DOI: 10.1002/ejoc.201101126

Synthesis of the Proposed Structure of Pentahydroxylated Pyrrolizidine Hyacinthacine C₅ and Its C₆,C₇ Epimer

Juan A. Tamayo,*^[a] Francisco Franco,*^[a] and Fernando Sánchez-Cantalejo^[a]

Keywords: Azasugars / Hyacinthacines / Enantioselectivity / Synthetic methods / Inhibitors

The synthesis of two pentahydroxylated pyrrolizidines, hyacinthacine C_5 (1) and 6,7-di-*epi*-hyacinthacine C_5 (12), from an orthogonally protected pyrrolidine of 2,5-dideoxy-2,5imino-D-mannitol configuration (i.e., 7) is described. The analytical and spectroscopic data of synthetic 1 present some differences with those of natural hyacinthacine C5 that exceed the previously observed variations in similar com-

Introduction

Hyacinthacine C_5 (1) was first isolated by Kato et al.^[1] in 2007 from the bulbs of Scilla Socialis and has shown moderate inhibitory activity against rat intestinal maltase, Caldocellum saccharolyticum β -glucosidase, and Aspergillus niger amiloglucosidase with an IC₅₀ value of 77, 48, and 57 µM, respectively. This natural compound belongs to one of the five structural classes utilized to classify iminosugars:^[2] polyhydroxylated pyrrolizidines. Hyacinthacines C, a subclass of this group, present the characteristic five hydroxy groups along their bicyclic ring, one of them being a hydroxymethyl group adjacent to the ring nitrogen at C-3 (Figure 1). Not many hyacinthacines C have been described to date,^[1,3] and only six compounds presenting a substitution pattern similar to that of hyacinthacine $C_5(1)$ and hyacinthacine C_1 (2) have been published, including a very recent synthesis of proposed 1.^[1,3a,3b,3d-3f] Recently, we described the synthesis of two pentahydroxylated pyrrolizidines (compounds 4 and 5), which are epimers of hyacinthacine C_1 , by using suitably protected α , β -unsaturated ketone 6 as the starting material.^[3a] This ketone was in turn obtained from a 2,5-dideoxy-2,5-imino-D-altritol (DALDP) derivative.^[4] In our synthesis, a highly stereoselective bishydroxylation of ketone 6 was the key step to achieve highly functionalized hyacinthacine C_1 epimers 4 and 5.



pounds. On this respect, additional studies on chemical de-

rivatives of synthetic 1 support our initially proposed struc-

ture and establish the absolute configuration of putative (+)-

hyacinthacine C_5 as that shown in **1**. These studies lead us

to conclude that the natural compound originally labeled as

(+)-hyacinthacine C₅ might be a different isomer.

(+)-7a-epi-Hyacinthacine C_1 (4) (+)-5,7a-Di-epi-hyacinthacine C_1 (5)



Figure 1. Some examples of hyacinthacine C1 and C5-type alkaloids 1-5.

Related to this work, and as part of our studies concerned with the synthesis of polyhydroxylated pyrrolizidines alkaloids (PHPAs),^[3a,5] we decided to approach the synthesis of hyacinthacine C_5 (1). In this case, an orthogonally protected pyrrolidine of DMDP (2,5-dideoxy-2,5-imino-Dmannitol) configuration (i.e., 7),^[5e,6] previously used by our group for the synthesis of (+)-casuarine (8),^[5e] was the intermediate chosen as the starting material. Orthogonally

[[]a] Department of Medicinal and Organic Chemistry, Faculty of Pharmacy, University of Granada, Campus de Cartuja s/n, 18071 Granada, Spain Fax: +34-958-243845 E-mail: jtamayo@ugr.es ffranco@ugr.es

Supporting information for this article is available on the WWW under http://dx.doi.org/10.1002/ejoc.201101126.

protected α,β -unsaturated ketone 9, derived from 7, would be a desirable intermediate that, when subjected to dihydroxylation, could led us to pentahydroxylated pyrrolidines 10 and 11, which are precursors of the desired hyacinthacine C₅-type bicyclic ring when subjected to the appropriate chemical transformations (Scheme 1). In this article, we report on the synthesis of putative (+)-hyacinthacine C₅ (1)and (+)-6,7-di-*epi*-hyacinthacine C₅ (12) by using the orthogonally protected pyrrolidine of DMDP configuration (i.e., 7) as the starting chiral building block. Following the synthesis of putative hyacinthacine C_5 (1) and 6,7-di-epihyacinthacine C_5 (12), we describe an exhaustive structural study by NMR spectroscopic analysis of final compound 1 and its hydrochloride salt, as well as some of its chemical precursors and derivatives. Analytical and spectroscopic data for synthetic 1 and its hydrochloride salt, however, did not match those reported by Kato for the natural product,^[1] but did however match those reported by Yu in a recent synthesis of ent-1.^[3a] Moreover, NOE studies on synthetic 1 and 1·HCl together with those of the chemical precursors of synthetic 1 prompt us to conclude, together with the

ref [56

Cbz

9

dihydroxylation

cyclization

OBr

HC

0

н

BnO

OTBDPS

HO

CbzN

α,β-dihydroxylated

ketone 11

6,7-Di-epi-hyacinthacine C₅ (12)

Me

HO

Mé

(+)-Casuarine (8)

OBn

OH

OH

OH

OBn

OTBDPS

Scheme 1. Previous synthesis of (+)-casuarine (8) from DMDP precursor 7 and retrosynthetic analysis of hyacinthacine C_5 (1) and 6,7-di-*epi*- C_5 (12).



Results and Discussion

According to the retrosynthetic analysis shown in Scheme 1, the synthesis started with previously described pyrrolidine 7.^[5e,6] Oxidation of the free OH group in 7 produced aldehyde 13,^[5e] which was treated in situ with 1-triphenylphosphoranylidene-2-propanone to afford key intermediate α,β -unsaturated ketone 9 (Scheme 2). Dihydroxylation of 9 with aqueous 1% OsO₄ yielded diol intermediates 10/11 as a 1.4:1 mixture^[7] that was easily separated by flash chromatography. The configurations of the two new generated stereogenic centers were not established at this point but later in the synthesis, because their ¹H NMR spectra appeared as an irresolvable mixture of rotamers. Catalytic hydrogenation of 10 and 11 in the presence of Pd/ C yielded pyrrolizidines 14 and 15, respectively, as the only isomers (Scheme 3). The absolute configurations of 14 and 15 were established on the basis of spectroscopic analysis carried out on their corresponding O-desilylation derivatives 16 and 17, respectively (Scheme 3). Their proposed structures were confirmed by NOE experiments, which showed positive interactions between 1-H-7-H, 6-H-7a-H, 6-H-Me, 1-H-5-H, and 5-H-7-H in 16 and between 1-H-3-H, 1-H-5-H, 5-H-6-H, and 7-H-7a-H in 17 (Figure 2).



