Chem. Pharm. Bull. Note

Concise synthesis of hydroxy β -methyl fatty acid ethyl esters

Yuki Sakai,^a Yukiko Asakura,^a Mitsuhiro Morita^b and Takashi Takahashi^a*

^aDepartment of Pharmaceutical Science, Yokohama University of Pharmacy, 601 Matano-cho, Totsuka-ku, Yokohama, Kanagawa 245-0066, Japan; ^bDepartment of Biology, Kobe University Graduate School of Science, Kobe, Japan.

* E-mail address: ttak@yok.hamayaku.ac.jp

Summary

Hydroxy β -methyl fatty acid ethyl esters bearing different carbon chain lengths and varying hydroxyl group positions were successfully synthesized from symmetric diols. These fatty acid derivatives are useful intermediates of chemical probes for metabolic analyses of fatty acid.

Keywords: β-methyl fatty acid, chemical synthesis, Johnson-Claisen rearrangement, hydroxy fatty acid

Fatty acids are hydrocarbon compounds that contain various lengths of saturated carbon chains, unsaturated carbon chains, and substituted carbon chains. Fatty acids and their derivatives (including phospholipids and glycerolipids) are widely distributed in living organism and have various roles as structural and metabolic substrates.¹⁾ More specifically, methyl-substituted fatty acids are widespread in both prokaryotic and eukaryotic cells. For example, phytanic acid (1) (Figure 1), a tetramethyl-substituted fatty acid derived from phytol, is substituted at the β -position by one of the four methyl groups.²⁾ Although fatty acids are generally metabolized by β -oxidation, the presence of a methyl group on the β -position inhibits β -oxidation, and so metabolism via α -oxidation dominates. In terms of their biological influence, the accumulation of methyl-substituted fatty acids can cause a neurotic disorder known as Refsum disease. This disease results from a deficiency in peroxisomal α -oxidation enzymes, and manifests an accumulation of **1**.³⁾ The suppression of β -oxidation by a methyl group can also be employed for molecular imaging, as probe molecules can accumulate in the target tissue.⁴⁾ More specifically, a β -methyl palmitic acid derivative is an analog of $[^{123}I]$ -BMIPP (**2**, $[^{123}I]$ - β -methyl-p-iodophenyl pentadecanoic acid), which is a single-photon emission computed tomography (SPECT) probe for myocardial imaging.⁵⁾ Furthermore, β -methyl fatty acids substituted with hydroxyl groups have been reported as useful intermediates for positron emission tomography (PET) tracer candidates $(3^{6)}, 4^{7}$) for myocardial imaging. Based on this importance of hydroxy β -methyl fatty acids as valuable chemical probes in metabolic analyses, we herein report the concise synthesis of hydroxy β-methyl fatty acid ethyl esters bearing different carbon chain lengths and varying hydroxyl group positions.

Results and discussion

The synthesis of β -methylated fatty acid derivatives was carried out from the symmetric diols 1,6-hexanediol (**5a**) and 1,12-dodecandiol (**5b**) (see Chart 1). Initially, a single hydroxyl group from each diol was converted into the corresponding benzyl ether under basic conditions to give **6a** and **6b** in moderate yields, while the remaining hydroxyl group was efficiently converted to an aldehyde by a (2,2,6,6-tetramethyl-piperidin-1-yl)oxyl (TEMPO)-mediated oxidation reaction.⁸⁾ The resulting aldehydes (**7a**, **7b**) were then converted to allylic alcohols in good yields using propenylmagnesium bromide at -60 °C, and the resulting allylic alcohols (**8a**, **8b**) were heated under reflux in the presence of triethyl orthoacetate and catalytic amounts of pivalic acid to give the β -methylated γ , δ -unsaturated ethyl esters.⁹⁾ Finally, the γ , δ -unsaturated ethyl esters were reduced and the benzyl ether

Chemical and Pharmaceutical Bulletin Advance Publication

groups were removed under hydrogenation conditions to yield the desired ω -1-hydroxy- β -methyl fatty acids (**9a**, **9b**) in good yields.

