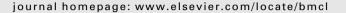
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Synthesis of new opioid derivatives with a propellane skeleton and their pharmacology. Part 2: Propellane derivatives with an amide side chain

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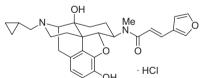
Keywords: Opioid Propellane derivative Shielding effect Lone electron pair

ABSTRACT

We designed and synthesized propellane derivatives with a 6- or 7-amide side chain on the basis of the active conformation of the κ selective agonist nalfurafine. The 6-amides showed high affinities for the κ receptor, and one of the 6 β -amides showed higher κ selectivity than nalfurafine. On the other hand, although the affinities of the 7-amides decreased compared to the 6-amides, some 7 α -amides showed the highest selectivities for the κ receptor among the tested compounds. The affinities of 7 β -isomers were extremely low, which was postulated to result from the shielding effect of the 7 β -amide side chain against the lone electron pair on the 17-nitrogen. This is the first conformational information about the 7-amide side chain in propellane derivatives.

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Three types of opioid receptors (μ , δ , κ) are now well established not only by pharmacological studies, but also by molecular biological studies.¹ Narcotic addiction is believed to be derived from the μ receptor type, and therefore the δ and κ types are promising drug targets for analgesics without addiction. To obtain ideal analgesics without addiction and other side effects derived from the µ receptor, we have synthesized various kinds of naltrexone derivatives and have reported selective ligands for the κ^{2-8} or δ^{9-13} receptors. Recently, one of our designed κ selective agonists, nalfurafine hydrochloride (TRK-820, Fig. 1),^{2,3,6,8} was launched in Japan as an antipruritic for patients undergoing dialysis.^{6,8} Although many arylacetamide derivatives such as U-50,488H^{14,15} and U-69,593 (Fig. 1)¹⁶ have been synthesized and developed as κ agonists, all of these derivatives were eliminated from clinical trials not only as analgesics but also as antipruritics because of their serious side effects like psychotomimetic and aversive reactions.^{17,18} On the other hand, nalfurafine has neither aversive nor addictive effects.¹⁹ We have been interested in the pharmacological differences between nalfurafine and arylacetamide derivatives with or without aversion side effects. We postulated that the differences in pharmacological effects between these two classes of compounds may derive from the differences in their affinities for κ receptor subtypes $(\kappa_1 \text{ and } \kappa_3)^{20-22}$ (arylacetamide derivatives: $\kappa_1^{21,22}$ and nalfurafine: κ_3^{23-27}). We were also interested in the pharmacological effects of the κ_2 subtype for which benzomorphans like (–)-pentazocine and bremazocine 20,22 are proposed to show high affinity.



nalfurafine hydrochloride (TRK-820)

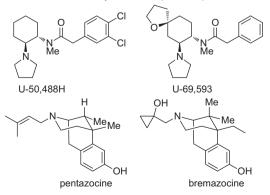


Figure 1. Structures of nalfurafine hydrochloride, U-50,488H, U-69,593, pentazocine, and bremazocine.

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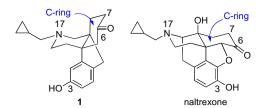


Figure 2. Structures of propellane 1²⁹ and naltrexone.

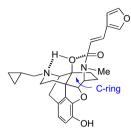
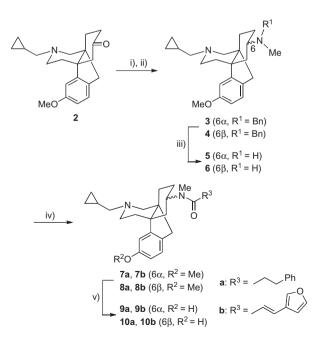


Figure 3. Proposed active conformation of nalfurafine binding to the κ receptor.