Scheme 2. Synthesis of pyrrolidines 10 and 11.

In this sense, it is worth noting the stereoselectivity observed in the hydrogenation reactions of 10 and 11. As suggested by Scheme 3, initial *N*-Cbz removal affords intermediates A and A', which upon intramolecular reductive amination form intermediate bicyclic iminium ions B and B'. Highly stereoselective hydrogenation of such intermediates through the α -face yields pyrrolizidines 14 and 15, which both present the *S* configuration at C-5 (Figure 3). According to this description, the stereochemistry at C-6 remains unaffected during the reaction, and consequently pyrrolizidine 14 shows a *trans* disposition between the OH group at C-6 and the Me group at C-5, and pyrrolizidine 15 adopts a *cis* disposition. These experimental facts

OBn

7

OBn

OBn

OH

OH

OH

OBn

OTBDPS

OTBDPS

HO L

Cbz

HO

0-

Cbz

Me

HQ H

Me 9

Hyacinthacine $C_5(1)$

HO

α,β-dihydroxylated

FULL PAPER



Scheme 3. Synthesis of pyrrolizidines 1 and 12 from pyrrolidines 10 and 11 respectively.



Figure 2. Main NOE interactions in 16 and 17.



Figure 3. Proposed iminium intermediates B and B' in the catalytic hydrogenation of 10 and 11.

prompt us to state that iminium intermediates **B** and **B**' are the chemical species subjected to hydrogenation and therefore to conclude that α,β -dihydroxylated ketone **10** is the precursor of **14** having the 1'*S*,2'*R* configuration, whereas **11**, with a 1'*R*,2'*S* configuration, is the precursor of **15**.

To finish the synthesis, the transformation of **16** and **17** into target molecules **1** and **12** was achieved by *O*-debenzylation to proposed hyacinthacine C_5 (**1**) and 6,7-di-*epi*-hyacinthacine C_5 (**12**, Scheme 3).

The analytical and spectroscopic data of compounds **1** and **12** were established by careful studies and confirmed by extensive ¹H, ¹³C, and NOE NMR experiments. As previously described by different authors,^[8] analytical and spectroscopic data of synthetic alkaloids can vary with those of natural compounds. Metal ions, ionic strength of the solvent, pH, and purification methodology can be responsible for these differences, especially in their NMR chemical shift values.

In our case, the analytical and spectroscopic data of synthetic 1 and 1·HCl are inconsistent with those data described for natural hyacinthacine C₅. ¹H and ¹³C NMR experiments carried out on synthetic 1 and 1·HCl under the same conditions as those described by Kato^[1] [D₂O and 3-(trimethylsilyl)propionate (TSP) as an internal standard, see the Supporting Information] do not match those reported for hyacinthacine C₅ (see Tables 1 and 2). Especially significant are the ¹³C NMR spectroscopic data. In both compounds, the chemical shifts of C-2 and C-8 appear upfield with respect to those of natural hyacinthacine C₅ (Table 2,

Table 1. ¹H NMR spectroscopic data of synthetic 1, 1·HCl, and natural 1 at 500 MHz in D_2O .^[a]

Entry	Proton	1·HCl	Synthetic 1	Natural 1
1	1 - H	4.42	4.19	4.01
2	2-H	4.15	3.99	3.80
3	3-H	3.73	2.97-2.93	3.01
4	5-H	3.64	2.97-2.93	2.81
5	6-H	3.93-3.96	3.72-3.66	3.60
6	7-H	4.38	4.15	3.99
7	7a-H	3.82-3.86	3.08	3.24
8	8-H	3.93-3.96	3.72-3.66	3.50
9	8' - H	3.82-3.86	3.72-3.66	3.55

[a] Chemical shifts are expressed in ppm downfield from sodium TSP.

Table 2. ^{13}C NMR spectroscopic data of synthetic 1, 1·HCl, and natural 1 at 125 MHz in $D_2O.^{[a]}$

Carbon	1.HC1		Synthetic 1		
Carbon	(x)	(x - z)	(y)	(y-z)	(<i>z</i>)
C-1	80.0	+1.8	79.6	+1.4	78.2
C-2	79.1	-1.9	78.7	-2.3	81.0
C-3	76.6	+11.5	65.3	+0.2	65.1
C-5	71.1	+9.7	70.4	+9	61.4
C-6	82.5	+0.8	82.3	+0.6	81.7
C-7	79.5	+1.7	79.0	+1.2	77.8
C-7a	76.1	+6.9	70.5	+1.3	69.2
C-8	60.9	-4.8	62.2	-3.5	65.7
C-9	17.0	+1.3	17.2	+1.5	15.7
	Carbon C-1 C-2 C-3 C-5 C-6 C-7 C-7a C-7a C-8 C-9	Carbon 1-HCl (x) C-1 80.0 C-2 79.1 C-3 76.6 C-5 71.1 C-6 82.5 C-7 79.5 C-7a 76.1 C-8 60.9 C-9 17.0	$\begin{array}{c cccc} Carbon & 1 \cdot HCl & & & \\ \hline (x) & (x-z) \\ \hline C-1 & 80.0 & +1.8 \\ C-2 & 79.1 & -1.9 \\ C-3 & 76.6 & +11.5 \\ C-5 & 71.1 & +9.7 \\ C-6 & 82.5 & +0.8 \\ C-7 & 79.5 & +1.7 \\ C-7a & 76.1 & +6.9 \\ C-8 & 60.9 & -4.8 \\ C-9 & 17.0 & +1.3 \\ \hline \end{array}$	Carbon 1·HCl Synthetic 1 (x) $(x-z)$ (y) C-1 80.0 +1.8 79.6 C-2 79.1 -1.9 78.7 C-3 76.6 +11.5 65.3 C-5 71.1 +9.7 70.4 C-6 82.5 +0.8 82.3 C-7 79.5 +1.7 79.0 C-7a 76.1 +6.9 70.5 C-8 60.9 -4.8 62.2 C-9 17.0 +1.3 17.2	$\begin{array}{c c c c c c c c c c c c c c c c c c c $

[a] Chemical shifts are expressed in ppm downfield from sodium TSP.

Entries 2 and 8), whereas the rest of the carbon atoms are moved downfield. In addition, C-5 in synthetic 1 and 1·HCl shows a 9 and 9.7 ppm downfield shift, respectively, to that of C-5 in natural hyacinthacine C₅ (Table 2, Entry 4), whereas the shift for the rest of the carbon atoms ranges from 0.2 to 6.9 ppm (see Table 2). As explained above, certain shifts can be expected between synthetic and natural alkaloids; still, those differences vary evenly for all the signals, which is not the case here. Also significant are the specific rotation values of both synthetic 1 { $[a]_D^{30} = +8 (c =$ 1, H₂O)} and 1·HCl { $[a]_D^{28} = +18 (c = 1, H_2O)$ }, which in concordance with the NMR results do not match those reported for natural 1 {ref.^[1] $[a]_D = +1.5 (c = 0.22, H_2O)$ }.

