

Synthetic Studies Directed toward the Assembly of the C-Glycoside Fragment of the Telomerase Inhibitor D8646-2-6

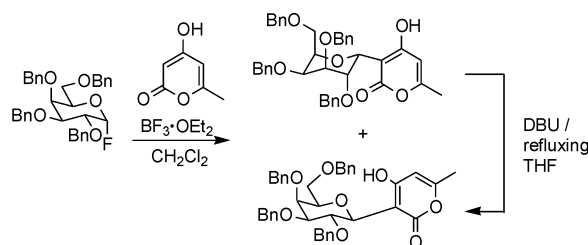
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ABSTRACT



Construction and characterization of the C-glycosidic moiety of telomerase inhibitor D8646-2-6 (**1**) are described. This is the first example of the C-glycosylation using electron-poor aromatics, 4-hydroxypyron-2-one, as a glycosyl acceptor. The glycosylation reaction and base-promoted isomerization affords desired beta-C-glycoside in a 61% overall yield.

D8646-2-6 (**1**) was isolated as an inhibitor of telomerase from the culture broth of *Epicoccum purpurascens* by the Mitsubishi Pharma Corporation group.¹ It has a unique C³-pyranosyl 4-hydroxypyron-2-one structure with a conjugated heptaene side chain. The relative and absolute stereochemistry of **1** have not been determined. Analysis of the reported ¹H NMR data of **1** strongly suggests a C³-beta-galactopyranosyl-4-hydroxypyron-2-one structure, although the chirality of the two centers in the side chain is not known. The galactopyranosyl-4-hydroxypyron-2-one structure is quite interesting when compared to C-glycosides containing phenol or naphthol derivatives as aryl units.² There are some examples of 4-hydroxypyrones connected to a 4-deoxyglucose as sugar moiety, and most of them have antifungal activity.³ The most challenging step in the synthesis of **1** involves the construction of the pyranosyl 4-hydroxypyron-2-one moiety. The alpha-pyrone moieties

of meroterpenoids (mixed polyketide-terpenoid metabolites) are usually constructed by the cyclization of the corresponding beta,d-diketo esters.⁴ A similar strategy involving the cyclization of alpha-C-pyranosyl beta,d-diketo ester, which could

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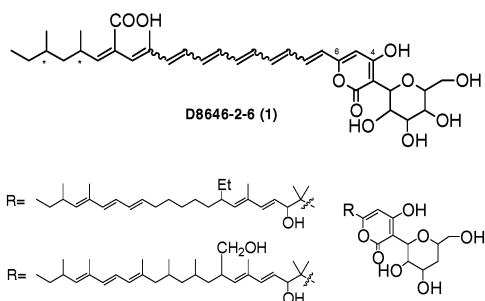
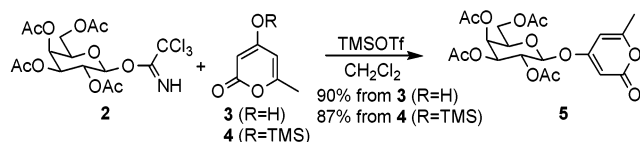


Figure 1. Structure of D8646-2-6 and some examples that contained deoxyglucose as a sugar moiety.

be readily prepared by C-glycosylation of β -ketoester or malonate,⁵ might seem like an obvious synthetic route. However, a direct coupling of 4-hydroxypyrene to the sugar moiety is desirable for establishing a convergent and efficient methodology. Despite extensive studies into the C-glycosylation of electron-rich aromatics,⁶ there have been no reports concerning C-glycosylation of 4-hydroxypyrene. During the course of our investigations into the synthesis of **1**, we studied the reaction of 4-hydroxypyrene and galactose derivatives. The results of these experiments are described in this paper.

