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## Design, Synthesis, Evaluation, and Structure of Vitamin D Analogues with Furan Side Chains

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Dedicated to Dr. Noburu Kubodera on the occasion of his retirement

**Abstract:** Based on the crystal structures of human vitamin D receptor (hVDR) bound to  $1\alpha$ ,25-dihydroxy-vitamin D<sub>3</sub> (1,25 D) and superagonist ligands, we previously designed new superagonist ligands with a tetrahydrofuran ring at the side chain that optimize the aliphatic side-chain conformation through an entropy benefit. Following a similar strategy, four novel vitamin D analogues with aromatic furan side chains (**3a**, **3b**, **4a**, **4b**) have now been developed. The triene system has been constructed by an efficient stereoselective intramolecular cyclization of an enol triflate (A-ring precursor) followed by a Suzuki-Miyaura coupling

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of the resulting intermediate with an alkenyl boronic ester (CD-side chain, upper fragment). The furan side chains have been constructed by gold chemistry. These analogues exhibit significant pro-differentiation effects and transactivation potency. The crystal structure of **3a** in a complex with the ligand-binding domain of hVDR revealed that the side-chain furanic ring adopts two conformations.

## Introduction

Vitamin  $D_3$  (1a), before eliciting its biological functions, must be dihydroxylated to its active metabolite 1 $\alpha$ ,25-dihydroxy-vitamin  $D_3$  (1b, 1,25D, calcitriol), which induces gene expression by signaling through the nuclear vitamin D receptor transcription factor (VDR). The *seco*-steroid hormone 1,25D regulates a plethora of biological functions, including calcium homeostasis, cell proliferation and differentiation, and immunomodulation.<sup>[1]</sup>

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Although the applicability of 1,25D is at present limited due to the parallel induction of hypercalcemia, its properties open perspectives for the treatment of several diseases, including osteoporosis, cancer, psoriasis, arthritis, and type 1 diabetes.<sup>[2,3]</sup> As a result, a great deal of attention is being focused on the synthesis of vitamin D analogues with selective properties, although few have proceeded beyond the preclinical stage.<sup>[4,5]</sup> Most of these analogues have been modified at the side chain, as exemplified by calcipotriol<sup>[6]</sup> (**2a**, MC903, Dovonex, Leo Pharmaceuticals, Denmark), used topically for the treatment of psoriasis<sup>[7]</sup> and seocalcitol (**2b**, EB1089,



Leo Pharmaceuticals, Denmark), a potent inhibitor of cell proliferation.<sup>[8]</sup>

All synthetic vitamin D analogues that crystallize in complexes with the VDR ligand-binding domain (LBD) have been shown to be anchored by the same residues in the ligand-binding pocket (LBP), with the hydroxyl moieties at the A-ring and the side chain located in identical positions and forming identical hydrogen bonds.<sup>[9,10]</sup> On the basis of docking studies using the crystal structure of an engineered active ligand-binding domain of the human vitamin D receptor (hVDR LBD) bound to 1,25 D<sup>[9]</sup> and to agonist ligands,<sup>[10]</sup> we have recently designed the new superagonist analogue 2c, the conformation of which in the binding pocket is stabilized by a side-chain tetrahydrofuran unit.<sup>[11]</sup> Attracted by the potent biological activities of 2b,<sup>[8]</sup> which possesses a conjugated double bond in the side chain, and superagonist 2c,<sup>[11]</sup> and to further study the correlation between in silico ligand design and biological activity, we designed the new analogues 3 and 4, which possess a rigid side-chain fragment in the form of a furan ring and bind significantly more efficiently to the hVDR in silico than the natural hormone 1,25 D.<sup>[12,13]</sup> We describe herein the synthesis of the four analogues **3a**,**b** and **4a**,**b**. In addition, we present the results of investigations of their biological behavior and structural studies.

### **Results and Discussion**

**General retrosynthesis of analogues 3 and 4**: The general synthetic strategy for the synthesis of vitamin D analogues **3a**, **3b**, **4a**, and **4b** (depicted as the general structure **5**) follows the convergent strategy recently developed in our laboratory (Scheme 1).<sup>[14]</sup> In this route, the triene system is constructed by an efficient stereoselective intramolecular cyclization of an enol triflate (A-ring precursor **7**)<sup>[15]</sup> followed in situ by a Suzuki–Miyaura coupling of the resulting palladium intermediate with an alkenyl boronic ester (CD-side chain upper fragment, **6**). We envisaged the preparation of the boronate ester **6** from the known protected aldehyde **8**<sup>[16]</sup> using gold chemistry to introduce the furan ring.



Scheme 1. General retrosynthesis of the target vitamin D3 analogues.

Preliminary studies on the conversion of furan 11 into ketone 13 were not very promising. Deprotonation of the furan ring with butyllithium and subsequent reaction of the resulting lithium anion with acetone furnished, after desilylation with tetrabutylammonium fluoride (TBAF),<sup>[22]</sup> diol **12** (75%, two steps, Scheme 2). Unfortunately, several attempts



Scheme 2. Synthesis of furan **11** and furanic ketone **16**. Reagents and conditions: a)  $HC_2CH_2ZnBr$ , THF, 0°C, 1 h, RT, 1.5 h, 98%; b) DMP,  $CH_2Cl_2$ , RT, 1.5 h, 84%; c)  $Et_3PAuCl$  (3 mol%), THF, then AgNO<sub>3</sub> (3 mol%), RT, 4 h, 98%; d) *n*BuLi, THF, RT, 2 h, -78°C, acetone, -78°C to RT, 80%; e) TBAF, THF, 50°C, 72 h, 94%; f) *n*BuLi, THF, 0°C, 2 h, -78°C, then  $ClCO_2Et$ , THF, -78°C to RT, 91%; g) HF (48%, 50 drops),  $CH_3CN$ , RT, 36 h, 92%; h) DMP,  $CH_2Cl_2$ , RT, 3.5 h, 93%. TBS=SiMe<sub>2</sub>*t*Bu. DMP=Dess-Martin periodinane. THF=tetrahydrofuran. TBAF= $nBu_4NF$ .

to selectively oxidize alcohol **12** to the desired ketone **13** under different reaction conditions were unsuccessful.<sup>[23]</sup> These results prompted us to attempt the installation of the hydroxylated side chain at the end of the synthesis. Considering that the ester group tolerates the reaction conditions chosen to introduce the vitamin D triene system,<sup>[14]</sup> we selected this functionality instead of the previous tertiary hydroxy group to continue the synthesis. The required ester **14** was prepared in 91 % yield by reaction of the lithium anion of **11** with ethyl chloroformate. Removal of the TBS group (HF/CH<sub>3</sub>CN) and oxidation of the resulting alcohol (DMP/CH<sub>2</sub>Cl<sub>2</sub>) afforded the desired upper ketone **16** (85 % yield over the two steps; 62 % yield from aldehyde **8**, six steps).

Completion of the synthesis of analogues **3a** and **3b** is illustrated in Scheme 3. Wittig reaction of **16** with ylide  $Ph_3P=CHBr^{[14]}$  afforded alkenyl bromide **17** (75%), which was treated with bis(pinacolato)diboron in the presence of [1,1'-bis(diphenylphosphino)ferrocene]dichloropalladiu-

m(II)-dichloromethane complex as catalyst and tricyclohexylphosphine as ligand to provide the desired boronate **6a** in 75% yield. With the requisite upper boronate **6a** in hand, we next proceeded to install the triene unit. Treatment of an equimolar mixture of **6a** and **7** in aqueous  $K_3PO_4(2m)/THF$ with a catalytic amount of  $[PdCl_2(Ph_3P)_2]$  (5 mol%) at RT for 1 h delivered, after desilylation with hydrogen fluoride,

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Scheme 3. Functionalization of the furan ring and synthesis of analogues **3a** and **3b**. Reagents and conditions: a)  $Ph_3P=CHBr$ , toluene, -15 to 0°C, 75%; b) [PdCl<sub>2</sub>(dppf)<sub>2</sub>]-CH<sub>2</sub>Cl<sub>2</sub>, Cy<sub>3</sub>P, KOAc, Pin<sub>2</sub>B<sub>2</sub>, DMSO, 80°C, 2.5 h, 75%; c) [PdCl<sub>2</sub>(Ph<sub>3</sub>P)<sub>2</sub>], K<sub>3</sub>PO<sub>4</sub>, THF/H<sub>2</sub>O, RT, 1 h; HF (48%, 5 drops), CH<sub>3</sub>CN, 15 min, 85%; d) MeLi, THF, -78°C to RT, 12 h, 88%; e) EtMgBr, THF, -78°C, 15 min, RT, 3 h, 82%. TES=SiEt<sub>3</sub>. dppf=1,1'-bis(diphenylphosphino)ferrocene.

the ester 5a (85% yield, two steps). The ester 5a is a suitable intermediate for further functionalization at the side chain with the possibility of isotopic labeling. Finally, the target analogue 3a was obtained in 88% yield by treatment of ester 5a with methyllithium in THF (33% overall yield from aldehyde 8, eleven steps). Replacing methyllithium by ethylmagnesium bromide furnished the vitamin D analogue 3b (31% overall yield from aldehyde 8, eleven steps).

Synthesis of vitamin D analogues 4a and 4b: Our synthetic efforts were first focused on alcohol 18, which contains the side chain of the target analogues 4 (Scheme 4). Unfortu-



Scheme 4. Retrosynthesis of 12a from aldehyde 8.

nately, our efforts to obtain **18** by treatment of the lithium or copper anion of **11** with isobutylene oxide resulted only in recovery of the starting material.<sup>[24]</sup> As an alternative, we next focused our attention on the Au<sup>III</sup>-assisted transformation of propargyl ketones into furans, as previously observed by Hashmi and co-workers.<sup>[20]</sup> Thus, furan **18** was envisaged as being accessible from ketone **19a**, which in turn would be prepared from aldehyde **8**.