These results prompted us to consider that the compound initially assigned as hyacinthacine C_5 could be a different isomer. To support this conclusion we decided to carry out additional experiments. In this sense, pyrrolizidines 14 and 16, chemical precursors of synthetic 1, and synthetic 1 itself were acetylated to 18, 19, and peracetylated 20, respectively (Scheme 4). NOE studies on these compounds showed positive interactions between 1-H–7-H, 1-H–5-H, and 5-H–7-H in all three derivatives 18, 19, and 20.^[9] These results definitely support the stereochemistry initially assigned by us to synthetic 1 confirming its structure as the one depicted in Scheme 3 and proving that the natural compound originally described as (+)-hyacinthacine C_5 is in fact a different isomer.^[3a]



Scheme 4. Synthesis of acetylated derivatives 18, 19, and 20.

Conclusions

We describe herein the synthesis of two pentahydroxylated pyrrolizidines, putative hyacinthacine C_5 (1) and 6,7di-*epi*-hyacinthacine C_5 (12), from an orthogonally protected pyrrolidine of DMDP configuration (i.e., 7) as starting chiral building block. The analytical and spectroscopic data of compounds 1 and 12 were established by extensive ¹H, ¹³C, and NOE NMR experiments. Some differences exist between the analytical and spectroscopic data of natural hyacinthacine C_5 (1) and synthetic 1, which, although to a certain point expected, exceed the previously observed variations in similar compounds.^[8] On this respect, additional studies on chemical derivatives of synthetic 1 support the initially proposed structure and establish the absolute configuration of putative hyacinthacine C_5 as that of structure

figuration of putative hyacinthacine C_5 as that of structure 1. These results, together with those recently published by Yu, lead us to conclude that the natural compound initially labeled as (+)-hyacinthacine C_5 might be a different isomer.

Experimental Section

General Remarks: Solutions were dried with MgSO4 before concentration under reduced pressure. The ¹H and ¹³C NMR spectra were recorded with Varian Direct Drive 400 and 500 MHz spectrometers for solutions in CDCl3 (internal Me4Si). Splitting patterns are designated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet, and br., broad. IR spectra were recorded with a Perkin-Elmer FTIR Spectrum One instrument, and mass spectra were recorded with a Hewlett-Packard HP-5988-A and Fisons mod. Platform II and VG Autospec-Q mass spectrometers or a NALDI ionization-time-of-flight (NALDI-TOF) mass spectrometer and EI mass spectrometer. Optical rotations were measured for solutions in CHCl₃ (1-dm tube) with a Jasco DIP-370 polarimeter. TLC was performed on precoated silica gel 60 F254 aluminum sheets and detection by employing a mixture of 10% ammonium molybdate (w/v) in 10% aqueous sulfuric acid containing 0.8% cerium sulfate (w/v) and heating. Column chromatography was performed on silica gel (Merck, 7734). All compounds were shown to be homogeneous by chromatographic methods and characterized by NMR spectroscopy and MS and HRMS.

(E)-4-[(2'R,3'R,4'R,5'R)-3',4'-Dibenzyloxy-N-benzyloxycarbonyl-5'-tert-butyldiphenylsilyloximethylpyrrolidin-2'-yl]but-3-en-2-one (9): To a solution of 7 (1.01 g, 1.4 mmol) in dry DCM (15 mL) was added activated powdered 4 Å molecular sieves (200 mg), Nmethylmorpholine N-oxide (250 mg, 2.1 mmol), and tetrapropylammonium perruthenate (TPAP, 100 mg), and the reaction mixture was kept at room temperature for 1 h. TLC (Et₂O/hexane, 3:2) then showed a faster-running compound. The reaction was diluted with Et₂O (30 mL), filtered through a bed of Silica gel 60 (230-400 mesh), and thoroughly washed with Et₂O. The combined filtrate and washings were concentrated to aldehyde 13 (955 mg, 95%), which was used in the next step. IR (KBr, neat): $\tilde{v} = 3063$ and 3029 (Ph), 1735 (CHO), 1724 (C=O, Cbz) cm⁻¹. A solution of 13 (955 mg, 1.34 mmol) and 1-(triphenylphosphoranylidene)-2propanone (1.35 g, 4.2 mmol) in dry toluene (15 mL) was heated at reflux for 24 h. TLC then showed a slower-running compound. The solvent was eliminated, and the residue was submitted to column chromatography (Et₂O/hexane, 3:2) to afford syrup 9 (870 mg, 86%, from 7). $[a]_{D}^{30} = +8$, $[a]_{405}^{30} + 7$ (c = 1, CHCl₃). IR (neat): $\tilde{v} =$ 3032 and 2930 (Ph), 1704 and 1677 (C=O) cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 7.73–7.12 (2 m, 25 H, 5Ph), 6.69 (ddd, $J_{3,4}$ = 16.1 Hz, $J_{4.5}'$ = 7.7 Hz, 1 H, 4-H, two rotamers), 6.04 (2 d, 1 H, 3-H, two rotamers), 5.22–4.87 (4 d, J = 12.1 Hz, 2 H, CH₂Ph, two rotamers), 4.74-3.72 (br. m, 10 H, 2 CH₂Ph, 2',3',4',5',2''a,2''b-H,), 2.19 and 2.01 (2 s, 3 H, 1,1,1-H, two rotamers), 1.11 and 1.06 (2 s, 9 H, CMe₃, two rotamers) ppm. ¹³C (125 MHz, CDCl₃): δ = 198.2 (C-2), 154.5 (Cbz), 145.2, 144.9, 137.5, 135.5, 133.3, 131.4,

FULL PAPER

129.7, 127.9, 86.4, and 85.5 (C-3', two rotamers), 82.4 and 81.4 (C-4', two rotamers), 71.8, 71.7, 71.4 and 71.2 (2 CH₂Ph, two rotamers), 67.3, 67.1 (Cbz, two rotamers), 65.4 and 65.4 (C-2', two rotamers), 65.1 (C-5'), 62.1 and 61.4 (C-5'', two rotamers), 27.0, 26.9 (CMe₃, two rotamers), 19.4 and 19.2 (Me, two rotamers) ppm. HRMS (NALDI-TOF): calcd. for $C_{47}H_{51}NO_6NaSi [M + H]^+$ 776.3383; found 776.3381 (deviation +0.3 ppm).

