Tetrahedron 67 (2011) 8229-8234

Contents lists available at SciVerse ScienceDirect

Tetrahedron

journal homepage: www.elsevier.com/locate/tet

Stereochemical investigation of conjugate additions of carbon- and heteronucleophiles to ring-substituted nitrosocyclohexenes

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ARTICLE INFO

Article history: Received 8 July 2011 Received in revised form 19 August 2011 Accepted 22 August 2011 Available online 28 August 2011

Keywords: Stereoselectivity Michael-type additions Enolonium ion equivalents Organocuprate additions

ABSTRACT

Intermolecular Michael-type conjugate additions of some in situ-generated ring-substituted nitrosocyclohexenes with both carbon- and heteronucleophiles have been found to be highly stereoselective, leading predominantly (or exclusively) to products resulting from axial attack on a half-chair conformation of the nitrosoalkene substrate.

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1. Introduction and background

We have recently been involved in exploring synthetic methodology based on inter-¹ and intramolecular² conjugate additions of nucleophiles to highly reactive, transient nitrosoalkenes.³ In particular, we have been interested in using this type of transformation to evaluate the synthetic potential of nitrosoalkenes as enolonium ion equivalents.⁴ At the outset of these investigations it became evident to us that there are a number of fundamental stereochemical issues relevant to this type of conjugate addition that have never been addressed. For example, in a recent publication we described stereochemical studies on nucleophilic additions to acyclic nitrosoalkenes bearing a γ -stereogenic center, a process found to be highly diastereoselective, leading exclusively to the *anti* adducts.⁵ We now report some new stereochemical investigations of conjugate additions of various carbon- and heteronucleophiles to cyclic systems.

2. Results and discussion

Our current studies focussed on probing the stereochemical outcome of nucleophilic additions to substituted nitrosocyclohexenes **3** to form α -substituted oxime products **4** (Scheme 1). For comparison purposes in the exploratory stages of this investigation, the requisite nitrosoalkenes **3** were generated and reacted with the nucleophile via two different procedures: (1) treatment of the α -chlorooxime **1**, formed from a ring-substituted α -chlorocyclohexanone, with 2.3 equiv of the potassium salt of the nucleophile in THF at -78 °C (Method A) and (2) the Denmark procedure⁶ involving treatment of an α -chloro-*O*-silyloxime **2** with tetrabutylammonium fluoride in THF in the presence of 1.2 equiv of the potassium salt of the nucleophile (Method B).



Scheme 1. Addition of nucleophiles to substituted nitrosocyclohexenes.

Initial experiments involved examining the addition of various malonate ester enolates to in situ-produced 4-*tert*-butylnitrosocyclohexene. Thus, the known^{6a,7} racemic *cis-* α -chloro-4-*tert*butylcyclohexanone oxime **5a** and the corresponding *trans*-isomer **5b** were combined with the potassium enolate of diethyl malonate (**6a**) under the conditions of Method A (see Experimental section for details) to afford exclusively *trans*-adduct **8a** (R'=H) in good



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^{0040-4020/\$ —} see front matter \odot 2011 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2011.08.054

vields (Table 1, >20:1 8/9 estimated by ¹H NMR). In both experiments only one oxime geometric isomer of 8a was produced. Although we cannot definitively assign geometry in this case, the fact that we have generally observed formation of the (E)-oximes as the exclusive (or predominant) product in other examples (vide infra) has led us to tentatively propose this geometry for adduct 8a. The configuration and chair conformation of **8a**. as shown in Scheme 2. were established by NMR analysis.⁸ Exposure of *cis-a*-chloro-Osilvloxime $5c^{6a}$ to the Denmark conditions in the presence of diethyl malonate potassium enolate (Method B) also afforded a comparable yield of the same *trans*-adduct **8a** as the sole product. It seems reasonable that adduct 8a is formed in all three cases via preferred axial attack⁹ on the half-chair conformation of 4-tertbutylnitrosocyclohexene (7). These experiments also indicated that the stereoselectivity and yield of the addition products are independent of the precursor and method of generation of the nitrosoalkene.^{6a}



Scheme 2. Configurations and conformations of malonate adducts of 4-tertbutylnitrosocyclohexene.

Table 1

Malonate additions to 4-tert-butylnitrosocyclohexene

RO _N + R	'─ <mark>CO₂Et</mark> CO₂Et		eq (<i>cis</i>) ↓ ↓ ↓ ↓ H ax (<i>trans</i>)	HO_N_CO2Et	$HO_N CO_2Et$ $Et + A_R^{(1)}CO_2Et$
5	6	-	7 -	8	9
				Total Yield (8 + 9)	Isomer Ratio (8:9)
a R = H, α-Cl	a R' = H	Method A		85%	
b $R = H$, β -Cl		Method A		93%	only 8
c R = TBS, α -Cl		Method B		86%	
a R = H, α-Cl	b R' = Me	Method A		89%	
b R = H, β-Cl		Method A		85%	6.2:1
$\textbf{c}~\textbf{R}=\textbf{TBS},~\alpha\text{-CI}$		Method B		81%	
a R = H, α-Cl	c R' = Et	Method A		76%	
$\mathbf{b} \mathbf{R} = \mathbf{H}, \beta - \mathbf{CI}$		Method A		88%	only 8
c R = TBS, α-Cl		Method B		72%	
a R = H, α-Cl	d R' = allyl	Method A		89%	
$\textbf{c}~\textbf{R}=\textbf{TBS},~\alpha\text{-CI}$		Method B		72%	9.1:1
-					