As shown in Scheme 5, our initial efforts to obtain a diastereomeric mixture of propargylic alcohols 25 a by reaction of aldehyde 8 with propargyl zinc 20 produced a mixture of



FULL PAPER

Scheme 5. Synthesis of the upper ketone **26a**. Reagents and conditions: a) MeLi addition to **8** in ClCH<sub>2</sub>I, THF, -78 °C, 30 min, RT, 1.5 h, 98%; b) **22** and F<sub>3</sub>B·OEt<sub>2</sub>, THF, addition to **23a**, -78 °C to RT, 12 h, 98%; c) DMP, CH<sub>2</sub>Cl<sub>2</sub>, RT, 1.5 h, 91%; d) AuCl<sub>3</sub>, THF, RT, 15 min; TBAF, THF, 50 °C, 4 days, 91%; e) DMP, CH<sub>2</sub>Cl<sub>2</sub>, RT, 1.5 h, 91%.

the unwanted allene **21** and dimer **22**. However, the desired alcohols **25a** could be obtained in 96 % yield through a twostep sequence, which involved treatment of a mixture of chloroiodomethane and **8** with methyllithium, followed by opening of the resultant epoxide  $23^{[25]}$  with the lithium alkynide **24a** in the presence of boron trifluoride diethyl etherate. Oxidation of alcohols **25a** with Dess-Martin periodinane proceeded smoothly to produce ketone **26a** (97% yield) accompanied by a trace amount of the corresponding isomeric allenone. As previously planned, exposure of ketone **26a** to gold(III) chloride furnished, after desilylation (TBAF), the desired diol **27a** (91% yield over the two steps). Diol **27a** was then oxidized with Dess-Martin periodinane to generate ketone **28a** (91% yield).

Scheme 6 depicts the completion of the synthesis of **4a** following the same sequence of reactions as above. Olefination of ketone **28a** with Ph<sub>3</sub>P=CHBr provided the alkenyl bromide **29a** (72%), exposure of which to Pin<sub>2</sub>B<sub>2</sub> in the presence of [PdCl<sub>2</sub>(dppf)<sub>2</sub>]·CH<sub>2</sub>Cl<sub>2</sub> as catalyst and Cy<sub>3</sub>P as a ligand, furnished the expected boronate **30a** (87%). The cyclization-coupling cascade between enol triflate **7** and boronate **30a** proceeded smoothly under the standard reaction conditions indicated in Scheme 3 to provide, after desilylation (HF/CH<sub>3</sub>CN), the desired analogue **4a** in 76% yield (nine steps, 36.7% overall yield from aldehyde **8**). Analogue **4b** was synthesized in 26.6% yield from aldehyde **8** employing the same sequence of reactions.<sup>[26]</sup>

**Biological activity**: The biological activities of the four vitamin D analogues (**3a**, **3b**, **4a**, **4b**) were assayed in human SW480-ADH colon cancer cells. These cells express endoge-



Scheme 6. Synthesis of **4a**. Reagents and conditions: a)  $Ph_3P=CHBr$ , toluene, -15 to 0°C, 72%; b)  $[PdCl_2(dppf)_2]\cdot CH_2Cl_2$ ,  $Cy_3P$ , KOAc,  $Pin_2B_2$ , DMSO, 80°C, 3 h, 87%; c)  $[PdCl_2(Ph_3P)_2]$ ,  $K_3PO_4$ , THF/H<sub>2</sub>O, RT, 1 h; d) HF (48%, 5 drops), CH<sub>3</sub>CN, 20 min, 76% (two steps).

nous VDR and respond to 1,25D treatment by inhibition of proliferation and differentiation to an epithelial phenotype.<sup>[27]</sup> The ability of the four derivatives to activate VDR was first examined by their capacity to induce in SW480-ADH cells a morphological change towards a differentiated adhesive phenotype. All four compounds displayed a similar effect to that of 1,25D when assayed at 10<sup>-7</sup>M, leading to the formation of compact epitheloid islands (Figure 1A). We subsequently studied the effect of the analogues in increasing the cellular content of VDR in comparison with that exerted by 1,25 D, an effect that is believed to be a consequence of both transcriptional and post-transcriptional mechanisms.<sup>[28]</sup> Western blot analyses showed that they were all able to increase the VDR protein level with comparable potency to that of 1,25 D (Figure 1B). Moreover, and in line with the induction of intercellular adhesion, all four compounds caused an increase in the amount of E-cadherin, the crucial component of adherens junctions structures at the plasma membrane that is mainly responsible for intercellular adhesion in epithelia. E-Cadherin has been shown to be induced by 1,25D through the activation of a rapid extranuclear signaling pathway that is required for CDH1/E-cadherin gene transcription.<sup>[29]</sup> Thus, our results indicate that compounds 3a, 3b, 4a, and 4b are able to activate both non-genomic and genomic VDR actions.

Finally, we examined the ability of the compounds to induce VDR transcriptional activity in transactivation experiments using cells that were transfected with a plasmid encoding the luciferase reporter gene under the control of a vitamin D response element (VDRE; Figure 1C). Analogues **3a** and **4a** showed potencies slightly lower than but comparable to that of 1,25D, whereas analogues **3b** and **4b** were an order of magnitude less potent. All in all, these results show that our compounds efficiently bind VDR in vivo, inducing its ability to interact with target genes and to activate transcription from a consensus VDRE, with the var-



Figure 1. Biological assays of **3a**, **3b**, **4a**, and **4b**. Numbers below tracks correspond to the values of E-cadherin and VDR protein expression levels normalized to that of  $\beta$ -tubulin in 1,25D- and analogues-treated versus vehicle-treated cells. RLU=relative luciferase units.

iable potency observed in the assays being attributable to gene- or sequence-specific differences.

**Crystal structure of 3a bound to the hVDR ligand-binding domain (PDB ID: 3TKC):** To gain insight into structure– function relationships, we solved the crystal structure of the

606 -

hVDR LBD bound to **3a** at a resolution of 1.75 Å (Table S1 in the Supporting Information). The complex hVDR LBD/ **3a** displays the canonical agonist conformation of all previously reported structures of VDR bound to agonist and superagonist ligands, with helix H12 folded in the agonistic position.<sup>[9-11]</sup> The ligand is buried in the predominantly hydrophobic pocket and adopts the same orientation as the natural ligand. In contrast to all previous structures of hVDR LBD complexes, the side chain of the bound **3a** ligand adopts two conformations, X and Y (Figure 2). Their relative occupancies refined by PHENIX.REFINE were evaluated as 60% and 40% for conformations **3a**-X and **3a**-Y, respectively.



Figure 2. Conformations of **3a** bound to VDR. A) Two views of **3a** (in white) in the  $2F_{o}-F_{c}$  electron density omit map contoured at 1.0  $\sigma$ . B) Superimposition of the two conformations X and Y of **3a** (light grey) with that of 1,25D (grey). C) Intermolecular and intramolecular hydrogen bonds around the side chain of **3a**. X and Y are the two conformations of the side chain of **3a**.

In the two conformations, the A, seco-B, and C/D rings form identical hydrogen-bonding interactions as previously described for the hVDR LBD/1 $\alpha$ ,25(OH)<sub>2</sub>D<sub>3</sub> complex. The presence of an oxygen atom in the tetrahydrofuran ring in the vicinity of the 25-OH group of the side chain of **3a** favors an intramolecular hydrogen bond (darker grey central dashed lines in Figure 2C; distances are 3.07 Å in the **3a-X** conformation and 2.81 Å in the **3b-Y** conformation). Consequently, the side chain of **3a** is more flexible in the LBP and the hydrophobic and electrostatic environment stabilizes two distinct conformations. The distances between the N $\epsilon$ 2 of His305 and the 25-OH of the ligand are 2.73, 3.02, and 2.82 Å for **3a-X**, **3a-Y**, and 1 $\alpha$ ,25(OH)<sub>2</sub>D<sub>3</sub>, respec-

# **FULL PAPER**

tively, and the distances between the N $\epsilon$ 2 of His397 and 25-OH are 2.91, 2.68, and 2.81 Å for **3a**-X, **3a**-Y, and  $1\alpha$ ,25(OH)<sub>2</sub>D<sub>3</sub>, respectively. Additional van der Waals interactions stabilize each conformation. Thus, the flexibility of compound **3a**, illustrated by the presence of two distinct conformations of the furan ring in the LBP, may explain the slightly lower agonist property of this molecule.

### Conclusion

We have designed by docking and efficiently synthesized four novel 1a,25-dihydroxy-vitamin D<sub>3</sub> analogues with furan side chains. Key features of our work include the use of a Pd<sup>0</sup>-catalyzed cyclization-coupling cascade for the stereoselective construction of the triene unit from an enol triflate (A-ring precursor) and an alkenyl boronate (CD-side chain fragment), and an Au<sup>III</sup>-catalyzed intramolecular construction of the side-chain furan unit. These analogues efficiently bind the vitamin D receptor (VDR) in vivo to induce VDR transcriptional activity. Analogues 3a and 4a showed comparable potency to that of 1,25 D, whereas analogues 3b and 4b were an order of magnitude less potent. All four compounds exerted a similar effect in human SW480-ADH colon cancer cells as the natural hormone at concentrations of  $10^{-7}$  M. In contrast to all previous structures of hVDR LBD complexes, the side chain of the bound ligand 3a adopts two conformations. The slightly lower agonist potency of 3a in comparison with 1,25D may be explained in terms of intramolecular hydrogen-bonding between the hydroxy group of the side chain and the oxygen of the furan ring, which diminishes the intermolecular interaction of the analogue with His305 and His397 of the VDR.

### **Experimental Section**

**General**: Details of the general materials and methods have been provided elsewhere.<sup>[12,14]</sup> For details concerning the preparation of compounds **8**, **23**, **28a**, **29a**, **30a**, **4a**, **26b**, **17b**, **28b**, **29b**, **30b**, and **4b**, see the Supporting Information.

