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# **Bioorganic & Medicinal Chemistry Letters**

journal homepage: www.elsevier.com/locate/bmcl

# Synthesis and biological evaluation of oxazole derivatives as T-type calcium channel blockers

Jie Eun Lee<sup>a,b</sup>, Hun Yeong Koh<sup>b</sup>, Seon Hee Seo<sup>a</sup>, Yi Yeon Baek<sup>a</sup>, Hyewhon Rhim<sup>a</sup>, Yong Seo Cho<sup>a</sup>, Hyunah Choo<sup>a,\*,†</sup>, Ae Nim Pae<sup>a,\*,†</sup>

<sup>a</sup> Life/Health Division, Korea Institute of Science and Technology, PO Box 131, Cheongryang, Seoul 130-650, Republic of Korea <sup>b</sup> Department of Chemistry, Inha University, Nam-gu, Incheon 402-751, Republic of Korea

#### ARTICLE INFO

Article history: Received 16 March 2010 Revised 11 May 2010 Accepted 12 May 2010 Available online 15 May 2010

Keywords: T-type calcium channel HTS assay  $\alpha_{1G}$  Subtype Neuropathic pain Oxazole

## ABSTRACT

T-type calcium channel is one of therapeutic targets for the treatment of cardiovascular diseases and neuropathic pain. In this study, as a part of our ongoing efforts to develop potent T-type calcium channel blockers, we designed oxazole derivatives substituted with arylpiperazinylalkylamines. The oxazoles were synthesized in a convenient convergent synthetic method, and biologically evaluated against  $\alpha_{1G}$  (Ca<sub>v</sub>3.1) T-type calcium channel. Among total 41 oxazole compounds synthesized, the most active one was the compound **10-35** with an IC<sub>50</sub> value of 0.65  $\mu$ M, which is comparable with that of mibefradil. © 2010 Elsevier Ltd. All rights reserved.

Calcium is an essential signaling molecule with fundamental physiological roles that range from activation of calcium-dependent enzymes to the secretion of neurotransmitters and hormones.<sup>1,2</sup> The most abundant route of calcium entry into electrically excitable cells is by opening of voltage-gated calcium channels.<sup>1,2</sup> The highest organisms express several different types of voltage-gated calcium channels, which are grossly divided into two categories: high-voltage activated (HVA) and low-voltage activated (LVA) calcium channels.<sup>3</sup> Members of the HVA channel family include the N-, P-, Q-, L-, and R-types and typically require large membrane depolarizations to open.<sup>3</sup> LVA calcium channels, also known as T-types, are widely expressed in various types of neurons and can be activated by a weak depolarization near the resting membrane potential and regulate the excitability and electroresponsiveness of neurons under physiological conditions near resting states.<sup>3</sup>

Since mibefradil, a selective T-type calcium channel blocker, approved for the treatment of angina pectoris and hypertension (Fig. 1), was withdrawn from the market in 1998 due to unfavorable drug-drug interaction,<sup>4,5</sup> there have been efforts to discover novel T-type calcium channel blockers.<sup>6</sup> According to accumulation of new findings on T-type calcium channels, it has been

reported that T-type calcium channels play crucial roles in the control of pain which are caused by hyperexcitable neurons.<sup>7</sup> The role of T-type calcium channels in pain has been addressed using specific genetic modulation of T-type calcium channel isoforms. Compared with wild-type mice, Ca<sub>v</sub>3.1 knockout ( $\alpha_{1G}^{-I-}$ ) mice were observed, in which, after L5 spinal nerve ligation, spontaneous pain responses were reduced and a threshold for paw withdrawal was increased in response to mechanical stimulation.<sup>8</sup> Ca<sub>v</sub>3.2 antisense treatment resulted in major anti-nociceptive and anti-hyperalgesic effect, suggesting that Ca<sub>v</sub>3.2 plays a major pronociceptive role in acute and chronic pain states.<sup>9</sup> Together, the results of these two studies suggest that blocking T-type calcium channels should reduce nociceptive pain and neuropathic pain. T-type calcium channel blockers such as ethosuximide and mibefradil were reported to reverse neuropathic pain in animal models.<sup>10</sup>



Figure 1. Structure of mibefradil.