Further studies were subsequently carried out with α -alkylsubstituted malonate nucleophiles. It was found that repeating the above experiments with oxime derivatives **5a**–**c** using α -methyl diethyl malonate (**6b**) in most runs afforded an inseparable 6.2:1 mixture of adducts (*E*)-*trans*-**8b** and (*E*)-*cis*-**9b** in good total yield. However, in one reaction conducted with oxime **5a**, only the *trans*product **8b** was formed, thereby providing a pure sample of this material. Interestingly, the major *trans*-product **8b** apparently exists in the twist boat conformation shown in Scheme 2 as determined by 2D NMR NOE analysis of the pure isomer. We have again assumed the oxime geometry to be (*E*) in this system since the stereochemistry could not be determined unambiguously by spectral methods. The fact that **8b** is a twist boat is not too surprising since Allinger has reported that *trans*-1,3-di-*tert*-butyl-cyclohexane exists in this conformation.¹⁰

Addition of the potassium enolate of α -ethyl diethyl malonate (**6c**) to the nitrosocycloalkene derived from the three oxime derivatives **5a**–**c** as was done with malonates **6a** and **6b** also afforded a single *trans*-adduct **8c** with the (*E*)-oxime configuration. As was the case with **8b**, NMR analysis indicated that this system exists in a twist boat conformation (Scheme 2).

Finally, addition of α -allyl diethyl malonate (**6d**) to α -chlorooximes **5a** via Method A and **5c** via Method B led to an inseparable 9.1:1 mixture of (*E*)-*trans*-adduct **8d** and (*E*)-*cis*-compound **9d**. It was possible in this case to crystallize the major (*E*)-*trans* isomer **8d** from the mixture, and X-ray analysis of this compound showed it indeed to exist in a twist boat conformation and confirmed the (*E*)oxime geometry.¹¹

It is also possible to add heteronucleophiles to vinylnitroso compound **7** with a high degree of stereoselectivity. Therefore, *N*-methyl-*p*-toluenesulfonamide undergoes smooth axial addition to the nitrosoalkene formed from α -chloro-*O*-TBS-oxime **5c** under the conditions of Method B to afford exclusively the *trans*-(*E*)-oxime product **10** (Scheme 3). The structure and stereochemistry of this adduct were firmly established by X-ray analysis.



Scheme 3. Addition of heteronucleophiles to 4-tert-butylnitrosocyclohexene.

Similarly, potassium thiophenoxide undergoes highly axialstereoselective addition to the nitrosoalkene derived from α chlorooxime **5a** via Method A, as well as using the Denmark procedure from O-silyloxime **5c** (Method B).¹² Both procedures led to a chromatographically separable ~ 19:1 mixture of the (*E*)-transproduct **11** and the (*E*)-*cis* isomer **12** in high yields. The structure of the minor product **12** was confirmed by X-ray analysis.

We have also investigated the stereochemistry of the addition of organocuprates to 4-tert-butylnitrosocyclohexene (7). For example, treatment of α -chlorooxime **5a** with 2 equiv of dimethylcopper lithium afforded a chromatographically separable 4:1 mixture of *trans* α -methyl product **13a** derived from axial attack on nitrosoalkene 7, and the corresponding cis isomer 14a in excellent total yield (Scheme 4). A small amount ($\sim 10\%$) of the reductive dechlorination product of starting oxime 5a was also formed in this reaction. In addition, the major isomer 13a was found to be an inseparable 1:1 mixture of (E/Z)-oxime geometric isomers,⁸ whereas the minor product **14a** was exclusively (E) as determined by X-ray analysis. Similar but improved stereoselectivity was observed in the conjugate addition reactions with aryl cuprates. Therefore, the phenyl cuprate addition gave a 12:1 mixture of trans-13b and cis-14b in high overall yield, whereas the p-tolyl cuprate cleanly afforded a 9:1 mixture of 13c and 14c. It should also be noted that in the aryl cuprate additions



Scheme 4. Addition of cuprates to 4-tert-butylnitrosocyclohexene.

diastereomer and (E/Z)-oxime ratios were observed to be quite variable from run to run, probably due to equilibration during workup and/or purification. However, we have been unable to determine if similar isomerizations occurred in the malonate reactions (cf. Table 1).

Finally, we have briefly investigated the stereochemistry of the addition of diethyl malonate to some other nitrosocyclohexenes. For example, treatment of α -chloro-O-TBS-oxime **15b**, prepared from the corresponding known α -chloroketone **15a** (mixture of isomers),¹³ with the potassium salt of diethyl malonate, followed by TBAF led to a single diastereomeric adduct **17** (yield unoptimized) (Scheme 5). This product, which is a 5:1 (*E*/*Z*) mixture of oxime isomers, has the cis-stereochemistry and conformation as shown based upon NMR analysis. The adduct **17** probably arises via axial attack by the malonate enolate on the half-chair vinylnitroso intermediate **16**.

generally favored by varying degrees using a wide variety of carbon- and heteronucleophiles. It should be noted that due to the high electrophilicity of nitrosoalkenes, as well as the methods of generation of these species, we are limited to use of nonnucleophilic solvents, such as THF.

