Glycocinnasperimicin D Synthetic Studies: Synthesis of Cinnamoyl Glycosides via Iodination-Heck Reaction Sequence Starting from Phenyl Glycosides

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Received 9 September 2003

Abstract: A new approach for the synthesis of cinnamoyl glycoside has been developed via iodination-Heck reaction sequence starting from phenyl glycoside. Successful application of this procedure accomplished the construction of the right core structure of aminosugar antibiotic, glycocinnasperimicin D.

Key words: antibiotics, cinnamoyl glycoside, glycosylation, Heck reaction, palladium

In 1985, Umezawa et al. reported the isolation of a novel antibiotic, glycocinnasperimicin D, from the fermentation broth of Nocardia strain.¹ Its structure was elucidated by spectroscopic studies as represented in Figure 1, which clarified that this aminosugar antibiotic belongs to a member of glycocinnamoylspermidine. The more complex relative with an additional 2,4-diaminopentose is LL-BM123 β [cinodine β , (2)], which was isolated and identified at Lederle Laboratories.² Glycocinnasperimicin D(1)has demonstrated broad antibacterial spectrum and several unique structural features; the left portion is 2-ureidopentose and the right part is 2,4-diamino-2,4,6-trideoxy- α -D-glucopyranose with *p*-cinnamoyl spermidine aglyson. These two amino sugars are joined with urea glycosyl linkage. The interesting structure and small availability of 1 from natural sources promoted us to pursue the synthetic work of $1.^3$ In this manuscript, we describe a new approach to cinnamoyl glycosides and its application for the synthesis of right segment of 1.



SYNLETT 2004, No. 1, pp 0089–0092 Advanced online publication: 4.12.2003 DOI: 10.1055/s-2003-43376; Art ID: U17903ST © Georg Thieme Verlag Stuttgart · New York During our synthetic endeavor, we soon recognized that glycosylation of a highly functionalized pyranose, such as 2,4-diamino-2,4,6-trideoxy-D-glucopyranose or its equivalents, with *p*-hydroxycinnamic acid derivatives is quite problematic due to a poor nucleophilic nature of the phenol bearing an electron-withdrawing group.⁴ To circumvent this problem, we developed a synthetic plan which involves a new approach for the synthesis of cinnamoyl glycosides (Scheme 1).



Scheme 1

Starting from α -phenyl glycoside I, initial efforts focused on the deoxygenation of C-6 hydroxy group.⁵ Iodination of the resulting phenyl 6-deoxy-glycoside II followed by the Heck reaction with acrylamide containing spermidine moiety would offer a highly convergent tactic to the right subunit of **1**. In our initial attempts to explore this strategy, we focused on the synthesis of cinnamoyl α -D-galactopyranoside (Scheme 2).

Phenyl α -D-galactopyranoside **4** was readily prepared by Lewis acid catalyzed glycosylation of pentaacetate **3** with phenol in 47% yield after recrystallization.⁶ A screen of several iodination agents revealed that cerium (IV)-promoted iodination is an optimal solution.⁷ In practice, treatment of **4** with ceric ammonium nitrate (CAN) and iodine salt (*n*-Bu₄NI) in MeCN at 70 °C for 24 h furnished iodophenyl galactoside **5** in 95% yield. The Heck reaction of **5** with methyl acrylate [Pd(OAc)₂, P(*o*-tol)₃, CH₃CN, 70 °C] proceeded smoothly to furnish α -cinnamoyl galactoside **6** in 88% yield.⁸

We examined the generality of this cinnamoyl glycoside synthesis using a variety of phenyl glycosides (Table 1).



Scheme 2

Ready availability of each phenyl glycoside and good overall yield over two steps make this approach highly acceptable.⁹

With an efficient route to cinnamoyl glycoside established, we turned our attention to the right core synthesis of glycocinnasperimicin D (Scheme 3).

Glycosylation of glucosamine derivative **12** with phenol catalyzed with tin (IV) chloride in CH_2Cl_2 proceeded smoothly to afford a 4:1 anomeric mixture of phenyl glycosides (87%).¹⁰ Following purification of α -anomer **13** by chromatography, hydrolysis of acetyl groups in **13** (Et₃N, MeOH, H₂O) and selective tosylation (TsCl, pyridine) of the triol **14** furnished tosylate **15** (61% in two





 Table 1
 Cinnamoyl Glycoside Synthesis Using a Variety of Phenyl Glycosides







^a CAN (3.0 equiv), *n*-Bu₄NI (1.5 equiv), CH₃CN, 70 °C, 24 h. ^b Methyl acrylate (3.0 equiv), 30 mol% Pd(OAc)₂, 50 mol% P(*o*-tol)₃, Et₃N (5.0 equiv), CH₃CN, 70 °C, overnight.

