

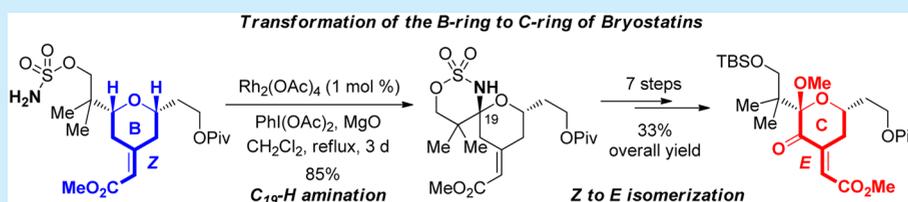
# Transformation of the B Ring to the C Ring of Bryostatins by Csp<sup>3</sup>–H Amination and Z to E Isomerization

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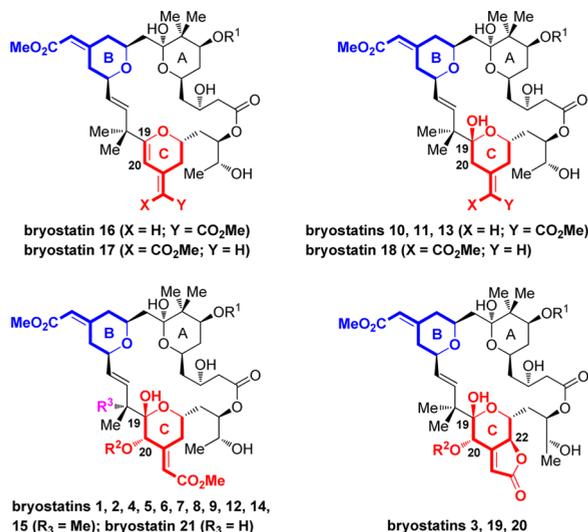
**S** Supporting Information



**ABSTRACT:** An interesting approach to transform the B ring of bryostatins to the C ring has been developed. The key tactics of the approach feature an intramolecular Csp<sup>3</sup>–H bond amination to form spirocyclic hemiaminal, which undergoes ring opening to afford the C ring found in bryostatins 17. The subsequent epoxidation/oxidation sequence results in Z to E isomerization of the *exo*-cyclic enoate, delivering the common precursor, which could be transformed into the C ring found in bryostatins 1, 2, 4–9, 12, 14, and 15.

The bryostatins<sup>1</sup> are a family of 21 complex macrolides produced by a bacterial symbiont of the marine bryozoan *Bugula neritina* (Scheme 1). Bryostatins 1, the first member in the

**Scheme 1. Structure of Bryostatins 1–21**



family, was isolated and structurally characterized by Pettit and co-workers in 1982.<sup>2</sup> It has been recognized that the activity of bryostatins is mediated by binding to the C1 domains of protein kinase C (PKC) isoforms.<sup>3</sup> Extensive biological studies on bryostatins 1 have discovered its remarkable activity against a wide range of cancers. It is attractive that bryostatins 1 also shows

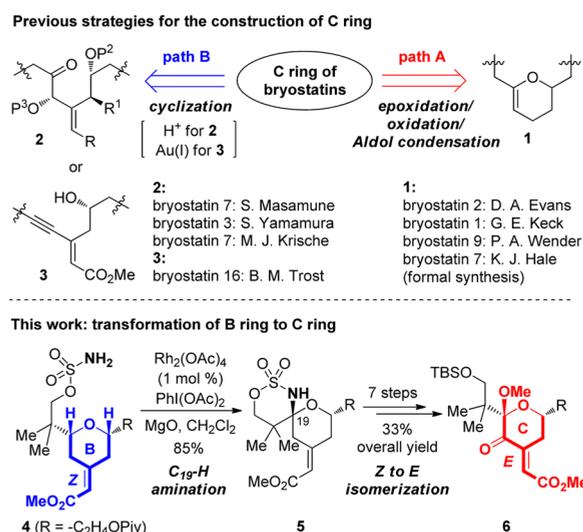
synergism with established oncolytic agents such as Taxol, which has led to numerous clinical trials for cancers.<sup>4</sup> In addition, bryostatins 1 exhibits promising potential in the treatment of other diseases such as ischemic stroke,<sup>5</sup> Alzheimer's disease,<sup>6</sup> and HIV.<sup>7</sup>