8β-[(tert-Butyldimethylsilyl)oxy]-de-A,B-26,27-dinor-24-cholestyn-22-ol (9): A suspension of Zn (0.197 g, 3.01 mmol, 7 equiv) in dry THF (10 mL) was heated at reflux for 10 min. 1,2-Dibromoethane (0.03 mL, 0.345 mmol, 0.8 equiv) was then added. The suspension was heated at reflux for 10 min and then cooled to 0°C, whereupon propargyl bromide (0.16 mL, 1.5 mmol, 3.5 equiv) was added dropwise. The resulting mixture was stirred in the dark for 1 h at 0°C. A solution of aldehyde 8 (0.156 g, 0.43 mmol, 1 equiv) in THF (4 mL) was then added via a cannula. The mixture was stirred for 1 h at 0°C and for 1.5 h at RT. The reaction was then guenched with saturated aqueous NH<sub>4</sub>Cl solution (10 mL). The aqueous phase was extracted with Et2O (3×10 mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO2, 1×12 cm, 5% EtOAc/hexanes) to give a mixture of isomers 9 [0.154 g, 0.42 mmol, 98%,  $R_{\rm f}$ =0.44 (20% EtOAc/hexanes), colorless oil]. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta =$ 4.00 (m, 1H; H-8), 3.90 (m, 1H; H-22), 2.44 (ddd, J=16.6, 8.1, 2.6 Hz, 1 H: H-23), 2.24 (ddd, J = 16.6, 5.8, 2.6 Hz, 1 H: H-23), 2.03 (t, J = 2.6 Hz, 1H; H-25), 0.92 (s, 3H; H-18), 0.89-0.88 (m, 12H; Me<sub>3</sub>CSi and H-21), 0.00 (s, 3H; MeSi), -0.01 ppm (s, 3H; MeSi); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): *δ* = 81.8 (C; C-24), 71.8 (CH; C-22), 70.2 (CH; C-25), 69.4 (CH;

## CHEMISTRY

A EUROPEAN JOURNAL

C-8), 53.2 (CH; C-17), 53.0 (CH; C-14), 42.1 (C; C-13), 40.7 (CH<sub>2</sub>; C-12), 39.1 (CH; C-20), 34.4 (CH<sub>2</sub>; C-9), 26.8 (CH<sub>2</sub>; C-11), 25.8 (3×CH<sub>3</sub>; Me<sub>3</sub>CSi), 25.4 (CH<sub>2</sub>; C-23), 22.9 (CH<sub>2</sub>; C-15), 18.0 (C; CSi), 17.6 (CH<sub>2</sub>; C-16), 13.6 (CH<sub>3</sub>; C-18), 11.3 (CH<sub>3</sub>; C-21), -4.8 (CH<sub>3</sub>; MeSi), -5.2 ppm (CH<sub>3</sub>; MeSi); IR (film):  $\tilde{\nu}$ =3446, 3313 cm<sup>-1</sup>; HRMS (EI<sup>+</sup>): *m/z*: calcd for [C<sub>22</sub>H<sub>39</sub>OSi]<sup>+</sup>: 347.2770 [*M*]<sup>+</sup>; found 347.2773.

 $\$\beta$ -[(*tert*-Butyldimethylsilyl)oxy]-de-A,B-26,27-dinor-23(24),24(25)-cholesdien-22-one (10): Dess-Martin periodinane (DMP, 0.641 g, 1.51 mmol, 1.2 equiv) was added to a solution of 9 (0.460 g, 1.26 mmol, 1 equiv) in

dry CH<sub>2</sub>Cl<sub>2</sub> (15 mL). After stirring at RT for 1.5 h, the reaction was quenched with saturated aqueous NaCl solution (15 mL). The aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×15 mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 3×7 cm, hexanes) to give 10 [0.403 g, 1.07 mmol, 84%,  $R_f = 0.68$  (20% EtOAc/hexanes), pale-yellow solid]. M.p. 99-101 °C (CH<sub>2</sub>Cl<sub>2</sub>/hexanes); <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>): δ=5.72 (t, J=6.5 Hz, 1H; H-23), 5.22 (m, 2H; H-25), 3.99 (m, 1H; H-8), 3.08 (dq, J=10.5, 6.8 Hz, 1H; H-20), 1.06 (d, J=6.6 Hz, 3H; H-21), 0.92 (s, 3H, H-18), 0.87 (s, 9H; Me<sub>3</sub>CSi), -0.01 (s, 3H; MeSi), -0.03 ppm (s, 3H; MeSi); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta = 216.7$  (C; C-24), 204.7 (C; C-22), 96.6 (CH; C-23), 79.3 (CH<sub>2</sub>; C-25), 69.2 (CH; C-8), 53.0 (CH; C-17), 52.3 (CH; C-14), 43.9 (CH; C-20), 42.3 (C; C-13), 40.5 (CH<sub>2</sub>; C-12), 34.3 (CH<sub>2</sub>; C-9), 26.5 (CH<sub>2</sub>; C-15), 25.8 (3×CH<sub>3</sub>; Me<sub>3</sub>CSi), 23.1 (CH<sub>2</sub>; C-16), 18.0 (C; CSi), 17.6 (CH<sub>2</sub>; C-11), 17.1 (CH<sub>3</sub>; C-21), 13.9 (CH<sub>3</sub>; C-18), -4.8 (CH<sub>3</sub>; MeSi), -5.2 ppm (CH<sub>3</sub>; MeSi); IR (film):  $\tilde{\nu} = 1967$ ,  $1677 \text{ cm}^{-1}$ ; HRMS (CI<sup>+</sup>): m/z: calcd for  $[C_{22}H_{39}O_2Si]^+$ : 363.2719  $[M+H]^+$ ; found 363.2729.

8β-[(tert-Butyldimethylsilyl)oxy]-(20S)-de-A,B-(2-furyl)pregnane (11): A solution of 10 (0.082 g, 0.225 mmol, 1 equiv) in dry THF (5 mL) was added dropwise via a cannula to a solution of Et<sub>3</sub>PAuCl (2.4 mg, 0.007 mmol, 3 mol%) in THF (5 mL). AgNO<sub>3</sub> (1.2 mg, 0.007 mmol, 3mol%) was added and the reaction mixture was stirred in the dark at RT for 4 h. Et<sub>2</sub>O (15 mL) was then added, and the mixture was filtered through Celite, washing with further Et<sub>2</sub>O (50 mL). The collected organic phase was washed with saturated aqueous NaCl solution (40 mL), dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 1×10 cm, hexanes) to afford **11** [0.08 g, 0.22 mmol, 98%,  $R_{\rm f}$ =0.72 (10% EtOAc/hexanes), colorless oil]. <sup>1</sup>H NMR (250 MHz,  $CDCl_3$ ):  $\delta = 7.26$  (d, J = 1.3 Hz, 1H; H-25), 6.25 (dd, J = 3, 1.9 Hz, 1H; H-24), 5.90 (d, J=3 Hz, 1H; H-23), 4.02 (m, 1H; H-8), 2.70 (dq, J=10.6, 6.9 Hz, 1H; H-20), 1.24 (d, J=6.8 Hz, 3H; H-21), 1.00 (s, 3H; H-18), 0.90 (s, 9H; Me<sub>3</sub>CSi), 0.03 (s, 3H; MeSi), 0.01 ppm (s, 3H; MeSi); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta = 161.6$  (C; C-22), 140.0 (CH; C-25), 109.7 (CH; C-24), 103.2 (CH; C-23), 69.4 (CH; C-8), 55.8 (CH; C-17), 53.0 (CH; C-14), 42.1 (C; C-13), 40.6 (CH<sub>2</sub>; C-12), 35.8 (CH; C-20), 34.4 (CH<sub>2</sub>; C-9), 27.6 (CH<sub>2</sub>; C-15), 25.8 (3×CH<sub>3</sub>; Me<sub>3</sub>CSi), 23.0 (CH<sub>2</sub>; C-16), 19.7 (CH<sub>3</sub>; C-21), 18.0 (C; CSi), 17.7 (CH<sub>2</sub>; C-11), 13.7 (CH<sub>3</sub>; C-18), -4.8 (CH<sub>3</sub>; MeSi), -5.1 ppm (CH<sub>3</sub>; MeSi); IR (film):  $\tilde{\nu} = 2933$ , 2858 cm<sup>-1</sup>; elemental analysis calcd (%) for C<sub>22</sub>H<sub>38</sub>O<sub>2</sub>Si: C 72.87, H 10.56; found: C 72.60, H 10.83.

(20S)-De-A,B-[5-(2-hydroxypropan-2-yl)furan-2-yl]-8β-pregnanol (12): A solution of nBuLi in hexanes (0.25 mL, 2.24 M, 0.56 mmol, 1.2 equiv) was added dropwise to a solution of 11 (0.170 g, 0.47 mmol, 1 equiv) in dry THF (5 mL) at -78 °C. The mixture was allowed to warm to 0 °C, stirred for 2 h, and then cooled to -78°C once more. Dry acetone (0.14 mL, 1.86 mmol, 4 equiv) was added dropwise, and the reaction mixture was allowed to warm slowly to RT. The reaction was quenched with saturated aqueous NH<sub>4</sub>Cl solution (10 mL) and Et<sub>2</sub>O (10 mL), and the aqueous phase was extracted with Et<sub>2</sub>O ( $3 \times 15$  mL). The combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 2×10 cm, 10% EtOAc/hexanes) to afford 12-TBS  $[0.156 \text{ g}, 0.37 \text{ mmol}, 80\%, R_f = 0.26 (10\% \text{ EtOAc/hexanes}), \text{ colorless oil}].$ <sup>1</sup>H NMR (250 MHz, CDCl<sub>2</sub>):  $\delta = 6.02$  (d, J = 3.1 Hz, 1H; H-24), 5.80 (d, *J*=3.1 Hz, 1H; H-23), 4.00 (m, 1H; H-8), 2.70 (dq, *J*=10.6, 6.9 Hz, 1H; H-20), 1.56 (s, 6H; H-27 and H-28), 1.23 (d, J=6.9 Hz, 3H; H-21), 0.98 (s, 3H; H-18), 0.89 (s, 9H; Me<sub>3</sub>CSi), 0.01 (s, 3H; MeSi), -0.01 ppm (s, 3H; MeSi); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta = 160.8$  (C; C-25), 157.5 (C; C-22), 103.6 (CH; C-24), 103.5 (CH; C-23), 69.4 (CH; C-8), 68.7 (C; C-

