Chromene-Based Synthetic Chalcones as Potent Antileishmanial Agents: Synthesis and Biological Activity

Alireza Foroumadi^{1,2}, Saeed Emami³, Maedeh Sorkhi¹, Maryam Nakhjiri¹, Zohreh Nazarian², Samaneh Heydari², Sussan K. Ardestani⁴, Fatemeh Poorrajab⁴ and Abbas Shafiee^{1,*}

¹Department of Medicinal Chemistry, Faculty of Pharmacy and Pharmaceutical Sciences Research Center, Tehran University of Medical Sciences, Tehran 14174, Iran

²Drug Design & Development Research Center, Tehran University of Medical Sciences, Tehran 14174, Iran

³Department of Medicinal Chemistry and Pharmaceutical Sciences Research Center, Faculty of Pharmacy, Mazandaran University of Medical Sciences, Sari, Iran

⁴Institute of Biochemistry and Biophysics, Department of Biochemistry, University of Tehran, PO Box 13145-1384, Tehran, Iran *Corresponding author: Abbas Shafiee, ashafiee@ams.ac.ir

Two types of regioisomeric chromene-based chalcones namely, 1-(6-methoxy-2*H*-chromen-3-yl)-3-phenylpropen-1-ones and 3-(6-methoxy-2*H*-chromen-3-yl)-1-phenylpropen-1-ones were prepared and investigated for their antileishmanial activity against promastigotes form of *Leishmania major*. The obtained results from *in vitro* biological assays indicated that chloro-substituted 1-(6-methoxy-2*H*-chromen-3-yl)-3-phenylpropen-1-ones exhibited excellent activity against *Leishmania major* at non-cytotoxic concentrations.

Key words: antileishmanial activity, chalcones, chromene, *Leishmania major*, promastigotes

Received 13 August 2009, revised 4 February 2010, accepted for publication 6 February 2010

Leishmaniasis comprises a group of parasitic diseases typically transmitted by the bite of female phlebotomine sandfly and manifest with visceral, cutaneous and mucutaneous forms (1). Human leishmaniasis is mainly distributed in the tropic and subtropic area with a prevalence of 12 million cases and approximate incidence of 0.5 million cases of visceral leishmaniasis and 1.5 million cases of cutaneous leishmaniasis. Overall, 350 million individuals are at risk of leishmaniasis infection in the 88 countries (2). Whereas no vaccine exists, the current chemotherapy for leishmaniasis possesses a set of problems because of the emergence of drug resistant strains,

limited efficacy, long-term treatment, cost expensive and severe side effects (3–5). Thus, the necessity for the development of new, efficient, affordable and safe drug is felt. Also, because of the high toxicity associated with the currently used antileishmanial drugs, efforts are being made to identify new structures from natural and synthetic compounds.

Chalcones (1,3-diaryl-2-propen-1-ones) are natural or synthetic compounds belonging to the flavonoid family with widespread distribution in vegetables, fruits, spices and tea and are present in a variety of plant species (6). Chemically, they consist of open-chain flavonoids in which the two aromatic rings are joined by a threecarbon α , β -unsaturated carbonyl system. Chalcones have been reported to possess many pharmacological activities (7), including anti-inflammatory (8), immunomodulatory (9), antifungal (10), anticancer (11,12), antioxidant (13), antibacterial (14), antimalarial (15,16) and antileishmanial (17,18) properties.

In the past decade, synthetic or naturally occurring chalcones emerged as a new class of antileishmanial compounds. The most studied antileishmanial chalcones are licochalcone A (1) isolated from the roots of Chinese liquorice, licochalcone C (2) and oxygenated chalcones **3** (19). Recently, Narender *et al.* (20,21) have described the promising antileishmanial activity of few naturally occurring chromenochalcones **4**, including crotaramosmin, crotaramin and crotin (Figure 1). These compounds contain a benzopyran system, which is frequently found in many natural products.

In the search for new synthetic chromene-based chalcones (22), we prepared two types of novel chromenochalcones **5a–e** and **6a–e**, to investigate their antileishmanial activity against promastigotes form of *Leishmania major* (Figure 1). The structure **5a–e** possessing the carbonyl group close to chromene ring is called Type A chalcones. The structure **6a–e** possessing the carbonyl group away from chromene ring is called Type B chalcones (Figure 1). The synthesis and *in vitro* antileishmanial activity of both regioisomeric chromene-based chalcones are reported in this article.