The studies outlined here further demonstrate the potential for stereospecific alkylation and arylation of nitrosoalkenes, which can act as effective enolonium ion equivalents. The product α -substituted oximes can be converted back to the parent carbonyl compounds via a plethora of methods.¹⁶ Alternatively, these oximes could potentially be reduced to the corresponding amines or subjected to Beckmann rearrangements to afford ring-expanded lactams. We are continuing to investigate the potential of nitrosoalkenes in organic synthesis.

4. Experimental

4.1. General methods

All non-aqueous reactions were carried out under an inert argon atmosphere in flame dried glassware. Liquid reagents sensitive to air were added via a dry syringe. All solvents and reagents were obtained from commercial sources and used without further purification. EM Science silica gel 60 (230–400 mesh) was used for flash column chromatography, and silica gel 60 PF₂₅₄ plates were used for analytical and preparative thin layer chromatography. ¹H and ¹³C NMR experiments were performed on Bruker CDPX 300, DRX 400 or AV-III-600 MHz



Scheme 5. Addition of diethyl malonate to ring-substituted nitrosocyclohexenes.

Similarly, addition of malonate anion to the nitrosoalkene derived from known α -chloro-O-silyloxime **18**^{6c} led to the *trans*-product **20** as an 11:1 (*E*/*Z*) oxime mixture. It seems reasonable that this adduct is formed via axial attack on the preferred (due to A^{1,2}-strain)¹⁴ quasi-axial-ethyl half-chair nitrosoalkene **19**.

3. Conclusion

Analogous to the work described here, a number of studies have appeared probing the stereochemistry of Michael additions to cyclohexenes bearing electron-withdrawing groups, such as cyano, phenylazo, acetyl, carboxylate, etc.¹⁵ Interestingly, with many of these Michael acceptors the preference for axial or equatorial attack varies widely depending on the particular nucleophile, solvent, and reaction conditions, as well as steric factors. In the case of the nitrosocyclohexene systems described here, axial attack is spectrometers. All high and low resolution mass spectral data were obtained using a Waters LCT Premier time of flight mass spectrometer (Waters Corporation, Micromass Ltd., Manchester, UK). IR spectra were measured on a Perkin–Elmer 1600 Series FTIR.

4.2. Synthesis and characterization

4.2.1. General procedures for intermolecular Michael additions of nucleophiles to in situ-generated 4-tert-butylnitrosocyclohexene. Method A: Addition of malonate and thiophenoxide nucleophiles to 4-tert-butylnitrosocyclohexene generated from α -chlorooximes **5a** and **5b**. A flame dried 25 mL round-bottomed flask containing a stirring bar was purged with Ar. The flask was cooled to -78 °C and charged with dry THF (3 mL). At this temperature, KHMDS (0.5 M in PhMe, 0.9 mL, 0.45 mmol) was added, followed by the nucleophile (0.44 mmol). The mixture was stirred at -78 °C for 45 min. A solution of the α -chlorooxime **5a** or **5b** (39 mg, 0.19 mmol) in THF (1 mL) was added slowly and the mixture was stirred at -78 °C for 20 min. The reaction was quenched with saturated NH₄Cl solution and was extracted with Et₂O. The combined organics were dried over anhydrous MgSO₄. The residue obtained after solvent removal under reduced pressure was purified by flash column chromatography on silica gel (gradient elution using 10–20% ethyl acetate in hexanes).

Method B: Addition of malonate, sulfonamide, and thiophenoxide nucleophiles to 4-tert-butylnitrosocyclohexene generated from α chloro-O-TBS-oxime 5c. A flame dried 25 mL round-bottomed flask containing a stirring bar was purged with Ar. The flask was cooled to -78 °C and charged with dry THF (2 mL), followed by KHMDS (0.5 M in PhMe, 0.45 mL, 0.23 mmol). The nucleophile (0.22 mmol) was added and the mixture was stirred at $-78 \degree \text{C}$ for 45 min. A solution of the α -chloro-O-TBS-oxime **5c** (57 mg, 0.18 mmol) in THF (1 mL) was added slowly to the reaction mixture at -78 °C, followed by the dropwise addition of TBAF (1.0 M in THF, 0.56 mL, 0.56 mmol) at this temperature. The temperature of the reaction bath was raised to -60 °C and maintained there for 20 min. The reaction was guenched with saturated NH₄Cl solution at -60 °C, removed from the cold bath, and allowed to warm to rt. The reaction mixture was extracted with Et₂O and the combined organics were dried over anhydrous MgSO₄. The residue obtained after solvent removal under reduced pressure was purified by flash column chromatography on silica gel (gradient elution using 10-20% ethyl acetate in hexanes).