26), 56.0 (CH; C-17), 53.0 (CH; C-14), 42.1 (C; C-13), 40.6 (CH<sub>2</sub>; C-12), 35.8 (CH; C-20), 34.4 (CH<sub>2</sub>; C-9), 28.6 (CH<sub>3</sub>; C-27 and C-28), 27.3 (CH<sub>2</sub>, C-15), 25.8 (3×CH<sub>3</sub>; Me<sub>3</sub>CSi), 23.0 (CH<sub>2</sub>; C-16), 19.7 (CH<sub>3</sub>; C-21), 18.0 (C; C-Si), 17.7 (CH<sub>2</sub>; C-11), 13.7 (CH<sub>3</sub>; C-18), -4.8 (CH<sub>3</sub>; MeSi), -5.2 ppm (CH<sub>3</sub>; MeSi). A solution of TBAF in THF (3.1 mL, 1 M, 3 mmol, 5 equiv) was added to a solution of 12-TBS (0.260 g, 0.619 mmol, 1 equiv) in dry THF (10 mL), and the mixture was heated at 50 °C for 72 h. The reaction was then quenched at RT with saturated aqueous NH<sub>4</sub>Cl solution (10 mL) and Et<sub>2</sub>O (5 mL), and the aqueous phase was extracted with Et<sub>2</sub>O (3×15 mL). The combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>,  $2.5 \times 10$  cm, 10% EtOAc/hexanes) to give **12** [0.178 g, 0.583 mmol, 94%,  $R_f = 0.26$  (hexanes), colorless oil]. <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta = 6.01$  (d, J = 3.1 Hz, 1H; H-24), 5.80 (d, J = 3.1 Hz, 1H; H-23), 4.06 (m, 1H; H-8), 2.70 (dq, J=10.3, 6.9 Hz, 1H; H-20), 1.55 (s, 6H; H-27 and H-28), 1.23 (d, J=6.9 Hz, 3H; H-21), 0.99 ppm (s, 3H; H-18);  $^{13}\text{C}$  NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta\!=\!160.4$  (C; C-25), 157.6 (C; C-22), 103.6 (CH; C-23 and C-24), 69.2 (CH; C-8), 68.6 (COH; C-26), 55.8 (CH; C-17), 52.4 (CH; C-14), 41.7 (C; C-13), 40.2 (CH<sub>2</sub>; C-12), 35.7 (CH; C-20), 33.5 (CH<sub>2</sub>; C-9), 28.5 (CH<sub>3</sub>; C-27 and C-28), 27.1 (CH<sub>2</sub>; C-15), 22.4 (CH<sub>2</sub>; C-16), 19.7 (CH<sub>3</sub>; C-21), 17.4 (CH<sub>2</sub>; C-11), 13.4 ppm (CH<sub>3</sub>; C-18); HRMS (CI<sup>+</sup>): m/z: calcd for  $[C_{19}H_{31}O_3]^+$ : 307.2273  $[M+H]^+$ ; found 307.2271.

8β-[(tert-Butyldimethylsilyl)oxy]-(20S)-de-A,B-[(5-ethyloxycarbonyl)furan-2-yl]pregnane (14): A solution of nBuLi in hexanes (2.4 mL, 2.3 M, 5.51 mmol, 1.3 equiv) was added to a solution of **11** (1.53 g, 4.24 mmol, 1 equiv) in dry THF (20 mL) at 0°C. The mixture was stirred for 2 h and then cooled to -78 °C, whereupon ethyl chloroformate (2 mL, 21.2 mmol, 5 equiv) was slowly added dropwise. The reaction mixture was allowed to warm to RT overnight. The reaction was then quenched with saturated aqueous NaCl solution (20 mL) and Et<sub>2</sub>O (20 mL), and the aqueous phase was extracted with Et<sub>2</sub>O (3×20 mL). The combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 3×6 cm, 3% EtOAc/hexanes) to afford 14 [1.68 g, 3.87 mmol, 91%, R<sub>f</sub>=0.7 (10% EtOAc/hexanes), colorless oil]. <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta = 7.04$  (d, J = 3.3 Hz, 1H; H-24), 6.02 (d, *J*=3.3 Hz, 1H; H-23), 4.32 (q, *J*=7.1 Hz, 2H; -OCH<sub>2</sub>CH<sub>3</sub>), 3.97 (m, 1H; H-8), 2.80 (m, 1H; H-20), 1.35 (t, J=7.1 Hz, 3H; CH<sub>3</sub>CH<sub>2</sub>O-), 1.26 (d, J=7.1 Hz, 3H; H-21), 0.98 (s, 3H; H-18), 0.88 (s, 9H; Me<sub>3</sub>CSi), 0.00 (s, 3H; MeSi), -0.02 ppm (s, 3H; MeSi); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta =$ 166.5 (C; COOEt), 159.0 (C, C-22), 142.5 (C; C-25), 118.0 (CH; C-24), 106.1 (CH; C-23), 69.3 (CH; C-8), 60.5 (CH<sub>2</sub>; -OCH<sub>2</sub>CH<sub>3</sub>), 55.2 (CH; C-17), 52.8 (CH; C-14), 42.2 (C; C-13), 40.5 (CH<sub>2</sub>; C-12), 36.3 (CH; C-20), 34.3 (CH2;; C-9), 27.3 (CH2;; C-15), 25.8 (3×CH3;; Me3CSi), 22.9 (CH2;; C-16), 19.5 (CH<sub>3</sub>; C-21), 18.0 (C; C-Si), 17.6 (CH<sub>2</sub>; C-11), 14.4 (CH<sub>3</sub>; CH<sub>3</sub>CH<sub>2</sub>O-), 13.7 (CH<sub>3</sub>; C-18), -4.8 (CH<sub>3</sub>; MeSi), -5.2 ppm (CH<sub>3</sub>; MeSi); IR (film):  $\tilde{v} = 2934$ , 1729 cm<sup>-1</sup>; HRMS (CI<sup>+</sup>): m/z: calcd for [C<sub>25</sub>H<sub>43</sub>O<sub>4</sub>Si]<sup>+</sup>: 435.2931 [*M*+H]<sup>+</sup>; found 435.2935.

(20S)-De-A,B-[(5-ethyloxycarbonyl)furan-2-yl]-8β-pregnanol (15): Aqueous HF (48%, 50 drops) was added to a solution of 14 (0.430 g, 1 mmol, 1 equiv) in CH<sub>3</sub>CN (20 mL) and the mixture was stirred at RT for 36 h. The reaction was quenched by the slow addition of saturated aqueous NaHCO<sub>3</sub> solution (20 mL) (CAUTION: CO<sub>2</sub> evolution) and Et<sub>2</sub>O (20 mL). The aqueous phase was extracted with EtOAc (4  $\times 20$  mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 3.5×10 cm, 20%) EtOAc/hexanes) to give 15 [0.294 g, 0.92 mmol, 92%,  $R_{\rm f}$ =0.3 (20%) EtOAc/hexanes), colorless oil]. <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta = 7.03$  (d, J=3.1 Hz, 1H; H-24), 6.02 (d, J=3.1 Hz, 1H; H-23), 4.30 (q, J=7.1 Hz, 2H;  $-OCH_2CH_3$ ), 4.05 (s, 1H; H-8), 2.79 (m, 1H; H-20), 1.32 (t, J= 7.1 Hz, 3H; CH<sub>3</sub>CH<sub>2</sub>O-), 1.24 (d, J=7 Hz, 3H; H-21), 0.98 ppm (s, 3H; H-18); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>): δ=166.2 (C; COOEt), 158.9 (C; C-22), 142.5 (C; C-25), 118.8 (CH; C-24), 106.1 (CH; C-23), 69.0 (CH; C-8), 60.5 (CH<sub>2</sub>; -OCH<sub>2</sub>CH<sub>3</sub>), 55.0 (CH; C-17), 52.4 (CH; C-14), 41.9 (C; C-13), 40.1 (CH<sub>2</sub>; C-12), 36.2 (CH; C-20), 33.5 (CH<sub>2</sub>; C-9), 27.1 (CH<sub>2</sub>; C-15), 22.3 (CH<sub>2</sub>; C-16), 19.4 (CH<sub>3</sub>; C-21), 17.4 (CH<sub>2</sub>; C-11), 14.3 (CH<sub>3</sub>; CH<sub>3</sub>CH<sub>2</sub>O-), 13.4 ppm (CH<sub>3</sub>; C-18); IR (film):  $\tilde{\nu} = 3529$ , 1712 cm<sup>-1</sup>; HRMS (CI<sup>+</sup>): m/z: calcd for  $[C_{19}H_{29}O_4]^+$ : 321.2069  $[M+H]^+$ ; found 321.2066.

608 -

# **FULL PAPER**

(20S)-De-A,B-[(5-ethyloxycarbonyl)furan-2-yl]-8-pregnanone (16): Dess-Martin periodinane (0.528 g, 1.24 mmol, 1.5 equiv) was added to a solution of 15 (0.266 g, 0.83 mmol, 1 equiv) in dry  $CH_2Cl_2$  (12 mL) and the mixture was stirred at RT for 3.5 h. The reaction was then quenched with saturated aqueous NaCl solution (20 mL), and the resulting mixture was stirred for 30 min. The aqueous phase was extracted with Et<sub>2</sub>O (3× 20 mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>,  $2 \times$ 8 cm, 10% EtOAc/hexanes) to afford 16 [0.245 g, 0.77 mmol, 93%,  $R_{\rm f}$ = 0.47 (10% EtOAc/hexanes), colorless oil]. <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta = 7.03$  (d, J = 3.3 Hz, 1H; H-24), 6.05 (d, J = 3.3 Hz, 1H; H-23), 4.31 (q, J=7.1 Hz, 2H; -OCH<sub>2</sub>CH<sub>3</sub>), 2.81 (dq, J=10.5, 6.9 Hz, 1H; H-20), 2.47 (dd, J=11.5, 7.2 Hz, 1H; H-14), 1.34 (t, J=7.1 Hz, 3H, CH<sub>3</sub>CH<sub>2</sub>O-), 1.24 (d, J=7.2 Hz, 3H; H-21), 0.63 ppm (s, 3H; H-18); <sup>13</sup>C NMR (62.9 MHz,  $CDCl_3$ ):  $\delta = 211.3$  (C; C-8), 165.2 (C; COOEt), 158.8 (C; C-22), 142.7 (C; C-25), 118.7 (CH; C-24), 106.4 (CH; C-23), 61.6 (CH; C-14), 60.6 (CH<sub>2</sub>; -OCH2CH3), 55.0 (CH; C-17), 49.6 (C; C-13), 40.8 (CH2; C-12), 38.6 (CH<sub>2</sub>; C-9), 36.3 (CH<sub>2</sub>; C-20), 27.3 (CH<sub>2</sub>; C-15), 23.9 (CH<sub>2</sub>; C-16), 19.5 (CH<sub>3</sub>; C-21), 18.9 (CH<sub>2</sub>; C-11), 14.3 (CH<sub>3</sub>; CH<sub>3</sub>CH<sub>2</sub>O-), 12.4 ppm (CH<sub>3</sub>; C-18); IR (film):  $\tilde{v} = 2961$ , 1714 cm<sup>-1</sup>; HRMS (CI<sup>+</sup>): m/z: calcd for  $[C_{19}H_{26}O_4]^+$ : 319.1909  $[M+H]^+$ ; found 319.1907.

