

Bioorganic & Medicinal Chemistry Letters 11 (2001) 2615–2617

Potent Peptide α-Ketohydroxamate Inhibitors of Recombinant Human Calpain I

Kurt A. Josef,* Frederick W. Kauer and Ron Bihovsky

Cephalon, Inc., 145 Brandywine Parkway, West Chester, PA 19380-4245, USA

Received 16 April 2001; accepted 24 July 2001

Abstract—A series of potent dipeptide and tripeptide α -ketohydroxamic esters was prepared as inhibitors of recombinant human calpain I. Compound **3c**, a Cbz-Leu-Phe hydroxamate, displayed the greatest potency against calpain I (IC₅₀=6 nM), while two compounds, **3l** and **3m**, both possessing the Cbz-Leu-Phe sequence, were the most potent (IC₅₀=0.2 μ M) in a MOLT-4 cell assay. © 2001 Elsevier Science Ltd. All rights reserved.

Calpains, calcium-activated cysteine proteases widely distributed in mammalian cells, have been identified in two major forms: calpain I and calpain II. It is believed that calpain I is activated during a biochemical cascade that leads to a delayed degeneration of neurons following ischemia.¹ Calpain is implicated in many nervous system disorders such as stroke, severe head trauma, and other neurological diseases. This understanding has led to the search for potent, membrane permeable inhibitors as pharmacological targets. A variety of inhibitor structures, both reversible (aldehyde, hydroxyoxazolidine and α -ketocarbonyl)² and irreversible (diazomethyl ketone, fluoromethyl ketone, epoxysuccinate, and acyloxymethyl ketone),³ have been reported. The most potent reversible inhibitors show IC₅₀ values less than 10 nM against calpain I.² We report on a novel series⁴ of peptide α -ketohydroxamates,⁵ compounds **3a–n**, and their inhibitory activities against recombinant human calpain I and in a whole-cell MOLT-4 assay.

The title compounds were synthesized following one of two routes. In General Method A (Scheme 1), the P₁ *N*-BOC- β -amino- α -hydroxy acid 1⁶ was esterified and *N*deprotected in a single step with thionyl chloride in methanol. This was then coupled to the desired *N*-protected P₂ amino acid (or P₃–P₂ dipeptide) using standard BOP/HOBt procedures. Hydrolysis of the resulting ester with LiOH gave the α -hydroxy acid 2 with the appropriate P₂ (or P₃–P₂) moiety of the peptide in place. Formation of the hydroxamate was achieved with BOP/HOBt and the appropriate hydroxylamine. Subsequent oxidation with the Dess–Martin periodinane⁷ afforded α -ketohydroxamates, compounds 3a–f,⁸ 3h, 3i, 3l, and 3m.

For General Method B (Scheme 2), the P₁ *N*-BOC- β amino- α -hydroxy acid 4 was first coupled to the hydroxylamine with BOP/HOBt and then *N*-deprotected with HCl to give the corresponding β -amino- α -hydroxyhydroxamate 5. Standard peptide coupling reagents

General Method A



Scheme 1. (a) SOCl₂, MeOH; (b) Cbz-Leu-OH, BOP, HOBt, DMF; (c) LiOH, MeOH; (d) H₂NOBn, BOP, HOBt, DMF; (e) Dess-Martin.

0960-894X/01/\$ - see front matter \odot 2001 Elsevier Science Ltd. All rights reserved. P11: S0960-894X(01)00526-1

^{*}Corresponding author. Fax: +1-610-738-6643; e-mail: kjosef@cephalon.com

General Method B



Scheme 2. (a) H₂NOBn, BOP, HOBt, DMF; (b) HCl, dioxane; (c) CH₃SO₂-D-Ser(Bn)-OH, EDCI, HOBt, NMM, DMF; (d) Dess-Martin.

(EDCI/HOBt) were used to add the desired *N*-protected amino acid to the P_2 portion. Oxidation under Dess-Martin conditions yielded the α -ketohydroxamates, compounds **3g**, **3j**, **3k**, and **3n**.