Experimental Section

All chemical and solvent used in this study were purchased from Merck AG and Aldrich. Melting points were determined on a Kofler hot stage apparatus and are uncorrected. The IR spectra were obtained on a Shimadzu 470 spectrophotometer (potassium bromide

Chromene-Based Chalcones as Antileishmanial Agents



Figure 1: Structures of naturally occurring and synthetic antileishmanial chalcones **1–4** and newly designed compounds (**5** and **6**) as chromene-based antileishmanial chalcones.

disks). ¹H NMR spectra was recorded using a Bruker 500 MHz spectrometer and chemical shifts are reported in parts per million (ppm) relative to tetramethylsilane as internal standard. The mass spectra were run on a Finigan TSQ-70 spectrometer (Finigan, San Jose, CA, USA) at 70 eV. Elemental analyses were carried out on CHN-O rapid elemental analyzer (Heraeus GmbH, Hanau, Germany) for C, H and N, and the results are within ±0.4% of the theoretical values. Merck silica gel F254 plates were used for analytical TLC.

Synthesis of 1-(6-methoxy-2H-chromen-3-yl) ethanone (9)

A mixture of 5-methoxy-2-hydroxybenzaldehyde (5 mmol) and potassium carbonate (5 mmol) in 1,4-dioxane (5 mL) was treated with methyl vinyl ketone (5 mmol). The mixture was heated at 100 °C for 4 h and allowed to cool. It was then diluted with water and extracted several times with ether. The combined ether extracts were dried (Na_2SO_4) and evaporated to give **9** as a yellow solid, which was crystallized from ethyl acetate-hexane (23).

Synthesis of 6-methoxy-2H-chromene-3-carbaldehyde (10)

A mixture of 5-methoxy-2-hydroxybenzaldehyde (7 mmol) and potassium carbonate (7 mmol) in 1,4-dioxane (12.5 mL) was treated with acrolein (0.5 mL). The mixture was heated at 100 °C for 8 h and allowed to cool. It was then diluted with water and extracted several times with ether. The combined ether extracts were dried (Na₂SO₄) and evaporated to give **10** as a yellow solid, which was crystallized from ethyl acetate-hexane (23).

General procedure for the synthesis of (E)-1-(6methoxy-2H-chromen-3-yl)-3-phenyl prop-2-en-1one derivatives 5a–e

To a solution of compound **9** (1 mmol) and appropriate aldehyde (1 mmol) in absolute ethanol (5 mL), NaOH solution (3.5 M, 2 mL)

was added and stirred overnight in an ice-bath. The reaction mixture was diluted with water and the precipitate was filtered and crystallized from ethanol to give the corresponding chalcones **5a-e**.

(E)-1-(6-Methoxy-2H-chromen-3-yl)-3-phenylprop-2-en-1-one (5a)

IR (KBr, cm⁻¹) ν_{max} : 1649 (C=0), 1224 (C-0). ¹H NMR (CDCl₃) δ : 7.74 (d, 1H, J = 15.6 Hz, H₃ propenone), 7.63 (dd, 2H, J = 7.5 and 2 Hz, H2, H₆ phenyl), 7.44 (s, 1H, H₄ chromene), 7.42 (m, 3H, H₃, H₄ and H₅ phenyl), 7.38 (d, 1H, J = 15.6 Hz, H₂ propenone), 6.84 (m, 2H, H₇ and H₈ chromene), 6.75 (d, 1H, J = 2.5 Hz, H₅ chromene), 5.08 (s, 2H, OCH₂), 3.80 (s, 3H, OCH₃). MS (m/z, %): 293 (M + 1, 38), 291 (60), 273 (38), 270 (16), 159 (38), 130 (50), 100 (98), 74 (100). Anal. Calcd for C₁₉H₁₆O₃: C, 78.06; H, 5.52. Found: C, 77.87; H, 5.69.

(E)-3-(2-Chlorophenyl)-1-(6-methoxy-2H-chromen-3-yl)prop-2-en-1-one (5b)

IR (KBr, cm⁻¹) ν_{max} : 1639 (C=0), 1219 (C-0). ¹H NMR (CDCl₃) δ : 8.1 (d, 1H, J = 15.6 Hz, H₃ propenone), 7.73 (dd, 1H, J = 7 and 2 Hz, H₆ phenyl), 7.44 (m,1H, H₄ phenyl), 7.43 (s, 1H, H₄ chromene), 7.33 (d, 1H, J = 15.6 Hz, H₂ propenone),7.32 (m, 2H, H₃ and H₅ phenyl), 6.84 (m, 2H, H₇ and H₈ chromene), 6.74 (d, 1H, J = 2.5 Hz, H₅ chromene), 5.07 (s, 2H, OCH₂), 3.79 (s, 3H, OCH₃). MS (m/z, %): 328 (M + 2, 15), 326 (M⁺, 45), 309 (40), 199 (11), 185 (100), 143 (33), 137 (94), 109 (80), 73 (46). Anal. Calcd for C₁₉H₁₅ClO₃: C, 69.84; H, 4.63. Found: C, 70.03; H, 4.77.