(2R,3R,4R,5R)-3,4-Dibenzyloxy-N-benzyloxycarbonyl-2-tertbutyldiphenylsilyloxymethyl-5-[(1'S,2'R)-1'-2'-dihydroxy-3'-oxobutyl]pyrrolidine (10) and (2R,3R,4R,5R)-3,4-Dibenzyloxy-N-benzyloxycarbonyl-2-tert-butyldiphenylsilyloxymethyl-5-[(1'R,2'S)-1'-2'dihydroxy-3'-oxobutyl]pyrrolidine (11): To a stirred solution of 9 (855 mg, 1.13 mmol) in acetone/water (8:1, 13.5 mL) was added NMO (332 mg, 2.83 mmol) and aqueous 1% OsO₄ (2.5 mL). The mixture was left at room temperature for 20 h. TLC (Et₂O/hexane, 3:2) then revealed the presence of two new products of lower mobility. The mixture was concentrated to a residue that was submitted to chromatography (Et₂O/hexane, 1:1) to afford first 10 as a syrup. Yield: 328 mg (37%). $[a]_D^{24} = -25$ (c = 1, CHCl₃). IR (neat): $\tilde{v} = 3435$ (OH), 1702 and 1679 (C=O) cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ = 7.73–7.07 (2 m, 25 H, 5Ph), 5.06–4.85 (m, 2 H, CH₂Ph), 4.69–4.02 (2 br. m, 11 H, 2CH₂Ph, 3',4',5',2''a,2''b,3,4-H), 3.92 (dd, 1 H, OH, two rotamers), 3.82-3.66 (m, 1 H, 2'-H), 3.53 (dd, 1 H, OH, two rotamers), 2.25 and 1.79 (2 s, 3 H, 1,1,1-H, two rotamers), 1.08 and 1.01 (2 s, 9 H, CMe₃, two rotamers) ppm. ¹³C NMR (125 MHz, CDCl₃): δ = 208.1 (C-2), 155.4 (Cbz), 137.7, 136.7, 135.9, 135.5, 133.3, 129.8, 128.2, 127.8, 82.1 (C-3'), 81.3 (C-4'), 78.3 (C-3), 71.6 (C-4), 69.8 (C-2'), 68.0 (C-5'), 67.3, 65.2 and 61.8 (2CH₂Ph and Cbz), 26.8 (CMe₃), 25.9 (Me) and 19.2 (CMe₃) ppm. HRMS (NALDI-TOF): calcd. for C₄₇H₅₄NO₈Si [M + H]⁺ 788.3619; found 788.3607 (deviation -1.5 ppm). Eluted second was 11 (246 mg, 28%) as a syrup. $[a]_D^{26} = -5$, $[a]_{546}^{27} = -6$ (c 1, CHCl₃). IR (neat): $\tilde{v} = 3389$ (OH), 1721 and 1672 (C=O) cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 7.61–7.12 (2 m, 25 H, 5Ph), 5.00 (m, 2 H, CH₂Ph), 4.63 (m, 2 H, CH₂Ph), 4.50 (m, 2 H, 3,4-H), 4.40 (m, 2 H, CH₂Ph), 4.17 (m, 3 H, 2"a,2"b,3'-H), 4.04 (m, 1 H, 2'-H), 3.78 (m, 2 H, 4',5'-H), 2.01 (s, 3 H, Me), 1.04 (s, 9 H, CMe₃) ppm. ¹³C NMR (125 MHz, CDCl₃): δ = 207.0 (C-2), 157.9 (Cbz), 137.6, 137.5, 135.6, 135.4, 133.4, 133.3, 129.8, 128.5, 127.7, 82.6 (C-3'), 81.7 (C-4'), 78.4 (C-3), 74.6 (C-4), 71.5 and 71.1 (C-2', two rotamers), 67.8 (C-5'), 67.6, 65.2 and 61.7 (2 CH₂Ph, Cbz), 26.8 (CMe₃), 25.0 (Me) and 19.2 (CMe₃) ppm. HRMS (NALDI-TOF): calcd. for C47H54NO8Si [M + H]+ 788.3619; found 788.3616 (deviation -0.4).

(1R,2R,3R,5S,6S,7S,7aR)-1,2-Dibenzyloxy-3-tert-butyldiphenylsilyloxymethyl-6,7-dihydroxy-5-methylpyrrolizidine (14): Compound 10 (230 mg, 0.29 mmol) in MeOH (4 mL) was hydrogenated (60 psi H₂) in the presence of 10% Pd/C (35 mg) overnight. TLC (AcOEt/hexane, 3:1) then showed a slower-running compound. The catalyst was filtered off, washed with MeOH, and the combined filtrate and washings were concentrated to a residue that was submitted to chromatography (AcOEt/hexane, 1:1) to afford pure 14 (121 mg, 65%) as a colorless viscous syrup. $[a]_{D}^{25} = -5$ (c = 1, CHCl₃). IR (neat): $\tilde{v} = 3368$ (OH) cm⁻¹. ¹H NMR (400 MHz, CDCl₃): *δ* = 7.70–7.22 (2 m, 20 H, 4Ph), 4.52 (2 d, 4 H, 2CH₂Ph), 4.23 (t, $J_{1,2} = J_{2,3} = 2.3$ Hz, 1 H, 2-H), 4.13–4.07 (m, 1 H, 1-H), 3.99 (t, $J_{6,7} = J_{7,7a} = 7.5$ Hz, 1 H, 7-H), 3.72–3.56 (m, 3 H, 6,8,8'-H), 3.40 (m, 1 H, 7a-H), 3.18 (m, 1 H, 3-H), 2.95 (m, 1 H, 5-H), 1.08 (d, $J_{Me,5} = 6.2$ Hz, 3 H, Me), 1.04 (s, 9 H, CMe₃) ppm. ¹³C NMR (125 MHz, CDCl₃): δ = 138.0, 135.6, 135.6, 133.6, 133.5, 129.6, 129.6, 128.4, 128.4, 127.7, 127.6, 127.6, 127.5, 87.1 (C-1), 86.0 (C-2), 83.0 (C-6), 80.1 (C-7), 72.8 (C-7a), 71.6 (CH₂Ph), 71.5 (CH₂Ph), 71.3 (C-3), 65.2 (C-5), 65.1 (C-8), 26.9 (CMe₃), 19.2 (Me), 19.2 (*C*Me₃) ppm. HRMS (NALDI-TOF): calcd. for $C_{39}H_{48}NO_5Si [M + H]^+$ 638.3302; found 638.3307 (deviation +0.5 ppm).

