine fragment in Part B exhibited higher efficacy as compared to the aryl piperazine fragment. Further structural optimization studies are presently in progress and will be reported in due course.

Acknowledgment

The work was supported by Program for Innovative Research Team of the Ministry of Education of China and Program for Liaoning Innovative Research Team in University. Authors wish to thank to the Innovation and entrepreneurship training program for college students in Liaoning Province (No. 201410163009).

References

- [1] J.A. Williams, O.M. Guicherit, B.I. Zaharian, et al., Identification of a small molecule inhibitor of the hedgehog signaling pathway: Effects on basal cell carcinoma-like lesions, *Proc. Natl. Acad. Sci. USA* 100 (2003) 4616-4621.
- [2] N. Mahindroo, C. Punchihewa, N. Fujii, Hedgehog-gli signaling pathway inhibitors as anticancer agents, *J. Med. Chem.* 52 (2009) 3829-3845.
- [3] H. Onishi, M. Katano, Hedgehog signaling pathway as a therapeutic target in various types of cancer, *Cancer Sci.* 102 (2011) 1756-1760.
- [4] M.K. Hadden, Hedgehog pathway inhibitors : a patent review (2009 - present), *Expert Opin. Ther. Pat.* 23 (2013) 345-361.
- [5] A. Pimentel, M. Velez, L.J. Barahona, et al., New prospects for drug development: the hedgehog pathway revealed. Focus on hematologic malignancies, *Future Oncol.* 9 (2013) 681-697.
- [6] J. M. Ruch, E. J. Kim, Hedgehog signaling pathway and cancer therapeutics: progress to date, *Drugs* 73 (2013) 613-623.
- [7] C. Dockendorff, M.M. Nagiec, M. Weïwer, Macrocyclic hedgehog pathway inhibitors: optimization of cellular activity and mode of action studies, *ACS Med. Chem. Lett.* 3 (2012) 808-813.
- [8] S.X. Atwood, M. Li, A. Lee, J.Y. Tang, A.E. Oro, GLI activation by atypical protein kinase C α regulates the growth of basal cell carcinomas, *Nature* 494 (2013) 484-488.
- [9] Y. Wang, A.C. Arvanites, L. Davidow, et al., Selective Identification of Hedgehog Pathway Antagonists By Direct Analysis of Smoothed Ciliary Translocation, *ACS Chem. Biol.* 7 (2012) 1040-1048.
- [10] F. Manetti, H. Faure, H. Roudaut, et al., Virtual screening-based discovery and mechanistic characterization of the acylthiourea MRT-10 family as smoothed antagonists, *Mol. Pharmacol.* 78 (2010) 658-664.
- [11] S.E. Gould, J.A. Low, J.C. Marsters, et al., Discovery and preclinical development of vismodegib, *Expert Opin. Drug Discov.* 9 (2014) 969-984.
- [12] A.E. Proctor, L.A. Thompson, C.L. O'Bryant, Vismodegib: An inhibitor of the hedgehog signaling pathway in the treatment of basal cell carcinoma, *Ann. Pharmacother.* 48 (2014) 99-106.
- [13] S.F. Pan, X. Wu, J.Q. Jiang, et al., Discovery of NVP-LDE225, a potent and selective smoothed antagonist, *ACS Med. Chem. Lett.* 1 (2010) 130-134.
- [14] M. Zollinger, F. Lozac'h, E. Hurh, et al., Absorption, distribution, metabolism, and excretion (ADME) of ^{14}C -sonidegib (LDE225) in healthy volunteers, *Cancer Chemother. Pharmacol.* 74 (2014) 63-75.
- [15] S. Peukert, F. He, M. Dai, et al., Discovery of NVP-LEQ506, a second-generation inhibitor of smoothed, *Chem. Med. Chem.* 8 (2013) 1261-1265.
- [16] A.R. Hajipour, M. Seddighi, Application of [Hcpy] HSO_4 brønsted acidic ionic liquid for the synthesis of aryl iodides from aromatic amines, *Org. Prep. Procd. Inter.* 43 (2011) 292-296.
- [17] Y.H. Yang, Z.L. Wang, J.Z. Yang, et al., Design, synthesis and evaluation of novel molecules with a diphenyl ether nucleus as potential antitubercular agents, *Bioorg. Med. Chem. Lett.* 22 (2012) 954-957.
- [18] H.W. Ding, Z. Chen, C.L. Zhang, et al., Synthesis and cytotoxic activity of some novel *N*-pyridinyl-2-(6-phenylimidazo[2,1-*b*]thiazol-3-yl) acetamide derivatives, *Molecules* 17 (2012) 4703-4716.
- [19] B. Yang, A.W. Hird, D.J. Russell, et al., Discovery of novel hedgehog antagonists from cell-based screening: isosteric modification of p38 bisamides as potent inhibitors of SMO, *Bioorg. Med. Chem. Lett.* 22 (2012) 4907-4911.
- [20] Y.B. Chen, J.L. Li, X.S. Shao, et al., Design, synthesis and insecticidal activity of novel anthranilic diamides with benzyl sulfide scaffold, *Chin. Chem. Lett.* 24 (2013) 673-676.
- [21] K.M. Jung, K.H. Kim, J.L. Jin, M.J. Cho, D.H. Choi, Deep-red light-emitting phosphorescent dendrimer encapsulated tris-[2-benzo[*b*]thiophen-2-yl-pyridyl] iridium (III) core for light-emitting device applications, *J. Polym. Sci. Part A: Polym. Chem.* 46 (2008) 7517-7533.