(E)-3-(3-Chlorophenyl)-1-(6-methoxy-2H-chromen-3-yl)prop-2-en-1-one (5c)

IR (KBr, cm⁻¹) ν_{max} : 1644 (C=0), 1229 (C-0). ¹H NMR (CDCl₃) δ : 7.81 (d, 1H, J = 15.6 Hz, H₃ propenone), 7.73 (m, 1H, H₆ phenyl), 7.58 (m, 2H, H₂ and H₄ phenyl), 7.43 (s,1H, H₄ chromene), 7.32 (d, 1H, J = 15.6 Hz, H₂ propenone), 7.18 (m, 1H, H₅ phenyl), 6.84 (m, 2H, H₇ and H₈ chromene), 6.73 (d, 1H, J = 2.5 Hz, H₅ chromene), 5.08 (s, 2H, OCH₂), 3.80 (s, 3H, OCH₃). MS (m/z, %): 328 (M + 2, 33), 326 (M⁺, 90), 309 (50), 201 (8), 189 (14), 159 (60), 146 (43), 101 (100), 88 (30). Anal. Calcd for C₁₉H₁₅ClO₃: C, 69.84; H, 4.63. Found: C, 69.71; H, 4.80.

(E)-3-(4-Chlorophenyl)-1-(6-methoxy-2H-chromen-3-yl)prop-2-en-1-one (5d)

IR (KBr, cm⁻¹) ν_{max} : 1649 (C=0), 1224 (C-0). ¹H NMR (CDCl₃) δ : 7.68 (d, 1H, J = 15.6 Hz, H₃ propenone), 7.56 (d, 2H, J = 8.2 Hz, H₂ and H₆ phenyl), 7.43 (s, 1H, H₄ chromene), 7.39 (d, 2H, J = 8.2 Hz, H₃ and H₅ phenyl), 7.35 (d, 1H, J = 15.6 Hz, H₂ propenone), 6.84 (m, 2H, H₇ and H₈ chromene), 6.75 (d, 1H, J = 2.5 Hz, H₅ chromene), 5.07 (s, 2H, OCH₂), 3.80 (s, 3H, OCH₃). MS (m/z, %): 328 (M + 2, 30), 326 (M⁺, 90), 307 (42), 275 (17), 227 (8), 165 (50), 159 (63), 135 (67), 101 (100), 89 (58), 45 (50). Anal. Calcd for C₁₉H₁₅ClO₃: C, 69.84; H, 4.63. Found: 69.52; H, 4.49.

Foroumadi et al.

(E)-3-(2,4-Dichlorophenyl)-1-(6-methoxy-2H-chromen-3-yl)prop-2-en-1-one (5e)

IR (KBr, cm⁻¹) ν_{max} : 1647 (C=0), 1222 (C-0). ¹H NMR (CDCl₃) δ : 8.02 (d, 1H, J = 15.6 Hz, H₃ propenone), 7.66 (d, 2H, J = 8.4 Hz H₆ phenyl), 7.47 (d, 1H, J = 2 Hz, H₃ phenyl), 7.42 (s, 1H, H₄ chromene), 7.32 (d, 1H, J = 15.6 Hz, H₂ propenone), 7.28 (dd, 2H, J = 8.4 and 2 Hz, H₅ phenyl), 6.86 (dd, 1H, J = 8.8 and 2.5 Hz, H₇ chromene), 6.83 (d, 1H, J = 8.8 Hz, H₈ chromene), 6.74 (d, 1H, J = 2.5 Hz, H₅ chromene), 5.07 (s, 2H, OCH₂), 3.79 (s, 3H, OCH₃). MS (m/z, %): 364 (M + 4, 10), 362 (M + 2, 65), 361 (M⁺, 21), 360 (100), 325 (58), 309 (17), 260 (8), 202 (10), 198 (35), 161 (50), 118 (62), 89 (45), 63 (20). Anal. Calcd for C₁₉H₁₄Cl₂O₃: C, 63.18; H, 3.91. Found: C, 62.96; H, 4.05.

General procedure for the synthesis of 3-(6-methoxy-2H-chromen-3-yl)-1-phenyl prop-2-en-1-one derivatives 6a–e

To a solution of compound **10** (1 mmol) and appropriate acetophenone (1 mmol) in absolute ethanol (5 mL), NaOH solution (3.5 m, 2 mL) was added and stirred overnight in an ice-bath. The reaction mixture was diluted with water and the precipitate was filtered and crystallized from ethanol to give the corresponding chalcones **6a–e**.

(E)-3-(6-Methoxy-2H-chromen-3-yl)-1-phenylprop-2-en-1-one (6a)

IR (KBr, cm⁻¹) ν_{max} : 1649 (C=O), 1229 (C-O). ¹H NMR (CDCl₃) δ : 7.96 (d, 2H, J = 7.5 Hz, H₂ and H₆ phenyl), 7.58 (m, 1H, H₄ phenyl), 7.52 (d, 1H, J = 15.6 Hz, H₃ propenone), 7.50 (m, 2H, H₃ and H₅ phenyl), 6.86 (d, 1H, J = 15.6 Hz, H₂ propenone), 6.82 (s, 1H, H₄ chromene), 6.80 (d, 1H, J = 8.7 Hz, H₈ chromene), 6.76 (dd, 1H, J = 8.7 and 2.5 Hz, H₇ chromene), 6.65 (d, 1H, J = 2.5 Hz, H₅ chromene), 5.03 (s, 2H, OCH₂), 3.78 (s, 3H, OCH₃). MS (m/z, %): 292 (M⁺, 60), 277 (30), 187 (39), 142 (23), 113 (24), 105 (75), 75 (88), 45 (100). Anal. Calcd for C₁₉H₁₆O₃: C, 78.06; H, 5.52. Found: 78.32; H, 5.51.