Table 1 summarizes the potency of the peptide α -ketohydroxamates **3** evaluated in this study. The activity as inhibitors of purified human recombinant calpain I at 20 °C was determined utilizing Suc-Leu-Tyr-MNA (0.2 mM; $K_m = 0.4$ mM) as substrate, as previously described.⁹ The cell permeability was evaluated by measuring the spectrin breakdown product in intact MOLT-4 cells (human leukemia T-cell line) in which calpain was activated by ionomycin.¹⁰

Hydroxamates with the Cbz-Leu-Phe peptide sequence, compounds **3a–f**, are generally better inhibitors of the calpain I enzyme than the other hydroxamates investigated. The activity does not vary greatly as the relative size of the *O*-substituent of the hydroxamic ester is increased from methyl to pentafluorobenzyl. In contrast, in a series of Cbz-Leu-Phe α -ketoamides described by Li et al.,² the phenethyl amide ($K_i = 0.052 \,\mu$ M) is approximately 300-fold more potent than the *n*-propyl analogue ($K_i = 15.0 \,\mu$ M). Compound **3a**, the methyl hydroxamate, is approximately isosteric with the corresponding α -ketoamide Cbz-Leu-Phe-CONHEt. The activity of these two compounds are nearly identical in our calpain I enzyme assay ($IC_{50} = 10$ and 11 nM, respectively) and in the MOLT-4 cell assay ($IC_{50} = 0.9$ and $0.8 \mu M$, respectively).

The three dipeptides with the unnatural D-Ser-benzyl ether at P₂, **3g**–i, were prepared to further investigate this template previously reported from our laboratories.¹¹ The hydroxamates **3h** and **3i** are about 2–3 times more potent than their corresponding α -keto-amides, CH₃SO₂-D-Ser(Bn)-Phe-CONHBn and CH₃SO₂-D-Ser(Bn)-Phe-CONHEt (IC₅₀ = 28 and 56 nM vs. IC₅₀ = 56 and 180 nM, respectively). Compound **3g**, bearing a Ser-methyl ether at P₁, is five times less potent than **3h**, containing Phe P₁, which is consistent with P₁ site affinity criteria for the calpain enzyme.^{1,2}

Compounds **3j** and **3k**, both with Val at P_2 , are potent in both the calpain I enzyme assay and the MOLT-4 cell assay. The Nle at P_1 for **3k** results in only a minor loss of activity compared to **3j** containing P_1 Phe. Compound **3n**, with P_2 - P_1 Phe-Nle, is 10-fold less active than **3k**, which is consistent with previous observations that peptides with P_2 Phe are not preferred for calpain binding.^{1,2} The N-capping group in **3n** is PhCO rather than Cbz, but this should not significantly affect the potency.^{1,2}

Table 1.	Calpain inhibitory activity	
Table 1.	Calpain inhibitory activity	

B1	\mathbb{A}^2	H I N	0 1	H N	B^4
'' `N 	(*) C	/"\ ;) F		\int_{0}^{∞}	0/11

Compd	\mathbb{R}^1	\mathbb{R}^2	R ³	\mathbb{R}^4	Calpain I IC50 (nM)	MOLT-4 IC50 (µM)
3a ^a	BnOCO	L-CH ₂ CH(CH ₃) ₂	Bn	CH ₃	10	0.9
3b	BnOCO	$L-CH_2CH(CH_3)_2$	Bn	CH ₂ CH ₃	19	1.1
3c	BnOCO	$L-CH_2CH(CH_3)_2$	Bn	Ēn	6	0.6
3d	BnOCO	$L-CH_2CH(CH_3)_2$	Bn	$CH_2C_6F_5$	17	2.9
3e	BnOCO	$L-CH_2CH(CH_3)_2$	Bn	t-Bu	26	3.9
3f	BnOCO	$L-CH_2CH(CH_3)_2$	Bn	(4-Methyl-cyclohexyl)	21	26% ^b
3g	CH_3SO_2	D-CH ₂ OBn	CH ₂ OCH ₃	Bn	152	ND ^c
3h	CH_3SO_2	D-CH ₂ OBn	Bn	Bn	28	0.9
3i	CH_3SO_2	D-CH ₂ OBn	Bn	CH_2CH_3	56	ND
3j	BnOCO	$D-CH(CH_3)_2$	Bn	Bn	12	0.6
3k	BnOCO	$L-CH(CH_3)_2$	$(CH_2)_3CH_3$	Bn	21	0.5
31	Cbz-Leu	$L-CH_2CH(CH_3)_2$	Bn	CH ₃	20	0.2
3m	Cbz-Leu	$L-CH_2CH(CH_3)_2$	Bn	Bn	17	0.2
3n	PhCO	L-Bn	$(CH_2)_3CH_3$	CH ₂ CH ₃	193	ND