steps). Acetylation of two hydroxy groups in **15** (Ac₂O, pyridine) followed by the $S_N 2$ displacement (*n*-Bu₄NI, NaI, DME) produced iodide **17** (91% yield over two steps). Reductive hydrogenolysis of **17** (H₂, 10% Pd-C, Et₃N, EtOAc) then furnished 6-deoxy pyranose **18** in good yield (90%). Finally, iodination of phenyl glycoside **18** followed by the Heck reaction with acrylylspermidine successfully constructed the right core structure of glyco-cinnasperimicin D, and the product, 2-amino-6-deoxy- α -D-glucocinnamoyl spermidine **19**, was isolated in 75% yield.¹¹

During these synthetic works, we recognized that iodophenyl glycosides, prepared from readily available phenyl glycosides by high-yielding iodination process, are particularly attractive synthons for the synthesis of substituted phenyl glycosides (Scheme 4). In fact, introduction of 1-dodecyne onto the phenyl ring in **20** was achieved efficiently by the Sonogashira cross-coupling reaction [1-dodecyne, PdCl₂(PPh₃)₂, CuI, Et₃N, benzene] to provide a useful amphilic carbohydrate derivative **21** for the nonionic detergents in 75% yield.¹² This example demonstrates synthetic potential of iodophenyl glycosides.¹³



Scheme 4

In conclusion, a new approach for the synthesis of cinnamoyl glycoside was established, and successful application of this method accomplished the construction of the right core structure found in glycocinnasperimicin D. Furthermore, palladium-catalyzed reaction successfully demonstrated synthetic potential of iodophenyl glycosides.

General Procedure for the Synthesis of Cinnamoyl Glycosides via Iodination-Heck Reaction Sequence

A solution of phenyl galactoside **4** (500 mg, 1.18 mmol), tetra-*n*-butylammonium iodide (654 mg, 1.77 mmol) and CAN (1.94 g, 3.54 mmol) in MeCN (20 mL) was heated at 70 °C for 24 h. The resulting yellow solution was poured into sat. aq NaHCO₃, and the aq layer was extracted with EtOAc. The combined organic extracts were washed with water and brine, dried over anhyd Na₂SO₄, and then concentrated. The resulting residue was purified by silica gel chromatography (stepwise gradient of 25% to 30% EtOAc–hexane) to afford the iodophenyl galactoside **5** (613 mg, 1.11 mmol, 95%) as a white solid.

A dry Schlenk flask was charged with **5** (80 mg, 0.15 mmol), methyl acrylate (40 μ L, 0.44 mmol), tri-*o*-tolylphosphine (18 mg, 0.073 mmol), Et₃N (0.10 mL, 0.73 mmol), and MeCN (2.0 mL). The solution was degassed by two freeze-thaw cycles. After cooling the Schlenk flask at –78 °C, palladium(II) acetate (9.0 mg, 0.040 mmol) was placed on the resulting solidified reaction mixture. The Schlenk flask was evacuated and then heated at 70 °C overnight. After cooling to r.t., the reaction mixture was poured into sat. aq NH₄Cl, and the aq layer was extracted with EtOAc. The combined organic extracts were washed with brine, dried over Na₂SO₄, concentrated, and purified by silica gel chromatography (stepwise gradient of 25% to 30% EtOAc–hexane) to furnish α -cinnamoyl galactoside **6** (65 mg, 88%) as a white crystalline mass.

Acknowledgment

This research was financially supported by a Grant-In-Aid for Scientific Research from the Ministry of Education, Science, Sports and Culture.

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(11) Spectroscopic data of **19**; $[\alpha]_D^{26} = +98.9 (c \ 1.09, CHCl_3)$. ¹H NMR (CDCl₃, 300 MHz): $\delta = 1.17 (d, J = 6.5 Hz, 3 H), 1.40-1.90 (m, 6 H), 1.44 (s, 9 H), 1.48 (s, 9 H), 2.05 (s, 3 H), 2.07 (s, 3 H), 3.08-3.25 (br s, 4 H), 3.25-3.40 (br s, 4 H), 3.64 (s, 3 H), 3.93 (dq, <math>J = 10.5, 6.5 Hz, 1 H$), 4.18 (td, J = 10.5, 3.5 Hz), 4.50-4.70 (br s, 1 H), 4.94 (dd, J = 10.5, 9.5 Hz, 1 H), 5.11 (d, J = 10.0 Hz, 1 H), 5.37 (dd, J = 10.5, 9.5 Hz, 1 H),

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5.54 (d, J = 3.5 Hz, 1 H), 6.37 (d, J = 15.5 Hz, 1 H), 7.04– 7.10 (3 H), 7.44–7.52 (2 H), 7.57 (d, J = 15.5 Hz, 1 H). ¹³C NMR (CDCl₃, 75 MHz): $\delta = 17.1$, 20.47, 20.53, 25.5, 27.2, 27.5, 28.19, 28.23, 35.6, 39.9, 43.2, 46.5, 52.3, 53.9, 66.3, 70.7, 73.1, 79.0, 79.7, 95.6, 116.4, 120.2, 129.2, 129.8, 139.4, 156.0, 156.5, 157.0, 166.1, 169.7, 171.2.

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