We examined coupling of *O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)trichloroacetimidate **2**⁷ to 4-hydroxy-6-methylpyrone **3** or its TMS ether **4**⁸ (Scheme 1). In both

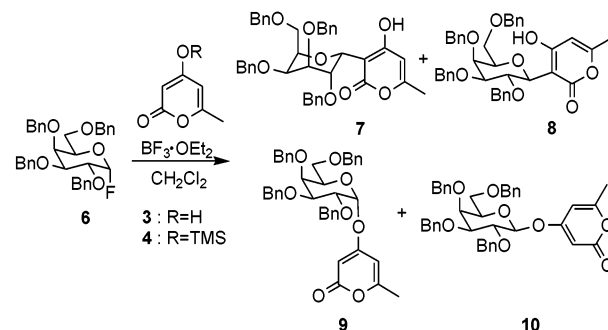
Scheme 1. Glycosylation Reaction between the Trichloroacetimidate **2** and Pyrones **3** and **4**



cases O-glycosylation proceeded smoothly in the presence of TMSOTf in CH₂Cl₂ affording β -*O*-glycoside **5**⁹ in high yield (90% for **3** and 87% for **4**), but we could not detect the formation of the desired C-glycoside. With an excellent methodology developed by Suzuki in mind, we also at-

tempted an *O*→*C* glycosyl rearrangement¹⁰ of **5** by treatment with Lewis acid. Despite varying the reaction conditions (TMSOTf in CH₂Cl₂, BF₃·OEt₂ in CH₂Cl₂, AgClO₄ in CH₂Cl₂, etc.), none of the desired C-glycoside was obtained and, in most cases, almost all of the starting material **5** was recovered. The low reactivity of the *O*→*C* rearrangement could be due to electron-withdrawing acetoxy group at C-2 in **5**. This led us to employ an *O*-benzyl-protected galactose derivative as the glycosyl donor. After a systematic survey of glycosyl acceptors (4-OBn pyrene, 4-OMe pyrene, 4-OTMS pyrene), Lewis acids, and solvents (TMSOTf in CH₂Cl₂/CH₃CN, BF₃·OEt₂ in CH₂Cl₂/CH₃CN, SnCl₄ in CH₂Cl₂, AlCl₃ in CH₂Cl₂), we obtained a mixture of both C-glycosides (**7** and **8**) and *O*-glycosides (**9** and **10**) by treatment of tetra-*O*-benzyl-D-galactopyranosyl fluoride **6**¹¹ with 4-hydroxy-6-methylpyrone **3** in the presence of BF₃·OEt₂ in CH₂Cl₂ (Table 1, entry 1). The structures of the C-gly-

Table 1. Glycosylation Reaction between the Galactosyl Fluoride **6** and Pyrones **3** and **4**



entry	pyrone (equiv)	BF ₃ ·OEt ₂ (equiv)	solvent	yield (%)		
1	3 (1)	1	CH ₂ Cl ₂	7 (15)	8 (8)	9, 10 (15)
2	3 (3)	1	CH ₂ Cl ₂	7 (47)	8 (1)	9, 10 (7)
3	3 (5)	1	CH ₂ Cl ₂	7 (56)	8 (5)	9, 10 (20)
4	3 (10)	1	CH ₂ Cl ₂	7 (54)	8 (4)	9, 10 (21)
5	3 (5)	1	CH ₃ CN	7 (9)	8 (14)	9, 10 (2)
6	3 (10)	3	CH ₃ CN	7 (21)	8 (16)	9, 10 (8)
7	3 (10)	3	CH ₂ Cl ₂	7 (36)	8 (7)	9, 10 (37)
8	4 (5)	1	CH ₂ Cl ₂	7 (40)	8 (3)	9, 10 (38)
9	4 (10)	1	CH ₂ Cl ₂	7 (32)	8 (trace)	9, 10 (46)

cosides **7** and **8** were established by ¹H NMR analysis. The anomeric proton of the minor glycoside **8** appeared at δ 4.71 with $J = 9.4$ Hz allowing us to determine the β -C-glycoside structure. The α -C-glycoside structure of the major isomer **7** could not be unambiguously determined from the chemical shift and the coupling constant of the anomeric proton (δ 5.25 as a singlet). However, the strong NOE (7%) between

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(9) Anomeric proton of **5** appeared at δ 5.20 with $J = 7.9$ Hz.

