The Synthesis of Alisamycin, Nisamycin, LL-C10037 α and Novel Epoxyquinol and Epoxyquinone Analogues of Manumycin A

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Dedicated to Professor Sandy McKillop to mark his retirement from the University of East Anglia, Norwich and the many achievements, both in research and teaching, during his career there.

Abstract: Versatile synthetic routes are described for the preparation of a range of epoxyquinol and epoxyquinone analogues of the antitumour antibiotic manumycin A lacking the lower side chain, and these procedures have also been applied to prepare the bioactive natural product LL-C10037 α . The extension of this methodology to provide a general synthetic route to the manumycin family of antibiotics is discussed and exemplified by the first total synthesis of aliasmycin and *ent*-aliasmycin. This route includes the novel, stereoselective organometallic addition of the Corey– Wollenberg reagent (*E*-2-tributylstannylethenyllithium) to the manumycin nucleus and palladium catalysed Stille coupling technology for the introduction of the polyunsaturated 2-amino-3hydroxycyclopentenone derived amide. Similar methodology has also been employed to complete the first total synthesis of the antibiotic nisamycin.

Key words: LL-C10037 α , alisamycin, nisamycin, manumycin analogues, Stille coupling

The manumycin family of antibiotics has grown steadily since the isolation of manumycin A(1) from *Streptomyces* Parvalus in 1963.^{1,2} Other members (Figure 1) include manumycin B (2),³ manumycin C (3),³ manumycin E (4),⁴ manumycin F (5),⁴ manumycin G (6),⁴ asukamycin (7),⁵ alisamycin (8),^{6–8} *ent*-alisamycin (9),⁹ El-1625-2 (10),⁹ El-1511-3 (11)⁹ and El-1511-5 (12).⁹ All of these compounds have the same E,E,E-trienyl lower side chains terminating in a 2-amino-3-hydroxycyclopentenone derived amide. U-56407 $(13)^{10}$ is reported to possess a Z,Z,E-trienyl lower side chain (although this stereochemical assignment must be treated with caution^{5b}) whereas colabomycin (14)¹¹ is a higher vinylogue, nisamycin $(15)^{12}$ lacks the terminal amide linkage, and U-62162 $(16)^{13}$ has a saturated lower side chain. The upper side chains of the family members are more distinctive but all of these compounds are based on the epoxyaminocyclohexenone nucleus which is also found in the anti-tumour antibiotic LL-C10037 α (17),¹⁴ and the broad spectrum an-tibiotic MM-14201 (18).¹⁵ Most remarkably, all diastereoisomers of the central nucleus are represented in the manumycin family. Thus, manumycin A (1), together with 2 and 10 have the *anti*-(4R, 5R, 6S) configuration whereas manumycin C (3) and 6, 7, 9 and 11-14 all have the isomeric syn-hydroxy epoxide (4S, 5R, 6S) configuration found in LL-C10037 α (17). Manumycin F (5) is in the enantiomeric anti-series (4R, 5S, 6R); alisamycin (8) and nisamycin (15) are in the enantiomeric syn (4R, 5S, 6R) series along with MM 14201 (18). The related diols, manumycin D (19), 3,16 TMC-1 A (20), 16 TMC-1 B (21), 16 TMC-1 C (22), 16 and TMC-1 D (23) 16 (Figure 2) have also been isolated.

The manumycin natural products possess a range of bioactivities in addition to their anti-bacterial properties. These include antifungal activity and inhibitory activity against polymorphonuclear leukocyte elastase and interleukin-1 β converting enzyme. Of greatest interest, however, is the discovery that the manumycins A–C act as selective inhibitors of *ras* farnesyltransferase and as such have potential in cancer chemotherapy.¹⁷ It is noteworthy that the oxidative degradation products of the manumycins, epoxyquinones **24–26** (Figure 3), retain their anti-tumour activity indicating that the lower side chain is not too important in this regard. The dihydroxy manumycin analogues **19–23** were also shown to retain cytotoxicity but were ineffective antibiotics, indicating the importance of the epoxide group for antibacterial activity.¹⁶

There have been a number of studies by Floss, Gould, Zeeck and others^{3b,5b,18} to elucidate the biosynthetic pathway leading to the manumycin family and related compounds. It has been established that the polyene side chains are of polyketide origin and that the C₅N unit which terminates the lower side chain is derived from 5-aminolevulinic acid. The biosynthesis of the mC_7N nucleus, however, proved surprising in that it is not shikimate derived but rather originates from 3-amino-4-hydroxybenzoic acid^{18a} which in turn is constructed from succinate and glycerol.^{3b}