In the course of our synthetic and pharmacological studies with naltrexone derivatives, we discovered the synthesis of propellane derivatives.²⁸ In contrast to nalfurafine, the propellane derivative **1** (Fig. 2) lacks an amide side chain, thought to be a crucial structural determinant for binding to the κ receptor (κ address). Surprisingly, **1** showed κ agonist activity,³⁰ indicating that the propellane skeleton would have a potential to bind to the κ receptor. We also proposed that in its binding to the κ receptor, nalfurafine would acquire an active conformation in which the C-ring assumes the boat form to orient the 6-side chain toward the upper side of the C-ring (Fig. 3).^{4,5,31,32} The 6-keto group in the C-ring of propellane **1** could therefore exist in a relatively higher position than that observed for naltrexone (Fig. 2). If an amide group could



Scheme 1. Reagents and conditions: (i) BnMeNH, PhCO₂H, *p*-TsOH-H₂O, PhH reflux; (ii) NaBH₃CN, MeOH, 0 °C, **3**: 40%, **4**: 25%; (iii) H₂, Pd/C, MeOH, rt; (iv) R³COCl, Et₃N, CH₂Cl₂, 0 °C to rt, **7a**: 88% from **3**, **7b**: 76% from **3**, **8a**: 77% from **4**, **8b**: 80% from **4**; (v) BBr₃, CH₂Cl₂, -78 °C to rt, **9a**: 70%, **9b**: 35%, **10a**: 43%, **10b**: 56%.

be introduced in the 6- or 7-position of **1**, the amide group would be expected to orient at a position that was closer to the upper side as compared to that of nalfurafine. Therefore, we attempted to synthesize propellane derivatives with an amide side chain in the 6- or 7-position. Herein, we report the synthesis of more κ selective propellane derivatives with an amide side chain and their pharmacologies.

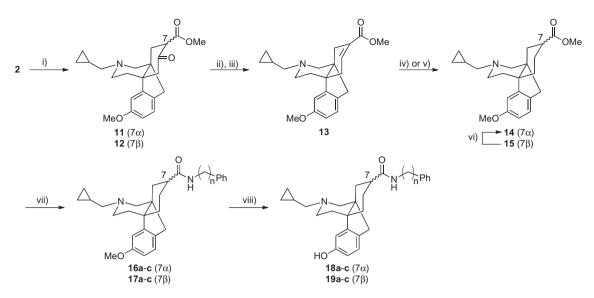
As the 17-cyclopropylmethyl (CPM) group was known to promote κ selectivity,³⁰ we synthesized only the derivatives with a 17-CPM substituent. The synthesis of 6-amide derivatives commenced with propellane **2** (Scheme 1). Propellane **2** was converted to methylamines **5** and **6** by reductive amination followed by debenzylation. Obtained methylamines **5** and **6** were treated with acyl chlorides to provide **7** and **8** and then the 0-methyl group in **7** and **8** was removed by boron tribromide treatment.

The 7-amides **18** and **19** were synthesized from propellane **2** (Scheme 2). Condensation of **2** with dimethyl carbonate gave ketoesters **11** and **12**. The keto group in **11** and **12** was removed by reduction, and then subsequent dehydration and hydrogenation gave 7-esters **14** and **15** as a diastereomixture. 7-Esters **14** and **15** were alternatively prepared from **13** by reduction using magnesium.³³ The epimerization of the major 7 β -isomer **15** gave the minor 7 α -isomer **14** in 24% yield. 7-Amides **18** and **19** were prepared by ester-amide exchange reaction of **14** and **15** followed by demethylation. α , β -Unsaturated amides **21** were provided from the intermediate **13** by the same method shown in Scheme 2 (Scheme 3).