Limited availability of bryostatins has motivated great efforts toward their total synthesis, leading to impressive achievement of six members in this family.<sup>8</sup> In addition, extensive studies have focused on discovering functional analogues<sup>9</sup> by simplifying the northern part containing the A and B rings ("spacer domain") because the southern half containing the C ring ("recognition domain") is suggested to govern binding and thus be "recognized" by PKC.<sup>10</sup> Previous strategies for constructing the C ring can be categorized into two types according to the order of the formation of pyran ring and the *exo*-cyclic alkene (Scheme 2, upper). Path A features formation of dihydropyran 1 first.<sup>8a,c,g,h</sup> Epoxidation, oxidation, and aldol condensation with methyl glyoxylate then install the *exo*-cyclic *E*-enoate. Path B relies on formation of the corresponding ketone precursor 2 or enyne precursor 3, which contains the configurationally defined alkene moiety. The subsequent Brønsted acid<sup>8d,e</sup> or Au(I)-catalyzed cyclization leads to formation of the pyran.<sup>8i,j</sup> Despite these successes, new strategies for constructing the C ring are still needed to provide diverse analogues, which are important for SAR studies and discovery of promising drug candidates.

It is noteworthy that both B and C rings contain an *exo*-cyclic enoate at the 4-position of pyran. This structural similarity implies that the C ring might be biogenetically formed from the

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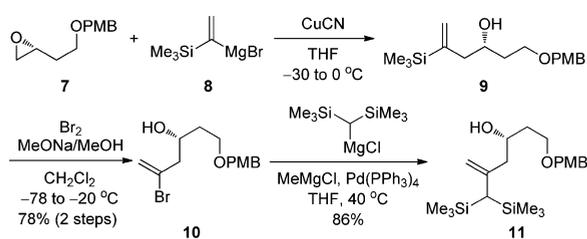
### Scheme 2. Summary of Previous Strategies To Form the C-Ring (Top); Transformation of the B-Ring to C-Ring (Bottom)



ring by oxidation with oxidase. Inspired by the hypothesis and based on the Prins cyclization developed previously by us to construct the B ring,<sup>11</sup> we launched the studies on the transformation of the B ring to C ring. The key steps involve intramolecular Csp<sup>3</sup>-H bond amination<sup>12</sup> of **4** to form spirocyclic hemiaminal **5** and *Z* to *E* isomerization of the *exo*-cyclic enoate to give **6** (Scheme 2, bottom). Pyranone **6** contains the ring core, which has been used as the common precursor by four groups to construct the C ring in the synthesis of bryostatins.<sup>8a,c,g,h</sup> Herein, we report the details of our studies.

The synthesis commenced with formation of the B ring from the known epoxide **7** (Scheme 3).<sup>13</sup> Epoxide ring opening by the

### Scheme 3. Synthesis of Prins Cyclization Precursor 11



1-trimethylsilyl magnesium bromide **8** provided homoallylic alcohol **9**. Bromination of the vinyl silane moiety in **9** afforded vinyl bromide **10** in 78% overall yield.<sup>14</sup> The subsequent Pd(PPh<sub>3</sub>)<sub>4</sub>-catalyzed Kumada cross-coupling<sup>15</sup> of **10** with bis(trimethylsilyl) magnesium chloride installed the desired bis(silyl) moiety successfully, leading to the Prins cyclization precursor **11** in 86% yield.

Prins cyclization to construct the B ring was examined using **11** and aldehydes **12a–e** possessing an  $\alpha$ -quaternary carbon center. The desired tetrahydropyrans **13a–e** were obtained with complete 2,6-*cis* diastereoselectivity. However, the protecting groups on the hydroxyl moiety in **12** show great impacts on both yields and *Z/E* ratios. As shown in Table 1, partial conversion was observed in the reaction of **11** and Bn-substituted **12a**, giving **13a** in 45% yield with a *Z/E* ratio of 83:17 (entry 1). Similar results were observed when PMB-substituted **12b** was used (35%, *Z/E* = 83:17, entry 2). Switching the R group from benzyloxy to silyl