<sup>\*</sup> Corresponding authors. Tel.: +82 2 958 5157; fax: +82 2 958 5189 (H.C.); tel.: +82 2 958 5185; fax: +82 2 958 5189 (A.N.P.).

*E-mail addresses:* hchoo@kist.re.kr (H. Choo), anpae@kist.re.kr (A.N. Pae).

<sup>0960-894</sup>X/\$ - see front matter @ 2010 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2010.05.030

Previously, we reported isoxazole derivatives substituted with arylpiperazinylalkylamines as T-type calcium channel blockers.<sup>6e</sup> Their activities depended on the substituents on isoxazoles, aryl moieties of arylpiperazine, and alkyl chains. In this study, to obtain better pharmacological activities against T-type calcium channel, we introduced an oxazole group instead of the isoxazole group and more diverse aryl moieties at arylpiperazines. By using an oxazole scaffold, structurally unaccessible positions of the isoxazole ring can be substituted with various functional groups, which would allow extensive structure–activity relationship study. Herein, we report the synthesis and T-type calcium channel inhibitory

activities of oxazole derivatives with arylpiperazinylalkylamines substituents.

The strategy to synthesize the designed oxazole derivatives is a convergent synthesis, where the separately prepared arylpiperazinylalkylamine part (Scheme 1) and the oxazole part (Scheme 2) were combined through reductive amination to afford the desired oxazole derivatives (Scheme 3).

Arylpiperazinylalkylamines **4** were synthesized starting from arylpiperazines **1** in two steps. Various arylpiperazines underwent alkylation reaction with *N*-(2-bromoethyl)phthalimide (n = 1) or *N*-(3-bromopropyl)phthalimide (n = 2) to afford compounds **3** in



Scheme 1. Synthesis of arylpiperazinylalkylamines.







Scheme 3. Convergent synthesis of desired oxazole derivatives 10.



Figure 2. Benzhydrylpiperazine derivatives 11.

40–80% yields. The compounds **3** were converted to the corresponding amines **4** by treatment with hydrazine in refluxing EtOH in over 90% yields (Scheme 1).

#### Table 1

Inhibitory activities of oxazole derivatives 10 and 11 against  $\alpha_{1G}$  (Ca<sub>V</sub>3.1) T-type calcium channel

The oxazole parts, oxazole aldehydes **9**, were synthesized starting from ethyl benzoylacetate **5** in four steps. The benzoylacetate **5** was treated with hydroxyl(2,4-dinitrobenzensulfonyloxy)iodobenzene (HDNIB), which was easily prepared from diacetoxyiodobenzene and 2,4-dinitrobenzensulfonic acid, to afford an intermediate **6** installed with a good leaving group, 2,4-dinitrobenzenesulfonate (ODNs) (Scheme 2).<sup>11</sup> The intermediate **6** was condensed with acetamide in acetonitrile under reflux conditions to give one oxazole **7a** with R<sup>2</sup> as methyl group in 45% yield, while reaction with butyramide under microwave irradiation afforded the other oxazole **7b** with R<sup>2</sup> as propyl group in 48% yield. The oxazoles **7** underwent reduction by treating with DIBAL-H to afford the corresponding alcohols **8a** and **8b** in 91–95% yields. Swern oxidation of the alcohols **8** provided the corresponding aldehydes **9a** and **9b** in 44–93% yields.



