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Research Article

Asymmetric epoxidation of digeranyl by cultured cells of *Nicotiana tabacum*

Osamu Nakagawa, Kei Shimoda, Sunsuke Izumi and Toshifumi Hirata*

Department of Mathematical and Life Sciences, Graduate School of Science, Hiroshima University, 1-3-1 Kagamiyama, Higashi-Hiroshima 739-8526, Japan

Summary

Asymmetric epoxidation of digeranyl, which is a squalene analog, with cultured cells of *Nicotiana tabacum* was investigated. Feeding of [8- 3 H]-digeranyl into the cultured cells of *N. tabacum* resulted in the formation of (3*S*)-2,3-epoxydigeranyl and 6,7-epoxydigeranyl. It was found that the epoxidation of digeranyl with *N. tabacum* was highly stereoselective. Copyright © 2003 John Wiley & Sons, Ltd.

Key Words: Asymmetric epoxidation; Biotransformation; Digeranyl; Cultured cells of *Nicotiana tabacum*

Introduction

The epoxy group plays an important role in organic synthesis,^{1–4} with the development of asymmetric epoxidation being particularly noteworthy.^{5–8} However, many asymmetric epoxidation reactions can only be performed under restricted conditions, e.g. by using organometallic reagents, acting under rigorous exclusion of air, and act only when specific substituents are present in the substrates.

*Correspondence to: Professor T. Hirata, Department of Mathematical and Life Sciences, Graduate School of Science, Hiroshima University, 1-3-1 Kagamiyama, Higashi- Hiroshima 739-8526, Japan. E-mail: thirata@sci.hiroshima-u.ac.jp

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Recently, biological transformations has enabled the accomplishment of asymmetric epoxidations. Such studies revealed that several bacteria, such as *Pseudomonas* and *Corynebacterium*, transformed stereoselectively the unsaturated hydrocarbons into the corresponding epoxides. Ne also found that plant cultured cells, such as *Nicotiana tabacum* and *Catharanthus roseus*, have the ability to introduce enantioselectively an epoxy group into monoterpenes. Asymmetric epoxidation of squalene into (3*S*)-2,3-epoxysqualene is one of the most fascinating reactions, because epoxysqualene is a key precursor for the syntheses of steroids and triterpenoids. If such a reaction can be adapted to squalene analogs, it may be extremely useful from the viewpoint of formation of many cyclic compounds.

Here we have investigated the epoxidation of digeranyl (1), which is a C_{20} squalene analog, with the cultured cells of *N. tabacum*.

Results and discussion

Biotransformation of digeranyl with cultured cells of N. tabacum

[8- 3 H]-Digeranyl (1) was administered to the cultured suspension cells of *N. tabacum*, and the cultures were incubated at 25°C for 2 days. Two products were found by **TLC** autoradiography and **HPLC** analyses of the ether extract of the reaction mixture in 15.4 and 13.6% yields. These products were identified as 2,3-epoxydigeranyl (2) and 6,7-epoxydigeranyl (3), respectively, by comparison of their TLC and GLC with the synthetic specimens. Identification of these products was confirmed by GC-MS analyses of the products in the incubation of non-labeled digeranyl with the cultured cells of *N. tabacum*. The absolute configuration and enantiomeric purity of the 2,3-epoxydigeranyl obtained by the biotransformation were determined by comparison of the chiral GLC with those of synthetic (3*S*)-2,3-epoxydigeranyl and (*RS*)-2,3-epoxydigeranyl. The stereochemistry of the 2,3-epoxydigeranyl was shown to be S in >99% ee (Scheme 1).

Thus, it was found that the cultured cells of *N. tabacum* are capable of transforming digeranyl into 2,3-epoxy- and 6,7-epoxydigeranyls stereoselectively.