### Table 1. Optimization of Prins Cyclization To Form the B Ring<sup>a</sup>

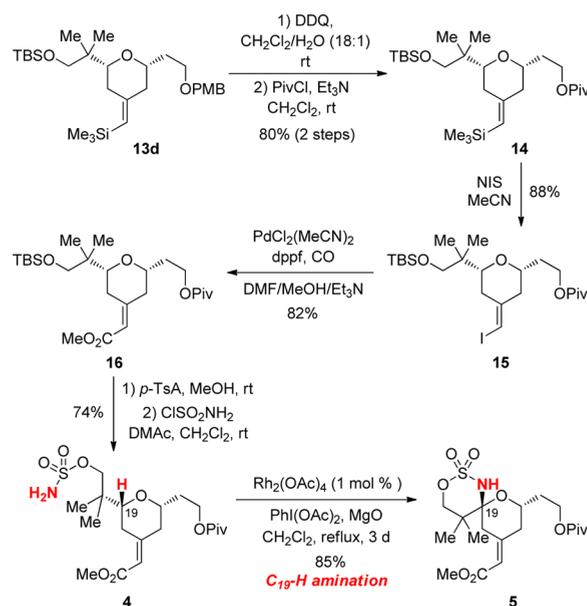
entry	12	R	t (h)	13 (yield, %) <sup>b</sup>	Z/E <sup>c</sup>
1	12a	Bn	0.5	13a (45)	83:17
2	12b	PMB	0.5	13b (38)	80:20
3	12c	TES	1.0	13c (50)	≥95:5
4	12d	TBS	1.0	13d (80)	≥95:5
5	12e	TBDPS	1.0	13e (32)	86:14

<sup>a</sup>Reaction conditions: **11** (1.0 mmol), **12** (2.0 mmol), and TMSOTf (1.5 mmol) in 20 mL of Et<sub>2</sub>O at -78 °C. <sup>b</sup>Isolated yields after purification by silica gel column chromatography. <sup>c</sup>The ratios were determined by the <sup>1</sup>H NMR of the crude products.

groups increased the *Z/E* ratios remarkably, giving **13d** or **13e** as a single *Z*-isomer (entries 3 and 4). Moreover, the TBS group was better than the TES group at affording **13e** in 80% yield (entry 4). Bulker silyl protecting groups such as TBDPS, however, lowered the yield to 32% and the *Z/E* ratio to 86:14 (entry 5).

With **13d** in hand, the synthesis was continued to transform the B ring to the C ring. Removal of the PMB group with DDQ and the subsequent protection with PivCl afforded **14** in 80% overall yield (Scheme 4). Iodination of the *exo*-cyclic vinylsilane

### Scheme 4. C<sub>19</sub>-H Amination of 4 To Form 5

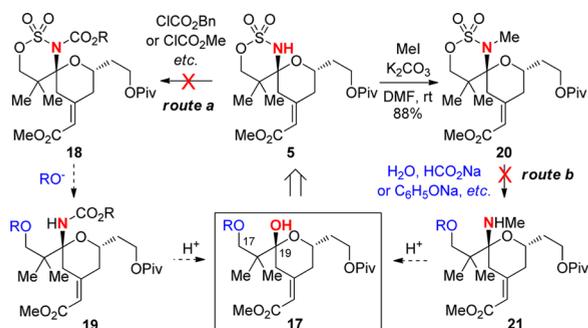


in **14** with NIS gave rise to vinyl iodide **15** in 88% yield with retention of the *Z*-configuration. Pyran **15** underwent Pd-catalyzed carbonylation to deliver *Z*-enoate **16** in 82% yield. Pyran **16** possesses the desired core structure of the B ring of bryostatins. Removal of the TBS group with *p*-TSA followed by sulfonylation of the resulting primary hydroxyl group with ClSO<sub>2</sub>NH<sub>2</sub> converted **16** into sulfamate ester **4** in 82% yield. According to the method developed by Du Bois and co-workers,<sup>16</sup> **4** was treated with 1 mol % of Rh<sub>2</sub>(OAc)<sub>4</sub>, 1.2 equiv of

PhI(OAc)<sub>2</sub>, and 2.3 equiv of MgO in CH<sub>2</sub>Cl<sub>2</sub> by refluxing for 3 days. The desired Csp<sup>3</sup>–H bond amination via metallonitrene insertion occurred smoothly at the C<sub>19</sub> position, giving the spirocyclic hemiaminal **5** in 85% yield as a single regio- and diastereomer. The competitive Csp<sup>3</sup>–H bond amination was not observed at either one of the geminal methyl groups or at the methylene moiety at the C<sub>20</sub> position.