Subsequently, the corresponding ω -7-hydroxy- β -methyl fatty acids were synthesized from alcohols **9a** and **9b**, as depicted in Chart 2. This was achieved through the conversion of the hydroxyl groups of **9a** and **9b** to their corresponding aldehydes (**10a**, **10b**) by TEMPO oxidation, and subsequent treatment of the aldehydes with hexyl magnesium bromide at -60 °C to give the desired products **11a** and **11b** in moderate to good yields.¹⁰

Using the above-described synthetic method, hydroxy β -methyl fatty acids containing different carbon chain lengths and varying hydroxyl group positions were successfully synthesized. We expect that the obtained fatty acids will be suitable precursors for the preparation of PET/SPECT tracer candidates, in addition to being valuable chemical probes for use in chemical biology investigations. Investigations into the development of chemical probes fatty acid metabolism studies are currently underway.

Experimental

General Methods and Materials

¹H and ¹³C NMR spectra were recorded on a JEOL ECA-500 instrument. Chemical shifts are reported in parts per million (ppm) relative to a tetramethylsilane internal standard (0.0 ppm) or to the solvent peak (CDCl₃, δ_H 7.26 ppm, δ_C 77.1 ppm). ¹H NMR data are reported as follows: Chemical shift (δ ppm), integration, multiplicity, and coupling constant (J, Hz). ¹³C NMR data are reported as follows: chemical shift (δ ppm), multiplicity, and coupling constant (Hz, where applicable). Multiplicities are reported using the following abbreviations: s, singlet; d, doublet; t, triplet; q, quartet; quint, quintet; sp, septet; m, multiplet; and br, broad. Only the strongest and/or structurally relevant IR peaks are reported (cm⁻¹). All mass spectra were measured on a JEOL JMS-700 MStation mass spectrometer. Column chromatography was performed using silica gel (CHROMATOREX PSQ 100B, Fuji Silysia chemical Ltd.). Analytical thin layer chromatography was performed on glass plates pre-coated with silica gel (Merck, Kieselgel 60 F₂₅₄, 0.25 mm).

Synthetic procedures

6-(Benzyloxy)hexan-1-ol (6a)

To a stirred solution of 1,6-hexanediol (**5a**, 5.91 g, 50.0 mmol) in tetrahydrofuran (THF, 200 mL) at 0 °C was added NaH (2.00 g, 50.0 mmol). After 20 min, BnBr (5.20 mL, 43.8 mmol) was added, and the reaction mixture heated under reflux for 5 h. After this time,

Chemical and Pharmaceutical Bulletin Advance Publication

a saturated aqueous NH₄Cl solution was added, and the reaction mixture was extracted with ethyl acetate. The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated in vacuo, and the resulting crude product was purified by silica gel column chromatography (hexane/ethyl acetate, 3:1–1:1) to afford **6a** as a clear colorless oil (4.96 g, 23.8 mmol, 54% yield based on BnBr). **6b** (white wax, 8.94 g, 30.5 mmol, 44%) was synthesized from 1,12-dodecanediol (**5b**, 14.2 g, 70.0 mmol) according to the above procedure. The spectroscopic data obtained for **6a** and **6b** corresponded with literature values.^{11, 12})

6-(Benzyloxy)hexanal (7a)

To a stirred solution of **6a** (4.96 g, 23.8 mmol) in CH₂Cl₂ (120 mL) at 0 °C were added iodobenzene diacetate (11.5 g, 35.7 mmol) and TEMPO (743 mg, 4.76 mmol). After warming to 20 °C and stirring for 15 h, saturated aq. Na₂S₂O₃ was added, and the resulting mixture was extracted with CH₂Cl₂. The combined organic layer was then washed with saturated aq. NaHCO₃ and brine, dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated in vacuo, and the resulting crude product was purified by silica gel column chromatography (hexane/ethyl acetate, 1:0–5:1) to afford **7a** as a clear colorless oil (4.52 g, 21.9 mmol, 92%). **7b** (white wax, 3.50 g, 12.1 mmol, 93%) was synthesized from **6b** (3.80 g, 13.0 mmol) according to the above procedure. The spectroscopic data obtained for **7a** and **7b** corresponded with literature values.^{11, 13}