4.2.1.1. 2-(5-tert-Butyl-2-hydroximinocyclohexyl)-malonic acid diethyl ester (**8a**). Method A: cis-**5a** (yield: 85%), trans-**5b** (yield: 93%). Method B: cis-**5c** (yield: 86%). ¹H NMR (300 MHz, CDCl₃) δ 7.95 (br s, 1H), 4.28–4.13 (m, 4H), 3.77 (d, *J*=11.5 Hz, 1H), 3.26 (dt, *J*=4.7, 11.5 Hz, 1H), 3.15 (dq, *J*=2.1, 15.6 Hz, 1H), 2.07–1.75 (m, 3H), 1.53–1.18 (m, 9H), 0.87 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 168.4, 168.3, 160.2, 62.0, 53.7, 42.6, 40.3, 32.9, 30.0, 27.7, 25.5, 22.5, 14.6, 14.4; IR (thin film) 3460, 3307, 2954, 2872, 1731, 1655, 1249, 1179, 1149, 1102, 1032, 926 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₇H₃₀NO₅: 328.2124, found: 328.2127.

4.2.1.2. 2-(5-tert-Butyl-2-hydroxyiminocyclohexyl)-2methylmalonic acid diethyl ester (**8b**). Method A: cis-**5a** (yield: 89%), trans-**5b** (yield: 85%). Method: B: cis-**5c** (yield: 81%). Product was isolated as an inseparable 6.2:1 mixture of trans-**8b**/cis-**9b**. ¹H NMR (300 MHz, CDCl₃, cis isomer **9b**) δ 7.38 (br s, 1H), 4.25–4.07 (m, 4H), 3.41 (dm, J=15.6 Hz, 1H), 3.17 (dd, J=3.3, 12.4 Hz, 1H), 1.93–1.58 (m, 4H), 1.54 (s, 3H), 1.43–1.16 (m, 8H), 0.85 (s, 9H). ¹H NMR (300 MHz, CDCl₃, trans isomer **8b**) δ 7.60 (br s, 1H), 4.25–4.07 (m, 4H), 3.33 (t, J=8.2 Hz, 1H), 2.71 (dm, J=17.4 Hz, 1H), 2.33–2.20 (m, 1H), 1.83–1.57 (m, 3H), 1.52 (s, 3H), 1.43–1.16 (m, 8H), 0.85 (s, 9H); ¹³C NMR (100 MHz, CDCl₃, trans isomer **8b**) δ 172.0, 171.9, 160.2, 61.8, 61.7, 56.2, 43.8, 42.1, 33.5, 27.4, 26.4, 25.1, 22.0, 18.0, 14.4, 14.3; IR (thin film) 3448, 2954, 2872, 1725, 1655, 1296, 1243, 1179, 1108, 1020, 932 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₈H₃₂NO₅: 342.2280, found: 342.2281.

4.2.1.3. 2-(5-tert-Butyl-2-hydroxyiminocyclohexyl)-2ethylmalonic acid diethyl ester (**8**c). Method A: cis-**5a** (yield: 76%), trans-**5b** (yield: 88%). Method B, cis-**5c** (yield: 72%). ¹H NMR (300 MHz, CDCl₃) δ 7.44 (br s, 1H), 4.31–4.14 (m, 4H), 3.20 (t, J=6.6 Hz, 1H), 2.89 (br d, J=14.5 Hz, 1H), 2.24–2.06 (m, 3H), 1.97–1.85 (m, 1H), 1.79–1.76 (m, 1H), 1.69–1.61 (m, 1H), 1.32–1.22 (m, 8H), 0.96–0.87 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 172.1, 171.2, 160.3, 61.2, 59.7, 43.0, 42.3, 32.9, 28.3, 28.0, 27.3, 23.8, 14.2, 14.0, 9.5; IR (thin film) 3460, 2954, 2872, 1725, 1655, 1296, 1232, 1114, 1026, 955 cm $^{-1}$; HRMS-ES $[M+H]^+$ calcd for $C_{19}H_{34}NO_5$: 356.2437, found: 356.2444.

4.2.1.4. 2-Allyl-2-(5-tert-butyl-2-hydroxyiminocyclohexyl)-malonic acid diethyl ester (8d). Method A: cis-5a (vield: 89%). Method B: *cis*-**5c** (vield: 72%). Product was isolated as an inseparable 9.1:1 mixture of 8d/9d. X-ray quality crystals of the major trans isomer 8d were obtained from the mixture by slow evaporation from acetone/ ethyl acetate. ¹H NMR (400 MHz, CDCl₃, *cis* isomer **9d**) δ 7.44 (br s, 1H), 5.99–5.89 (m, 1H), 5.05 (br d, *J*=8.3 Hz, 1H), 5.03 (br s, 1H), 4.30-4.12 (m, 4H), 3.42 (br d, *I*=13.2 Hz, 1H), 2.97 (dd, *I*=5.8, 14.0 Hz, 1H), 2.91–2.65 (m, 3H), 2.09 (br d, *J*=12.4 Hz, 1H), 1.91 (br d, *I*=10.5 Hz, 1H), 1.65–1.54 (m, 1H), 1.30–1.18 (m, 8H), 0.88 (s, 9H). ¹H NMR (400 MHz, CDCl₃, trans isomer **8d**) δ 7.52 (br s, 1H), 5.99–5.89 (m, 1H), 5.05 (br d, *J*=8.3 Hz, 1H), 5.03 (br s, 1H), 4.30–4.12 (m, 4H), 3.16 (t, *I*=7.4 Hz, 1H), 2.91–2.65 (m, 3H), 2.29–2.17 (m, 2H), 1.75–1.57 (m, 2H), 1.30–1.18 (m, 8H), 0.87(s, 9H); ¹³C NMR (100 MHz, CDCl₃, *trans* isomer **8d**) δ 171.7, 171.0, 160.3, 134.5, 118.6, 61.6, 61.5, 59.8, 43.7, 42.6, 39.8, 33.3, 27.8, 27.5, 24.6, 23.1, 14.5, 14.4; IR (thin film, 8d/9d mixture) 3448, 2954, 2873, 1725, 1655, 1249, 1167, 1108, 1020, 930 cm⁻¹; HRMS-ES [M+H]⁺ (**8d/9d** mixture) calcd for C₂₀H₃₄NO₅: 368.2437, found: 368.2422.