- [22] J. Wehbe, V. Rolland, A. Fruchier, M.L. Roumestant, J. Martinez, Enantioselective synthesis of new 4-substituted glutamic acid derivatives, *Tetrahedron: Asym.* 15 (2004) 851-858.
- [23] T. You, K. Chen, F.H. Wang, et al., Design, synthesis, and biological evaluation of *N*-hydroxycinnamamide/salicylic acid hybrids as histone deacetylase inhibitors, *Chin. Chem. Lett.* 25 (2014) 474-478.
- [24] J.K. Chen, J. Taipale, K.E. Young, T. Maiti, P.A. Beachy, Small molecule modulation of Smoothened activity, *Proc. Nat. Acad. Sci. USA* 99 (2002) 14071-14076.
- [25] M.H. Xin, L.D. Zhang, F. Tang, et al., Design, synthesis, and evaluation of pyrrolo[2,1-*f*][1,2,4]triazine derivatives as novel hedgehog signaling pathway inhibitors, *Bioorg. Med. Chem.* 22 (2014) 1429-1440.

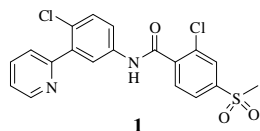


Fig. 1 The structure of vismodegib (**1**)

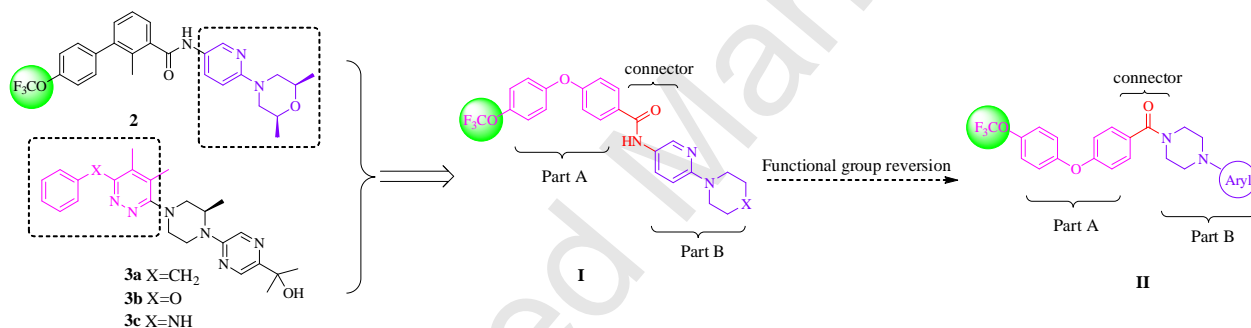
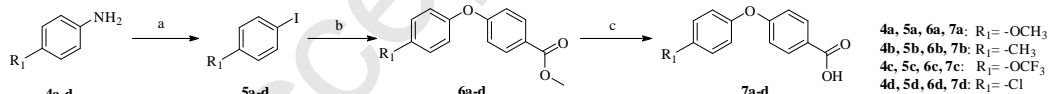
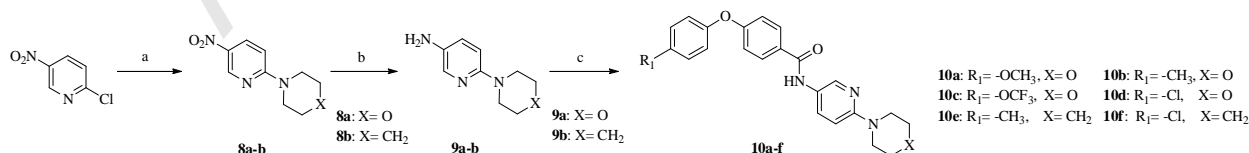