(EZ)-9-(Benzyloxy)non-2-en-4-ol (8a)

To a stirred solution of **7a** (3.50 g, 17.0 mmol) in THF (85.0 mL) was added propenylmagnesium bromide (0.5M in THF, 40.8 mL, 20.4 mmol) at -60 °C. After stirring for 4 h, a 1N HCl solution was added, and the reaction mixture was extracted with ethyl acetate. The combined organic layer was washed with 1N HCl and brine, dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated in vacuo, and the resulting crude product was purified by silica gel column chromatography (hexane/ethyl acetate, 1:0–3:1) to afford **8a** as a clear colorless oil (3.64 g, 14.7 mmol, 86%) and as an EZ mixture. **8b** (pale yellow oil, 2.59 g, 7.79 mmol, 64%) was synthesized from **7b** (3.50 g, 12.1 mmol) according to the above procedure. Data for **8a**: ¹H-NMR (CDCl₃) δ : 1.27–1.69 (11H, m), 3.46 (2H, t, J = 6.0 Hz), 4.01 (0.5H, q, J=6.5 Hz), 4.44 (0.5H, q, J=6.5 Hz), 4.49 (2H, s), 5.35–5.40 (0.5H, m), 5.43–5.49 (0.5H, m), 5.52–5.67 (1H, m), 7.25–7.35 (5H, m). ¹³C-NMR (CDCl₃) δ : 13.26, 17.62, 25.10, 25.26, 26.08, 26.11, 29.63, 37.15, 37.31, 67.21, 70.29, 72.78,

72.98, 126.15, 126.69, 127.42, 127.56, 128.28, 133.52, 134.28, 138.57. IR (ATR) cm⁻¹: 3403, 2935, 2858, 1453. FAB-MS m/z: 231.1774 (Calcd for $C_{16}H_{25}O_2-H_2O$: 231.1743). Data for **8b:** ¹H-NMR (CDCl₃) δ : 1.27–1.70 (23H, m), 3.46 (2H, t, J=6.0 Hz), 4.02 (0.6H, q, J=6.5 Hz), 4.46 (0.4H, q, J=6.0 Hz), 4.50 (2H, s), 5.36–5.41 (0.4H, m), 5.44–5.50 (0.6H, m), 5.53–5.68 (1H, m), 7.27–7.35 (5H, m). ¹³C-NMR (CDCl₃) δ : 13.30, 17.68, 25.29, 25.46, 26.14, 29.44, 29.54, 29.72, 37.24, 37.40, 67.33, 70.47, 72.79, 73.14, 126.15, 126.70, 127.42, 127.58, 128.30, 133.57, 134.34, 138.62. IR (ATR) cm⁻¹: 3364, 2923, 2852, 1454. FAB-MS m/z: 315.2712 (Calcd for $C_{22}H_{37}O_2-H_2O$: 315.2682).

Ethyl 10-hydroxy-3-methyldecanoate (9a)