4.2.1.5. N-(5-tert-Butyl-2-hydroxyiminocyclohexyl)-4,N-dimethylbenzenesulfonamide (**10**). Method B: cis-**5c** (yield: 83%). X-ray quality crystals were obtained by slow evaporation from acetone/ ethyl acetate. ¹H NMR (300 MHz, CDCl₃) δ 8.59 (s, 1H), 7.72 (d, *J*=8.1 Hz, 2H), 7.31 (d, *J*=8.1 Hz, 2H), 4.11 (t, *J*=6.0 Hz, 1H), 3.05 (dq, *J*=2.2, 15.5 Hz, 1H), 2.75 (s, 3H), 2.42 (s, 3H), 2.34–2.22 (m, 1H), 2.15–2.07 (m, 1H), 1.85 (d, *J*=10.7 Hz, 1H), 1.65–1.52 (m, 1H), 1.50–1.40 (m, 1H), 1.35–1.21 (m, 1H), 0.86 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 160.9, 143.8, 135.0, 130.0, 128.3, 57.7, 42.4, 33.2, 33.0, 31.1, 27.5, 24.1, 23.7, 22.0; IR (thin film) 3460, 3284, 2954, 2872, 1655, 1337, 961 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₈H₂₉N₂O₃S: 353.1899, found: 353.1890.

4.2.1.6. trans-4-tert-Butyl-2-phenylsulfanylcyclohexanone oxime (**11**). Method A: cis-**5a** (isolated yield: 87%). Method B: cis-**5c** (isolated yield: 89%). ¹H NMR (400 MHz, CDCl₃) δ 8.05 (br s, 1H), 7.43 (d, *J*=7.0 Hz, 2H), 7.31–7.23 (m, 3H), 4.05 (br s, 1H), 3.25 (br d, *J*=14.6 Hz, 1H), 2.32–2.19 (m, 2H), 2.01–1.97 (m, 1H), 1.79 (br t, *J*=12.4 Hz, 1H) 1.70 (dq, *J*=4.5, 12.7 Hz, 1H), 1.20 (qd, *J*=4.2, 13.0 Hz, 1H), 0.90 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 160.0, 134.8, 133.1, 129.3, 127.8, 50.4, 42.5, 33.9, 32.6, 27.8, 26.9, 20.8; IR (thin film) 3248, 3070, 2954, 2860, 1708, 926 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₆H₂₄NOS: 278.1579, found: 278.1571.

4.2.1.7. *cis*-4-*tert*-*Butyl*-2-*phenylsulfanylcyclohexanone* oxime (**12**). Method A: *cis*-**5a** (isolated yield: 5%). Method B: *cis*-**5c** (isolated yield: 5%). X-ray quality crystals were obtained by slow evaporation from dichloromethane/acetone, mp 142–144 °C. ¹H NMR (600 MHz, CDCl₃) δ 7.88 (br s, 1H), 7.39 (d, *J*=7.5 Hz, 2H), 7.27 (t, *J*=7.4 Hz, 2H), 7.27 (t, *J*=7.4 Hz, 2H), 7.27 (t, *J*=7.4 Hz, 1H), 3.77 (dd, *J*=4.4, 11.7 Hz, 1H), 3.47 (dt, *J*=3.9, 14.7 Hz, 1H), 1.58–1.19 (m, 3H), 0.82 (s, 9H); ¹³C NMR (150 MHz, CDCl₃) δ 159.0, 134.8, 131.6, 128.8, 126.8, 49.8, 47.6, 36.0, 32.6, 27.4, 26.1, 24.6; IR (thin film) 3260, 2943, 2872, 932 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₆H₂₄NOS: 278.1579, found: 278.1571.