(1R,2R,3R,5S,6S,7S,7aR)-1,2-Dibenzyloxy-6,7-dihydroxy-3hydroxymethyl-5-methylpyrrolizidine (16): To a stirred solution of 14 (51 mg, 0.08 mmol) in THF (2.5 mL) was added TBAF·3H₂O (76 mg, 0.24 mmol). The mixture was kept at 50 °C for 16 h. TLC (AcOEt/H₃CCN/MeOH/H₂O, 70:10:10:5) then showed a new slower-running compound. The reaction mixture was evaporated and subjected to chromatography (AcOEt/H₃CCN/MeOH/H₂O, 95:10:4:2) to afford pure **16** (30 mg, 94%). $[a]_{D}^{28} = -14$ (c = 1, CHCl₃). IR (neat): $\tilde{v} = 3368$ (OH) cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 7.37–7.23 (m, 10 H, 2 Ph), 4.63 and 4.60 (2 d, J = 3.1 Hz, 2 H, CH₂Ph), 4.52 and 4.47 (2 d, J = 11.8 Hz, 2 H, CH₂Ph), 4.10 (t, $J_{1.7a} = J_{1.2} = 3.7$ Hz, 1 H, 1-H), 4.00 (t, $J_{6.7} = J_{7.7a} =$ 7.55 Hz, 1 H, 7-H), 3.96 (t, $J_{2,3}$ = 3.7 Hz, 1 H, 2-H), 3.86 (br. s, 1 H, OH), 3.73 (t, $J_{5.6} = 7.55$ Hz, 1 H, 6-H), 3.59–3.43 (m, 3 H, 7a,8,8'-H), 3.19 (m, 1 H, 3- H), 2.96 (m, 1 H, 5-H), 1.21 (d, J_{Me.5} = 6.22 Hz, 3 H, Me) ppm. ¹³C NMR (125 MHz, CDCl₃): δ = 131.2, 131.2, 130.6, 130.5, 130.5, 130.3 (Ph), 89.4 (C-1), 87.9 (C-2), 85.2 (C-6), 82.7 (C-7), 74.9 (CH₂Ph), 74.6 (C-7a), 74.5 (CH₂Ph), 73.7 (C-3), 67.5 (C-5), 63.3 (C-8), 20.9 (Me) ppm. HRMS (NALDI-TOF): calcd. for C₂₃H₃₀NO₅ [M + H]⁺ 400.2124; found 400.2115 (deviation -2.2 ppm).

(1R,2R,3R,5S,6S,7S,7aR)-1,2,6,7-Tetrahydroxy-3-hydroxymethyl-5-methylpyrrolizidine [(+)-hyacinthacine C₅, 1]: Compound 16 (41 mg, 0.10 mmol) in MeOH (4.5 mL) and conc. HCl (four drops) was hydrogenated (70 psi H₂) in the presence of 10% Pd/C (16 mg) for 27 h. The catalyst was filtered off, washed with MeOH, and the combined filtrate and washings were treated with Amberlite IRA-400 resin (OH⁻ form). Evaporation of the solvent afforded pure 1 (16 mg, 73%) as a colorless viscous syrup. $[a]_{D}^{28} = +8$ ($c = 1, H_{2}O$). IR (neat): $\tilde{v} = 3363$ (OH) cm⁻¹. ¹H NMR (500 MHz, D₂O): $\delta =$ 4.19 (t, $J_{1,2} = J_{1,7a} = 6.9$ Hz, 1 H, 1-H), 4.15 (t, $J_{6,7} = J_{7,7a} = 6.9$ Hz, 1 H, 7-H), 3.99 (t, $J_{2,3}$ = 6.9 Hz, 1 H, 2-H), 3.72–3.66 (m, 3 H, 6,8,8-H), 3.08 (t, 1 H, 7a-H), 2.97-2.93 (m, 2 H, 3,5-H), 1.24 (d, $J_{\text{Me},5}$ = 6.3 Hz, 3 H, Me) ppm. ¹³C NMR (125 MHz, D₂O): δ = 82.3 (C-6), 79.6 (C-1), 79.0 (C-7), 78.7 (C-2), 70.5 (C-7a), 70.4 (C-5), 65.3 (C-3), 62.2 (C-8), 17.2 (Me) ppm. HRMS (NALDI-TOF): calcd. for C₉H₁₈NO₅ [M + H]⁺ 220.1185; found 220.1182 (deviation -1.4 ppm).

(1*R*,2*R*,3*R*,5*S*,6*S*,7*S*,7*aR*)-1,2,6,7-Tetrahydroxy-3-hydroxymethyl-5-methylpyrrolizidine Hydrochloride [1·HCl]: Conc. HCl (three drops) was added to a solution of 1 (16 mg, 0.073 mmol) in H₂O (4 mL). Evaporation of the solvent afforded 1·HCl (18 mg, quantitative). $[a]_{12}^{28} = +18$ (c = 1, H₂O). ¹H NMR (500 MHz, D₂O): $\delta =$ 4.42 (t, $J_{1,2} = J_{1,7a} = 5.1$ Hz, 1 H, 1-H), 4.38 (t, $J_{6,7} = J_{7,7a} = 6.6$ Hz, 1 H, 7-H), 4.15 (t, $J_{2,3} = 5.2$ Hz, 1 H, 2-H), 3.96–3.93 (m, 2 H, 6,8-H), 3.86–3.82 (m, 2 H, 7a,8'-H), 3.73 (m, 1 H, 3-H), 3.64 (quint., $J_{5,6} = 6.9$ Hz, 1 H, 5-H), 1.50 (d, $J_{Me,5} = 6.8$ Hz, 3 H, Me) ppm. ¹³C NMR (125 MHz, D₂O): $\delta = 82.5$ (C-6), 80.0 (C-1), 79.5 (C-7), 79.1 (C-2), 76.6 (C-3), 76.1 (C-7a), 71.1 (C-5), 60.9 (C-8), 17.0 (Me) ppm.

(1*R*,2*R*,3*R*,5*S*,6*R*,7*R*,7*aR*)-1,2-Dibenzyloxy-3-*tert*-butyldiphenylsilyloxymethyl-6,7-dihydroxy-5-methylpyrrolizidine (15): Compound 11 (200 mg, 0.25 mmol) in MeOH (3.5 mL) was hydrogenated (60 psi H₂) in the presence of 10% Pd/C (40 mg) overnight. TLC (AcOEt/hexane, 3:1) then showed a slower-running compound. The catalyst was filtered off, washed with MeOH, and the combined filtrate and washings were concentrated to a residue that was submitted to chromatography (AcOEt/hexane, 3:2) to afford pure 15 (91 mg, 56%) as a colorless viscous syrup. $[a]_D^{25} = +14$ (*c*



= 1, CHCl₃). IR (neat): \tilde{v} = 3459 (OH) cm⁻¹. ¹H NMR (400 MHz): δ = 7.70–7.24 (2 m, 20 H, 4 Ph), 4.58 and 4.56 (2 d, *J* = 12.0 Hz, 2 H, CH₂Ph), 4.50 (t, *J* = 12.0 Hz, 2 H, CH₂Ph), 4.36 (m, 1 H, 2-H), 4.28 (m, 1 H, 1-H), 4.01 (m, 1 H, 7-H), 3.97 (m, 1 H, 7a-H), 3.86 (m, 1 H, 6-H), 3.76 (t, *J* = 10.0 Hz, 1 H, 8-H), 3.61 (m, 1 H, 8'-H), 3.41 (d, 1 H, OH), 3.19 (m, 1 H, 3-H), 3.13 (m, 1 H, 5-H), 1.06 (s, 9 H, CMe₃), 1.01 (d, *J*_{Me,5} = 6.5 Hz, 3 H, Me) ppm. ¹³C NMR (125 MHz): δ = 140.5, 139.5, 138.2, 136.4, 136.2, 132.3, 132.2, 131.3, 131.1, 130.8, 130.5, 130.3, 130.3, 130.2, 87.7 (C-2), 85.1 (C-1), 83.8 (C-6), 79.7 (C-7), 75.8 (C-7a), 74.5 and 74.5 (2 CH₂Ph), 73.4 (C-3), 67.9 (C-8), 65.5 (C-5), 29.5 (C*Me*₃), 21.9 (Me), 17.6 (*CMe*₃) ppm. HRMS (NALDI-TOF): calcd. for C₃₉H₄₈NO₅Si [M + H]⁺ 638.3302; found 638.3311 (deviation +1.4 ppm).