To a stirred solution of 8a (3.64 g, 14.7 mmol) in triethyl orthoacetate (15.0 mL) was added pivalic acid (300 mg, 3.00 mmol) and the reaction mixture heated to reflux. After stirring for 19 h, the reaction mixture was concentrated in vacuo to give the crude target ethyl ester, which was used in the following stage without further purification. Thus, to a stirred solution of this crude mixture in ethanol (45.0 mL) was added Pd/C (10%, 2.40 g, 2.60 mmol). After stirring under an H₂ atmosphere for 22 h at 20 °C, the resulting mixture was filtered through a celite pad and concentrated in vacuo. The crude product was purified by silica gel column chromatography (hexane/ethyl acetate, 1:0-4:1) to afford 9a as a clear colorless oil (2.54 g, 11.0 mmol, 75% over 2 steps). 9b (clear colorless oil, 1.68 g, 5.34 mmol, 68%) was synthesized from 8b (2.59 g, 7.79 mmol) according to the above procedure. Data for **9a**: ¹H-NMR (CDCl₃) δ: 0.93 (3H, d, J=6.0 Hz), 1.24–1.36 (13H, m), 1.57 (2H, quint. J=7.0 Hz), 1.89-1.99 (1H, m), 2.07-2.12 (1H, m), 2.26-2.31 (1H, m), 3.64 (2H, t, J=6.5 Hz), 4.13 (2H, q, J=7.0 Hz). ¹³C-NMR (CDCl₃) δ: 14.23, 19.67, 25.65, 26.75, 29.32, 29.62, 30.29, 32.71, 36.61, 41.88, 60.06, 62.94, 173.40. IR (ATR) cm⁻¹: 3456, 2855, 1734, 1463. FAB-MS m/z: 231.1989 (Calcd for C₁₃H₂₇O₃: 231.1955). Data for 9b: ¹H-NMR (CDCl₃) δ: 0.93 (3H, d, J=6.0 Hz), 1.24–1.37 (25H, m), 1.57 (2H, quint. J=7.0 Hz), 1.89–1.99 (1H, m), 2.07–2.11 (1H, m), 2.27–2.31 (1H, m), 3.64 (2H, t, J=6.5 Hz), 4.13 (2H, q, J=7.0 Hz). ¹³C-NMR (CDCl₃) δ: 14.26, 19.69, 25.71, 26.88, 29.40, 29.60, 29.72, 30.34, 32.78, 36.70, 41.93, 60.05, 63.05, 173.43. IR (ATR) cm⁻¹: 3369, 2853, 1736, 1464. FAB-MS m/z: 315.2919 (Calcd for C₁₉H₃₉O₃: 315.2894).

Ethyl 3-methyl-10-oxodecanoate (10a)

To a stirred solution of **9a** (1.54 g, 6.70 mmol) in CH_2Cl_2 (33.0 mL) at 0 °C were added iodobenzene diacetate (3.24 g, 10.0 mmol) and TEMPO (0.21 g, 1.34 mmol). After stirring

for 8 h at this temperature, saturated aq. Na₂S₂O₃ was added, and the reaction mixture extracted with CH₂Cl₂. The combined organic layer was washed with saturated aq. NaHCO₃, dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated in vacuo, and the crude product was purified by silica gel column chromatography (hexane/ethyl acetate, 1:0–10:1) to afford 10a as a clear colorless oil (981 mg, 4.30 mmol, 64%). 10b (clear colorless oil, 986 mg, 3.15 mmol, 93%) was synthesized from **9b** (1.08 g, 3.40 mmol) according to the above procedure. Data for 10a: 0.93 (3H, d, J=6.5 Hz), 1.24-1.37 (m, 9H), 1.63 (2H, quint. J=7.0 Hz), 1.89–1.99 (1H, m), 2.07–2.12 (1H, m), 2.26–2.30 (1H, m), 2.43 (2H, td, J=7.0, 2.0 Hz), 4.13 (2H, q, J = 7.0 Hz), 9.77 (1H, t, J=2.0 Hz). ¹³C-NMR (CDCl₃) δ : 14.24, 19.65, 21.97, 26.63, 29.05, 29.34, 30.24, 36.53, 41.84, 43.84, 60.04, 173.29, 202.85. IR (ATR) cm⁻¹: 2927, 2856, 2719, 1727, 1462. FAB-MS m/z: 229.1776 (Calcd for $C_{13}H_{25}O_{3}$: 229.1798). Data for **10b**: ¹H-NMR (CDCl₃) δ : 0.93 (3H, d, J=6.5 Hz), 1.24–1.35 (21H, m), 1.63 (2H, quint., J=7.0 Hz), 1.90-1.99 (1H, m), 2.07-2.11 (1H, m), 2.42 (2H, td, J=7.0, 1.5 Hz), 4.13 (2H, q, J=7.0 Hz), 9.77 (1H, t, J=1.5 Hz). ¹³C-NMR (CDCl₃) δ: 11.94, 17.33, 19.68, 24.46, 26.75, 26.93, 27.00, 27.14, 27.18, 27.32, 27.91, 34.29, 39.53, 41.59, 57.69, 170.95, 200.66. IR (ATR) cm⁻¹: 2924, 2853, 2711, 1730, 1464. FAB-MS m/z: 313.2738 (Calcd for C₁₉H₃₇O₃: 313.2737).