Entry	Compd	R <sup>1</sup>	R <sup>2</sup>		%Inhbition at 10 µM	IC50 in µM
1	10-1	Н	Propyl	1	54.26	10.25 ± 0.91
2	10-2	2-F	Propyl	1	46.49	b
3	10-3	3-F	Propyl	1	62.34	3.35 ± 0.26
4	10-4	4-F	Propyl	1	54.08	7.08 ± 0.29
5	10-5	2-Cl	Propyl	1	60.07	4.37 ± 0.30
6	10-6	3-Cl	Propyl	1	51.24	$1.45 \pm 0.9^{b}$
7	10-7	4-Cl	Propyl	1	55.84	$2.82 \pm 0.04$
8	10-8	2-CH <sub>3</sub>	Propyl	1	51.34	$4.30 \pm 0.41$
9	10-9	3-CH <sub>3</sub>	Propyl	1	43.13	b
10	10-10	4-CH <sub>3</sub>	Propyl	1	48.06	b
11	10-11	2-0CH <sub>3</sub>	Propyl	1	39.73	b
12	10-12	3-OCH <sub>3</sub>	Propyl	1	40.17	b
13	10-13	4-0CH <sub>3</sub>	Propyl	1	51.49	$9.20 \pm 0.46$
14	10-14	2-CF3	Propyl	1	54.62	$2.10 \pm 0.09$
15	10-15	3-CF <sub>3</sub>	Propyl	1	56.30	$1.94 \pm 0.12$
16	10-16	4-CF <sub>3</sub>	Propyl	1	58.70	$1.30 \pm 0.08$
17	10-17	2,3-DiCH <sub>3</sub>	Propyl	1	66.68	1.71 ± 0.16
18	10-18	$2,4-DiCH_3$	Propyl	1	64.28	$2.22 \pm 0.11$
19	10-19	3,4-DiCH <sub>3</sub>	Propyl	1	67.87	$2.22 \pm 0.27$
20	10-20	2-Pyridyl <sup>a</sup>	Propyl	1	48.13	b
21	10-21	Н	Propyl	2	43.69	b
22	10-22	2-F	Propyl	2	54.41	16.13 ± 3.48
23	10-23	3-F	Propyl	2	68.94	$3.03 \pm 0.22$
24	10-24	4-F	Propyl	2	62.53	$3.21 \pm 0.20$
25	10-25	2-Cl	Propyl	2	64.72	$2.80 \pm 0.39$
26	10-26	3-Cl	Propyl	2	65.88	$1.75 \pm 0.04$
27	10-27	4-Cl	Propyl	2	63.25	$1.31 \pm 0.05$
28	10-28	2-CH3	Propyl	2	54.45	8.75 ± 5.4
29	10-29	3-CH <sub>3</sub>	Propyl	2	47.78	$1.19 \pm 0.6$
30	10-30	4-CH <sub>3</sub>	Propyl	2	53.92	130.01 ± 10.4
31	10-31	2-0CH <sub>3</sub>	Propyl	2	41.03	b
32	10-32	3-OCH <sub>3</sub>	Propyl	2	36.35	b
33	10-33	4-0CH <sub>3</sub>	Propyl	2	42.13	b
34	10-34	2-CF3	Propyl	2	44.99	b
35	10-35	3-CF <sub>3</sub>	Propyl	2	51.63	0.65 ± 0.3
36	10-36	4-CF <sub>3</sub>	Propyl	2	51.62	7.25 ± 3.7
37	10-37	2,3-DiCH <sub>3</sub>	Propyl	2	55.24	$1.78 \pm 0.14$
38	10-38	3,4-DiCH <sub>3</sub>	Propyl	2	54.59	$3.02 \pm 0.18$
39	10-39	4-Cl	Methyl	2	38.08	b
40	11-1	b	Propyl	b	71.85	7.47 ± 1.69
41	11-2	b	Methyl	b	57.32	b
42		Mibefrad	lil		80.00	$0.83 \pm 0.19$

<sup>a</sup> Pyridyl instead of R<sup>1</sup>-phenyl.

<sup>b</sup> Not available or determined.

Arylpiperazinylalkylamines **4** and the oxazole aldehydes **9** were combined by reductive amination to afford compounds **10** in 11–81% yields (Scheme 3). Instead of  $R^1$ -substituted phenyl, benzhydryl group was also introduced with as same procedures as shown here to afford compounds **11** in Figure 2.