The hydrolytic ring-opening of **5** to give hemiketal **17** proved more difficult than we expected. Introducing an electron-withdrawing group on the nitrogen is typically required to promote the hydrolytic ring-opening of the cyclic sulfamate ester.<sup>17</sup> Unfortunately, the steric congestion around C<sub>19</sub> totally inhibited acylation of **5** with either CbzCl or MeOCOCl to give carbamate **18** (Scheme 5, route a). Methylation with MeI was

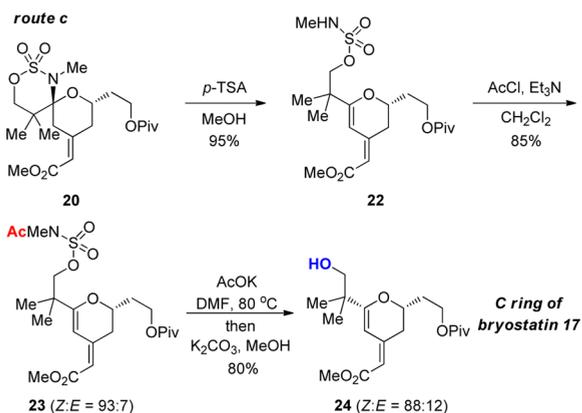
Scheme 5. Attempts To Hydrolyze the Sulfamate Ester



feasible to afford **20** in 88% yield. However, the resulting methyl-substituted ester did not perform well as a leaving group, as the subsequent hydrolytic ring-opening with H<sub>2</sub>O, HCO<sub>2</sub>Na, or C<sub>6</sub>H<sub>5</sub>ONa all failed to give **21** (Scheme 5, route b). The inefficiency might be attributed to the fact that the leaving group at C<sub>17</sub> locates on the neopentyl position, which is embedded in a conformationally fixed ring system. Based on the above analysis, we turned our attention to the strategy of opening the cyclic sulfamate ester first.

As shown in route c (Scheme 6), treatment of **5** with *p*-TSA in MeOH indeed led to ring opening. But dihydropyran **22** instead

Scheme 6. Ring Opening of the Cyclic Sulfamate Ester in 5

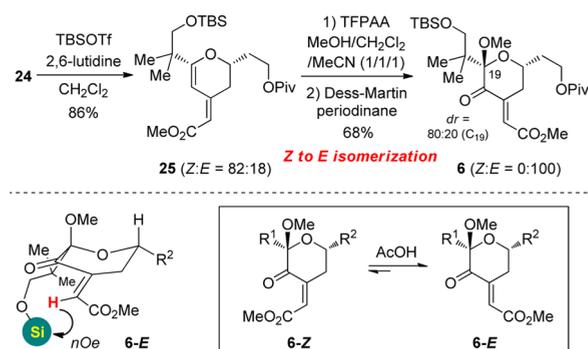


of the expected hemiketal was obtained in 95% yield with the configurational retention of the *exo*-cyclic enoate. Compound **22** contains the C ring core structure of bryostatin **17**.<sup>18</sup> It is noteworthy that this type of C ring has not been achieved previously, despite the successful construction of its *E*-analogue

in the synthesis of bryostatin **16** by Trost and co-workers.<sup>8i,j</sup> Dihydropyran **22** was subsequently transformed into **23** in 85% yield by acylation with AcCl. In this step, *Z* to *E* isomerization of the *exo*-cyclic enoate occurred slightly, leading to a *Z/E* ratio of 93:7. As expected, acylated sulfamate ester moiety at the C<sub>17</sub> position was readily substituted with AcOK in DMF at 80 °C. The subsequent methanolysis of the resulting acetate moiety provided dihydropyran **24** in 80% yield with a *Z/E* ratio of 88:12.

The constantly decreased *Z/E* ratios from **22** to **24** led us to consider the following interesting possibility (Scheme 7). We

Scheme 7. *Z* to *E* Isomerization Leading to **6**



envisioned that we might utilize this *Z* to *E* isomerization to construct the *E*-enoate-substituted C ring, which are found in all 21 members except bryostatins **17** and **18**. To this end, **24** was first protected by TBS to give silyl ether **25** in 86% yield with a *Z/E* ratio of 82:18. According to the protocol developed by Trost,<sup>19</sup> TFPPA-mediated epoxidation of the C<sub>19</sub>–C<sub>20</sub> double bond in **25** followed by in situ epoxide ring opening at C<sub>19</sub> provided a complex mixture containing at least four isomers. However, we were delighted to find that oxidation of the reaction mixture with Dess–Martin periodinane cleanly gave rise to ketone **6** in 68% overall yield as a single *E*-isomer (*dr* = 80:20 at C<sub>19</sub>). We reasoned that AcOH formed in Dess–Martin oxidation might facilitate isomerization of the *Z*-vinylogous ketoester in **6-Z** to thermodynamically more stable **6-E**.<sup>20</sup> Ketone **6** contains the core, which has been used as the common precursor by four groups to construct the C ring in the synthesis of bryostatins and their analogues.<sup>8a,c,g,h</sup> Thus, **6-E** could be principally transformed into the C ring found in bryostatins **1**, **2**, **4–9**, **12**, **14**, and **15**.