(1R,2R,3R,5S,6R,7R,7aR)-1,2-Dibenzyloxy-6,7-dihydroxy-3hydroxymethyl-5-methylpyrrolizidine (17): To a stirred solution of 15 (86 mg, 0.13 mmol) in THF (3 mL) was added TBAF \cdot 3H₂O (106 mg, 0.34 mmol). The mixture was kept at 55 °C for 4 h. TLC (AcOEt/MeCN/MeOH/H2O, 70:10:5:5) then showed a new slowerrunning compound. The reaction mixture was evaporated and subjected to chromatography (AcOEt/MeCN/MeOH/H₂O, 70:10:5:5) to afford pure 17 as a syrup (41 mg, 76%). $[a]_{D}^{28} = +39$ (c = 1, CHCl₃). IR (neat): $\tilde{v} = 3391$ (OH) cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 7.39–7.22 (m, 10 H, 2 Ph), 4.73 and 4.62 (2 d, J = 11.8 Hz, 2 H, CH₂Ph), 4.58 and 4.52 (2 d, *J* = 11.5 Hz, 2 H, CH₂Ph), 4.45 (t, $J_{1,2} = J_{1,7a} = 5.9$ Hz, 1 H, 1-H), 4.15 (m, 1 H, 2-H), 4.00 (m, 1 H, 7-H), 3.94 (m, 1 H, 6-H), 3.71 (t, J_{7.7a} = 5.9 Hz, 1 H, 7a-H), 3.65 (dd, $J_{3,8}$ = 3.9 Hz, $J_{8,8'}$ = 11.4 Hz, 1 H, 8-H), 3.56 (dd, $J_{3,8'}$ = 4.1 Hz, 1 H, 8'-H), 3.20 (m, 1 H, 5-H), 2.99 (m, 1 H, 3-H), 1.19 (d, $J_{Me,5}$ = 6.7 Hz, 3 H, Me) ppm. ¹³C (125 MHz, $CDCl_3$): $\delta = 138.2, 127.9, 127.6, 127.5, 127.3, 127.2$ (Ph), 84.8 (C-2), 80.7 (C-6), 80.4 (C-1), 75.9 (C-7), 72.2 (CH₂Ph), 71.8 (CH₂Ph), 71.1 (C-7a), 70.6 (C-3), 63.4 (C-5), 60.3 (C-8), 13.4 (Me) ppm. HRMS (NALDI-TOF): calcd. for $C_{23}H_{30}NO_5 [M + H]^+ 400.2124$; found 400.2116 (deviation -0.8 ppm).

(1R,2R,3R,5S,6R,7R,7aR)-1,2,6,7-Tetrahydroxy-3-hydroxymethyl-5-methylpyrrolizidine [(+)-6,7-Di-epi-hyacinthacine C₅, 12]: Compound 17 (44 mg, 0.11 mmol) in MeOH (4 mL) and conc. HCl (four drops) was hydrogenated (70 psi H_2) in the presence of 10% Pd/C (20 mg) for 20 h. The catalyst was filtered off, washed with MeOH, and the combined filtrate and washings were treated with Amberlite IRA-400 resin (OH⁻ form). Evaporation of the solvent afforded pure 12 (24 mg, 99%) as a colorless viscous syrup. $[a]_{D}^{30} =$ +8 ($c = 1, H_2O$) {ref.^[1] [a]_D = +1.5 ($c = 0.22, H_2O$)}. IR (neat): \tilde{v} = 3363 (OH) cm⁻¹. ¹H NMR (500 MHz, D₂O): δ = 4.31 (t, J_{1,2} = $J_{1,7a} = 7.5$ Hz, 1 H, 1-H), 4.24 (m, 1 H, 7-H), 4.15 (m, 1 H, 6-H), 3.99 (br. t, $J_{2,3}$ = 7.5 Hz, 1 H, 2-H), 3.80 (br. dd, $J_{3,8}$ = 4.0 Hz, $J_{8,8'}$ = 11.9 Hz, 1 H, 8-H), 3.71 (br. dd, $J_{3,8'}$ = 4.7 Hz, 1 H, 8'-H), 3.49 (m, 1 H, 7a-H), 3.16 (m, 1 H, 5-H), 2.81 (m, 1 H, 3-H), 1.19 (d, $J_{\text{Me},5} = 6.7 \text{ Hz}, 1 \text{ H}, \text{ Me}$ ppm. ¹³C NMR (125 MHz, D₂O): $\delta =$ 79.7 (C-6), 78.7 (C-2), 74.3 (C-7), 73.1 (C-1), 70.6 (C-3), 69.1 (C-7a), 62.7 (C-5), 61.6 (C-8), 14.0 (Me) ppm. HRMS (NALDI-TOF): calcd. for C₉H₁₇NO₅Na [M + Na]⁺ 242.1004; found 242.1012 (deviation +3.3 ppm).