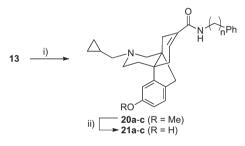
The binding affinities of the prepared propellane derivatives for the opioid receptors were evaluated with a competitive binding assay (Table 1). The assays were performed by a previously reported procedure.¹³

All the 6-amide derivatives 9 and 10 showed higher affinities for all three receptor types than did propellane 1 which lacked the amide side chain. The κ selectivity over the μ receptor of **10b** was improved as compared to those of nalfurafine and propellane **1**. Interestingly, 6β -isomers **10** were more κ selective over the μ receptor than the corresponding 6α -isomers **9**. This result with the 6^B-isomers may be caused by the orientation of the side chain toward the upper side of the C-ring, as opposed to the orientation of the side chain in the 6α -isomers. Meanwhile, the affinities of 7amide derivatives 18, 19, and 21 were lower than those of the 6amide derivatives **9** and **10**. Especially, the affinities of 7β -isomers **19** were low and **19b** showed almost no interaction with the δ and κ receptors. On the other hand, 7α -isomers **18** and α , β -unsaturated amides 21 exhibited somewhat higher affinities for the three receptor types as compared to the β -isomers **19**. The α , β -unsaturated amides **21** showed μ selectivity, whereas the α -isomers **18** except for **18c** exhibited κ selectivity. The κ selectivities of **18a** and **18b** were higher than those of nalfurafine and 6β-amide **10b**. These outcomes may result from the different orientation of the amide side chain (Fig. 4). The amide side chain of **18** may be able to locate in a direction that enhances binding to the κ receptor, whereas that of 21 cannot. The extreme decrease in the affinities of the 7β-isomer **19** is difficult to explain by an unfavorable orientation of the amide side chain, that is, the side chain is incapable of functioning as the κ address. So, we postulated that the flexible 7 β side chain could locate over the 17-nitrogen to interfere with the ionic interaction between the 17-nitrogen and the opioid receptor. The ionic interaction of the 17-nitrogen, along with a π - π interaction with the phenol ring and formation of a hydrogen bond with the phenolic hydroxy group are three important processes that have been identified in opioid ligand-receptor binding.³⁵⁻³⁷

To confirm our hypothesis, we carried out conformational analyses of three 7-amides **18b**, **19b**, and **21b** using Conformational Analyzer with Molecular Dynamics And Sampling (CAMDAS) 2.1 program³⁸ (Fig. 5). The benzene ring of the 7β -amide side chain



Scheme 2. Reagents and conditions: (i) NaH, CO(OMe)₂, 50 °C; (ii) NaBH₄, MeOH, 0 °C; (iii) POCl₃, pyridine, 60 °C, 61% from 2; (iv) H₂, Pd/C, MeOH, rt, 14: 5%, 15: 77%; (v) Mg, MeOH, rt, 14: 30%, 15: 53%; (vi) LDA, THF, -78 °C, then BHT, 14: 24%, 15: 65% (recovery); (vii) Ph(CH₂)_nNH₂, n-BuLi, THF, -78 °C, 16a (n = 0): 90%, 16b (n = 1): 85%, 16c (n = 2): 99%, 17a (n = 0): 72%, 17b (n = 1): 83%, 17c (n = 2): 65%; (viii) BBr₃, CH₂Cl₂, -78 °C to rt, 18a: 80%, 18b: 88%, 18c: 75%, 19a: 64%, 19b: 83%, 19c: 92%. BHT: 2,6-di-tert-butyl-4methylphenol(butylated hydroxytoluene).



Scheme 3. Reagents and conditions: (i) Ph(CH₂)_nNH₂, n-BuLi, THF, -78 °C, 20a (n = 0): 87%, **20b** (n = 1): 80%, **20c** (n = 2): 85%; (ii) BBr₃, CH₂Cl₂, -78 °C to rt, **21a**: 32%, 21b: quant, 21c: 84%.

in 19b located over the 17-nitrogen to almost completely shield the lone electron pair of the 17-nitrogen (Fig. 5C). On the other hand, the side chain in 7α -amide **18b** would not shield the lone electron pair of 17-nitrogen (Fig. 5A). The shielding effect of the side chain in the α . β -unsaturated amide **21b** would be insufficient (Fig. 5B). However, the longer the length of the side chain, the lower the affinity (phenyl **21a** > benzyl **21b** > phenethyl **21c**). The results of conformational analyses supported our hypothesis and indicated that the ionic interaction between the 17-nitrogen and the opioid receptor would be especially important among the three known opioid ligand-receptor interactions.^{35–37} The identification of the stereospecific shielding effects against the lone electron pair of the 17-nitrogen would be the foremost significant outcome of the flexible propellane derivatives. Furthermore, the higher κ selectivity of 7α -amides **18a** and **18b** compared to that of nalfurafine supports the active conformation of nalfurafine for κ receptor binding (Fig. 3).