In summary, we have developed an interesting approach to transform the B ring of bryostatins to the C ring. The approach was initiated by geminal bis(silyl) Prins cyclization to construct the B ring. Intramolecular Csp<sup>3</sup>–H bond amination gave the spirocyclic hemiaminal, which underwent ring opening to afford the C ring found in bryostatin **17**. *Z* to *E* isomerization of the *exo*-cyclic enoate provided the common precursor, which could be used to construct the C ring found in bryostatins **1**, **2**, **4–9**, **12**, **14**, and **15**. Applications of this approach in the synthesis of bryostatins and their analogues are underway.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.7b02510.

Experimental procedures and spectra data for products (PDF)

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## Notes

The authors declare no competing financial interest.

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## REFERENCES

- (1) For selected reviews on bryostatins, see: (a) Hale, K. J.; Hummerson, M. G.; Manaviazar, S.; Frigerio, M. *Nat. Prod. Rep.* **2002**, *19*, 413. (b) Kortmansky, J.; Schwartz, G. K. *Cancer Invest.* **2003**, *21*, 924. (c) Wender, P. A.; Baryza, J. L.; Hilinski, M. K.; Horan, J. C.; Kan, C.; Verma, V. A. *Beyond Natural Products: Synthetic Analogues of Bryostatin I*. In *Drug Discovery Research: New Frontiers in the Post-Genomic Era*; Huang, Z., Ed.; Wiley-VCH: Hoboken, NJ, 2007; pp 127–162. (d) Hale, K. J.; Manaviazar, S. *Chem. - Asian J.* **2010**, *5*, 704. (e) Wender, P. A.; Loy, B. A.; Schrier, A. J. *Isr. J. Chem.* **2011**, *51*, 453. (f) Yu, L.; Krische, M. J. In *Total Synthesis: At the Frontier of Organic Chemistry*; Li, J. J., Corey, E. J., Eds.; Springer: Heidelberg, 2013; pp 103–130. For isolation of the latest member of bryostatin **21**, see: (g) Yu, H. B.; Yang, F.; Li, Y. Y.; Gan, J. H.; Jiao, W. H.; Lin, H. W. *J. Nat. Prod.* **2015**, *78*, 1169.
- (2) Pettit, G. R.; Herald, C. L.; Doubek, D. L.; Herald, D. L.; Arnold, E.; Clardy, J. *J. Am. Chem. Soc.* **1982**, *104*, 6846.
- (3) (a) Kazanietz, M. G. *Mol. Pharmacol.* **2002**, *61*, 759. (b) Stang, S. L.; Lopez-Campistrous, A.; Song, X.; Dower, N. A.; Blumberg, P. M.; Wender, P. A.; Stone, J. C. *Exp. Hematol.* **2009**, *37*, 122.
- (4) For current clinical information, see : <http://clinicaltrials.gov>.
- (a) Kortmansky, J.; Schwartz, G. K. *Cancer Invest.* **2003**, *21*, 924. (b) Wang, S.; Wang, Z.; Dent, P.; Grant, S. *Blood* **2003**, *101*, 3648. (c) Schwartz, G. K.; Shah, M. A. *J. Clin. Oncol.* **2005**, *23*, 9408. (d) Boije af Gennas, G.; Talman, V.; Yli-Kauhaluoma, J.; Tuominen, R.; Ekokoski, E. *Curr. Top. Med. Chem.* **2011**, *11*, 1370.
- (5) Sun, M.-K.; Hongpaisan, J.; Nelson, T. J.; Alkon, D. L. *Proc. Natl. Acad. Sci. U. S. A.* **2008**, *105*, 13620.