4.2.2. Addition of organocuprates to 4-tert-butylnitrosocyclohexene generated from cis- α -chlorooxime **5a**. Cul powder (75 mg, 0.40 mmol) in a round-bottomed flask was dried by heating at 150 °C for 2 h under vacuum. The heating was discontinued and the flask was allowed to cool to rt under Ar. The flask was then transferred to an ice bath and charged with dry THF (1 mL). The mixture was maintained at 0 °C and a solution of the

organolithium reagent (0.80 mmol) was added slowly. A turbid green coloration was observed after addition of the first equivalent of the organolithium reagent, but the mixture turned transparent or clear tan after the addition of the second equivalent. The mixture was stirred at this temperature for 2 min and then the reaction flask was transferred to an acetone/drv-ice bath at -78 °C. A solution of the α -chlorooxime *cis*-**5a** (41 mg, 0.20 mmol) in THF (2 mL) was added slowly to this mixture at -78 °C whereby the solution turned bright yellow. The reaction mixture was stirred for 30 min at -78 °C and guenched with concentrated HCl (12.1 M, 1 mL, 12.1 mmol) at this temperature. The dry-ice bath was removed and ethylenediamine (3 mL, 45 mmol) was added to the cold mixture followed by the addition of ice-cold water (10 mL). The addition of ethylenediamine was exothermic and the flask was properly vented to prevent the buildup of pressure. The reaction mixture turned turbid blue on the initial addition of ethylenediamine and later changed to clear Prussian blue on the addition of water. Stirring was discontinued after 2 min and the organic layer was extracted with ether, dried over anhydrous MgSO₄ and the solvent was removed under vacuum. The product was purified by flash column chromatography on silica gel eluting with dichloromethane (100%) followed by ethyl acetate/hexanes (gradient, 10%–20% ethyl acetate in hexanes).

4.2.2.1. trans-4-tert-Butyl-2-methylcyclohexanone oxime (**13a**). Isolated yield: 71%. ¹H NMR (300 MHz, CDCl₃, inseparable ~1:1 mixture of oxime *Z*/*E* isomers) δ 8.75 (br s, 1H), 3.65–3.61 (m, 0.5H, *Z*), 3.15 (br d, *J*=8.7 Hz, 0.5H), 2.67–2.64 (m, 0.5H, *E*), 2.34–2.22 (m, 1H), 1.95–1.85 (m, 1.5H), 1.67–1.63 (m, 1H), 1.49–1.36 (m, 2H), 1.20–1.11 (m, 4H), 0.86 (s, 9H); ¹³C NMR (100 MHz, CDCl₃, inseparable ~1:1 mixture of oxime *Z*/*E* isomers) δ 164.7, 164.6, 41.9, 41.8, 34.8, 34.0, 33.2, 32.7, 32.6, 29.0, 28.1, 27.9, 27.8, 27.0, 26.3, 21.5, 19.0, 17.2; IR (thin film) 3237, 2954, 2872, 1660, 944 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₁H₂₂NO: 184.1701, found: 184.1709.

4.2.2.2. cis-4-tert-Butyl-2-methylcyclohexanone oxime (**14a**). Isolated yield: 18%. X-ray quality crystals were obtained by slow evaporation from chloroform/ethyl acetate, mp 143–145 °C. ¹H NMR (300 MHz, CDCl₃) δ 9.62 (br, s, 1H), 3.44 (dm, *J*=14.0 Hz, 1H), 2.25 (m, 1H), 1.92 (dm, *J*=12.3 Hz, 2H), 1.62 (td, *J*=5.1, 14.0 Hz, 1H), 1.30 (tt, *J*=2.9, 12.0 Hz, 1H), 1.25–1.10 (m, 4H), 1.01 (q, *J*=9.4 Hz, 1H), 0.87 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 163.5, 47.8, 37.9, 37.7, 32.8, 28.0, 27.3, 24.8, 17.0; IR (thin film) 3260, 2943, 2872, 1660, 932 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₁H₂₂NO: 184.1701, found: 184.1708.

4.2.2.3. trans-4-tert-Butyl-2-phenylcyclohexanone oxime (**13b**). Isolated yield: 85% as an inseparable 36:1 mixture of oxime *E/Z* diastereomers. ¹H NMR (300 MHz, CDCl₃, *Z* isomer) δ 8.92 (br s, 1H), 7.40–7.24 (m, 5H), 4.93 (br s, 1H), 2.58–2.53 (m, 1H), 2.45 (br d, *J*=14.0 Hz, 1H), 1.92–1.21 (m, 5H), 0.94 (s, 9H). ¹H NMR (300 MHz, CDCl₃, *E* isomer) δ 8.92 (br s, 1H), 7.40–7.24 (m, 5H), 3.88 (br s, 1H), 3.33 (dm, *J*=12.2 Hz, 1H), 2.60 (dm, *J*=13.7 Hz, 1H), 1.92–1.6 (m, 3H), 1.50 (tm, *J*=9.3 Hz, 1H), 1.36–1.21 (m, 1H), 0.94 (s, 9H); ¹³C NMR (75 MHz, CDCl₃, *E* isomer) δ 162.7, 141.0, 129.0, 127.7, 126.6, 44.3, 42.3, 32.9, 31.2, 27.8, 27.0, 22.5; IR (thin film) 3260, 2954, 2872, 1666, 932 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₆H₂₄NO: 246.1858, found: 246.1855.