### (20S),(8E)-Bromomethylene-de-A,B-[(5-ethyloxycarbonyl)furan-2-yl]-

pregnane (17): A suspension of (Ph<sub>3</sub>PCH<sub>2</sub>Br)Br (2.023 g, 4.640 mmol, 8 equiv) in dry toluene (15 mL) was sonicated for 30 min and then cooled to -15°C. KOtBu (0.454 g, 4.060 mmol, 7 equiv) was added and the mixture was stirred for 2 h. A solution of ketone 16 (0.185 g, 0.58 mmol, 1 equiv) in toluene (15 mL) was then added, and the resulting mixture was allowed to warm slowly to 0°C. The reaction was quenched with saturated aqueous  $NH_4Cl$  solution (1 mL), and the mixture was filtered through a layer of silica gel. The silica was washed with Et<sub>2</sub>O (3×20 mL) and the resulting filtrate was concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 2×12 cm, 1% EtOAc/hexanes) to afford 17  $[0.172 \text{ g}, 0.435 \text{ mmol}, 75\%, R_f = 0.58 (10\% \text{ EtOAc/hexanes}), \text{ yellow oil}].$ <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta = 7.04$  (d, J = 3.3 Hz, 1H; H-24), 6.04 (d, J=3.3 Hz, 1H; H-23), 5.62 (s, 1H; H-7), 4.31 (q, J=7.1 Hz, 2H; -OCH<sub>2</sub>CH<sub>3</sub>), 2.86 (m, 1H; H-9), 2.81 (dq, J=10.7, 7.1 Hz, 1H; H-20), 1.83 (q, J=9.4 Hz, 1H; H-17), 1.33 (t, J=7.1 Hz, 3H; CH<sub>3</sub>CH<sub>2</sub>O-), 1.28 (d, J=6.9 Hz, 3H; H-21), 0.60 ppm (s, 3H; H-18); <sup>13</sup>C NMR (62.9 MHz,  $CDCl_3$ ):  $\delta = 165.9$  (C; COOEt), 159.0 (C; C-22), 144.7 (C; C-8), 142.7 (C; C-25), 118.8 (CH; C-24), 106.4 (CH; C-23), 97.8 (CH; C-7), 60.6 (CH<sub>2</sub>; -OCH2CH3), 55.7 (CH; C-14), 54.3 (CH; C-17), 45.5 (C; C-13), 40.0 (CH<sub>2</sub>; C-12), 36.8 (CH<sub>2</sub>; C-20), 31.0 (CH<sub>2</sub>; C-9), 27.4 (CH<sub>2</sub>; C-11), 22.5 (CH<sub>2</sub>; C-16), 22.0 (CH<sub>2</sub>; C-15), 19.7 (CH<sub>3</sub>; C-21), 14.4 (CH<sub>3</sub>; CH<sub>3</sub>CH<sub>2</sub>O-), 11.9 ppm (CH<sub>3</sub>; C-18); IR (film):  $\tilde{\nu}$ =2947, 1726 cm<sup>-1</sup>; HRMS (CI<sup>+</sup>): *m*/*z*: calcd for [C<sub>20</sub>H<sub>28</sub>O<sub>3</sub>Br]<sup>+</sup>: 398.1235 [M+H]<sup>+</sup>; found 398.1221.

(20S)-De-A,B-[(5-ethyloxycarbonyl)furan-2-yl]-(8E)-[(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)methylene]pregnane (6a): Cy<sub>3</sub>P (5.2 mg, 0.06 equiv) and [PdCl<sub>2</sub>(dppf)<sub>2</sub>]·CH<sub>2</sub>Cl<sub>2</sub> (7.7 mg, 0.0186 mmol, 0.0094 mmol, 0.03 equiv) were dissolved in dry DMSO (3 mL) and the mixture was stirred for 20 min. A solution of **17** (0.120 g, 0.31 mmol, 1 equiv) in DMSO (2 mL) was then added via a cannula. KOAc (0.091 g, 0.930 mmol, 3 equiv) and  $Pin_2B_2$  (0.157 g, 0.62 mmol, 2 equiv) were then successively added. The reaction mixture was heated at 80°C for 2.5 h, then cooled to RT, and the reaction was guenched by the addition of  $H_2O\ (10\ mL)$  and  $Et_2O\ (10\ mL).$  The aqueous phase was extracted with Et<sub>2</sub>O (3×30 mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 2×7 cm, 2-5% EtOAc/hexanes) to give **6a** [0.103 g, 0.233 mmol, 75%,  $R_{\rm f}$ =0.42 (10% EtOAc/hexanes), yellow oil]. <sup>1</sup>H NMR (250 MHz,  $CDCl_3$ ):  $\delta = 7.04$  (d, J = 3.4 Hz, 1H; H-24), 6.04 (d, J = 3.4 Hz, 1H; H-23), 4.89 (s, 1H; H-7), 4.32 (q, J=7.1 Hz, 2H; -OCH<sub>2</sub>CH<sub>3</sub>), 3.18 (dd, J=13.2, 2.9 Hz; H-9), 2.82 (dq, J=10.5, 6.9 Hz, 1H; H-20), 1.34 (t, J=7.1 Hz, 3H; CH<sub>3</sub>CH<sub>2</sub>O-), 1.29 (d, J=6.9 Hz, 3H; H-21), 1.25 (s, 12H; CH<sub>3</sub>-pinacol), 0.60 ppm (s, 3H; H-18);  ${}^{13}$ C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta$  = 166.1 (C; COOEt), 165.4 (C; C-8), 158.9 (C; C-22), 142.5 (C; C-25), 118.8 (CH; C-24), 106.2 (CH; C-23), 82.5 (C; COB), 60.5 (CH<sub>2</sub>; -OCH<sub>2</sub>CH<sub>3</sub>), 57.7 (CH; C-14), 55.2 (CH; C-17), 46.2 (C; C-13), 40.2 (CH<sub>2</sub>; C-12), 36.9 (CH<sub>2</sub>; C-20), 33.1 (CH<sub>2</sub>; C-9), 27.3 (CH<sub>2</sub>; C-16), 24.8 (2×CH<sub>3</sub>; CH<sub>3</sub>-pinacol), 24.7 (2×CH<sub>3</sub>; CH<sub>3</sub>-pinacol), 24.2 (CH<sub>2</sub>; C-11), 22.1 (CH<sub>2</sub>; C-15), 19.6 (CH<sub>3</sub>; C-21), 14.4 (CH<sub>3</sub>; *C*H<sub>3</sub>CH<sub>2</sub>O-), 12.0 ppm (CH<sub>3</sub>; C-18); IR (film):  $\tilde{\nu}$ =2975, 1727 cm<sup>-1</sup>; HRMS (CI<sup>+</sup>): *m*/*z*: calcd for [C<sub>26</sub>H<sub>40</sub>O<sub>5</sub>B]<sup>+</sup>: 443.2969 [*M*+H]<sup>+</sup> : found 443.2954.