- (6) (a) Hongpaisan, J.; Sun, M. K.; Alkon, D. L. *J. Neurosci.* **2011**, *31*, 630. (b) Williams, P.; Sorribas, A.; Howes, M.-J. R. *Nat. Prod. Rep.* **2011**, *28*, 48. (c) Xu, C.; Liu, Q.-Y.; Alkon, D. L. *Neuroscience* **2014**, *268*, 75.
- (7) (a) DeChristopher, B. A.; Loy, B. A.; Marsden, M. D.; Schrier, A. J.; Zack, J. A.; Wender, P. A. *Nat. Chem.* **2012**, *4*, 705. (b) Bullen, C. K.; Laird, G. M.; Durand, C. M.; Siliciano, J. D.; Siliciano, R. F. *Nat. Med.* **2014**, *20*, 425. (c) Archin, N. M.; Margolis, D. M. *Curr. Opin. Infect. Dis.* **2014**, *27*, 29.
- (8) For total synthesis of bryostatins, see the following. Bryostatin 1: (a) Keck, G. E.; Poudel, Y. B.; Cummins, T. J.; Rudra, A.; Covell, J. A. *J. Am. Chem. Soc.* **2011**, *133*, 744. Bryostatin 2: (b) Evans, D. A.; Carter, P. H.; Carreira, E. M.; Prunet, J. A.; Charette, A. B.; Lautens, M. *Angew. Chem., Int. Ed.* **1998**, *37*, 2354. (c) Evans, D. A.; Carter, P. H.; Carreira, E. M.; Charette, A. B.; Prunet, J. A.; Lautens, M. *J. Am. Chem. Soc.* **1999**, *121*, 7540. Bryostatin 3: (d) Ohmori, K.; Ogawa, Y.; Obitsu, T.; Ishikawa, Y.; Nishiyama, S.; Yamamura, S. *Angew. Chem., Int. Ed.* **2000**, *39*, 2290. Bryostatin 7: (e) Kageyama, M.; Tamura, T.; Nantz, M. H.; Roberts, J. C.; Somfai, P.; Whritenour, D. C.; Masamune, S. *J. Am. Chem. Soc.* **1990**, *112*, 7407. (f) Lu, Y.; Woo, S. K.; Krische, M. J. *J. Am. Chem. Soc.* **2011**, *133*, 13876. Formal synthesis of bryostatin 7: (g) Aliev, A. E.; Hale, K. J. *Org. Lett.* **2006**, *8*, 4477. Bryostatin 9: (h) Wender, P. A.; Schrier, A. J. *J. Am. Chem. Soc.* **2011**, *133*, 9228. Bryostatin 16: (i) Trost, B. M.; Dong, G. *Nature* **2008**, *456*, 485. (j) Trost, B. M.; Dong, G. *J. Am. Chem. Soc.* **2010**, *132*, 16403.
- (9) For the latest studies on bryostatin analogues, see: (a) Trost, B. M.; Yang, H.; Thiel, O. R.; Frontier, A. J.; Brindle, C. S. *J. Am. Chem. Soc.* **2007**, *129*, 2206. (b) DeChristopher, B. A.; Fan, A. C.; Felsher, D. W.; Wender, P. A. *Oncotarget* **2010**, *1*, 58. (c) Wender, P. A.; Baryza, J. L.; Brenner, S. E.; DeChristopher, B. A.; Loy, B. A.; Schrier, A. J.; Verma, V. A. *Proc. Natl. Acad. Sci. U. S. A.* **2011**, *108*, 6721. (d) Trost, B. M.; Yang, H.; Dong, G. *Chem. - Eur. J.* **2011**, *17*, 9789. (e) Kedei, N.; Telek, A.; Michalowski, A. M.; Kraft, M. B.; Li, W.; Poudel, Y. B.; Rudra, A.; Petersen, M. E.; Keck, G. E.; Blumberg, P. M. *Biochem. Pharmacol.* **2013**, *85*, 313. (f) Wender, P. A.; Nakagawa, Y.; Near, K. E.; Staveness, D. *Org. Lett.* **2014**, *16*, 5136. (g) Wender, P. A.; Staveness, D. *Org. Lett.* **2014**, *16*, 5140. (h) Kraft, M. B.; Poudel, Y. B.; Kedei, N.; Lewin, N. E.; Peach, M. L.; Blumberg, P. M.; Keck, G. E. *J. Am. Chem. Soc.