A Biosynthetic Proposal for Ring Formation in the Antitumor Agent Halichomycin. Asymmetric Synthesis of the AB-Carbon Backbone of Halichomycin

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ABSTRACT

A biosynthetic proposal for ring formation in the antitumor agent halichomycin is presented in which macrocyclization of the putative prehalichomycin intermediate 1 is the first step. Compound 2 then undergoes dehydration to the α -keto *N*-acylimine 3 followed by tandem nucleophilic addition of the C(16)-hydroxyl to form the hemimacrolactam. A stereospecific Michael ring closure and enol protonation complete C-ring assembly. So far, synthetic efforts toward 1 have resulted in 8.

Halichomycin is a structurally unprecedented tricyclic hemimacrolactam produced by a strain of *Streptomyces hygroscopicus*, obtained from the gastrointestinal tract of *Halichoeres bleekeri*, a well-known marine fish.¹ Halichomycin displays powerful antitumor effects in vitro, exhibiting an ED₅₀ of 0.13 μ g/mL against a murine P388 lymphocytic leukemia cell line. As such, it is of potential interest for the future treatment of human cancer.

The extraordinary molecular complexity of halichomycin naturally raises questions about its biosynthesis. While much

of its skeleton looks propionate- and acetate-derived, it is not at all clear how the bonds adjacent to the C(8)-C(9)bond are formed by such a pathway. It is also not readily apparent how nature closes the three intersecting ring systems that are present, which include a fully functionalized 11membered ether ring and a structurally unique 13/11membered bicyclic hemimacrolactam. These combined conceptual difficulties recently led Kobayashi and Ishibashi to comment that the biosynthetic provenance of halichomycin "appeared to be strange".² Our biosynthetic proposal for ring assembly in halichomycin invokes the branched precursor **1**

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as an intermediate and postulates internal macrocyclic N-acylcarbinolamine formation³ as the first step in ring formation (Scheme 1). Intermediate **2** is then thought to



undergo dehydration to the α -keto-*N*-acylimine **3**. Molecular models of **3** show that it can adopt a conformation appropriate for stereospecific internal attack of the C(16)-hydroxyl upon the *N*-acylimine carbon,⁴ which would lead to the hemimacrolactam **4** after C(14)-ketone reduction and *O*methylation with *S*-adenosylmethionine. Models further show that the C(7)-hydroxyl of **4** can readily approach the C(22)position of the dienone by a trajectory suitable for acidcatalyzed Michael ring closure, a process that could later be followed by stereospecific enol protonation at C(21). To experimentally test this simple biogenetic hypothesis, by both chemical and biological means, we embarked on an asymmetric total synthesis of the putative halichomycin precursor 1, and now report a synthetic strategy for the key AB-carbon backbone intermediate 8 needed for this venture.

We envisaged constructing the C(18)-C(24) dienone sector of **1** through a Kishi-Nozaki-Hiyama-Takai coupling⁵ between **6** and **7**, followed by oxidation (Scheme 2).



Aldehyde **6** would be derived from **8** by ammonolysis, *O*-debenzylation, and primary alcohol oxidation. A double

⁽³⁾ It is well-known that primary amides add reversibly to the carbonyl group of aldehydes and ketones in acidic, neutral, or basic media. In fact, for many aldehydes, the resulting *N*-acylcarbinolamines are quite stable and isolable. According to Weinreb, the equilibria of such additions usually lies toward the *N*-acylcarbinolamine to the extent of 5 kcal/mol. See: Auerbach, J.; Zamore, M.; Weinreb, S. M. J. Org. Chem. **1976**, *41*, 725. For reviews on this topic, see: (a) Challis, B. C.; Challis, J. A. Comprenensive Organic Chemistry; Barton, D., Ollis, W. D., Eds.; Pergamon: Oxford, 1979; Vol. 2, Chapter 9,9, p 957. (b) Zaugg, H. E.; Martin, W. B. Org. React. **1965**, *14*, 52. For other pertinent papers on this topic, see: (a) Vail, S. L.; Moran, C. M.; Barker R. H. J. Org. Chem. **1965**, *30*, 1195. (b) Feuer, H.; Lynch, U. E. J. Am. Chem. Soc. **1953**, *75*, 5027. (c) Overkleeft, H. S.; van Wittenberg, J.; Pandit, U. K. Tetrahedron **1994**, *50*, 4215.