(E)-1-(2-Chlorophenyl)-3-(6-methoxy-2H-chromen-3-yl)prop-2-en-1-one (6b)

IR (KBr, cm⁻¹) ν_{max} : 1659 (C=0), 1219 (C-0). ¹H NMR (CDCl₃) δ : 7.45 (m, 2H, H₄, H₆ phenyl), 7.41 (m, 1H, H₃ phenyl), 7.36 (m, 1H, H₅ phenyl), 7.18 (d, 1H, J = 16 Hz, H₃ propenone), 6.77 (m, 2H, H₇, H₈ chromene), 6.75 (s, 1H, H₄ chromene), 6.61 (d, 1H, J = 2.5 Hz, H₅ chromene), 6.48 (d, 1H, J = 16 Hz, H₂ propenone), 4.97 (s, 2H, OCH₂), 3.76 (s, 3H, OCH₃). MS (m/z, %): 328 (M + 2, 22), 326 (M⁺, 66), 311 (48), 236 (11), 201 (11), 185 (100), 141 (40), 139 (80), 109 (80), 67 (38), 55 (55). Anal. Calcd for C₁₉H₁₅ClO₃: C, 69.84; H, 4.63. Found: C, 69.83; H, 4.70.

(E)-1-(3-Chlorophenyl)-3-(6-methoxy-2H-chromen-3-yl)prop-2-en-1-one (6c)

IR (KBr, cm⁻¹) v_{max} : 1654 (C=O), 1219 (C-O). ¹H NMR (CDCI₃) δ : 7.93 (s, 1H, H₂ phenyl), 7.83 (d, 1H, J = 7.75 Hz, H₄ phenyl), 7.56 (m, 1H, H₆ phenyl), 7.53 (d, 1H, J = 15.5 Hz, H₃ propenone), 7.44 (t, 1H, J = 7.75 Hz, H₅ phenyl), 6.84 (s, 1H, H₄ chromene), 6.80 (d, 1H,

 $\begin{array}{l} J=15.5 \mbox{ Hz, } H_2 \mbox{ propenone}), \ 6.78 \ (m, \ 2H, \ H_7, \ H_8 \ chromene), \ 6.65 \ (d, \ 1H, \ J=2.5 \ Hz, \ H_5 \ chromene), \ 5.02 \ (s, \ 2H, \ OCH_2), \ 3.78 \ (s, \ 3H, \ OCH_3). \ MS \ (m/z, \ \%): \ 328 \ (M+2, \ 28), \ 326 \ (M^+, \ 84), \ 309 \ (76), \ 189 \ (11), \ 159 \ (60), \ 146 \ (45), \ 101 \ (100), \ 87 \ (28). \ Anal. \ Calcd \ for \ C_{19}H_{15}ClO_3: \ C, \ 69.84; \ H, \ 4.63. \ Found: \ C, \ 69.99; \ H, \ 4.48. \end{array}$

(E)-1-(4-Chlorophenyl)-3-(6-methoxy-2H-chromen-3-yl)prop-2-en-1-one (6d)

IR (KBr, cm⁻¹) v_{max} : 1649 (C=O), 1229 (C-O). ¹H NMR (CDCl₃) δ : 7.90 (d, 2H, J = 8.5 Hz, H₂ and H₆ phenyl), 7.52 (d, 1H, J = 15.6 Hz, H₃ propenone), 7.47 (d, 2H, J = 8.5 Hz, H₃ and H₅ phenyl), 6.83 (s,1H, H₄ chromene), 6.80 (m, 2H, H₇ and H₈ chromene), 6.79 (d, 1H, J = 15.6 Hz, H₂ propenone), 6.65 (d, 1H, J = 2.5 Hz, H₅ chromene), 5.02 (s, 2H, OCH₂), 3.78 (s, 3H, OCH₃). MS (m/z, %): 328 (M + 2, 20), 326 (M⁺, 93), 323 (68), 311 (46), 308 (25), 245 (11), 202 (11), 187 (95), 184 (32), 139 (100), 111 (80), 109 (46), 75 (28). Anal. Calcd for C₁₉H₁₅ClO₃: C, 69.84; H, 4.63. Found: C, 70.21; H, 4.49.