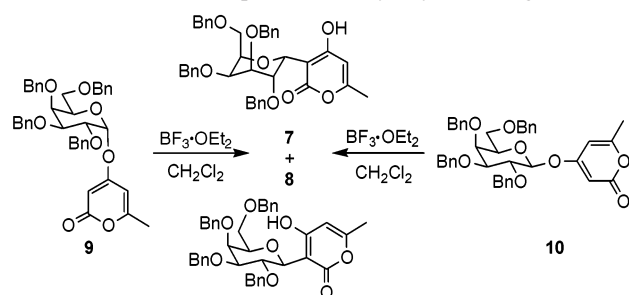
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H-1' and H-2' and the NOE (8%) between H-1' and H-6' unambiguously established the stereochemistry, which indicates that the α -C-glycoside **7** adopts the 1C_4 conformation with the equatorially oriented pyrone moiety. The yield of C-glycoside was improved by using an excess amount of pyrone **3** (Table 1, entry 1–4). In acetonitrile, C-glycosylation did not proceed well, but the desired β -C-glycoside was obtained in higher yield than in CH_2Cl_2 . We think that this selectivity is due to the nitrile effect.¹² When TMS-protected pyrone **4** was used, a considerable amount of O-glycosylation products were formed (Table 1, entry 8–9).

To determine whether C-glycoside was formed via O-glycoside, **9** and **10** were separately treated with $\text{BF}_3 \cdot \text{OEt}_2$ in CH_2Cl_2 (Scheme 2). Interestingly, both **9** and **10** afforded

Scheme 2. Attempted O→C Glycosyl Rearrangement



S.M.	Yield (%)	Recovered S.M.(%)
9	7 (9) 8 (0)	39
10	7 (trace) 8 (1)	66

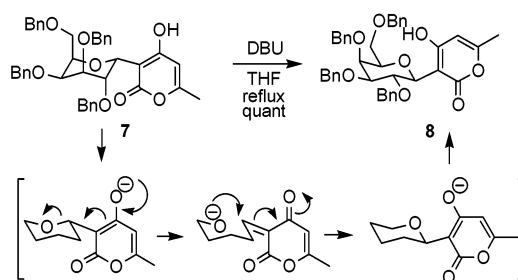
only a trace amount of C-glycoside. Therefore, C-glycosylation and O-glycosylation compete in the case of 4-hydroxypyrene. These results are quite different from glycosylation reactions involving electron-rich aromatics.

The results also show that the Fries-type rearrangement, which proceeded during acylation of 4-hydroxypyrene, did not occur.¹³

Although the desired β -C-glycoside was produced as a minor isomer, we found that the α -C-glycoside **7** underwent ready and quantitative isomerization^{5a,14} with DBU in refluxing THF. Isomerization probably proceeds via base-

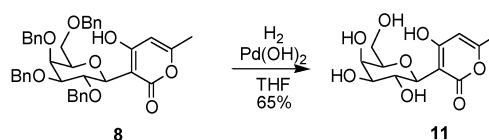
promoted ring-opening followed by recyclization, giving the thermodynamically more stable β -isomer (Scheme 3).

Scheme 3. Isomerization of **7** to **8**



Finally, it was necessary to deprotect **8**. Hydrogenolysis of **8** with $\text{Pd}(\text{OH})_2\text{--H}_2$ in THF provided **11** in 65% yield (Scheme 4). The ^1H NMR and ^{13}C NMR spectral data of **11**

Scheme 4. Hydrogenolysis of the C-Glycoside **8**



in CD_3OD were in good accordance with those of **1** except for the differences due to the conjugated side chain moiety.

In summary, glycosylation of 4-hydroxypyrene **3** and galactopyranosyl fluoride **6** provides the first reported synthetic route to C^3 -pyranosyl-4-hydroxypyrene. We also observed the base-promoted isomerization of α -C-glycoside into the desired β -C-glycoside **8**. This two-step process affords **8** in a 61% overall yield. Furthermore, the present investigation supports the proposed C^3 - β -galactopyranosyl-4-hydroxypyrene core structure of **1**.

Application of this method to the total synthesis of **1** is currently in progress.

Acknowledgment. We thank Dr. Tamotsu P. Niki (Gencom Corporation) for helpful discussions and Mitsubishi Pharma Corporation for providing the NMR spectra of D8646-2-6.

Supporting Information Available: Detail experimental procedure, full characterization, and copies of all new compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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