(20S)-[(5-Ethyloxycarbonyl)furan-2-yl]-22,23,24,25,26,27-hexanor-1α-hydroxy-vitamin D<sub>3</sub> (5a): Aqueous K<sub>3</sub>PO<sub>4</sub> (1.5 mL, 2M) was added to a solution of 6a (0.060 g, 0.135 mmol, 1 equiv) and 7 (0.077 g, 0.149 mmol, 1.1 equiv) in THF (3 mL). [PdCl<sub>2</sub>(Ph<sub>3</sub>P)<sub>2</sub>] (4.7 mg, 0.0068 mmol, 0.05 equiv) was added and the reaction mixture was stirred vigorously for 1 h in the dark. The reaction was then quenched by the addition of H<sub>2</sub>O (10 mL) and Et<sub>2</sub>O (10 mL). The aqueous phase was extracted with Et<sub>2</sub>O (3×10 mL), and the combined organic phases were dried, filtered, and concentrated. The residue was redissolved in CH<sub>3</sub>CN (3 mL), and then HF (48%, 5 drops) was added by means of a syringe. After stirring the solution for 20 min, the reaction was quenched with saturated aqueous NaHCO3 solution (10 mL) and EtOAc (10 mL). The aqueous phase was extracted with EtOAc (5×15 mL), and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 2.5×10 cm, 60% EtOAc/hexanes) to give 5a  $[0.052 \text{ g}, 0.114 \text{ mmol}, 85\%, R_f = 0.3 \text{ (60\% EtOAc/hexanes), white foam]}.$ <sup>1</sup>H NMR (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>):  $\delta = 7.05$  (d, J = 3.4 Hz, 1H; H-24), 6.36 (d, J=11.2 Hz, 1 H; H-6), 6.05 (d, J=3.4 Hz, 1 H; H-23), 6.00 (d, J=11.2 Hz, 1H; H-7), 5.30 (s, 1H; H-19), 4.97 (s, 1H; H-19), 4.41 (m, 1H; H-1), 4.32 (q, J=7.1 Hz, 2H; -OCH<sub>2</sub>CH<sub>3</sub>), 4.21 (m, 1H; H-3), 1.35 (t, J=7.1 Hz, 3H; CH<sub>3</sub>CH<sub>2</sub>O), 1.28 (d, J=7 Hz, 3H; H-21), 0.59 ppm (s, 3H; H-18); <sup>13</sup>C NMR (126 MHz, CD<sub>2</sub>Cl<sub>2</sub>):  $\delta = 166.2$  (CO; C-26), 158.9 (C; C-22), 147.5 (C; C-10), 142.4 (C; C-25), 142.2 (C; C-8), 133.4 (C; C-5), 124.5 (CH; C-6), 118.8 (CH; C-24), 117.3 (CH; C-7), 111.7 (CH<sub>2</sub>; C-19), 106.2 (CH; C-23), 70.7 (CH; C-1), 66.6 (CH; C-3), 60.5 (CH<sub>2</sub>; OCH<sub>2</sub>CH<sub>3</sub>), 56.0 (CH; C-17), 54.7 (CH; C-14), 45.7 (C; C-13), 45.0 (CH<sub>2</sub>; C-4), 42.7 (CH<sub>2</sub>; C-2), 40.1 (CH<sub>2</sub>; C-12), 38.9 (CH; C-20), 29.0 (CH<sub>2</sub>; C-9), 27.3 (CH<sub>2</sub>; C-16), 23.3 (CH<sub>2</sub>; C-11), 22.1 (CH<sub>2</sub>; C-15), 19.7 (CH<sub>3</sub>; C-21), 14.3 (CH<sub>3</sub>; CH<sub>3</sub>CH<sub>2</sub>O), 11.9 ppm (CH<sub>3</sub>; C-18); IR (CHCl<sub>3</sub>):  $\tilde{\nu} = 3412$ , 2933, 1712 cm<sup>-1</sup>; HRMS (ESI-TOF): m/z: calcd for  $[C_{28}H_{38}O_5Na]^+$ : 477.2611 [*M*+Na]<sup>+</sup>; found 477.2604.

(20S)-[5-(2-Hydroxypropan-2-yl)furan-2-yl]-22,23,24,25,26,27-hexanor-1αhydroxy-vitamin D<sub>3</sub> (3a): A solution of MeLi in Et<sub>2</sub>O (1.15 mL, 1.6 M, 1.85 mmol, 5 equiv) was added by means of a syringe to a solution of 5a (0.168 g, 0.37 mmol, 1 equiv) in dry THF (20 mL) at -78 °C. The resulting solution was stirred at -78°C for 6 h and then allowed to warm to RT overnight. The reaction was quenched by the slow addition of saturated aqueous NH<sub>4</sub>Cl solution (10 mL) followed by EtOAc (10 mL). The aqueous phase was extracted with EtOAc (5×15 mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>,  $2 \times 8$  cm, 70% EtOAc/hexanes) to afford **3a** [0.143 g, 0.325 mmol, 88%, R<sub>f</sub>=0.15 (60% EtOAc/hexanes), amorphous solid]. M.p. 84–86 °C (Et<sub>2</sub>O/hexanes);  $[\alpha]_D^{25}=0.88$  (c=1.3 in 96% EtOH); <sup>1</sup>H NMR (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>):  $\delta = 6.38$  (d, J = 11.2 Hz, 1H; H-6), 6.04 (d, J=3.1 Hz, 1H; H-24), 6.02 (d, J=11.2 Hz, 1H; H-7), 5.83 (d, J=3 Hz, 1H; H-23), 5.31 (s, 1H; H-19E), 4.99 (s, 1H; H-19Z), 4.42 (m, 1H; H-1), 4.22 (m, 1H; H-3), 2.72 (dq, J=10.1, 6.9 Hz, 1H; H-20), 1.56 (s, 6H; H-27 and H-28), 1.26 (d, J=6.8 Hz, 3H; H-21), 0.59 ppm (s, 3H; H-18); <sup>13</sup>C NMR (CD<sub>2</sub>Cl<sub>2</sub>, 126 MHz):  $\delta = 160.4$  (C; C-25), 157.6 (C; C-22), 147.6 (C; C-10), 142.8 (C; C-8), 133.1 (C; C-5), 124.8 (CH; C-6), 117.2 (CH; C-7), 111.8 (CH<sub>2</sub>; C-19), 104.0 (CH; C-24), 103.8 (CH; C-23), 70.8 (CH; C-1), 69.9 (COH; C-26), 66.8 (CH; C-3), 56.2 (CH; C-17), 55.6 (CH; C-14), 45.8 (C; C-13), 45.2 (CH<sub>2</sub>; C-4), 42.8 (CH<sub>2</sub>; C-2), 40.2 (CH<sub>2</sub>; C-12), 36.3 (CH; C-20), 29.0 (CH<sub>2</sub>; C-9), 28.6 (2×CH<sub>3</sub>; C-27 and C-28), 27.4 (CH<sub>2</sub>; C-16), 23.5 (CH<sub>2</sub>; C-11), 22.2 (CH<sub>2</sub>; C-15), 20.0 (CH<sub>3</sub>; C-21), 12.0 ppm (CH<sub>3</sub>; C-18); IR (CHCl<sub>3</sub>):  $\tilde{\nu} = 3405$ , 2961 cm<sup>-1</sup>; UV (96%) EtOH):  $\lambda_{\text{max}}$  ( $\epsilon$ ) = 220 nm (18500),  $\lambda$  = 266 nm (14500 mol<sup>-1</sup>m<sup>3</sup> cm<sup>-1</sup>),  $\lambda_{\min} = 250 \text{ nm}; \text{ HRMS} \text{ (ESI-TOF): } m/z: calcd for [C_{28}H_{40}O_4Na]^+:$ 463.2819 [M+Na]+; found 463.2822.

(20S)-[5-(3-Hydroxypentan-3-yl)furan-2-yl]-22,23,24,25,26,27-hexanor-1 $\alpha$ -hydroxy-vitamin D<sub>3</sub> (3b): A solution of EtMgBr in Et<sub>2</sub>O (0.22 mL, 3 M, 0.66 mmol, 6 equiv) was added by means of a syringe to a solution of 5a (0.052 g, 0.11 mmol, 1 equiv) in dry THF (8 mL) at -78 °C. The mixture was stirred at -78 °C for 15 min and at RT for 3 h. The reaction was then quenched by the slow addition of saturated aqueous NH<sub>4</sub>Cl solution

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(10 mL) followed by EtOAc (10 mL). The aqueous phase was extracted with EtOAc (5×15 mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>,  $1.5 \times 6$  cm, 70% EtOAc/hexanes) to give **3b** [0.042 g, 0.089 mmol, 82%,  $R_f = 0.2$  (60% EtOAc/hexanes), amorphous solid]. M.p. 66–68 °C (Et<sub>2</sub>O/hexanes);  $[\alpha]_D^{25} = 1.06$  (c=2.1 in 96% EtOH); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta = 6.36$  (d, J = 11.2 Hz, 1H; H-6), 6.03 (d, *J*=3 Hz, 1H; H-24), 6.00 (d, *J*=11.2 Hz, 1H; H-7), 5.80 (d, *J*=3 Hz, 1H; H-23), 5.30 (s, 1H; H-19E), 4.97 (s, 1H; H-19Z), 4.40 (m, 1H; H-1), 4.20 (m, 1H; H-3), 2.69 (dq, J = 10.1, 6.8 Hz, 1H; H-20), 1.24 (d, J = 6.9 Hz, 3H; H-21), 0.81 (t, J=7.4 Hz, 6H; 2×CH<sub>3</sub>CH<sub>2</sub>), 0.58 ppm (s, 3H; CH<sub>3</sub>-18); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>):  $\delta = 160.1$  (C; C-25), 155.9 (C; C-22), 147.6 (C; C-10), 142.7 (C; C-8), 133.2 (C; C-5), 124.7 (CH; C-6), 117.2 (CH; C-7), 111.8 (CH<sub>2</sub>; C-19), 105.5 (CH; C-24), 103.7 (CH; C-23), 74.9 (COH; C-26), 70.7 (CH; C-1), 66.7 (CH; C-3), 56.1 (CH; C-17), 55.6 (CH; C-14), 45.7 (C; C-13), 45.1 (CH<sub>2</sub>; C-4), 42.8 (CH<sub>2</sub>; C-2), 40.2 (CH<sub>2</sub>; C-12), 36.3 (CH; C-20), 31.7 (2×CH<sub>2</sub>; C-27 and C-29), 29.0 (CH<sub>2</sub>; C-9), 27.6 (CH<sub>2</sub>; C-16), 23.5 (CH<sub>2</sub>; C-11), 22.2 (CH<sub>2</sub>; C-15), 20.0 (CH<sub>3</sub>; C-21), 12.0 (CH<sub>3</sub>; C-18), 7.9 ppm (2×CH<sub>3</sub>; -CH<sub>2</sub>CH<sub>3</sub>); IR (CHCl<sub>3</sub>):  $\tilde{\nu}$  = 3386, 2925 cm<sup>-1</sup>; UV (96% EtOH):  $\lambda_{max}$  ( $\epsilon$ ) = 218 nm (18900),  $\lambda$  = 266 nm (14600 mol<sup>-1</sup> m<sup>3</sup> cm<sup>-1</sup>),  $\lambda_{min}$ =245 nm; HRMS (ESI-TOF): *m*/*z*: calcd for [C<sub>30</sub>H<sub>43</sub>O<sub>3</sub>]<sup>+</sup>: 451.3194 [M-OH]<sup>+</sup>; found 451.3207.