^aFor comparison: Cbz-Leu-Phe-CONHEt calpain I IC₅₀ = 11 nM and MOLT-4 IC₅₀ = 0.8 M. ^b% Inhibition at 1 μ M.

^cNot determined.

The tripeptides in the series, **31** and **3m**, are also potent inhibitors of the calpain I enzyme, but more importantly, are the most potent inhibitors identified in our MOLT-4 cell assay to date. Therefore, these compounds have greater cell permeability than the dipeptide α -keto-hydroxamates in our assay.¹² This result is consistent with α -ketoamide and α -ketoester tripeptides described by Li et al.² in a platelet membrane permeability assay.

We have described a series of novel peptide α -ketohydroxamates that are potent inhibitors of recombinant human calpain I. The activity of this series compares favorably to that of previously described α -ketoamides.² However, substituent changes on the P' hydroxamates do not affect the potency as significantly as changes to the P' amide substituents. These α -ketohydroxamates are highly membrane permeable as demonstrated in a whole-cell MOLT-4 assay.

Acknowledgements

The authors would like to thank John Mallamo, Sankar Chatterjee, and Derek Dunn for their support, and helpful discussions. We would also like to thank Mark Ator, Donna Bozyczko-Coyne, Satish Mallya, Beth Ann McKenna, Teresa M. O'Kane, and Shobha E. Senadhi for performing the biological assays. We would also like to thank SmithKline Beecham for partial support in this project.

References and Notes

(a) Wells, G. J.; Bihovsky, R. Exp. Opin. Ther. Patents
1998, 8, 1707. (b) Chatterjee, S. Drugs Future 1998, 23, 1217.
(c) Donkor, I. O. Curr. Med. Chem. 2000, 7, 1171.

 (a) Medhi, S. Trends Biol. Sci. 1991, 16, 50. (b) Harbeson, S. L.; Abelleira, S. M.; Akiyama, A.; Barrett, R., III; Carroll, R. M.; Straub, J. A.; Tkacz, J. N.; Wu, C.; Musso, G. F. J. Med. Chem. 1994, 37, 2918. (c) Li, Z.; Patil, G.; Golubski, Z. E.; Hori, H.; Tehrani, K.; Foreman, J. E.; Eveleth, D. D.; Bartus, R. T.; Powers, J. C. J. Med. Chem. 1993, 36, 3742. (d) Peet, N. P.; Kim, H. O.; Marquart, A. L.; Angelastro, M. R.; Nieduzak, T. R.; White, J. N.; Friedrich, D.; Flynn, G. A.; Webster, M. E.; Vaz, R. J.; Linnik, M. D.; Koehl, J. R.; Mehdi, S.; Bey, P.; Emary, W. B.; Hwang, K.-K. Bioorg. Med. Chem. Lett. 1999, 9, 2365. (e) Iqbal, M.; Messina, P. A.; Freed, B.; Das, M.; Chatterjee, S.; Tripathy, R.; Tao, M.; Josef, K. A.; Dembofsky, B.; Dunn, D.; Griffith, E.; Siman, R.; Senadhi, S. E.; Biazzo, W.; Bozyczko-Coyne, D.; Meyer, S. I.; Ator, M. A.; Bihovsky, R. *Bioorg. Med. Chem. Lett.* **1997**, 7, 539. (f) Conroy, J. L.; Seto, C. T. *J. Org. Chem.* **1998**, *63*, 2367.