4.2.2.4. cis-4-tert-Butyl-2-phenylcyclohexanone oxime (**14b**). Isolated yield: 7%. ¹H NMR (400 MHz, CDCl₃) δ 7.36–7.21 (m, 5H), 3.50 (dm, *J*=13.2 Hz, 1H), 3.40 (dd, *J*=4.1, 12.9 Hz, 1H), 2.12–1.98 (m, 2H), 1.79 (td, *J*=5.4, 13.9 Hz, 1H), 1.64 (q, *J*=12.4 Hz, 1H), 1.49–1.43 (m, 1H), 1.37–1.22 (m, 2H), 0.92 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 162.9, 141.3, 129.1, 128.7, 127.1, 49.8, 48.1, 36.1, 33.0, 28.0, 26.9, 24.9; IR (thin film) 3247, 2950, 2868, 1668, 928 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₆H₂₄NO: 246.1858, found: 246.1855.

4.2.2.5. trans-4-tert-Butyl-2-p-tolylcyclohexanone oxime (**13c**). Isolated yield: 85% as an inseparable 5.5:1 mixture of oxime *E/Z* isomers. ¹H NMR (400 MHz, CDCl₃, *Z* isomer) δ 8.77 (br s, 1H), 7.19–7.14 (m, 4H), 4.84 (br s, 1H), 2.51–2.47 (m, 1H), 2.41 (dm, *J*=14.2 Hz, 1H), 2.34 (s, 3H), 2.16 (td, *J*=4.8, 13.9 Hz, 1H), 1.87–1.20 (m, 4H), 0.91 (s, 9H). ¹H NMR (400 MHz, CDCl₃, *E* isomer) δ 8.77 (br s, 1H), 7.19–7.14 (m, 4H), 3.80 (br s, 1H), 3.29 (br d, *J*=14.4 Hz, 1H), 2.55 (dm, *J*=13.7 Hz, 1H), 2.34 (s, 3H), 1.87–1.73 (m, 2H), 1.66 (td, *J*=5.1, 12.2 Hz, 1H), 1.51–1.40 (m, 1H), 1.27 (td, *J*=4.4, 12.5 Hz, 1H), 0.91 (s, 9H); ¹³C NMR (100 MHz, CDCl₃, *E* isomer) δ 162.8, 137.9, 136.1, 129.8, 127.5, 43.9, 42.2, 32.9, 31.2, 27.8, 27.0, 22.4, 21.4; IR (thin film) 3237, 3107, 2942, 2872, 1655, 932 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₇H₂₆NO: 260.2014, found: 260.2011.

4.2.2.6. *cis*-4-*tert*-*Butyl*-2-*p*-*tolylcyclohexanone* oxime (**14c**). Isolated yield: 9%. ¹H NMR (400 MHz, THF- d_8) δ 10.82 (s, 1H), 7.01 (dd, *J*=7.9, 9.2 Hz, 4H), 3.49 (br d, *J*=14.4 Hz, 1H), 3.30 (dd, *J*=4.05, 12.8 Hz, 1H), 2.26 (s, 3H), 2.00–1.89 (m, 2H), 1.69–1.64 (m, 1H), 1.56 (t, *J*=12.0 Hz, 1H), 1.48–1.42 (m, 1H), 1.25–1.21 (m, 1H), 0.91 (s, 9H); ¹³C NMR (100 MHz, THF- d_8 , compound has extremely low solubility leading to reduced signal intensity and therefore the oxime carbon peak could not be observed) δ 140.8, 136.2, 130.3, 129.4, 50.6, 49.4, 37.7, 33.7, 31.2, 28.5, 28.1, 21.7; IR (thin film) 3232, 2954, 2922, 2859, 1664, 930 cm⁻¹; HRMS-ES [M+H]⁺ calcd for C₁₇H₂₆NO: 260.2014, found: 260.2011.

4.2.2.7. 2-Chloro-5-methylcvclohexanone O-(tert-butyldimethylsilvl) oxime (**15b**). To a stirred solution of the α -chloroketone **15a** (3.41 mmol) in CH₂Cl₂ (7 mL) and a spatula of 4 Å molecular sieves were added H₂NOTBS (529 mg, 3.41 mmol) and PPTS (spatula tip full). The reaction mixture was stirred at rt for 12 h and then filtered through a pad of Celite. The solution was concentrated in vacuo and the residue was purified by flash column chromatography on silica gel eluting with a mixture of ethyl acetate in hexanes (5-30%) to provide the product **15b** as a clear oil (632 mg) in 67% yield as a mixture of (E/Z)-isomers and diastereomers: ¹H NMR (360 MHz, CDCl₃) δ 5.64 (br s, 0.4H), 5.50 (br s, 0.1H), 4.72 (br s, 0.4H), 4.57 (br s, 0.1H), 3.31 (t, J=5.7 Hz, 1H), 2.40-2.30 (m, 1H), 2.23-2.18 (m, 1H), 2.10-2.05 (m, 1H), 1.94-1.76 (m, 3H), 1.69-1.45 (m, 5H), 1.11-1.04 (m, 6H), 0.96–0.94 (m, 18H), 0.22–0.19 (m, 12H); ¹³C NMR (90 MHz, CDCl₃) ô 162.7, 161.9, 160.7, 159.3, 66.3, 59.1, 54.5, 49.3, 47.4, 47.0, 38.7, 37.1, 35.4, 34.9, 33.6, 33.5, 32.4, 31.4, 31.3, 31.2, 29.7, 29.0, 28.4, 28.2, 27.6, 27.5, 26.6, 26.1, 25.8, 24.5, 23.5, 22.4, 22.1, 21.9, 21.2, 19.9, 18.9, 18.8, 18.2, 16.8; IR (thin film) 2955, 2858, 1744, 1461, 1252, 945 cm⁻¹; LRMS-ES⁺ m/z (relative intensity) 276 (MH⁺, 10); HRMS-ES [M+H]⁺ calcd for C₁₃H₂₇NOSCI: 276.1550, found: 276.1543.