(E)-1-(2,4-Dichlorophenyl)-3-(6-methoxy-2H-chromen-3-yl)prop-2-en-1-one (6e)

IR (KBr, cm⁻¹) ν_{max} : 1662 (C=O), 1224 (C-O). ¹H NMR (CDCl₃) δ : 7.47 (d, 1H, J = 2 Hz, H₃ phenyl), 7.40 (d, 1H, J = 8 Hz, H₆ phenyl), 7.35 (dd, 1H, J = 8 and 2 Hz, H₅ phenyl), 7.19 (d, 1H, J = 16 Hz, H₃ propenone), 6.77 (m, 2H, H₇ and H₈ chromene), 6.61 (d, 1H, J = 2 Hz, H₅ chromene), 6.46 (d, 1H, J = 16 Hz, H₂ propenone), 4.96 (s, 2H, OCH₂), 3.77 (s, 3H, OCH₃). MS (m/z, %): 365 (M + 4, 7), 363 (M + 2, 42), 361 (M⁺, 63), 325 (46), 279 (60), 261 (18), 206 (8), 189 (23), 187 (100), 149 (100), 113 (48), 71 (64), 57 (70). Anal. Calcd for C₁₉H₁₄Cl₂O₃: C, 63.18; H, 3.91. Found: C, 63.20; H, 4.04.

Biological activity

Parasite and culture

The strain of *L. major* used in this study was the vaccine strain (MRHO/IR/75/ER), obtained from Pasteur Institute, Tehran (Iran). The infectivity of the parasites was maintained by regular passage in susceptible BALB/c mice. The promastigote form of parasite was grown in blood agar cultures at 25 °C. The stationary parasite inoculation was 2×10^6 cells/mL. For the experiments described here, the stationary phase of promastigotes was washed with phosphate-buffered saline and recultured in RPMI 1640 medium (Sigma, St. Louis, MO, USA) at 2×10^6 cells/mL density, supplemented with 10% of heat-inactivated fetal bovine serum, 2-mM glutamine (Sigma), pH approximately 7.2, 100 U/mL penicillin (Sigma) and 100 μ g/mL streptomycin (Sigma).

In vitro antileishmanial activity

The antileishmanial evaluation of compounds **5a–e** and **6a–e** was performed using direct counting and MTT assay (24,25). The growth curve of the *L. major* strain was determined daily under light microscope and counting in a Neubauer's chamber. Then, parasites $(2 \times 10^6/\text{mL})$ in the logarithmic phase were incubated with a serial range of drug concentrations for 24 h at 25 °C. To determine 50%

Chromene-Based Chalcones as Antileishmanial Agents

inhibitory concentrations (IC₅₀), the tetrazolium bromide salt (MTT) assay was used. Briefly, promastigotes from early log phase of growth were seeded in 96-well plastic cell culture trays, containing serial dilution of drug and phenol red free RPMI 1640 medium, supplemented with 10% of FCS, 2-mM glutamine, pH approximately 7.2 and antibiotics, in a volume of 200 μ L. After 24 h of incubation at 25 °C, the media was renewed with 100 μ g/well of MTT (0.5 mg/mL) and plates were further incubated for 4 h at 37 °C. The plates were centrifuged (700 g × 5 min) and the pellets were dissolved in 200 μ L of DMSO. The samples were read using an ELISA plate reader at a wavelength of 492 nm. Two or more independent experiments in triplicate were performed for determination of sensitivity to each drug, the IC₅₀ was calculated by linear regression analysis, expressed in mean ± SD. Control cells were incubated with culture medium plus DMSO.

Cytotoxicity against macrophages

In vitro toxicity against mouse peritoneal macrophages was assessed with cells plated in 96-well plates at 2×10^5 cells/well. After cell adherence, the medium was removed and replaced by the media containing IC₅₀ concentration of each compound. The plates were incubated for 24 h at 37 °C in a humidified incubator with 5% CO₂. Control cells were incubated with culture medium plus DMS0. Cell viability was determined using MTT colorimetric assay.

Table 1: Structures and physicochemical data of chromenebased chalcones 5a-e and 6a-e

H ₃ CO	5a-e (Type A		H ₃ CO	fa-e (Type	
Compound	R	m.p. (°C)	M.W.	Yield	Formula
5a 5b 5c 5d 5e 6a 6b 6c 6d	H 2-Cl 3-Cl 4-Cl 2,4-Cl ₂ H 2-Cl 3-Cl 4-Cl	73–75 87–89 54–56 111–113 124–125 156–158 91–93 146–148 187–189	292.33 326.77 326.77 361.22 292.33 326.77 326.77 326.77 326.77	76 62 59 34 89 55 61 64 78	$\begin{array}{c} C_{19}H_{16}O_3\\ C_{19}H_{15}ClO_3\\ C_{19}H_{15}ClO_3\\ C_{19}H_{15}ClO_3\\ C_{19}H_{15}ClO_3\\ C_{19}H_{14}Cl_2O_3\\ C_{19}H_{16}O_3\\ C_{19}H_{15}ClO_3\\ C_{19}H_$