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Wittig sequence involving aldehyde **10** and ylide **11** was envisioned for stereospecific elaboration of the dienoate array in **8**, while a Stille reaction⁶ with β -stannylenone **9** was planned for fashioning the dienone perimeter. The *syn*relationship between the C(6) and C(7) stereocenters of **10** could potentially be controlled through an Evans asymmetric aldol reaction⁷ between **12** and **13**, while a face-selective alkylation reaction between **15** and **16** could set the C(8) stereocenter. We imagined deriving lactone **16** from the known homoallylic alcohol **17**, which would be available from the Roush asymmetric crotylboration of **19** with (*R*,*R*)-**18**.⁸

The synthesis of vinyl iodide 26 is shown in Scheme 3. O-Benzvlation of the anti-alcohol 17 with O-benzvl trichloroacetimidate and catalytic TfOH9 procured the doubly protected alkene 20, which was converted to the primary alcohol by rhodium-catalyzed hydroboration¹⁰ and oxidation. Further oxidation to acid 21 was accomplished with PDC in DMF. Hydrogenation of acid **21** over a 20% $Pd(OH)_2$ on carbon catalyst effected a clean, but rather slow, deprotection of the O-benzyl ether to permit in situ butyrolactonization. The stereospecific C-alkylation of butyrolactone 16 was achieved by low-temperature enolization with LDA and addition of the allylic bromide 15.¹¹ The total stereocontrol observed in this reaction is attributable to the stereodirecting influence of the C(25)-Me group (which hinders synapproach of the bulky electrophile to the enolate) and preservation of the reaction temperature at -78 °C throughout. In this regard, premature warming markedly lowered the selectivity levels that could be attained. The configuration of the newly induced stereocenter in 14 was verified by NOE analysis. An OPMB for OTBDPS protecting group interchange was now effected to permit C(19)-C(24) side-chain elaboration later on in the synthesis; this delivered the PMBether 22.

Having fulfilled its role in stereospecific attachment of the C(8)-methallyl unit, the butyrolactone ring of **22** was reductively ring-opened with lithium borohydride and diol **23** differentially *O*-silylated to obtain **24**. Selective cleavage of the primary OTES group now permitted oxidation to the aldehyde **12** with TPAP/NMO.¹² The Evans aldol addition between **12** and **13** required the use of a significant excess of the propionimide enolate (4 equiv) to drive the reaction to completion, which made the purification of **25** exceedingly difficult. The subsequent *O*-triethylsilylation reaction rem-



^a Reagents and conditions: (a) BnOC(NH)CCl₃ (1 equiv), TfOH (0.05 equiv), CH₂Cl₂ (0.4 M), rt, 3.5 h; (b) catecholborane (1.1 equiv), (Ph₃P)₃RhCl (0.05 equiv), THF (0.2 M), 0 °C for 5 min, then rt for 14 h; 27.5% H₂O₂/MeOH/2N NaOH, 0 °C, 2 h; (c) PDC (7 equiv), DMF (0.3 M), rt, 48 h; (d) H₂, 20% Pd(OH)₂/C, MeOH (0.4 M), 3-7 d; (e) 16, LiN(Pr-i)₂ (1.3 equiv), THF-HMPA (10: 1) (0.2 M), -78 °C, 1 h, then add 15 (1.2 equiv) in THF at -78 °C dropwise and stir at -78 °C for 2 h; (f) 40% aqueous HF/THF/ MeCN (1:2:1) (concentration of 14 ca. 0.09 M), rt, 24-27 h; (g) PMBOC(NH)CCl₃ (2 equiv), PPTS (0.5 equiv), CH₂Cl₂ (0.1 M), rt, 7 h; (h) LiBH₄ (10 equiv), THF/MeOH (100:1) (0.2 M), Δ, 3 h; (i) 23, Imid (2.2 equiv), DMF (concentration of 23, ca. 0.1 M), 0 °C, add Et₃SiCl (1.2 equiv) over 5 min, then stir at 0 °C for 1.5 h; (j) 2,6-lutidine (20 equiv), CH₂Cl₂, -50 °C, add TBSOTf (3 equiv) over 5 min, then stir for 0.5 h; (k) 2% aqueous HF, THF/MeCN (1:1), rt, 1.5 h; (l) TPAP (0.05 equiv), NMO (2 equiv), CH₂Cl₂ (0.01 M), 4A MS, rt, 40 min; (m) 13 (4 equiv), (n-Bu)₂BOTf (4 equiv), Et₃N (4.2 equiv), CH₂Cl₂, 0 °C, 0.5 h, then cool to -78°C, add 12 (1 equiv) in CH₂Cl₂, stir for 35 min, then warm to rt for 1 h; (n) add Et₃SiOTf (5 equiv) over 5 min to 25 in CH₂Cl₂ (0.02M), 2,6-lutidine (20 equiv), at -50 °C, then warm to rt for 45 min.