In conclusion, we have synthesized propellane derivatives with 6- or 7-amide side chains based on the concept of an active conformation of nalfurafine. The 6-amide derivatives 9 and 10 showed high affinities for the κ receptor, and 6β -isomer **10b** showed higher κ selectivity over the μ receptor than nalfurafine. On the other hand, although the affinities of 7-amides 18, 19, and 21 decreased compared to 6-amides 9 and 10, the 7\alpha-amides 18a and 18b

Table 1 Binding affinities of propellane derivatives 1, 9, 10, 18, 19, 21, and nalfurafine for opioid receptorsa,b

Compound	$K_{\rm i} ({\rm nM})$			Selectivity	
	μ ^c	$\delta^{\mathbf{d}}$	ĸ	μ/κ	δ/κ
Nalfurafine	0.431	51.3	0.178	2.42	288
1 ^f	58.2	448	17.4	3.34	25.7
9a	1.23	44.3	7.10	0.17	6.24
9b	2.20	35.0	0.83	2.65	42.2
10a	31.9	65.2	16.4	1.95	3.98
10b	12.2	121	3.43	3.56	35.3
18a	79.7	282	13.4	5.95	21.0
18b	106	>1000	19.2	5.52	g
18c	54.1	>1000	169	0.32	g
19a	80.8	207	108	0.75	1.92
19b	112	>1000	>1000	h	_h
19c	453	>1000	641	0.71	g
21a	12.4	192	46.7	0.27	4.11
21b	33.9	298	72.7	0.47	4.10
21c	35.9	412	253	0.14	1.63

Binding assays were carried out in duplicate (κ : cerebellum of guinea pig, μ and δ : whole brain without cerebellum of mouse).

^b See Ref. 34.

с ^{[3}H] DAMGO was used

d [³H] DPDPE was used.

e [³H] U-69,593 was used.

f Ref. 30.

^g Selectivity was not calculated because the K_i value for the δ receptor was over 1000 nM.

^h Selectivity was not calculated because the K_i value for the κ receptor was over 1000 nM.

showed higher selectivities for the κ receptor than nalfurafine and 6β -amide **10b**. Among the 7-amides, the affinities of 7β -isomers 19 were extremely low. We postulated that the low affinities of the 7^β-isomers **19** would result from the shielding of the lone electron pair of the 17-nitrogen conferred by the 7_β-amide side chain. The conformational analysis by CAMDAS also supported this postulation. This is the first conformational information about the 7-amide side chain in propellane derivatives. These finding afforded important information that may be applied to the design of new types of opioid ligands.

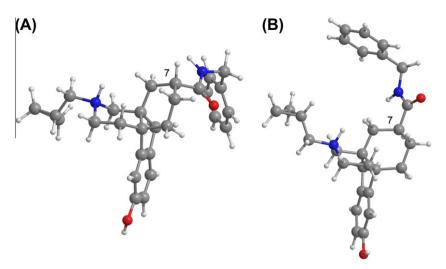


Figure 4. Comparison of 3D-alignments of (A) 7α-amide 18b and (B) 7β-amide 19b.

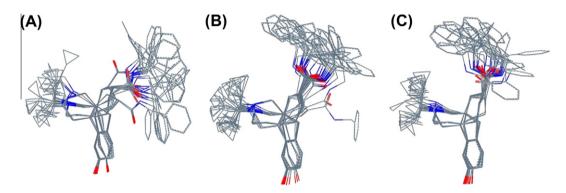


Figure 5. Results of conformational analyses of (A) 7α-amide 18b, (B) α,β-unsaturated amide 21b, and (C) 7β-amide 19b. Structures within 10 kcal/mol of the most stable conformer were collected.