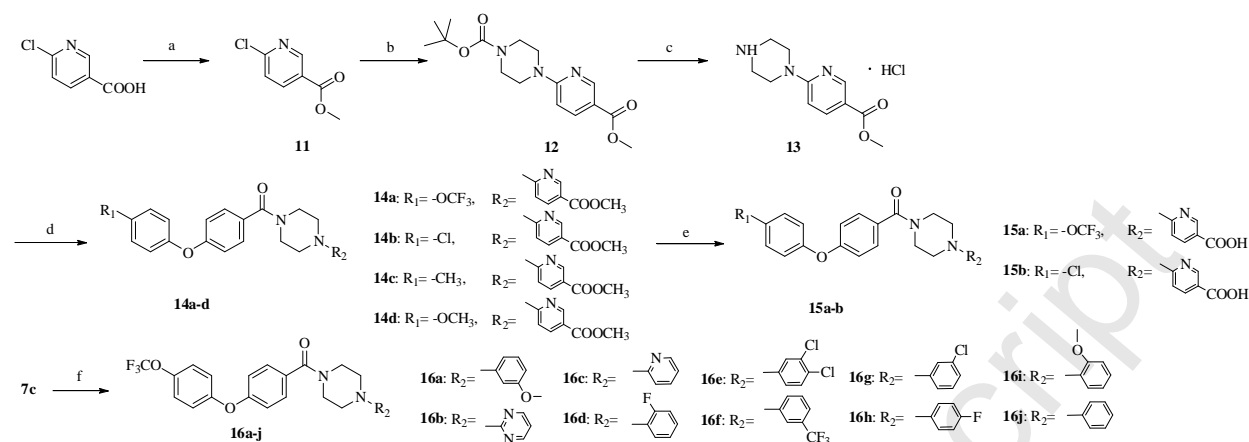
Fig. 2 Sonidegib (**2**), LEQ506 analogues (**3a-c**) and general structure of the target compounds (**I**, **II**).



Scheme 1 Reagents and conditions: (a) (1) NaNO_2 , H_2SO_4 , 0°C , 2h; (2) KI , dichloromethane, 0°C , 6h; (b) 4-methyl-1-iodobenzene, CuI , *N,N*-dimethylglycine, Cs_2CO_3 , 1,4-dioxane, reflux under nitrogen atmosphere, 24h; (c) NaOH , ethanol solution, reflux, 3h



Scheme 2 Reagents and conditions: (a) morpholine or piperidine, K_2CO_3 , THF, reflux, 4h; (b) $\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$, ethanol, HCl , reflux, 8h; (c) acyl chloride of **7a-d**, Et_3N , dichloromethane, 0°C , 12h.



Scheme 3 Reagents and conditions: (a) Methanol, concentrated sulfuric acid, 65 °C; (b) 1-boc-piperazine, K₂CO₃, DMAP, DMF, 110 °C, 24h; (c) concentrated hydrochloric acid in dioxane, dichloromethane 30 °C, 2h; (d) chloride of **7a-d**, Et₃N, dichloromethane, 0 °C, 4h; (e) NaOH, ethanol solution, reflux, 3h; (f) chloride of **7a-d**, aryl piperazine or heteroaryl piperazine, Et₃N, dichloromethane, 0 °C, 4h.

Table 1 *In vitro* cell growth inhibition by the target compounds and vismodegib (1).