Scheme 1. Biotransformation of digeranyl by cultured cells of N. tabacum

Scheme 2. Reagents and conditions: (a) AC_2O , Pyridine, DMAP, rt; (b) PYDZ Ligand, K_2OsO_4 , $K_3Fe(CN)_6$, K_2CO_3 , Methanesulfonamide, t-BuOH: H_2O , 4 °C; (c) 2,2-dimethoxypropane, PPTS, CH_2Cl_2 , rt; (d) Pd(PPh₃)₄, TsNa-4 H_2O , THF, rt; (e) n-BuLi, THF, 0 °C; (f) PdCl₂(dppp), LiHBEt₃, THF, 0 °C; (g) TsOH, MeOH, rt; (h) MsCl, TEA, CH_2Cl_2 –40 °C; (i) K_2CO_3 , MeOH, rt

Synthesis of authentic (3S)-2,3-epoxydigeranyl

(3*S*)-2,3-Epoxydigeranyl (**2a**) was synthesized from geraniol (**4**) as shown in Scheme 2. The asymmetric dihydroxylation of geranyl acetate (**5**) using PYDZ ligand, ¹⁹ K_2OsO_4 , K_2CO_3 and $K_3Fe(CN)_6$ in 1:1 t-BuOH/H₂O gave a diol **6** in 70% yield. Protection of the diol **6** followed by the sulfonylation of the resultant **7** with Pd(PPh₃)₄, TsNa-4H₂O in THF: MeOH furnished geranyl p-tolylsulfone (**8**). The product

8 was lithiated, followed by addition of geranyl bromide (9), which was prepared using N-bromosuccinimide and dimethyl sulfide in CH_2Cl_2 , to give **10** in 60% yield. **10** was reduced with $PdCl_2(dppp)$ and $LiHBEt_3$, in THF to give **11** in 79% yield. **11** was deprotected to give a diol **12** with TsOH in MeOH. The resulting diol **12** was converted to (3S)-2,3-epoxydigeranyl (**2a**) by sulfonylation of the secondary alcohol with methanesulfonyl chloride and pyridine followed by ring closure with K_2CO_3 in MeOH in 25% yield over two steps.

Thus, the synthesis of (3S)-2,3-epoxydigeranyl (2a) from geraniol (4) was achieved by asymmetric dihydroxylation, coupling of geranyl bromide (9) with geranyl p-tolylsulfone derivative (8), and reductive desulfonylation, as key steps.

Experimental

General: Analytical TLC was performed on precoated TLC plates (Merck 60 F₂₅₄ Column chromatography was conducted using silica-gel (Merck Silica gel 60, and Wakogel C-300 HG). ¹H and ¹³C NMR spectra were obtained using a JEOL JNM-LA500 spectrometer using tetramethylsilane as an internal standard. Mass spectra (MS) were obtained on JEOL JMS-SX102A and Hewlett Packard MSD-5971A mass spectrometers.

Substrates: The preparation of [8-³H]-digeranyl (1) was assayed by the synthetic method of [8-²H]-digeranyl reported in our previous paper. To a solution of 8-p-tolylsulfonated digeranyl (500 mg, 0.583 mmol), prepared from geranyl p-tolylsulfone and geranyl chloride, and 5 mol% 1,3-bis(diphenylphosphino)propane palladium(II) chloride complex [PdCl₂(dppp)] in THF (40 ml), LiB³HEt₃ (1.17 mmol in 2.3 ml THF; 1.48 GBq) was added at 0°C under a N₂ atmosphere. After stirring at 0°C for 1 h, 3 M NaOH and KCN (40 mg) was added to the reaction mixture, and then the mixture was extracted with hexane to give the crude product. The latter was subjected to column chromatography on silica gel and eluted with hexane to give [8-³H]-digeranyl (1) (145 mg, 670 MBq, 90%).

Feeding of [8-³H]-digeranyl (1) with the cultured cells of *N. tabacum*: The suspension cells of *N. tabacum* were prepared as reported previously.²¹ To the flask containing the suspension cells (about 40 g in 100 ml of Murashige and Skoog's medium²¹), [8-³H]-digeranyl (1) (58 µg; 131 kBq) was administered and incubated at 25°C for 2 days on a

rotary shaker. After incubation, the cells were collected by filtration and extracted with ether. The ether layer was evaporated and then subjected to short-column chromatography on silica gel with ether to give a crude product (36 kBq)

The crude product was traced by TLC autoradiography. A charged TLC was developed with hexane:EtOAc = 5:1 (v/v) as eluent and then analyzed using Fuji.BAS2000 imaging analyzer. The $R_{\rm f}$ values of the radioactive spots were 0.51, 0.57, and 0.65, which were identical with those of authentic 2,3-epoxydigeranyl, 6,7-epoxydigeranyl, and digeranyl, respectively. A part of the crude product was subjected to preparative HPLC (Inertsil SIL 5pm; 4.6×250 mm) using hexane:2-propanol=1000:1 (v/v) as eluent and the radioactivities of the isolated fractions were measured by liquid scintillation counting. Three radioactive fractions eluted at $R_{\rm t}$ 10 (12 Bq; 71%), 16 (2.6 Bq; 15.4%), and 19 min (2.3 Bq; 13.6%) were identical with those of authentic digeranyl, 6,7-epoxydigeranyl, and 2,3-epoxydigeranyl, respectively.