Acknowledgments

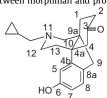
We acknowledge the financial supports from Shorai Foundation for Science and Uehara Memorial Foundation. We also acknowledge the Institute of Instrumental Analysis of Kitasato University, School of Pharmacy for its facilities.

References and notes

- 1. Dhawan, B. N.; Cesselin, F.; Raghubir, R.; Reisine, T.; Bradley, P. B.; Portoghese, P. S.; Hamon, M. *Pharmacol. Rev.* **1996**, *48*, 567.
- Nagase, H.; Hayakawa, J.; Kawamura, K.; Kawai, K.; Takezawa, Y.; Matsuura, H.; Tajima, C.; Endo, T. Chem. Pharm. Bull. 1998, 46, 366.
- Kawai, K.; Hayakawa, J.; Miyamoto, T.; Imamura, Y.; Yamane, S.; Wakita, H.; Fujii, H.; Kawamura, K.; Matsuura, H.; Izumimoto, N.; Kobayashi, R.; Endo, T.; Nagase, H. *Bioorg. Med. Chem.* **2008**, *16*, 9188.
- Nemoto, T.; Fujii, H.; Narita, M.; Miyoshi, K.; Nakamura, A.; Suzuki, T.; Nagase, H. Bioorg. Med. Chem. Lett. 2008, 18, 6398.
- Nagase, H.; Watanabe, A.; Nemoto, T.; Yamaotsu, N.; Hayashida, K.; Nakajima, M.; Hasebe, K.; Nakao, K.; Mochizuki, H.; Hirono, S.; Fujii, H. *Bioorg. Med. Chem. Lett.* 2010, 20, 121.
- 6. Nakao, K.; Mochizuki, H. Drugs Today 2009, 45, 323.
- Nemoto, T.; Yamamoto, N.; Watanabe, A.; Fujii, H.; Hasebe, K.; Nakajima, M.; Mochizuki, H.; Nagase, H. Bioorg. Med. Chem. 2011, 19, 1205.
- 8. Nagase, H.; Fujii, H. Top. Curr. Chem. 2011, 299, 29.
- 9. Nagase, H.; Kawai, K.; Hayakawa, J.; Wakita, H.; Mizusuna, A.; Matsuura, H.; Tajima, C.; Takezawa, Y.; Endo, T. *Chem. Pharm. Bull.* **1998**, 46, 1695.
- Nagase, H.; Yajima, Y.; Fujii, H.; Kawamura, K.; Narita, M.; Kamei, J.; Suzuki, T. Life Sci. 2001, 68, 2227.
- Nagase, H.; Osa, Y.; Nemoto, T.; Fujii, H.; Imai, M.; Nakamura, T.; Kanemasa, T.; Kato, A.; Gouda, H.; Hirono, S. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 2792.