Results and discussion

Chemistry

As illustrated in Scheme 1, the routes to synthesis of both target compounds **5a-e** and **6a-e** was started from phenolic



R= H; 2-Cl; 3-Cl; 4-Cl; 2,4-Cl₂

Scheme 1: Synthesis of chromene-based chalcones **5a–e** and **6a–e**. *Reagents and conditions*: (a) NaOH, CHCl₃, H₂O, reflux (b) methyl vinyl ketone, 1,4-dioxane, K₂CO₃, reflux (c) appropriate aldehyde, NaOH, EtOH, (d) acrolein, 1,4-dioxane, K₂CO₃, reflux (e) appropriate acetophenone, NaOH, EtOH.





5b	2-CI	100	100	100	5 ± 0.14
5c	3-CI	100	100	100	0.8 ± 0.26
5d	4-CI	100	100	100	0.7 ± 0.3
5e	2,4-Cl ₂	98.1	100	100	0.75 ± 0.23
6a	Н	0.0	12.8	62.8	45 ± 0.15
6b	2-CI	0.0	0.0	1.0	>50
6c	3-CI	0.0	0.0	75.2	45 ± 0.22
6d	4-CI	0.0	0.0	5.4	>50
6e	2,4-Cl ₂	4.3	4.3	8.4	>50
Glucantime®					81.97 ± 3.85 ^c

^aThe growth inhibitory effect of compounds on *Leishmania* parasite in three consecutive days.

^bThe values represent mean ± SD.

°IC₅₀ in mm.

compound 7. Compound 7 was converted to salicylaldehyde derivative **8** according to the general literature method (26.27). Then, compound 8 was reacted with methyl vinyl ketone, as a Michael acceptor, in refluxing dioxane in the presence of K₂CO₃ to give 3-acetylchromene 9. Claisen-Schmidt condensation of 3acetvlchromene 9 with different aldehvdes in ethanolic solution of NaOH yielded corresponding type A chalcones 5a-e. For obtaining type B chalcones **6a-e**, a slightly different strategy was used. Treatment of salicylaldehyde 8 with acrolein in refluxing dioxane in the presence of K₂CO₃ afforded chromene-3-carbaldehyde 10. Subsequently, condensation of aldehyde 10 with appropriate acetophenone in ethanolic solution of NaOH afforded type B chalcones 6a-e. The structures of desired products were established with IR, NMR, mass spectrometry and elemental analysis. ¹H NMR spectra showed that only (*E*)-isomer of chalcones were obtained. The structures and physicochemical data of target compounds are listed in Table 1.

Antileishmanial activity

The life cycle of *Leishmania* parasites consists of two evolutionary stages: promastigotes, flagellated extracellular parasites and amastigotes, non-flagellated, non-motile stages that is more sensitive and live in macrophages. In this study, the chromene-based chalcones **5a–e** and **6a–e** were evaluated for their *in vitro* activity against the promastigote form of the *Leishmania major* using MTT assay. In primary screening assay, the *Leishmania* parasite was affected by 10 μ M concentration of the synthesized compounds **5a–e** and **6a–e** for three consecutive days and the growth inhibitory effect of these compounds was monitored during Day 1, Day 2 and Day 3 and the results are reported in Table 2. The obtained

results indicate that compounds **5b–e** (Type A chalcones) exhibited excellent activity against *Leishmania* (100% inhibition) at the concentration of 10 μ M. Compound **5a** that showed no inhibition at the first day of exposure exhibited potent inhibitory activity after 3 days (% inhibition = 93%). In addition, remaining compounds (Type B chalcones) showed weak to moderate inhibitory activity at this level of concentration.

The IC₅₀ values of type A and type B chalcones against *L. major* in comparison with meglumine antimonate (Glucantime[®], Aventis, Paris, France) are presented in Table 2. The IC₅₀ values of the test compounds against *L. major* indicate that most compounds possessed good leishmanicidal activity (IC₅₀ \leq 50 μ M) with respect to reference drugs. The most potent compounds against the promastigote form of *L. major* were found to be chloro-substituted Type A chalcones **5c–e** with IC₅₀ values less than 1.0 μ M. The activity profile of these compounds (**5c–e**) against promastigotes demonstrated that there are no significant differences in their IC₅₀ values.

The effect of positional substitution was investigated by preparing all three possible chloro-substitutions (2-Cl, 3-Cl or 4-Cl) and 2,4dichloro-substitutions on phenyl ring attached to propenone scaffold. Although chlorine substitution on phenyl ring increases the activity in Type A chalcones, (compounds **5b-e** in comparison with **5a**) this alteration in Type B compounds cannot improve antileishmanial activity (compounds **6b-e** in comparison with **6a**). The better results were achieved with 3-chloro- or 4-chloro-containing analogues in the Type A series.