Ethyl 10-hydroxy-3-methylhexadecanoate (11a)

To a stirred solution of **10a** (400 mg, 1.80 mmol) in THF (9.00 mL) was added hexylmagnesium bromide (2.0M in THF, 1.05 mL, 2.10 mmol) at -60 °C. After warming to -40 °C and stirring at this temperature for 2 h, 1N HCl was added, and the reaction mixture was extracted with ethyl acetate. The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated in vacuo, and the crude product was purified by silica gel column chromatography (hexane/ethyl acetate, 1:0–15:1) to afford **11a** as a white wax (403 mg, 1.28 mmol, 73%). **11b** (white solid, 560 mg, 1.40 mmol, 47%) was synthesized from **10b** (935 mg, 3.00 mmol) according to the above procedure. Both compounds **11a** and **11b** were obtained and characterized as diastereomeric mixtures. Data for **11a**: ¹H-NMR (CDCl₃) δ : 0.89 (3H, t, J=7.5 Hz), 0.93 (3H, d, J=6.5 Hz), 1.24–1.46 (25H, m), 1.90–1.99 (1H, m), 2.07–2.13 (1H, m), 2.26–2.30 (1H, m), 3.57–3.59 (1H, m), 4.13 (2H, q, J=7.0 Hz). ¹³C-NMR (CDCl₃) δ : 14.06, 14.25, 19.68, 22.59, 25.59, 26.80, 29.35, 29.61, 29.67, 30.31, 31.82, 36.64, 37.43, 37.46, 41.90, 60.05, 71.95, 173.39. IR (ATR) cm⁻¹: 3322, 2925, 2854, 1735, 1460. FAB-MS m/z: 297.2781 (Calcd for C₁₉H₃₉O₃-H₂O: 297.2788). Data for **11b**: ¹H-NMR (CDCl₃) δ : 0.89 (3H, t, J=7.5 Hz), 0.93 (3H, d, J=6.5 Hz), 0.93 (3H, d, J=6.5 Hz), 1.24–1.46 (25H, 2.255, 2.55

J=6.5 Hz), 1.24–1.46 (37H, m), 1.90–1.99 (1H, m), 2.07–2.11 (1H, m), 2.27–2.31 (1H, m), 3.54–3.60 (1H, m), 4.13 (2H, q, J=7.0 Hz). ¹³C-NMR (CDCl₃) δ : 14.05, 14.24, 19.68, 22.58, 25.59, 25.63, 26.86, 29.35, 29.59, 29.68, 29.71, 30.32, 31.82, 36.69, 37.45, 41.90, 60.02, 71.94, 173.39. IR (ATR) cm⁻¹: 3341, 2926, 2849, 1732, 1463. FAB-MS m/z: 381.3724 (Calcd for C₂₅H₅₁O₃-H₂O: 381.3727).

Acknowledgments

We would like to thank Editage (www.editage.jp) for English language editing.

Conflict of Interest

The authors declare no conflict of interest.

References

Ibrguren M., Lopez D. J., Escriba P. V., Biochim. Biophys. Acta, **1838**, 1518–1528 (2014).
Roca-Saavedra P., Marino-Lorenzo P., Miranda J. M., Porto-Arias J. J., Lamas A., Vazquez B. I., Franco C. M., Cepeda A., Food Chem., **221**, 237–247 (2017).

3) Schönfeld P., Reiser G., Aging Dis. 7, 136–149 (2016).