* **2014**, *136*, 13202. (i) Andrews, I. P.; Ketcham, J. M.; Blumberg, P. M.; Kedei, N.; Lewin, N. E.; Krische, M. J. *J. Am. Chem. Soc.* **2014**, *136*, 13209. (j) Kelsey, J. S.; Cataisson, C.; Chen, J.; Herrmann, M. A.; Petersen, M. E.; Baumann, D. A.; McGowan, K. M.; Yuspa, S. H.; Keck, G. E.; Blumberg, P. M. *Mol. Cell. Carcinog.* **2016**, *55*, 2183. (k) Ketcham, J. M.; Volchkov, I.; Chen, T. Y.; Blumberg, P. M.; Kedei, N.; Lewin, N. E.; Krische, M. J. *J. Am. Chem. Soc.* **2016**, *138*, 13415.
- (10) (a) Berkow, R. L.; Kraft, A. S. *Biochem. Biophys. Res. Commun.* **1985**, *131*, 1109. (b) Kraft, A. S.; Smith, J. B.; Berkow, R. L. *Proc. Natl. Acad. Sci. U. S. A.* **1986**, *83*, 1334. (c) Kazanietz, M. G.; Lewin, N. E.; Gao, F.; Petit, G. R.; Blumberg, P. M. *Mol. Pharmacol.* **1994**, *46*, 374.
- (11) Lu, J.; Song, Z. L.; Zhang, Y. B.; Gan, Z. B.; Li, H. Z. *Angew. Chem., Int. Ed.* **2012**, *51*, 5367.
- (12) For selected reviews, see: (a) Díaz-Requejo, M. M.; Pérez, P. J. *Chem. Rev.* **2008**, *108*, 3379. (b) Collet, F.; Dodd, R. H.; Dauban, P. *Chem. Commun.* **2009**, *45*, 5061. (c) Collet, F.; Lescot, C.; Dauban, P. *Chem. Soc. Rev.* **2011**, *40*, 1926.
- (13) Reddy, Ch. R.; Rao, N. N. *Tetrahedron Lett.* **2010**, *51*, 5840.
- (14) Fürstner, A.; Flüggé, S.; Larionov, O.; Takahashi, Y.; Kubota, T.; Kobayashi, J. *Chem. - Eur. J.* **2009**, *15*, 4011.
- (15) White, J. D.; Kuntiyong, P.; Lee, T. H. *Org. Lett.* **2006**, *8*, 6039.
- (16) (a) Espino, C. G.; Wehn, P. M.; Chow, J.; Du Bois, J. *J. Am. Chem. Soc.* **2001**, *123*, 6935. (b) Espino, C. G.; Du Bois, J. *Angew. Chem., Int. Ed.* **2001**, *40*, 598. (c) Fleming, J. J.; Du Bois, J. *J. Am. Chem. Soc.* **2006**, *128*, 3926. (d) Zalatan, D. N.; Du Bois, J. *J. Am. Chem. Soc.* **2008**, *130*, 9220. (e) Du Bois, J. *Org. Process Res. Dev.* **2011**, *15*, 758. (f) Roizen, J. L.; Zalatan, D. N.; Du Bois, J. *Angew. Chem., Int. Ed.* **2013**, *52*, 11343.
- (17) Lyle, T. A.; Magill, C. A.; Pitzenger, S. M. *J. Am. Chem. Soc.* **1987**, *109*, 7890. (b) Meunier, N.; Vieth, U.; Jäger, V. *Chem. Commun.* **1996**, 331.
- (18) Pettit, G. R.; Gao, F.; Blumberg, P. M.; Herald, C. L.; Coll, J. C.; Kamano, Y.; Lewin, N. E.; Schmidt, J. M.; Chapuis, J. C. *J. Nat. Prod.* **1996**, *59*, 286.
- (19) Trost, B. M.; Yang, H.; Thiel, O. R.; Frontier, A. J.; Brindle, C. S. *J. Am. Chem. Soc.* **2007**, *129*, 2206.
- (20) The *E*-vinylogous ketoester is thermodynamically more stable than the *Z*-isomer probably by minimizing the dipole–dipole interaction between two carbonyl groups of keto and ester. For similar *Z* to *E*-isomerization of the *exo*-cyclic vinylogous ketoester, see: (a) Tanaka, K.; Ohta, Y.; Fuji, K. *Tetrahedron Lett.* **1993**, *34*, 4071. (b) Bürki, C.; Bonjoch, J.; Bradshaw, B.; Villa, G.; Renaud, P. *Chem. - Eur. J.* **2015**, *21*, 395.