Compd.	IC ₅₀ (μmol/L) ± SD ^a			Compd.	IC ₅₀ (μmol/L) ± SD ^a		
	SW620	HT29	MGC803		SW620	HT29	MGC803
10a	26.76 ± 3.56	1.87 ± 0.22	2.36 ± 0.35	15b	26.07 ± 3.64	16.20 ± 4.60	13.29 ± 2.43
10b	79.49 ± 10.28	2.82 ± 0.73	3.08 ± 0.63	16a	12.56 ± 2.87	1.95 ± 0.62	2.91 ± 0.57
10c	7.24 ± 0.80	1.15 ± 0.24	0.56 ± 0.11	16b	10.71 ± 1.98	2.14 ± 0.49	2.63 ± 0.61
10d	23.19 ± 3.03	1.53 ± 0.50	1.77 ± 0.48	16c	19.08 ± 3.62	4.33 ± 1.65	5.35 ± 1.40
10e	>100	4.41 ± 1.35	3.82 ± 1.02	16d	46.58 ± 7.41	14.58 ± 3.81	10.84 ± 2.59
10f	48.49 ± 6.17	3.45 ± 0.28	2.80 ± 0.91	16e	20.34 ± 3.53	8.26 ± 2.27	6.27 ± 0.82
14a	14.85 ± 3.12	2.36 ± 0.96	3.16 ± 0.87	16f	72.35 ± 9.60	34.93 ± 6.54	33.21 ± 5.88
14b	15.96 ± 4.25	3.88 ± 1.05	5.53 ± 1.66	16g	65.62 ± 9.75	26.31 ± 4.82	38.54 ± 5.76
14c	>100	16.20 ± 4.65	20.65 ± 2.90	16h	14.27 ± 2.82	2.52 ± 0.71	3.43 ± 0.94
14d	5.19 ± 1.21	11.74 ± 3.03	9.98 ± 1.78	16i	51.83 ± 8.36	13.27 ± 2.37	14.18 ± 2.52
15a	57.31 ± 8.65	5.00 ± 1.07	9.19 ± 1.32	16j	32.95 ± 4.51	26.08 ± 4.12	25.36 ± 3.35
Vismodegib ^b	10.92 ± 2.76	1.08 ± 0.16	2.39 ± 0.17				

Bold values show the IC₅₀ values of target compounds lower than the values of the positive control.

^a IC₅₀: concentration of the compound (μmol/L) producing 50% cell growth inhibition after 48 h of drug exposure, as determined by the MTT assay. Each experiment was carried out in triplicate.

^b Used as a positive control.

Table 2 The hedgehog pathway inhibition of compound **10c**, **10d**, **16c** and vismodegib (1).

Compd.	NIH3T3-Gli-luc IC ₅₀ (μmol/L) ^a
10c	0.082
10d	0.127
16c	0.364
vismodegib ^b	0.013

^a The values are an average of triplicate separate determinations.

^b Used as a positive control.

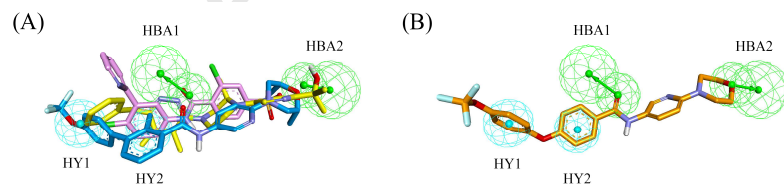


Fig. 3 A. Pharmacophore models for vismodegib, sonidegib and LEQ506. Graphical representation of vismodegib (pink), sonidegib (blue), LEQ506 (yellow) fitted to the proposed pharmacophoric model for hedgehog antagonists. Pharmacophoric features are color coded: green for hydrogen bond acceptor groups (HBA1-2) and cyan for hydrophobic regions (HY1-2). HBA features are constituted by a smaller sphere accommodating the hydrogen bond acceptor group, by a directionality vector represented by an arrow, and by a larger sphere intended to allocate the hydrogen bond donor group of the target macromolecule. B. Pharmacophore model for **10c**. The atoms are color coded: Orange, carbon; white, hydrogen; red, oxygen; blue, nitrogen