(1*R*,2*R*,3*R*,5*S*,6*S*,7*S*,7a*S*)-6,7-Di-*O*-acetyl-1,2-dibenzyloxy-3-*tert*butyldiphenylsilyloxymethyl-6,7-dihydroxy-5-methylpyrrolizidine (18): Compound 14 (43 mg, 0.067 mmol) was acetylated in dry pyridine (1.5 mL), acetic anhydride (32 μ L, 0.337 mmol), and DMAP (cat.) at room temperature for 1.5 h. TLC (AcOEt/hexane, 2:1) then showed a faster-running compound. The reaction mixture was supported on silica gel and chromatographed (AcOEt/hexane, 1:5) to afford pure 18 (17 mg, 35%) as a colorless viscous syrup. $[a]_{D}^{30} =$ $-3 (c = 1, CHCl_3)$. IR (neat): $\tilde{v} = 1743$ (C=O) cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 7.67–7.62 and 7.40–7.22 (2 m, 20 H, 4 Ph), 5.29 (dd, $J_{6,7}$ = 6.7 Hz, $J_{7,7a}$ = 5.6 Hz, 1 H, 7-H), 4.97 (dd, $J_{5,6}$ = 5.9 Hz, 1 H, 6-H), 4.59–4.46 (m, 4 H, 2CH₂Ph), 4.23 (br. t, $J_{1,2}$ = $J_{1,7a}$ = 3.3 Hz, 1 H, 1 H, 1-H), 4.12 (br. t, $J_{2,3}$ = 3.0 Hz, 1 H, 2-H), 3.68 (dd, $J_{8,8'}$ = 9.9 Hz, $J_{3,8}$ = 7.7 Hz, 1 H, 8-H), 3.58 (dd, $J_{3,8'}$ = 6.3 Hz, 1 H, 8'-H), 3.45 (dd, 1 H, 7a-H), 3.24 (quint., $J_{5,9}$ = 5.9 Hz, 1 H, 5-H), 3.19 (m, 1 H, 3-H), 2.04 and 1.99 (2 s, 6 H, 2Ac), 1.04 (d, $J_{Me,5}$ = 6.1 Hz, 3 H, Me), 1.03 (s, 9 H, CMe₃) ppm. ¹³C NMR (125 MHz, CDCl₃): δ = 173.2 and 173.0 (C=O), 140.8, 140.6, 138.3, 131.0, 130.3, 130.2 (Ph), 89.7 (C-1), 88.6 (C-2), 84.6 (C-6), 82.1 (C-7), 74.8 (C-7a), 74.5 (CH₂Ph), 74.4 (CH₂Ph), 73.3 (C-3), 68.2 (C-8), 66.4 (C-5), 29.5 (CMe₃), 23.6 (2 CH₃CO), 22.0 (Me), 21.9 (CMe₃) ppm. HRMS (NALDI-TOF): calcd. for C₄₃H₅₂NO₇Si [M + H]⁺ 722.3505; found 722.3513 (deviation +1.1 ppm).

(1R,2R,3R,5S,6S,7S,7aS)-6,7-Di-O-acetyl-3-acetyloxymethyl-1,2-dibenzyloxy-6,7-dihydroxy-5-methylpyrrolizidine (19): Compound 16 (46 mg, 0.115 mmol) was acetylated in dry pyridine (2 mL), acetic anhydride (87 µL, 0.92 mmol), and DMAP (cat.) at room temperature for 1 h. TLC (AcOEt/hexane, 2:1) then showed a faster-running compound. The reaction mixture was supported on silica gel and chromatographed (AcOEt/hexane, 1:3) to afford pure 19 (31 mg, 51%) as a colorless viscous syrup. $[a]_D^{27} = +2$ (c = 1, CHCl₃). IR (neat): $\tilde{v} = 1742$ (C=O) cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 7.35–7.29 (m, 10 H, 2 Ph), 5.29 (t, $J_{6,7} = J_{7,7a} = 5.9$ Hz, 1 H, 7-H), 5.02 (t, $J_{5.6}$ = 6.1 Hz, 1 H, 6-H), 4.61 and 4.51 (2 d, J = 11.9 Hz, 2 H, CH₂Ph) 4.55 and 4.47 (2 d, J = 11.8 Hz, 2 H, CH₂Ph), 4.23 (t, $J_{1,2} = J_{1,7a} = 3.5$ Hz, 1 H, 1-H), 4.02 (m, 2 H, 8,8'-H), 3.91 (t, J_{2,3} = 3.3 Hz, 1 H, 2-H), 3.53 (br. dd, 1 H, 7a-H), 3.28 (m, 1 H, 3-H), 3.20 (quint., 1 H, 5-H), 2.05, 2.01 and 2.00 (3 s, 9 H, 3Ac), 1.13 (d, $J_{Me,5}$ = 6.1 Hz, 3 H, Me) ppm. ¹³C NMR (150 MHz, CDCl₃): δ = 173.4, 173.2 and 172.9 (C=O), 140.4, 131.10, 131.08, 130.5, 130.4, 130.3 (Ph), 88.8 (C-1), 88.2 (C-2), 84.2 (C-6), 81.8 (C-7), 74.66 (CH₂Ph), 74.55 (C-7a), 74.51 (CH₂Ph), 70.5 (C-3), 67.9 (C-8), 66.5 (C-5), 23.62 and 23.60 (2 CH₃CO), 22.0 (Me) ppm. HRMS (NALDI-TOF): calcd. for C₂₉H₃₆NO₈ [M + H]⁺ 526.2441; found 526.2417 (deviation -4.6 ppm).

(1R,2R,3R,5S,6S,7S,7aS)-1,2,6,7-Tetra-O-acetyl-3-acetyloxymethyl-1,2,6,7-tetrahydroxy-5-methylpyrrolizidine (20): Compound 1 (40 mg, 0.812 mmol) was acetylated in dry pyridine (3 mL), acetic anhydride (258 µL, 2.74 mmol), and DMAP (cat.) at room temperature for 1.5 h. TLC (AcOEt/hexane, 2:1) then showed a fasterrunning compound. The solvent was evaporated, and the residue was supported on silica gel and chromatographed (AcOEt/hexane, 1:2) to afford pure 20 (61 mg, 78%) as a colorless viscous syrup. $[a]_{D}^{29} = +9.3$ (c = 1, CHCl₃). IR (neat): $\tilde{v} = 1743$ (C=O) cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 5.36 (t, $J_{1,2} = J_{1,7a} = 4.7$ Hz, 1 H, 1-H), 5.29 (t, $J_{7,7a} = J_{6,7} = 5.0$ Hz, 1 H, 7-H), 5.18 (t, $J_{2,3} = 4.6$ Hz, 1 H, 2-H), 5.04 (dd, $J_{5,6}$ = 6.5 Hz, 1 H, 6-H), 4.04 (d, $J_{3,8}$ = 6.6 Hz, 2 H, 8,8-H), 3.45 (t, 1 H, 7a-H), 3.29 (dt, 1 H, 3-H), 3.16 (quint., $J_{\text{Me},5} = 6.5 \text{ Hz}, 1 \text{ H}, 5\text{-H}$), 2.09, 2.08, 2.07, 2.07 and 2.06 (5 s, 15 H, 5Ac), 1.16 (d, 3 H, Me) ppm. ¹³C NMR (150 MHz, CDCl₃): δ = 170.8, 170.5, 170.2, 170.2 and 169.9 (C=O), 81.5 (C-6), 79.5 (C-1), 79.25 (C-2), 78.6 (C-7), 71.9 (C-7a), 67.5 (C-3), 64.8 (C-8), 64.5 (C-5), 21.08, 21.06 and 20.97 (5 CH₃CO), 19.4 (Me) ppm. HRMS (NALDI-TOF): calcd. for $C_{19}H_{28}NO_{10}$ [M + H]⁺ 430.1713; found 430.1707 (deviation -1.4 ppm).