Feeding of digeranyl (1) with the cultured cells of N. tabacum: To the flask containing the suspension cells (about 40 g in 100 ml of MS medium), non-labeled digeranyl (25 mg) was administered and the mixture was incubated at 25°C for 2 days on a rotary shaker. After incubation, the cells were collected by filtration and extracted with ether. The ether layer was evaporated and then subjected to shortcolumn chromatography on silica gel with ether to give a crude product (2 mg). Two products were identified by the comparison of their GLC [OV-17 on Chromosorb WAW-DMCS; 80-100 mesh; column temperature., 100–250°C (10°C/min, initial. time 5 min); inject temparature., 250°C] and GC-MS analysis with those of authentic 2,3- and 6,7-epoxydigeranyl. Retention times for synthetic 2,3- and 6, 7-epoxydigeranyl in the GLC were 18.4 and 18.9 min, respectively. GC-MS analyses of the peaks with retention times 18.4 and 18.9 min in the administered sample with N. tabacum showed identical fragmentation patterns to synthetic 2,3- and 6,7-epoxydigeranyl, respectively.

The absolute configuration and enantiomeric purity of the 2,3-epoxy product were determined by comparison of their chiral GLC (CP cyclodextrin β 236M-19 column; column temperature, 200°C; inject temperature, 220°C) of incubated products with those of synthetic samples, (3S)-2,3-epoxydigeranyl and racemic 2,3-epoxydigeranyl [R_t values of (3S)- and (3R)-epoxydigeranyl: 17.3 and 17.5 min, respectively].

Syntheses of racemic 2,3- and 6,7-epoxydigeranyl: Digeranyl (500 mg, 1.82 mmol) was dissolved in 52 ml of dry CH₂Cl₂. m-Chloroperbenzoic acid (377 mg, 2.18 mmol) was added to the stirred reaction mixture at 0°C. After stirring at room temperature for 12h, aqueous Na₂S₂O₃ and NaHCO₃ were added at 0°C. The reaction mixture was extracted with ether to give a crude product, which was purified by column chromatography on silica-gel with hexane-ether (95:5) to give 2.3epoxydigeranyl (2) (4.1 mg, 7%) [EI-MS m/z 290 (M⁺), 275, 166, 153, 135, 109, 95, 69; ¹H NMR (CDC1₃) $\delta = 5.14$ (m, 3H), 2.70 (t, 1H, J = 6.2 Hz), 2.07 (m, 10H), 1.68 (s, 3H), 1.64 (m, 2H), 1.62 (s, 3H), 1.60 (s, 6H), 1.30 (s, 3H) and 1.26 (s, 3H); 13 C NMR (CDC1₃) $\delta = 135.2$, 134.2, 131.3, 125.0, 124.4, 124.1, 64.2, 58.3, 39.7, 36.3, 28.3, 28.2, 27.5, 26.8, 25.7, 24.9, 18.7, 17.7 and 16.0] and 6,7-epoxydigeranyl (3) (9.4 mg, 16%) [EI-MS m/z 290 (M⁺); ¹H NMR (CDC1₃) $\delta = 5.14$ (m, 3H), 2.71 (t, 1H, J = 6.3 Hz), 2.07 (m, 8H), 1.68 (s, 6H), 1.64 (m, 2H), 1.61 (s, 9H),1.51 (m, 1H), 1.42 (m, 1H) and 1.25 (s, 3H); ¹³C NMR (CDC1₃) $\delta = 135.9, 131.8, 131.4, 124.2, 123.7, 123.3, 63.3, 60.7, 39.7, 38.9, 29.0,$ 26.7, 25.7, 24.8, 23.9, 17.6, 16.5, 16.0 and 15.3].