A total of 41 oxazole derivatives **10** and **11** were biologically evaluated against  $\alpha_{1G}$  (Ca<sub>v</sub>3.1) T-type calcium channel in HEK293 cells which stably express both T-type calcium channel Ca<sub>v</sub>3.1 and potassium channel Kir2.1.<sup>12</sup> All the synthesized compounds were screened by fluorescence-based high-throughput screening (HTS) FDSS6000 assay,<sup>13</sup> and the %-inhibitions of Ca<sup>2+</sup> current measured at 10  $\mu$ M concentration of the oxazole derivatives are summarized in Table 1. Among those, compounds with over 50% inhibition of Ca<sup>2+</sup> current in FDSS assay were selected for patch-clamp assays using single cells to evaluate IC<sub>50</sub> values.<sup>14,15</sup>

In general, the %-inhibition results from the fluorescence-based HTS FDSS6000 assay do not directly correlate with the IC<sub>50</sub> values obtained from patch-clamp assays. In this study, we also observed that the two compounds **10-23** and **10-19** with the highest %-inhibitions show inhibitory activities with moderate IC<sub>50</sub> values of 3.03  $\mu$ M and 2.22  $\mu$ M, respectively. On the other hand, two compounds **10-35** and **10-16** with the most potent IC<sub>50</sub> values have low inhibitions, respectively). Nevertheless, the filtering process using FDSS6000 assay is necessary because compounds active in both fluorescence-based and electrophysiological methods could have more possibility to be eventually effective in in vivo system. The %-inhibition results were used as primary screening tool to filter compounds for further laborious patch-clamp assays.

A total of 28 compounds were selected to obtain IC<sub>50</sub> values against T-type calcium channel, especially  $\alpha_{1G}$  subtype. The IC<sub>50</sub> values are between 0.65  $\mu$ M and 130.01  $\mu$ M as shown in Table 1. There is no huge difference in inhibitory activities according to the chain length (n = 1 or 2). However, the electronic nature of the R<sup>1</sup> substituents exerted subtle effects on the activities of the compounds. The compounds with electron-withdrawing groups such as F, Cl, and CF<sub>3</sub> are more potent than those with electrondonating groups such as OCH<sub>3</sub>. The position of the substituent also affects the biological activity. Generally, oxazole analogues with electron-withdrawing R<sup>1</sup> substituents at *meta* position show good inhibitory activities against T-type calcium channel. The compound 10-3 with meta-fluoro (3-F) substituent shows better inhibitory activity than 10-2 and 10-4 with ortho- and para-fluoro groups, respectively. By the same token, compounds 10-6, 10-23, and **10-35** are more potent than the corresponding regioisomers. Among those, the compound **10-35** with meta- $CF_3$  (3- $CF_3$ ) shows best inhibitory activity against  $\alpha_{1G}$  (Ca<sub>V</sub>3.1) T-type calcium channel with an IC<sub>50</sub> value of 0.65  $\mu$ M which is comparable to that of mibefradil.<sup>16</sup> In the case of compounds **10-15** and **10-26** with meta-substituents, their activities were only slightly lower than those of compounds 10-16 and 10-27 with para-substituents. On the other hand, it is noteworthy that the dimethyl substituted compounds 10-17, 10-18, 10-19, 10-37, and 10-38 show potent activities with  $IC_{50}$  values of 1.78–3.02  $\mu$ M, regardless of the substitution patterns. The compound 11-1 with a benzhydrylpiperazine group shows the highest %-inhibition with 71.85%, but shows only moderate activity with an IC<sub>50</sub> value of 7.47  $\mu$ M. When R<sup>2</sup> substituents are changed from a propyl group to a methyl group, activities are reduced (entries 39 and 41 in Table 1).