Supporting Information (see footnote on the first page of this article): Copies of the ¹H and ¹³C NMR spectra of compounds 9, 10, 11, 14, and 15 and copies of the ¹H, ¹³C, COSY, HSQC, and NOE of compounds 16, 17, 18, 19, 20, 1, and 12.

FULL PAPER

Acknowledgments

Major support for this project was provided by the Ministerio de Educación y Ciencia of Spain (Project Ref. No. CTQ2006-14043). Additional support was provided by Junta de Andalucía (Group BIO-250). We also acknowledge Fundación Ramón Areces for a grant (F.S.-C.).

- A. Kato, N. Kato, I. Adachi, J. Hollinshead, G. W. J. Fleet, C. Kuriyama, K. Ikeda, N. Asano, R. J. Nash, *J. Nat. Prod.* 2007, 70, 993–997.
- [2] For selected reviews, see: a) N. Asano, R. J. Nash, R. J. Molyneux, G. W. J. Fleet, Tetrahedron: Asymmetry 2000, 11, 1645-1680; b) M. S. M. Pearson, M. Mathe-Allainmat, V. Fargeas, J. Lebreton, Eur. J. Org. Chem. 2005, 2159-2191; c) P. V. Murphy, Eur. J. Org. Chem. 2007, 4177-4187; d) N. Asano, Cell Mol. Life Sci. 2009, 66, 1479-1492; e) R. Dwek, T. Butters, F. Platt, N. Zitzmann, Nat. Rev. Drug Discovery 2002, 1, 65–75; f) A. A. Watson, G. W. J. Fleet, N. Asano, R. J. Molyneux, R. J. Nash, Phytochemistry 2001, 56, 265-295; g) N. Asano, A. Kato, A. A. Watson, Mini-Rev. Med. Chem. 2001, 1, 145-154; h) T. D. Butlers, R. A. Dwek, F. M. Platt, Glycobiology 2005, 15, 43R-52R; i) P. Merino, I. Delso, E. Marca, T. Tejero, R. Matute, Curr. Chem. Biol. 2009, 3, 253-271; j) T. Ayad, Y. Genisson, M. Baltas, Curr. Org. Chem. 2004, 8, 1211-1233; k) F. Cardona, A. Goti, A. Brandi, Eur. J. Org. Chem. 2007, 1551-1565; 1) P. Compain, O. R. Martin (Eds.), Iminosugars: From Synthesis to Therapeutic Applications, Wiley-VCH, Weinheim 2007; m) B. L. Stocker, E. M. Dangerfield, A. L. Win-Mason, G. W. Haslett, M. S. M. Timmer, Eur. J. Org. Chem. 2010, 1615-1637; n) B. G. Davis, Tetrahedron: Asymmetry 2009, 20, 652-671; o) D. D'Alonzo, A. Guaragna, G. Palumbo, Curr. Med. Chem. 2009, 16, 473-505; p) T. M. Cox, F. M. Platt, J. M. F. G. Aerts, Iminosugars 2007, 295-326.
- [3] a) W. Zhang, K. Sato, A. Kato, Y.-M. Jia, X.-G. Hu, F.X. Wilson, R. van Well, G. Horne, G. W. J. Fleet, R. J. Nash, C.-Y. Yu, Org. Lett. 2011, 13, 4414-4417; b) J. A. Tamayo, F. Franco, F. Sánchez-Cantalejo, Tetrahedron 2010, 66, 7262– 7267; c) T. Sengoku, Y. Satoh, M. Takahashi, H. Yoda, Tetra-

hedron Lett. 2009, 50, 4937–4940; d) C. Yu, H. Gao, Faming Zhuanli Shenqing Gongkai Shuomingshu 2008; e) N. Asano, H. Kuroi, K. Ikeda, H. Kizu, Y. Kameda, A. Kato, I. Adachi, A. A. Watson, R. J. Nash, G. W. J. Fleet, *Tetrahedron: Asymmetry* 2000, 11, 1–8; f) A. Kato, I. Adachi, M. Miyauchi, K. Ikeda, T. Komae, H. Kizu, Y. Kameda, A. A. Watson, R. J. Nash, M. R. Wormald, G. W. J. Fleet, N. Asano, *Carbohydr. Res.* 1999, 316, 95–103.

- [4] I. Izquierdo, M. T. Plaza, M. Rodríguez, F. Franco, A. Martos, *Tetrahedron* 2005, 61, 11697–11704.
- [5] For selected articles, see: a) I. Izquierdo, M. T. Plaza, R. Robles, F. Franco, *Tetrahedron: Asymmetry* 2001, *12*, 2481–2487; b) I. Izquierdo, M. T. Plaza, F. Franco, *Tetrahedron: Asymmetry* 2002, *13*, 1581–1585; c) I. Izquierdo, M. T. Plaza, F. Franco, *Tetrahedron: Asymmetry* 2003, *14*, 3933–3935; d) I. Izquierdo, M. T. Plaza, F. Franco, *Tetrahedron: Asymmetry* 2004, *15*, 1465–1469; e) I. Izquierdo, M. T. Plaza, J. A. Tamayo, *Tetrahedron* 2005, *61*, 6527–6533; f) I. Izquierdo, M. T. Plaza, V. Yáñez, *Tetrahedron: Asymmetry* 2005, *16*, 3887–3891; g) I. Izquierdo, M. T. Plaza, J. A. Tamayo, *J. Carbohydr. Chem.* 2006, *25*, 281–295; h) I. Izquierdo, M. T. Plaza, J. A. Tamayo, M. Rodríguez, A. Martos, *Tetrahedron* 2006, *62*, 6006–6011; i) I. Izquierdo, M. T. Plaza, J. A. Tamayo, F. Franco, F. Sánchez-Cantalejo, *Tetrahedron* 2010, *66*, 3788–3794.
- [6] I. Izquierdo, M. T. Plaza, J. A. Tamayo, D. Lo Re, F. Sánchez-Cantalejo, Synthesis 2008, 1373–1378.
- [7] The *dr* of compounds **10** and **11** was determined by NMR spectroscopic analysis of the crude reaction.
- [8] a) T. J. Donohoe, R. E. Thomas, M. D. Cheeseman, C. L. Rigby, G. Bhalay, D. Linney, Org. Lett. 2008, 10, 3615–3618;
 b) T. J. Donohoe, H. O. Sintim, J. Hollinshead, J. Org. Chem. 2005, 70, 7297–7304; c) M. R. Wormald, R. J. Nash, P. Hrnciar, J. D. White, R. J. Molyneux, G. W. J. Fleet, Tetrahedron: Asymmetry 1998, 9, 2549–2558; d) T. Sengoku, Y. Satoh, M. Takahashi, H. Yoda, Tetrahedron Lett. 2009, 50, 4937–4940; e) C. W. G. Au, R. J. Nash, S. G. Pyne, Chem. Commun. 2010, 46, 713–715.
- [9] See the Supporting Information.

Received: July 31, 2011 Published Online: October 17, 2011