edied this situation, allowing the protected aldol adduct **26** to be isolated pure by simple flash chromatography. Significantly, no other aldol adducts were observed in the above addition. The structure of **26** was verified by X-ray crystallography.

Attention now shifted toward stereospecific elaboration of the two diene arrays present within 8 (Scheme 4).

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^{*a*} Reagents and conditions: (a) add LiBH₄ (10 equiv) in one portion to **26** in Et₂O/H₂O (160:1) (0.02 M), at 0 °C, then warm to rt and stir for 1.5 h; (b) TPAP (0.1 equiv), NMO (2 equiv), CH₂Cl₂ (0.02 M), 4A MS, rt, 1 h 10 min; (c) **11** (10 equiv), PhMe (0.01 M), Δ , 5 h; (d) *i*-Bu₂AlH (2.2 equiv), PhMe (0.048M), -78 °C, 0.5 h; (e) MnO₂ (20 equiv), CHCl₃ (0.02 M), Δ , 6 h; (f) **11** (15 equiv), PhMe (0.01 M), Δ , 16 h; (g) **9** (2 equiv), (CH₃CN)₂PdCl₂ (0.5 equiv), *i*-Pr₂NEt (10 equiv), DMF (0.01 M), 5 h.

Reductive removal¹³ of the oxazolidinone unit from **26** with LiBH₄ furnished the primary alcohol **27** in excellent yield (82%). A TPAP oxidation¹² converted **27** into the aldehyde



^{*a*} Reagents and conditions: (a) Bu₃SnH (1.2 equiv), AIBN (0.05 equiv), PhMe (0.3 M), Δ , 24 h; (b) PPTS (1.5 equiv), MeOH (0.3 M), rt, 24 h; (c) TBSCl (1 equiv) (0.03 M in CH₂Cl₂) added dropwise to **34** (1 equiv) and imidazole (2 equiv) in CH₂Cl₂ (concentration of **34** ca. 0.26 M) at 0 °C; stir 10 min; (d) TPAP (0.05 equiv), NMO (2.2 equiv), 4A MS, CH₂Cl₂ (0.26 M), rt, 1 h.

10, which reacted readily with 11 in PhMe at reflux to give 28 with complete stereocontrol.¹⁴ DIBAL reduction to 29 and allylic alcohol oxidation with MnO2 generated the aldehyde 30, which willingly engaged in a second Wittig reaction with 11. The (E,E)-dienoate 31 was formed as a single geometrical isomer in 97% yield. The dienone unit was fashioned by a Stille coupling⁶ between 31 and 9 (for the preparation of 9, see Scheme 5). The desired tetraene 8 was isolated as a single geometrical isomer in 33-40% yield, but was formed alongside a significant quantity of the stannane homocoupling product. Work is currently underway to improve the yield of 8 and to reduce the amount of dimerization that is occurring with stannylenone 9. Future reports will deal with the synthesis of isotopically labeled 1 from 8, and with our chemical and biological efforts to convert 1 into halichomycin itself.

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Supporting Information Available: High-resolution mass spectra, 500 MHz ¹H and 125 MHz ¹³C NMR spectra of all new compounds, and X-ray data for **26**. This material is available free of charge via the Internet at http://pubs.acs.org.

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