**8**β-[(*tert*-**Butyldimethylsilyl)oxy]-de-A,B-22,23-epoxy-24-norcolane** (23): A solution of MeLi in Et<sub>2</sub>O (1.92 mL, 1.6 $\times$ , 3.08 mmol, 2 equiv) was added dropwise over 30 min to a solution of **8** (0.500 g, 1.54 mmol, 1 equiv) and ClCH<sub>2</sub>I (0.223 mL, 3.08 mmol, 2 equiv) in dry THF (10 mL) at -78 °C. The reaction mixture was stirred at -78 °C for 30 min and then at RT for 1.5 h. The reaction was stopped by the addition of Et<sub>2</sub>O (10 mL) and brine (10 mL). The aqueous phase was extracted with Et<sub>2</sub>O (3×15 mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 3×10 cm, 5% EtOAc/hexanes) to afford a mixture of diastereoisomers **23**<sup>[25]</sup> [0.513 g, 1.51 mmol, 98%, *R*<sub>f</sub>=0.78 (2% Et<sub>2</sub>O/hexanes, two runs), white foam].

8β-[(tert-Butyldimethylsilyl)oxy]-de-A,B-26,27-dinor-25-[2-methyl-(2-triethylsilyloxy)prop-2-yl]-24-cholestin-22-ol (25a): A solution of 24a was prepared at -78°C by the addition of a solution of nBuLi in hexanes (0.82 mL, 2.5 m, 2.05 mmol, 2.5 equiv) to the corresponding alkyne (0.600 g, 2.83 mmol, 3.5 equiv) in dry THF (10 mL) followed by stirring for 40 min. F<sub>3</sub>B·OEt<sub>2</sub> (0.520 mL, 4.10 mmol, 5 equiv) and a solution of 23 (0.280 g, 0.82 mmol, 1 equiv) in THF (5 mL) were then successively added dropwise. The reaction mixture was allowed to warm to RT overnight. The reaction was then quenched by the addition of saturated aqueous NaCl solution (10 mL) and Et<sub>2</sub>O (10 mL). The aqueous phase was extracted with Et<sub>2</sub>O (4×15 mL), and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 3×10 cm, 2% EtOAc/hexanes) to give a mixture of alcohols 25a [0.441 g, 0.80 mmol, 98%,  $R_{\rm f}$ =0.39 (10% EtOAc/hexanes), oil]. <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta = 4.00$  (s, 1H; H-8), 3.84 (t, J =6.8 Hz, 1H; H-22), 2.41 (dd, J=16.2, 7.9 Hz, 1H; H-23), 2.31 (s, 2H; H-26), 2.23 (dd, J=16.2, 5.9 Hz, 1H; H-23), 1.29 (s, 6H; H-28 and H-29), 0.94 (t, J=7.9 Hz, 9H; CH<sub>3</sub>CH<sub>2</sub>Si), 0.91 (s, 3H; H-18), 0.89–0.87 (m, 12H; Me<sub>3</sub>CSi and H-21), 0.57 (q, J=7.9 Hz, 6H; CH<sub>3</sub>CH<sub>2</sub>Si), 0.01 (s, 3H; MeSi), 0.00 ppm (s, 3H; MeSi);  ${}^{13}$ C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta = 80.2$ (C; C-25), 78.6 (C; C-24), 73.1 (COH; C-27), 72.0 (CH; C-22), 69.4 (CH; C-8), 53.2 (CH; C-14), 52.9 (CH; C-17), 42.0 (C; C-13), 40.7 (CH<sub>2</sub>; C-12), 38.9 (CH; C-20), 35.4 (CH<sub>2</sub>; C-26), 34.4 (CH<sub>2</sub>; C-9), 29.4 (2×CH<sub>3</sub>; C-28 and C-29), 26.7 (CH<sub>2</sub>; C-16), 25.8 (3×CH<sub>3</sub>; Me<sub>3</sub>CSi and CH<sub>2</sub>; C-23), 22.9 (CH<sub>2</sub>; C-15), 18.0 (C; C-Si), 17.7 (CH<sub>2</sub>; C-11), 13.6 (CH<sub>3</sub>; C-18), 11.3 (CH<sub>3</sub>; C-21), 7.0 (2×CH<sub>3</sub>; CH<sub>3</sub>CH<sub>2</sub>Si), 6.6 (2×CH<sub>2</sub>; CH<sub>3</sub>CH<sub>2</sub>Si), -4.8 (CH<sub>3</sub>; MeSi), -5.2 ppm (CH<sub>3</sub>; MeSi); HRMS (CI<sup>+</sup>): m/z: calcd for [C<sub>34</sub>H<sub>67</sub>O<sub>3</sub>Si<sub>2</sub>]<sup>+</sup>: 579.4629 [*M*+H]<sup>+</sup>; found 579.4634.

 $8\beta$ -[(*tert*-Butyldimethylsilyl)oxy]-de-A,B-26,27-dinor-25-[2-methyl-(2-trie-thylsilyloxy)-prop-2-yl]-24-cholestin-22-one (26a): Dess–Martin periodinane (0.284 g, 0.66 mmol, 1.3 equiv) was added to a solution of 25a (0.280 g, 0.509 mmol, 1 equiv) in dry CH<sub>2</sub>Cl<sub>2</sub> (15 mL) and the reaction mixture was stirred at RT for 1.5 h. The reaction was then quenched by the addition of saturated aqueous NaCl solution (15 mL). The aqueous

phase was extracted with TBME (4×15 mL) and the combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO<sub>2</sub>, 2.5×8 cm, 2% EtOAc/hexanes) to give **26a** [0.270 g, 0.492 mmol, 97%,  $R_{\rm f}$ =0.43 (30% EtOAc/hexanes), paleyellow oil], slightly contaminated with its allenic ketone. <sup>1</sup>H NMR  $(250 \text{ MHz}, \text{CDCl}_3): \delta = 4.00 \text{ (m, 1H; H-8)}, 3.24 \text{ (t, } J = 2.2 \text{ Hz}, 1\text{ H}; \text{H-23}),$ 2.85 (dq, J=10.7, 6.8 Hz, 1 H; H-20), 2.35 (t, J=2.2 Hz, 2 H; H-26), 1.31 (s, 6H; H-28 and H-29), 1.10 (d, J=6.8 Hz, 3H; H-21), 0.97-0.91 (m, J= 7.9 Hz, 12 H; CH<sub>3</sub>CH<sub>2</sub>Si and H-18), 0.88 (s, 9 H; Me<sub>3</sub>Si), 0.61-0.54 (m, 6H; CH<sub>3</sub>CH<sub>2</sub>Si), 0.00 (s, 3H; MeSi), -0.02 ppm (s, 3H; MeSi); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta = 208.8$  (CO; C-22), 82.5 (C; C-25), 74.0 (C; C-24), 73.1 (COH; C-27), 69.2 (CH; C-8), 52.6 (CH; C-14), 52.4 (CH; C-17), 46.9 (CH; C-20), 42.2 (C; C-13), 40.5 (CH2;; C-12), 35.5 (CH2;; C-26), 34.3 (CH<sub>2</sub>; C-9), 33.8 (CH<sub>2</sub>; C-23), 29.4 (2×CH<sub>3</sub>; C-28 and C-29), 26.6 (CH<sub>2</sub>; C-16), 25.8 (3×CH<sub>3</sub>; Me<sub>3</sub>CSi), 23.2 (CH<sub>2</sub>; C-15), 18.0 (C; C-Si), 17.6 (CH<sub>2</sub>; C-11), 16.6 (CH<sub>3</sub>; C-21), 14.0 (CH<sub>3</sub>; C-18), 7.0 (2×CH<sub>3</sub>; CH<sub>3</sub>CH<sub>2</sub>Si), 6.6 (2×CH<sub>2</sub>; CH<sub>3</sub>CH<sub>2</sub>Si), -4.8 (CH<sub>3</sub>; MeSi), -5.2 ppm (CH<sub>3</sub>; MeSi); HRMS (CI<sup>+</sup>): m/z: calcd for [C<sub>36</sub>H<sub>70</sub>O<sub>3</sub>Si<sub>2</sub>]<sup>+</sup>: 564.4394 [M+H]<sup>+</sup>; found 564.4387.

(20S)-De-A,B-[5-(2-hydroxy-2-methylpropan-1-yl)furan-2-yl]-8\beta-pregnanol (27 a): AuCl<sub>3</sub> (4.4 mg, 0.014 mmol, 0.03 equiv) was added to a solution of 26 a (0.265 g, 0.48 mmol, 1 equiv) in dry THF (10 mL) and the reaction mixture was stirred at RT for 15 min. Saturated aqueous NaCl solution (10 mL) and TBME (10 mL) were then successively added, and the aqueous phase was extracted with TBME (4×10 mL). The combined organic phases were dried, filtered, and concentrated. The residue was redissolved in THF (15 mL), and then TBAF (1.5 mL, 1 M, 1.5 mmol, 3 equiv) was added. The reaction mixture was heated at 50 °C for 4 days. Saturated NH<sub>4</sub>Cl (15 mL) and EtOAc (15 mL) were then successively added, and the aqueous phase was extracted with EtOAc (3×15 mL). The combined organic phases were dried, filtered, and concentrated. The residue was purified by flash chromatography (SiO2, 2.5×8 cm, 20% EtOAc/hexanes) to give 27a [0.140 g, 0.44 mmol, 91%,  $R_{\rm f}$ =0.4 (40% EtOAc/hexanes), white amorphous solid]. M.p. 69–71  $^{\circ}\mathrm{C}$  (CH\_2Cl\_2);  $^{1}\mathrm{H}\,\mathrm{NMR}$ (250 MHz, CDCl<sub>3</sub>):  $\delta = 5.96$  (d, J = 2.9 Hz, 1H; H-24), 5.82 (d, J = 2.9 Hz, 1H; H-23), 4.06 (m, 1H; H-8), 2.73 (s, 2H; H-26), 2.67 (dq, J=10.5, 7.1 Hz, 1H; H-20), 1.22 (s, 9H; H-21, H-28 and H-29), 0.99 ppm (s, 3H; H-18); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta = 160.4$  (C; C-25), 150 (C; C-22), 107.9 (C; C-24), 103.9 (CH; C-23), 70.4 (COH; C-27), 69.9 (CH; C-8), 55.5 (CH; C-14), 52.4 (CH; C-17), 42.0 (CH<sub>2</sub>; C-26), 41.6 (C; C-13), 40.1 (CH<sub>2</sub>; C-12), 35.7 (CH; C-20), 33.4 (CH<sub>2</sub>; C-9), 28.8 ( $2 \times CH_3$ ; C-28 and C-29), 27.3 (CH<sub>2</sub>; C-16), 22.3 (CH<sub>2</sub>; C-15), 19.6 (CH<sub>3</sub>; C-21), 17.3 (CH<sub>2</sub>; C-11), 13.4 ppm (CH<sub>3</sub>; C-18); IR (film):  $\tilde{\nu} = 3377$ , 2935 cm<sup>-1</sup>; elemental analysis calcd (%) for C<sub>20</sub>H<sub>32</sub>O<sub>3</sub>: C 74.96, H 10.06; found: C 74.90, H 10.35.