Synthesis of (3S)-2,3-epoxydigeranyl. (6R)-6,7-Dihydroxygeranyl acetate (6): Following the method described, 23 a mixture of DHQD-PYDZ ligand¹⁹ (190 mg, 0.26 mmol), K₂OsO₄ (20 mg, 0.05 mmol), K₂Fe(CN)₆ (25 g, 76.5 mmol), K₂CO₃ (10.6 g, 76.5 mmol), methanesulfonamide (2.42 g, 25.5 mmol), geranyl acetate (5 g, 25.5 mmol) and 300 ml of 1:1 t-BuOH-H₂O was stirred for 12 h at 4°C. Sodium sulfite (30 g) was added, and the mixture was concentrated under reduced pressure. The mixture was extracted with CH₂Cl₂, washed with 2 M KOH and dried over Na₂SO₄. The crude product was purified by column chromatography on silica gel using ethyl acetate–hexane (2:1) as eluent to give a product **6** (2.6 g, 70%): EI-MS m/z 231 (MH⁺); ¹H-NMR (CDCl₃) $\delta = 5.33$ (t, 1H, J = 7.0 Hz), 4.53 (d, 2H, J = 7.0 Hz), 3.27 (dd, 1H, J = 10.5, 2.0 Hz), 2.27 (m, 1H), 2.04 (m, 1H), 2.00 (s, 3H), 1.66 (s, 3H), 1.55 (m, 1H), 1.39 (m, 1H), 1.15 (s, 3H) and 1.10 (s, 3H); ¹³C-NMR (CDCl₃) δ = 171.2, 142.0, 118.5, 77.9, 72.9, 61.3, 36.5, 29.4, 26.3, 23.1, 20.9 and 16.3.

(5R)-3-Methyl-5(2,2,5,5-tetramethyl-[1,3]dioxolan-4-yl)-pent-2-enyl acetate (7): To the suspension of **6** (4.8 g, 20.8 mmol) in 20 ml of CH₂Cl₂, 2,2-dimethoxypropane (4.8 g, 41.7 mmol) and pyridinium p-toluenesulfonate (10.3 g, 41.7 mmol) were added at 0°C under a N₂ atmosphere. After stirring at room temperature for 1 h, the reaction mixture was treated with Et₃N and concentrated. The crude product

was purified by column chromatography on silica-gel with hexane-ethyl acetate (2 : 1) to give a product 7 (5.3 g, 94%): EI-MS m/z 270 (M⁺); ¹H-NMR (CDCl₃) δ = 5.39 (bt, 1H), 4.59 (d, 2H), 3.65 (dd, 1H, J = 9.7, 3.4 Hz), 2.28 (m, 1H), 2.10 (m, 1H), 2.04 (s, 3H), 1.73 (s, 3H), 1.64 (m, 3H), 1.51 (m, 3H), 1.40 (s, 3H), 1.31 (s, 3H), 1.24 (s, 3H) and 1.09 (s, 3H); ¹³C-NMR (CDCl₃) δ = 170.4, 141.0, 118.6, 106.2, 82.4, 79.7, 60.9, 36.3, 28.3, 27.1, 26.5, 25.7, 22.7, 20.6 and 16.2.

(5R)-2,2,4,4-Tetramethyl-5-[3-methyl-5-(toluene-4-sulfonyl)-pent-3envl-[1, 3]dioxolane (8): To the solution of sodium p-tolylsulfonate tetrahydrate (5.8 g, 23.2 mmol) in THF (43 ml) and MeOH (14 ml) was added acetate 7 (5.7 g, 21.1 mmol) and 5 mol% Pd(PPh₃)₄ in THF (10 ml) under a N₂ atmosphere. After stirring at room temperature overnight, aqueous KCN was added dropwise at 0°C to deactivate the palladium catalyst. The reaction mixture was extracted with ether to give a crude product, which was purified by column chromatography on silica gel using hexane-ethyl acetate (4:1) as eluent to give a product 8 (5.3 g, 68%): FAB-MS m/z 367.8 (MH⁺); ¹H-NMR (CDCl₃) $\delta = 7.73$ (d, 2H, J=8.2 Hz), 7.32 (d, 2H, J=8.2 Hz), 5.24 (t, 1H, J=7.9 Hz), 3.80(d, 2H, J = 7.9 Hz), 3.63 (dd, 1H, J = 9.7, 3.0 Hz), 2.44 (s, 3H), 2.25 (m, 1H), 2.06 (m, lH), 1.55 (m, lH), 1.41 (s,3H), 1.40 (m, lH), 1.39 (s, 3H), 1.32 (s, 3H), 1.24 (s, 3H) and 1.08 (s, 3H); 13 C-NMR (CDC1₃) $\delta = 145.0$, 143.9, 135.6, 129.1, 127.9, 110.5, 106.0, 82.1, 79.5, 55.5, 36.3, 28.1, 27.0, 26.4, 25.6, 22.5, 21.1 and 15.8.