The cytotoxicity of target compounds was also assessed using MTT colorimetric assay on macrophage cells. Macrophage cells were treated with synthesized compounds at the concentration equal to IC_{50} values for 24h, side by side with the reference drug Glucan-time[®] (25). The results showed that these compounds display antileishmanial activity at non-cytotoxic concentrations.

Chemically, chalcones consist of open-chain flavonoids in which the two aromatic ring A and ring B are joined by a three-carbon α,β unsaturated carbonyl linker. Ring A is attached to the β -position respect to the carbonyl group and ring B is aryl moiety connected to the carbonyl group. Based on previous studies on antileishmanial chalcones, ring A and its substitution pattern are generally considered less important for antileishmanial activity compared to ring B (28,29). Our results with chromene-based chalcones revealed that very good antileishmanial activity was observed when ring B is 6methoxy-2H-chromen-3-yl and ring A is 3- or 4-chlorophenyl moiety (compounds 5c-e). In this work, the mechanisms by which the chromene-based chalcones showed antileishmanial activity were not addressed but based on the literature, it can be predicted that chalcones could potentially interfere with the function of parasite mitochondria and inhibit the activity of fumarate reductase, succinate dehydrogenase, NADH dehydrogenase, or succinate- and NADH-cytochrome c reductases (30-32).

In conclusion, we prepared two types of regioisomeric chromene-based chalcones namely, 1-(6-methoxy-2*H*-chromen-3-yl)-3-phenylpropen-1-ones and 3-(6-methoxy-2*H*-chromen-3-yl)-1-phenylpropen-1-ones and investigated their antileishmanial activity against promastigotes

Chromene-Based Chalcones as Antileishmanial Agents

form of *Leishmania major*. The obtained results from *in vitro* biological assays indicated that chloro-substituted 1-(6-methoxy-2*H*-chromen-3-yl)-3-phenylpropen-1-ones exhibited excellent activity against *Leishmania major* at non-cytotoxic concentrations. The marked activity and simple synthesis of these chalcones suggest that they are potential leads for the development of antileishmanial compounds and further work is in progress to improve the potency of these compounds.

Acknowledgment

This work was supported by grants from the research council of Tehran University of Medical Sciences and Iran National Science Foundation (INSF).

References

- Guerin P.J., Olliaro P., Sundar S., Boelaert M., Croft S.L., Desjeux P., Wasunna M.K., Bryceson A.D. (2002) Visceral leishmaniasis: current status of control, diagnosis, and treatment, and a proposed research and development agenda. Lancet Infect Dis;2:494–501.
- Piñero J., Temporal R.M., Silva-Goncalves A.J., Jiménez I.A., Bazzocchi I.L., Oliva A., Perera A., Leonb L.L., Valladares B. (2006) New administration model of trans-chalcone biodegradable polymers for the treatment of experimental leishmaniasis. Acta Trop;98:59–65.
- Croft S.L., Coombs G.H. (2003) Leishmaniasis-current chemotherapy and recent advances in the search for novel drugs. Trends Parasitol;19:502–508.
- Hemmateenejad B., Miri R., Niroomand U., Foroumadi A., Shafiee A. (2007) A mechanistic QSAR study on the leishmanicidal activity of some 5-substituted-1,3,4-thiadiazole derivatives. Chem Biol Drug Des;69:435–443.
- Singh S., Sivakumar R.J. (2004) Challenges and new discoveries in the treatment of leishmaniasis. J Infect Chemother;10: 307–315.
- Di Carlo G., Mascolo N., Izzo A.A., Capasso F. (1999) Flavonoids: old and new aspects of a class of natural therapeutic drugs. Life Sci;65:337–353.
- Go M.L., Wu X., Liu X.L. (2005) Chalcones: an update on cytotoxic and chemoprotective properties. Curr Med Chem;12:481– 499.
- Hsieh H.K., Lee T.H., Wang J.P., Wang J.J., Lin C.N. (1998) Synthesis and anti-inflammatory effect of chalcones and related compounds. Pharm Res;15:39–46.
- Barford L., Kemp K., Hansen M., Kharazmi A. (2002) Chalcones from Chinese liquorice inhibit proliferation of T cells and production of cytokines. Int Immunopharmacol;2:545–555.
- Sivakumar P.M., Muthu Kumar T., Doble M. (2009) Antifungal activity, mechanism and QSAR studies on chalcones. Chem Biol Drug Des;74:68–79.
- Nam N.H., Kim Y., You Y.J., Hong D.H., Kim H.M., Ahn B.Z. (2003) Cytotoxic 2',5'-dihydroxychalcones with unexpected antiangiogenic activity. Eur J Med Chem;38:179–187.