- Nagase, H.; Nemoto, T.; Matsubara, A.; Saito, M.; Yamamoto, N.; Osa, Y.; Hirayama, S.; Nakajima, M.; Nakao, K.; Mochizuki, H.; Fujii, H. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 6302.
- 13. Ida, Y.; Nemoto, T.; Hirayama, S.; Fujii, H.; Osa, Y.; Imai, M.; Nakamura, T.; Kanemasa, T.; Kato, A.; Nagase, H. *Bioorg. Med. Chem.* **2012**, *20*, 949.
- 14. Lahti, R. A.; Von Voigtlander, P. F.; Barsuhn, C. Life Sci. 1982, 31, 2257.
- 15. Szmuszkovicz, J.; Von Voigtlander, P. F. J. Med. Chem. 1982, 25, 1125.
- Lahti, R. A.; Mickelson, M. M.; McCall, J. M.; Von Voigtlander, P. F. Eur. J. Pharmacol. 1985, 109, 281.
- 17. Mucha, R. F.; Herz, A. Psychopharmacology 1985, 86, 274.
- 18. Millan, M. J. Trends Pharmacol. Sci. 1990, 11, 70.
- Tsuji, M.; Takeda, H.; Matsumiya, T.; Nagase, H.; Narita, M.; Suzuki, T. Life Sci. 2001, 68, 1717.
- Zukin, R. S.; Eghbali, M.; Olive, D.; Unterwald, E. M.; Tempel, A. Proc. Natl. Acad. Sci. U.S.A. 1988, 85, 4061.
- Clark, J. A.; Liu, L.; Price, M.; Hersh, B.; Edelson, M.; Pasternak, G. W. J. Pharmacol. Exp. Ther. 1989, 251, 461.
- Nock, B. In *The Pharmacology of Opioid Peptides*; Tseng, L. F., Ed.; Harwood Academic: Singapore, 1995; pp 29–56.
- Endoh, T.; Matsuura, H.; Tajima, A.; Izumimoto, N.; Tajima, C.; Suzuki, T.; Saitoh, A.; Suzuki, T.; Narita, M.; Tseng, L. F.; Nagase, H. *Life Sci.* **1999**, 65, 1685.
- Endoh, T.; Tajima, A.; Suzuki, T.; Kamei, J.; Suzuki, T.; Narita, M.; Tseng, L. F.; Nagase, H. Eur. J. Pharmacol. 2000, 387, 133.
- Tsuji, M.; Yamazaki, M.; Takeda, H.; Matsumiya, T.; Nagase, H.; Tseng, L. F.; Narita, M.; Suzuki, T. Eur. J. Pharmacol. 2000, 394, 91.
- Tsuji, M.; Takeda, H.; Matsumiya, T.; Nagase, H.; Yamazaki, M.; Narita, M.; Suzuki, T. Life Sci. 2000, 66, PL353.
- Mori, T.; Nomura, M.; Yoshizawa, K.; Nagase, H.; Narita, M.; Suzuki, T. Life Sci. 2004, 75, 2473.
- Nagase, H.; Yamamoto, N.; Nemoto, T.; Yoza, K.; Kamiya, K.; Hirono, S.; Momen, S.; Izumimoto, N.; Hasebe, K.; Mochizuki, H.; Fujii, H. J. Org. Chem. 2008, 73, 8093.
- 29. The numbering of propellane derivatives like **1** according to the IUPAC nomenclature is shown below. However, in this letter we used a tentative

numbering to the propellane derivatives, which would make it easy to compare the positions between morphinan and propellane skeletons.



 Yamamoto, N.; Fujii, H.; Nemoto, T.; Nakajima, R.; Momen, S.; Izumimoto, N.; Hasebe, K.; Mochizuki, H.; Nagase, H. Bioorg. Med. Chem. Lett. 2011, 21, 4104.

- 31. Yamaotsu, N.; Fujii, H.; Nagase, H.; Hirono, S. Bioorg. Med. Chem. 2010, 18, 4446.
- 32. Yamaotsu, N.; Hirono, S. Top. Curr. Chem. 2011, 299, 277.
- 33. Hudlicky, T.; Sinai-Zingde, G.; Natchus, M. G. Tetrahedron Lett. 1987, 28, 5287.
- 34. The selectivity obtained by our binding assay method was likely to be lower. Some groups have reported that the κ selectivity of nalfurafine was higher than our result. See the following reference for details. Fujii, H.; Hirayama, S.; Nagase, H. In *Pharmacology*; Gallelli, L., Ed.; InTech, 2012; pp 81–98.
- 35. Casy, A. F.; Beckett, A. H. J. Pharm. Pharmacol. 1954, 6, 986.
- 36. Beckett, A. H. J. Pharm. Pharmacol. 1956, 8, 848.
- Fries, D. S. In Foye's Principles of Medicinal Chemistry; Lemke, T. L., Williams, D. A., Eds., sixth ed.; Lippincott Williams & Wilkins: Philadelphia, 2008; pp 652–678.
- 38. Tsujishita, H.; Hirono, S. J. Comput. Aided Mol. Des. 1997, 11, 305.