2,2,4,4-Tetramethyl-5-[3,8,12-trimethyl-5-(toluene-4-sulfonyl)-trideca-3,7, 11-trienyl-[1,3]dioxolane (10): To the suspension of 8 (5.3 g, 14.5 mmol) in 20 ml of THF, 1.5 M n-BuLi (9.1 ml, 17.4 mmol) in THF and geranyl bromide (9) (3.0 g, 17.4 mmol) in THF (10 ml) were added dropwise at 0°C under a N₂ atmosphere. After stirring at 0°C for 10 min, the reaction mixture was treated with 20 ml of 10% NH₄C1 and extracted with ether to give a crude product, which was purified by column chromatography on silica-gel with hexane-ethyl acetate (4:1) to give a product 10 (4.3 g, 60%).

2,2,4,4-Tetramethyl-5-[3,8,12-trimethyltrideca-3,7,11-trienyl-[1,3] dioxolane (11): To the solution of 10 (2.6 g, 5.2 mmol) and 5 mol% $PdC_{12}(dppp)$ in THF (173 ml), LiHBEt₃, (1.2 ml, 10.2 mmol) was added at 0°C under N_2 atmosphere. After stirring at 0°C for 12 h, 3 M NaOH and KCN (50 mg) was added to the reaction mixture, and then the mixture was extracted with ether to give a crude product. The crude product was subjected to column chromatography on silica-gel and eluted with hexane-ethyl acetate (5:l) to give 11 (1.4 g, 79%): EI-MS m/z

348 (M⁺); ¹H NMR (CDC1₃) δ = 5.13 (m, 3H), 3.67 (dd, 1H, J = 9.3, 3.5 Hz), 2.12 (m, 1H), 1.68 (s, 3H), 1.62 (s, 3H), 1.60 (s, 6H), 1.48 (m, 2H), 1.42 (s, 3H), 1.33 (s, 3H), 1.24 (s, 3H) and 1.10 (s, 3H).

(3R)-Dihydroxydigeranyl (12): To the solution of 11 (23 mg, 66 μmol) in MeOH (2 ml) was added TsOH (500 mg, 79.2 μmol). After stirring at room temperature for 3 h, 20 ml of 5% NaHCO₃ was added, and washed with brine. The reaction mixture was extracted with ether to give a crude product, which was purified by column chromatography on silica gel using hexane–ethyl acetate (3:1) as eluent to give a product 12 (25 mg, quant): EI-MS m/z 308 (M⁺); ¹H-NMR (CDC1₃) δ = 5.13 (m, 3H), 3.34 (dd, lH, J=10.5, 1.7 Hz), 2.25 (m, lH), 2.02 (m, 9H), 1.68 (s, 3H), 1.62 (s, 3H), 1.60 (s, 6H), 1.55 (m, lH), 1.41 (m, lH), 1.19 (s, 3H) and 1.15 (s, 3H); ¹³C-NMR (CDCl₃) δ = 135.1, 134.9, 131.1, 124.9, 124.3, 124.1, 78.2, 73.0, 39.7, 36.8, 29.8, 28.2, 28.1, 26.7, 26.3, 25.6, 23.1, 17.6, 16.0 and 15.9.