3. (a) Crawford, C.; Mason, R. W.; Wickstrom, P.; Shaw, E. Biochem. J. 1988, 253, 751. (b) McGowan, E. B.; Becker, E.; Detwiler, T. C. Biochem. Biophys. Res. Commun. 1989, 158, 432. (c) Huang, Z.; McGowan, E. B.; Detwiler, T. C. J. Med. Chem. 1992, 35, 2048. (d) Harris, A. L.; Gregory, J. S.; Maycock, A. L.; Graybill, T. L.; Osifo, I. K.; Schmidt, S. L.; Dolle, R. E. Bioorg. Med. Chem. Lett. 1995, 5, 393. (e) Chatterjee, S.; Josef, K.; Wells, G.; Iqbal, M.; Bihovsky, R.; Mallamo, J. P.; Ator, M. A.; Bozyczko-Coyne, D.; Mallya, S.; Senadhi, S.; Siman, R. Bioorg. Med. Chem. Lett. 1996, 6, 1237.

4. After this manuscript was accepted for publication, a paper with similar subject matter was published. Donkor, I. O.; Zheng, X.; Han, J.; Lacy, C.; Miller, D. D. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 1753.

5. Mallamo, J. P.; Bihovsky, R.; Josef, K. A. WO 00/16767, 2000. *Chem. Abstr.* **2001**, *132*, 237376.

6. As described by Harbeson et al. in ref 2b.

7. Dess, D. B.; Martin, J. C. J. Org. Chem. 1983, 48, 4156.

8. For reference, **3a**: ¹H NMR (CDCl₃) δ 9.55 (br s, 1H), 7.20 (m, 10H), 6.82 (d, 1H), 5.40 (m, 1H), 5.03 (s, 2H), 4.95 (br s, 1H), 4.14 (m, 1H), 3.81 (s, 3H), 3.24 (dd, 1H), 2.96 (dd, 1H), 1.52 (m, 2H), 1.39 (m, 1H), 0. 83 (m, 6H). ¹H NMR spectra were recorded on a GE QE300 Plus spectrometer at 300 MHz using tetramethylsilane as internal standard.

9. Meyer, S. L.; Bozyczko-Coyne, D.; Mallya, S. K.; Spais, C. M.; Bihovsky, R.; Kawooya, J. K.; Lang, D. M.; Scott, R. W.; Siman, R. *Biochem. J.* **1996**, *314*, 511. In vitro calpain I enzymic activity was measured in a 96-well format using a continuous fluorimetric assay. In this assay, hydrolysis of succinyl-Leu-Tyr-4-methoxy-2-naphthylamine (Suc-Leu-Tyr-MNA; Enzyme Systems Products, Dublin, CA, USA; $K_m = 0.4 \text{ mM}$) was monitored by a Fluoroskan II fluorimeter (Labsystems, Helsinki, Finland). Enzyme activity was determined by measuring the calcium-dependant increase in fluorescence at 430 nm ($\lambda_{excitation} = 340 \text{ nm}$) of 0.2–1 mM substrate in 0.2 mL total volume of 50 mM Tris/HCl, pH 7.5, containing 50 mM NaCl, 1 mM EDTA, 1 mM EGTA, 5 mM β -mercaptoethanol, 2.5% DMSO, and 5 mM CaCl₂.

10. Chatterjee, S.; Ator, M. A.; Bozyczko-Coyne, D.; Josef, K.; Wells, G.; Tripathy, R.; Iqbal, M.; Bihovsky, R.; Senadhi, S. E.; Mallya, S.; O'Kane, T. M.; McKenna, B. A.; Siman, R.; Mallamo, J. P. J. Med. Chem. **1997**, 40, 3820.

11. Chatterjee, S.; Gu, Z. Q.; Dunn, D.; Tao, M.; Josef, K.; Tripathy, R.; Bihovsky, R.; Senadhi, S. E.; O'Kane, T. M.; McKenna, B. A.; Mallya, S.; Ator, M. A.; Bozyczko-Coyne, D.; Siman, R.; Mallamo, J. P. *J. Med. Chem.* **1998**, *41*, 2663. 12. One reviewer suggested there could be a correlation of the

MOLT-4 cell assay IC₅₀ values and some biophysical property such as lipophilicity. Calculation of the log P values gave a range of 4.5 to 7.3 ± 1 for compounds with IC₅₀ < 1 μ M. Although there appears to be no correlation with these data at this time, we gratefully thank the reviewer for bringing this to our attention.