- Lawrence N.J., Rannison D., McGown A.T., Ducki S., Gul L.A., Hadfield J.A., Khan N. (2001) Linked parallel synthesis and MTT bioassay screening of substituted chalcones. J Comb Chem; 3:421–426.
- Yaylı N., Üçüncü O., Yaşar A., Gök Y., Küçük M., Kolaylı S. (2004) Stereoselective photochemistry of methoxy chalcones in solution and their radical scavenging activity. Turk J Chem; 28:515–521.
- Sivakumar P.M., Priya S., Doble M. (2009) Synthesis, biological evaluation, mechanism of action and quantitative structure-activity relationship studies of chalcones as antibacterial agents. Chem Biol Drug Des;73:403–415.
- Xue C.X., Cui S.Y., Liu M.C., Hu Z.D., Fan B.T. (2004) 3D QSAR studies on antimalarial alkoxylated and hydroxylated chalcones by CoMFA and CoMSIA. Eur J Med Chem;39:745–753.
- Liu M., Wiliarat P., Go M.L. (2001) Antimalarial alkoxylated and hydroxylated chalones: structure-activity relationship analysis. J Med Chem;44:4443–4452.
- Liu M., Wilairat P., Croft S.L., Tan A.L., Go M.L. (2003) Structure–activity relationships of antileishmanial and antimalarial chalcones. Bioorg Med Chem;11:2729–2738.
- Chen M., Christensen S.B., Theander T.G., Kharazmi A. (1994) Antileishmanial activity of licochalcone A in mice infected with Leishmania major and in hamsters infected with *Leishmania donovani*. Antimicrob Agents Chemother;38:1339–1344.
- Nowakowska Z. (2007) A review of anti-infective and antiinflammatory chalcones. Eur J Med Chem;42:125–137.
- Kumar J.K., Narender T., Rao M.S., Rao P.S., Toth G., Balazs B., Duddeck H. (1999) Further Dihydrochalcones from *Crotolaria* ramosissima. J Braz Chem Soc;10:278–280.
- Narender T., Shweta, Gupta S. (2004) A convenient and biogenetic type synthesis of few naturally occurring chromeno dihydrochalcones and their in vitro antileishmanial activity. Bioorg Med Chem Lett;14:3913–3916.
- Nazarian Z., Emami S., Heydari S., Ardestani S.K., Nakhjiri M., Poorrajab F., Shafiee A., Foroumadi A. (2010) Novel antileishmanial chalconoids: synthesis and biological activity of 1- or 3-(6chloro-2*H*-chromen-3-yl)propen-1-ones. Eur J Med Chem; In press: doi:10.1016/j.ejmech.2009.12.046.
- Sorkhi M., Forouzani M., Dehghan G., Abdolahi M., Shafiee A., Foroumadi A. (2008) Synthesis and evaluation of antioxidant activity of 6-methoxy-2*H*-chromenes. Asian J Chem;20:2151– 2155.
- Behrouzi-Fardmoghadam M., Poorrajab F., Ardestani S.K., Emami S., Foroumadi A., Shafiee A. (2008) Synthesis and in vitro anti-leishmanial activity of 1-[5-(5-nitrofuran-2-yl]-1,3,4-thiadiazol-2-yl]- and 1-[5-(5-nitrothiophen-2-yl]-1,3,4-thiadiazol-2-yl]-4aroylpiperazines. Bioorg Med Chem;16:4509–4515.
- Dutta A., Bandyopadhyay S., Mandal C., Chatterjee M. (2005) Development of a modified MTT assay for screening antimonial resistant field isolates of Indian visceral leishmaniasis. Parasitol Int;54:119–122.
- Nielsen A.T., Houlihan W.J. (1968) The aldol condensation. Org React;16:1–438.
- Fine S.A., Pulaski P.D. (1973) Reexamination of the Claisen-Schmidt condensation of phenylacetone with aromatic aldehydes. J Org Chem;38:1747–1749.

Chem Biol Drug Des 2010; 75: 590-596

Foroumadi et al.

- Nielsen S.F., Christensen S.B., Cruciani G., Kharazmi A., Liljefors T. (1998) Antileishmanial chalcones: statistical design, synthesis, and three- dimensional quantitative structure-activity relationship analysis. J Med Chem;41:4819–4832.
- 29. Kayser O., Kiderlen A.F. (2001) In vitro leishmanicidal activity of naturally occurring chalcones. Phytother Res;15:148–152.
- Chen M., Zhai L., Brogger S., Christensen S.B., Theander T.G., Kharazmi A. (2001) Inhibition of fumarate reductase in *Leishmania major* and *L. donovani* by chalcones. Antimicrob Agents Chemother;45:2023–2029.
- 31. Zhai L., Blom J., Chen M., Christensen S.B., Kharazmi A. (1995) The antileishmanial agent licochalcone A interferes with the function of parasite mitochondria. Antimicrob Agents Chemother;39:2742–2748.
- Zhai L., Chen M., Blom J., Christensen S.B., Theander T.G., Kharazmi A. (1999) The antileishmanial activity of novel oxygenated chalcones and their mechanism of action. J Antimicrob Chemother;43:793–803.