(3S)-2,3-epoxydigeranyl (2a): A solution of 12 (63 mg, 0.20 mmol) and pyridine (0.03 ml, 0.40 mmol) in 3 ml of CH₂Cl₂ was treated at -40°C with methanesulfonyl chloride (34 mg, 0.30 mmol). After 30 min, the mixture was warmed to room temperature and stirred for 3 h. An additional 0.3 ml pyridine was added, and the mixture was stirred for 8 h. The mixture was poured into a suspension of 1 g of K₂CO₃ in 5 ml of MeOH and stirred for 6h at room temperature. The mixture was concentrated, diluted with water, extracted with ether, washed with 10% CuSO₄ and brine. The crude product was purified by column chromatography on silica gel using hexane-ether (95:5) as eluent to give a product **2a** (14.8 mg, 25%): EI-MS m/z 290 (M⁺), 275, 166, 153, 135, 109, 95, 69; ¹H NMR (CDC1₃) $\delta = 5.14$ (m, 3H), 2.70 (t, 1H, J =6.2 Hz), 2.07 (m, 10H), 1.68 (s, 3H), 1.64 (m, 2H), 1.62 (s, 3H), 1.60 (s, 6H), 1.30 (s, 3H) and 1.26 (s, 3H); 13 C NMR (CDCl₃) δ = 135.2, 134.2, 131.3, 125.0, 124.4, 124.1, 64.2, 58.3, 39.7, 36.3, 28.3, 28.2, 27.5, 26.8, 25.7, 24.9, 18.7, 17.7 and 16.0.

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References

- 1. Bartok M, Lang KL. In The Chemistry of Ethers, Crown Ethers, Hydroxyl Groups and Their Sulphur Analoges, Supplement E, Vol 2, Patai, S. (ed.), Wiley: New York, 1980; 609.
- 2. Chong JM, Sharpless KB. J Org Chem 1985; **50**: 1569.
- 3. Smith JG, Synthesis 1984; 629.
- 4. Rao AS, Paknikar SK, Kirtane JG. Tetrahedron 1983; 39: 2323.
- 5. Katsuki T, Sharpless KB. J Am Chem Soc 1980; **102**: 5874.
- 6. Zhang W, Loebach JL, Wilson SR, Jacobsen EN. J Am Chem Soc 1990; **112**: 2801.
- 7. Julia S, Masana J, Vega JC. Angew Chem Int Ed Engl 1980; 19: 929.
- 8. Yang D, Yip YC, Tang MW, Wong MK, Zheng JH, Cheung KK. J Am Chem Soc 1996; 118: 491.
- 9. Buhler M, Schindler J. In *Biotechnology*, Vol 6a, Rehm H-J, Reed G (eds). Verlag Chemie: Berlin, 1984; 329.
- 10. May SW, Schwartz RD. J Am Chem Soc 1974; 96: 4031.
- 11 Ohta H, Tetsukawa H. J Chem Soc Chem Commun 1978; 849.
- 12. Smet MJ, Witholt B, Wynberg H. J Org Chem, 1981; 46: 3128.
- 13. Smet MJ, Kingma J, Wynberg H, Witholt B. Enzyme Microb Technol 1983; **5**: 352 .
- 14. Habets-Crutzen AQH, Carlier SJN, Bont JAM, et al. Enzyme Microb Technol 1985; 7: 17
- 15. Hirata T, Izumi S, Kido T, Shimoi Y, Ikeda Y. Plant Tissue Culture Lett 1993: **10**: 89.
- 16. Hirata T, Ikeda Y, Izumi S, Shimoda K, Hamada H, Kawamura T Phytochemistry 1994; 37: 401.
- 17. Hirata T, Izumi M, Ogura M, Yawata T, Tetrahedron 1998; 54:15993.
- 18. Abe I, Prestwich PD. In Comprehensive Natural Products Chemistry, Vol 2 Barton SD, Nakanishi, K (eds). Elsevier: Amsterdam, 1999, 267.
- 19. Corey EJ, Noe MC, Sarshar S. J Am Chem Soc 1993; 115: 3828.
- 20. Nakagawa O, Shimoda K, Izumi S, Hirata T. J Label Compd. Radiopharm 2000; **43**: 1301.
- 21. Lee YS, Hirata T, Suga T. J Chem Soc Perkin Trans 1983; 1:2475.
- 22 Murashige T, Skoog F. Physiol Plant 1962; 15: 473.
- 23. Corey EJ, Noe MC, Shieh WC. Tetahedron Lett 1993; 34: 5995.