4.2.3. General procedure for conjugate additions to ring-substituted nitrosoalkenes 16 and 19. To a stirred solution of diethyl malonate (6a, 3 mmol, 481 mg) in THF (6.5 mL) was added KHMDS (6 mL, 0.5 M in PhMe, 3 mmol) at -78 °C. The resulting solution was then stirred for 45 min at that temperature. The O-TBS oxime (15b or 18, 1 mmol) dissolved in THF (600 µL) was added slowly over 1 min, followed by dropwise addition of TBAF (2 mL, 1.0 M in THF, 2 mmol) over 3 min. The resulting solution was immediately transferred to an ice bath and stirred for an additional 2 h. The reaction mixture was diluted with concentrated aqueous NH₄Cl and EtOAc. The organic layer was separated and the aqueous layer was extracted with EtOAc. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo to give a residue, which was purified by flash column chromatography on silica gel eluting with a mixture of ethyl acetate in hexanes (25-40%).

4.2.3.1. Diethyl 2-(2-(hydroxyimino)-4-methylcyclohexyl)malonate (17). The product 17 was obtained as a clear oil (47 mg, 45% yield) as a 5:1 mixture of (*E*/*Z*) oxime isomers: ¹H NMR (360 MHz, CDCl₃) δ 8.00–7.50 (br s, 1H), 4.23–4.10 (m, 4H), 3.79 (d, *J*=10.0 Hz, 0.2H), 3.71 (d, *J*=10.9 Hz), 3.20–3.13 (m, 0.3H), 3.11–3.04 (m, 1H), 2.86–2.72 (m, 0.3H), 2.56 (dd, *J*=14.1, 4.7 Hz, 1H), 2.33 (dd, *J*=14.0, 7.8 Hz, 1H), 1.93–1.74 (m, 2H), 1.72–1.61 (m, 4H), 1.44–1.37 (m, 2H), 1.30–1.19 (m, 6H), 1.01 (d, *J*=7.5 Hz, 1H), 0.96 (d, *J*=6.8 Hz, 3H); ¹³C NMR (90 MHz, CDCl₃) δ 168.7, 168.6, 159.3, 158.8, 61.9, 60.8, 53.8, 53.5, 41.4, 33.9, 32.0, 31.4, 30.7, 29.5, 27.7, 23.1, 23.0, 21.5, 20.6, 14.6, 14.5, 14.4, 11.8; IR (thin film) 3279, 2931, 2871, 1732, 1455, 1292, 1178, 1033, 947 cm⁻¹; LRMS-ES⁺ *m*/*z* (relative intensity) 286 (MH⁺, 36); HRMS-ES [M+H]⁺ calcd for C₁₄H₂₅NO₅: 286.1654, found: 286.1649.

4.2.3.2. Diethyl 2-(2-ethyl-6-(hydroxyimino)cyclohexyl)malonate (**20**). The product **20** was obtained as a clear oil (18 mg, 34% yield) as a ~11:1 mixture of (*E*/*Z*) oxime isomers: ¹H NMR (300 MHz, CDCl₃) δ 8.31 (s, 1H), 8.10 (s, 0.1H), 4.27–4.05 (m, 4H), 3.83 (d, *J*=8.5 Hz, 1H), 3.72 (d, *J*=8.1 Hz, 0.09H), 3.13 (d, *J*=10.6 Hz, 1H), 3.01 (d, *J*=8.5 Hz, 1H), 2.01–1.92 (m, 1H), 1.81–1.74 (m, 1H), 1.73–1.61 (m, 3H), 1.60–1.49 (m, 2H), 1.48–1.39 (m, 1H), 1.36–1.21 (m, 8H), 0.92 (t, *J*=5.5 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 168.3, 159.0, 158.6, 62.0, 61.9, 54.0, 53.5, 45.4, 39.7, 25.4, 25.3, 21.8, 20.5, 14.4, 12.1, 12.0, 11.7; IR (thin film) 3278, 2960, 2874, 1732, 1456, 1262, 1034, 951 cm⁻¹; LRMS-ES⁺ *m*/*z* (relative intensity) 300 (MH⁺, 26); HRMS-ES [M+H]⁺ calcd for C₁₅H₂₆NO₅: 300.1811, found: 300.1799.

Acknowledgements

We are grateful to the National Institutes of Health (GM-087733) and the National Science Foundation (CHE-0806807) for financial support of this research. We also thank Dr. H. Yennawar (Penn State Small Molecule X-Ray Cystallographic Facility) for the X-ray crystal structure determinations.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tet.2011.08.054.

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