4) Yamamichi Y., Kusuoka H., Morishita K., Shirakami Y., Kurami M., Okano K., Itoh O., Nishimura Y., J. Nucl. Med. **36**, 1043–1050 (1995).

5) Knapp F. F. (Russ), Jr., Franken, P., Kropp J., J. Nucl. Med., 36, 1022–1030 (1995).

6) Takahashi T., Ido T., Iwata R., Appl. Radiat. Isot. 42, 801-809 (1991).

7) Shoup T. M., Elmaleh D. R., Bonab A. A., Fischman A. J., J. Nucl. Med., 46, 297–304 (2005).

8) De Mico A., Margarita R., Parlanti L., Vescovi A., Piancatelli G., J. Org. Chem., **62**, 6974–6977 (1997).

9) Fernandes R. A., Chowdhury A. K., Kattanguru P., Eur. J. Org. Chem. **2014**, 2833–2871 (2014).

10) In the oxidation reaction of **9a**, **10a** was afforded in 64% yield due to generation of several unidentified by-products. On the other hand, the oxidation of **9b** smoothly proceeded.

11) Narayan R. S., Borhan B., J. Org. Chem., 71, 1416–1429 (2006).

12) Teunissen A. J. P., Paffen T. F. E., Ercolani G., de Greef T. F. A., Meijer E. W., J. Am. Chem. Soc., **138**, 6852–6860 (2016).

13) Koyanagi T., Leriche G., Onofrei D., Holland G. P., Mayer M., Yang J., Angew. Chem. Int. Ed., **55**, 1890–1893 (2016).

Chemical and Pharmaceutical Bulletin Advance Publication

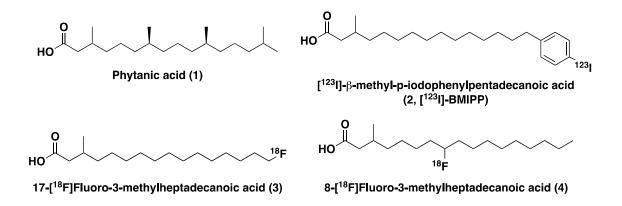


Figure 1. Structures of example β-methyl fatty acids.

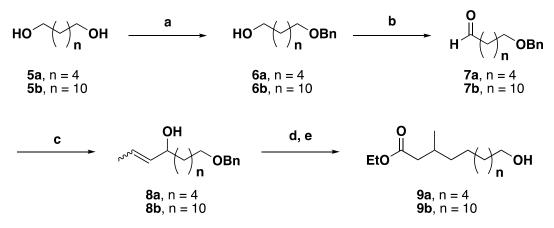


Chart 1. Synthesis of ω -1-hydroxy- β -methyl fatty acid ethyl esters. Reagents: a) NaH, BnBr, THF/DMF, 70 °C (**6a**, 54%; **6b**, 44%); b) TEMPO, PhI(OAc)₂, CH₂Cl₂, (**7a**, 92%; **7b**, 93%); c) Propenylmagnesium bromide, THF, -60 °C (**8a**, 86%; **8b**, 64%), d) CH₃C(OEt)₃, pivalic acid, reflux; e) H₂, Pd/C, EtOH (**9a**, 75%; **9b**, 68%; over 2 steps).

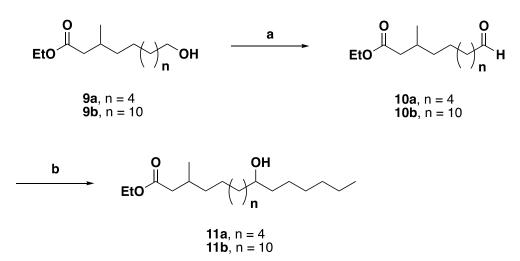


Chart 2. Synthesis of ω -7-hydroxy- β -methyl fatty acid ethyl esters. Reagents: a) TEMPO, PhI(OAc)₂, CH₂Cl₂ (10a, 64%; 10b, 93%); b) Hexylmagnesium bromide, THF (11a, 73%; 11b, 47%).