Previously, we reported isoxazole derivatives as T-type calcium channel blockers,<sup>6e</sup> in which compounds with 3-CF<sub>3</sub>-phenylarylpiperazinylalkylamine substituent showed the most potent inhibitory activity against T-type calcium channel. In this oxazole series, the most active compound **10-35** also has R<sup>1</sup> substituent as 3-CF<sub>3</sub>. For this reason, it is fair to assume that the aromatic *meta*-trifluoromethyl substituent may play an important role in inhibitory activity of oxazoles as well as isoxazole derivatives.

In summary, oxazole derivatives with arylpiperazinylalkylamines were designed, synthesized, and biologically evaluated against  $\alpha_{1G}$  (Ca<sub>V</sub>3.1) T-type calcium channel. Among total 41 oxazole compounds synthesized, the most active one was the compound **10-35** with an IC<sub>50</sub> value of 0.65  $\mu$ M, which is comparable to that of mibefradil. The compound **10-35** possesses 3-CF<sub>3</sub> as R<sup>1</sup> substituent, which correlates well to the results of previous isoxazole series. Further evaluations of the compound **10-35** such as selectivity for other calcium channels, pharmacokinetics and neuronal analgesic effect are in progress.

## Acknowledgement

This work was supported by Korea Institute of Science and Technology.

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- 15. Two methods are available: One is a manual patch-clamp assay (EPC 10<sup>®</sup>, HEKA, Germany) and the other is an automated patch-clamp assay (NPC<sup>®</sup>-16 patchliner, Nanion Technologies, Germany). Either of the patch-clamp assays was used to obtain IC<sub>50</sub> values of the selected oxazole derivatives, irrespective of kinds of the compounds, because both IC<sub>50</sub>s of mibefradil from two methods was practically same with IC<sub>50</sub> values of 0.86  $\mu$ M and 0.83  $\mu$ M, respectively.
- was practically same with IC<sub>50</sub> values of 0.86  $\mu$ M and 0.83  $\mu$ M, respectively. 16. *Spectral data:* Free form of **10-35**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.99 (t, *J* = 7.45 Hz, 3H), 1.70–1.89 (m, 4H), 2.48 (t, *J* = 7.07 Hz, 2H), 2.59 (br t, *J* = 5.1 Hz, 4H), 2.72–2.79 (m, 4H), 3.19 (br t, *J* = 5.1 Hz, 4H), 4.02 (s, 2H), 6.98–7.13 (m, 3H), 7.23–7.36 (m, 2H), 7.38–7.45 (m, 2H), 7.65 (dd, *J* = 8.3, 1.3 Hz, 2H); <sup>13</sup>C NMR (100 M Hz, CDCl<sub>3</sub>)  $\delta$  13.7, 20.6, 26.0, 30.1, 43.7, 78.2, 78.5, 53.0, 57.1, 112.1, 115.9, 118.7, 124.3 (q, *J* = 272.4 Hz), 127.1, 127.7, 128.7, 129.5, 131.4 (q, *J* = 31.7 Hz), 131.8, 136.5, 144.4, 151.3, 163.7; HCl salt form of **10-35**: <sup>1</sup>H NMR (400 MHz, MeOD)  $\delta$  1.06 (t, *J* = 7.45 Hz, 3H), 1.89 (sxt, *J* = 7.43 Hz, 2H), 2.21– 2.36 (m, 2H), 2.88 (t, *J* = 7.45 Hz, 2H), 3.24–3.34 (m, 8H), 3.62–3.75 (m, 2H), 3.84–4.00 (m, 2H), 4.61 (s, 2H), 7.20 (d, *J* = 7.58 Hz, 1H), 7.25–7.31 (m, 2H), 7.42–7.47 (m, 2H), 7.52 (t, *J* = 7.33 Hz, 2H), 7.71 (d, *J* = 7.33 Hz, 2H); HRMS (FAB, M+1) calcd for C<sub>27</sub>H<sub>34</sub>F<sub>3</sub>N<sub>4</sub>O 487.2679, found 487.2683.