#### (20S)-De-A,B-[5-(2-hydroxy-2-methylpropan-1-yl)furan-2-yl]-8b-pre-

**gnone (28a):** Procedure as for **16. 28a** [91%,  $R_f$ =0.4 (40% EtOAc/hexanes), colorless oil]. <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta$ =5.96 (s, 1 H; H-24), 5.84 (s, 1 H; H-23), 2.73 (s, 2 H; H-26), 2.69 (m, 1 H; H-20), 2.48 (dd, J= 11.5, 7 Hz; 1 H, H-14), 1.27 (d, J=6.3 Hz, 3 H; H-21), 1.21 (s, 6 H; H-28 and H-29), 0.68 ppm (s, 3 H; H-18); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta$ = 211.7 (C=O; C-8), 159.6 (C; C-25), 150.3 (C; C-22), 108.1 (C; C-24), 104.4 (CH; C-23), 70.4 (COH; C-27), 61.8 (CH; C-14), 55.5 (CH; C-17), 49.6 (C; C-13), 42.1 (CH<sub>2</sub>; C-26), 40.8 (CH<sub>2</sub>; C-12), 36.7 (CH<sub>2</sub>; C-9), 35.8 (CH; C-20), 28.9 (2×CH<sub>3</sub>, C-28 and C-29), 27.5 (CH<sub>2</sub>; C-16), 24.0 (CH<sub>2</sub>; C-15), 19.8 (CH<sub>3</sub>; C-21), 18.9 (CH<sub>2</sub>; C-11), 12.4 ppm (CH<sub>3</sub>; C-18). IR (film):  $\tilde{\nu}$ =3468, 2935, 1712 cm<sup>-1</sup>; HRMS (CI<sup>+</sup>): m/z: calcd for [C<sub>20</sub>H<sub>30</sub>O<sub>2</sub>]<sup>+</sup>: 302.2246 [M-H<sub>2</sub>O]<sup>+</sup>; found 302.2252.

(20S),(8*E*)-Bromomethylene-de-A,B-[5-(2-hydroxy-2-methylpropan-1-yl)furan-2-yl]pregnane (29 a): Procedure as for **17. 29a** [0.164 g, 0.417 mmol, 72%,  $R_f$ =0.4 (20% EtOAc/hexanes), yellow oil]. <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta$ =5.97 (d, *J*=2.9 Hz, 1 H; H-24), 5.84 (d, *J*=2.9 Hz, 1 H; H-23), 5.64 (s, 1 H; H-7), 2.74 (s, 2 H; H-26), 2.69 (m, 1 H; H-20), 1.25 (d, *J*= 6.9 Hz, 3 H; H-21), 1.22 (s, 6 H; H-28 and H-29), 0.61 ppm (s, 3 H; H-18); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta$ =160.2 (C; C-25), 150.2 (C; C-22), 144.8 (C; C-8), 108.1 (C; C-24), 104.2 (CH; C-23), 97.6 (CH; C-7), 70.5 (COH; C-27), 55.7 (CH; C-14), 54.7 (CH; C-17), 45.3 (C; C-13), 42.1 (CH<sub>2</sub>; C-26), 39.6 (CH<sub>2</sub>; C-12), 36.3 (CH; C-20), 31.0 (CH<sub>2</sub>; C-9), 29.0 (2×CH<sub>3</sub>; C-

610 -

28 and C-29), 27.6 (CH<sub>2</sub>; C-15), 22.5 (CH<sub>2</sub>; C-16), 21.9 (CH<sub>2</sub>; C-11), 20.0 (CH<sub>3</sub>; C-21), 11.8 ppm (CH<sub>3</sub>; C-18); IR (film):  $\tilde{\nu}$ =3418, 2965 cm<sup>-1</sup>; HRMS (CI<sup>+</sup>): *m*/*z*: calcd for [C<sub>21</sub>H<sub>31</sub>O<sub>2</sub>Br]<sup>+</sup>: 396.1487 [*M*+H]<sup>+</sup>; found 396.1485.

(20S),(8E)-De-A,B-[5-(2-hydroxy-2-methylpropan-1-yl)furan-2-yl]-

[(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)methylene]pregname (30a): Procedure as for 6a. 30a [87%,  $R_f$ =0.34 (20% EtOAc/hexanes), oil]. <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta$ =5.96 (d, J=2.9 Hz, 1H; H-24), 5.83 (d, J=2.9 Hz, 1H; H-23), 4.90 (s, 1H; H-7), 3.18 (dd, J=13.3, 3 Hz; 1H, H-9), 2.73 (s, 2H; H-26), 2.67 (dq, J=10.5, 7 Hz, 1H; H-20), 1.26–1.21 (m, 21H; H-21, H-28, H-29, and 4×Me-pinacol), 0.59 ppm (s, 3H; H-18); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta$ =165.7 (C; C-8), 160.5 (C; C-25), 150.0 (C; C-22), 108.0 (C; C-24), 104.1 (CH; C-23), 82.5 (CO; pinacol), 70.4 (COH; C-27), 57.8 (CH; C-14), 55.7 (CH; C-17), 46.0 (C; C-13), 42.1 (CH<sub>2</sub>; C-26), 40.2 (CH<sub>2</sub>; C-12), 36.4 (CH; C-20), 33.1 (CH<sub>2</sub>; C-9), 28.9 (2×CH<sub>3</sub>; C-28 and C-29), 27.4 (CH<sub>2</sub>; C-16), 24.9 (2×CH<sub>3</sub>; Me-pinacol), 24.8 (2×CH<sub>3</sub>; Me-pinacol), 24.2 (CH<sub>2</sub>; C-15), 22.1 (CH<sub>2</sub>; C-11), 20.0 (CH<sub>3</sub>; C-21), 12.0 ppm (CH<sub>3</sub>; C-18); IR (film):  $\tilde{\nu}$ =3459, 2974 cm<sup>-1</sup>; HRMS (CI<sup>+</sup>): m/z: calcd for [C<sub>27</sub>H<sub>44</sub>O<sub>4</sub>B]<sup>+</sup>: 443.3333 [*M*+H]<sup>+</sup>; found 443.3337.

### (20S)-[5-(2-Hydroxy-2-methylpropan-1-yl)furan-2-yl]-22,23,24,25,26,27-

hexanor-1 $\alpha$ -hydroxy-vitamin D<sub>3</sub> (4a): Procedure as for 3a. 4a [76%,  $R_{\rm f}$ = 0.11 (50% EtOAc/hexanes), amorphous white solid]; m.p. 77–79°C (Et<sub>2</sub>O/hexanes);  $[a]_D^{25}=1.2$  (c=2.9 in 96% EtOH; <sup>1</sup>H NMR (500 MHz,  $CDCl_3$ ):  $\delta = 6.36$  (d, J = 11.2 Hz, 1 H; H-6), 6.00 (d, J = 11.2 Hz, 1 H; H-7), 5.96 (d, J=2.9 Hz, 1 H; H-24), 5.83 (d, J=2.9 Hz, 1 H; H-23), 5.30 (s, 1 H; H-19E), 4.97 (s, 1H; H-19Z), 4.40 (m, 1H; H-1), 4.21 (m, 1H; H-3), 2.83 (dd, J=12.3, 3.4 Hz, 1H; H-9), 2.73 (s, 2H; H-26), 2.68 (m, 1H; H-20), 2.58 (dd, J=13.3, 2.5 Hz, 1H; H-4), 2.30 (dd, J=13.3, 6.5 Hz, 1H; H-4), 1.24 (d, J=6.9 Hz, 3 H; H-21), 1.21 (s, 6 H; H-28 and H-29), 0.59 ppm (s, 3H; CH<sub>3</sub>-18); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>):  $\delta = 160.4$  (C; C-25), 150.1 (C; C-22), 147.6 (C; C-10), 142.6 (C; C-8), 133.2 (C; C-5), 124.7 (CH; C-6), 117.2 (CH; C-7), 111.8 (CH<sub>2</sub>; C-19), 108.1 (CH; C-24), 104.1 (CH; C-23), 70.7 (CH; C-1), 70.5 (COH; C-27), 66.7 (CH; C-3), 56.2 (CH; C-17), 55.4 (CH; C-14), 45.7 (C; C-13), 45.2 (CH<sub>2</sub>; C-4), 42.8 (CH<sub>2</sub>; C-2), 42.1 (C; C-26), 40.2 (CH<sub>2</sub>; C-12), 36.4 (CH; C-20), 28.9 (CH<sub>2</sub>; C-9), 28.8 (2× CH3; C-28 and C-29), 27.6 (CH2; C-16), 23.5 (CH2; C-11), 22.1 (CH2; C-15), 20.0 (CH<sub>3</sub>; C-21), 12.0 ppm (CH<sub>3</sub>; C-18); IR (CHCl<sub>3</sub>):  $\tilde{\nu}$ =3346, 2954 cm<sup>-1</sup>; UV (96% EtOH):  $\lambda_{max}$  ( $\epsilon$ )=218 nm (18900),  $\lambda$ =266 nm  $(15000 \text{ mol}^{-1}\text{m}^3\text{cm}^{-1}), \lambda_{\min} = 246 \text{ nm}; \text{ HRMS (ESI-TOF): } m/z: \text{ calcd for}$  $[C_{29}H_{43}O_4]^+$ : 455.3156  $[M]^+$ ; found 455.3155.

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