Despite their promising biological properties and interesting biogenesis, synthetic approaches to the manumycins have not been reported (although Wipf et al. have prepared analogues lacking the lower side chain¹⁹). In designing a synthetic route to the manumycin antibiotics we adopted an approach which would also allow us to prepare a range of quinol 29 and quinone 30 analogues lacking the lower side chain for further structure-activity studies. Thus, as shown in Scheme 1, the protected amido quinone epoxide 28 was identified as a key intermediate for the synthesis of both novel manumycin analogues and the natural products themselves. We envisaged preparing 28 by acylation of 27 with the acid chloride corresponding to the upper side chain, the simplified target molecules then being obtained by straightforward functional group modification. It should be noted that the alternative sequence involving acylation before epoxidation, is not viable in these systems due to facile N-deacylation during epoxidation.^{19,22}

Amides **28** were also seen as the cornerstone of the synthetic route to the manumycin family, as is shown in retrosynthetic form in Scheme 2.

Biographical Sketches





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Richard Taylor gained his B. Sc. and Ph. D. from the University of Sheffield, carrying out his postgraduate research into steroid synthesis under the supervision of Dr. D. Neville Jones. He then carried out postdoctoral research with Dr. Ian Harrison at Syntex, Palo Alto and Professor Franz Sondheimer at University College, London. After academic jobs with the Open University (1975–79) and the University of East Anglia, Norwich (1979–1993) he moved to a Chair of Organic Chemistry at the University of York in 1993. Professor Taylor's research interests centre on the development of new organometallic and organosulfur based synthetic methodology and its application to the synthesis of compounds of biological importance. Natural product classes studied include steroids, prostaglandins, prostacyclins, thromboxanes, leukotrienes, insect pheromones, and a range of marine, fungal and plant metabolites.

Lilian Alcaraz was born in 1969 in France and received his Ph. D. degree in 1995 from the University of Strasbourg. His research was carried out on a novel synthetic approach to the total synthesis of Taxol as well as the development of new methodology using ylides under the direction of Dr. C. Mioskowski. In December 1995, Lilian joined the University of York as an ARC (France) Fellow and in 1996 was awarded a Marie-Curie (EC) postdoctoral fellowship. He is currently working as a research chemist for Astra Charnwood.

Isabelle Kapfer-Eyer was born in 1966 in France and received her Ph. D. degree in 1993 from the Université Louis Pasteur, Strasbourg. Her research was carried out in bio-organic chemistry under the direction of Professor M. Goeldner. In 1993, Isabelle joined the University of York as an EC TMR Fellow. In 1995 she took a teaching position (A. T. E. R.) in organic chemistry at the University of Strasbourg and is currently preparing for the agregation diploma in physical sciences.

Gregor Macdonald was born in Scotland in 1971 and received his B. Sc. from the University of Edinburgh in 1992. After two years with the Wellcome Foundation he joined the University of York to carry out his Ph. D. He was awarded the Kathryn Mary Stott Prize and a Pfizer Prize for his postgraduate research. He is currently working as a research chemist at SmithKline Beecham Pharmaceuticals.

Xudong Wei was born in 1965 in China and was awarded his Ph. D. degree in 1991 from Nanjing University. His research was carried out on new oxidative methodology and the addition reactions of iminoxy radicals under the supervision of Professor Hongwen Hu. Dr. Wei was appointed to a lectureship at Nanjing University in 1991 researching into heterocyclic chemistry. From 1994–1995 he carried out postdoctoral research with Professor Bernd Speiser at the University of Tübingen and in 1995–1996 was awarded a Royal Fellowship to study in York. Since then he has been supported by the University of York Innovation and Research Priming fund and the EPSRC.

Norman Lewis gained his B. Sc. and Ph. D. from the University of Bristol, under the supervision of Professor J. MacMillan. After carrying out postdoctoral research at the University of British Columbia (Professor J. Kutney) and the E. T. H., Zurich (Professor A. Eshenmoser), he joined Smith, Kline and French in 1982. He is now Associate Director in the the Synthetic Chemistry Department (part of Chemical Development) at Smith-Kline Beecham Pharmaceuticals, Tonbridge.



Figure 1



Thus, we planned to use organometallic elaboration of **28** with vinyllithium reagent 31^{20} to give the vinylstannane **32**. This organometallic addition was obviously the crucial (and most speculative) step in the sequence and is discussed in more detail later. The use of the Stille coupling between vinylstannane **32** and bromodienamide 33^{21} was central to the end-game of this succinct synthetic approach (see later).

Results and Discussion

(a) Preparation and Elaboration of Amine 27²²

The preparation of the key epoxycyclohexenylamine 27 was achieved using methodology based on that employed in our synthesis of the diepoxycyclohexanone antibiotic aranorosin²³ and the marine antitumour natural product

Scheme 1

OH (29)

bromoxone²⁴ (and to that employed by Wipf and Kim¹⁹ in their synthesis of LL-C10037 α). The BOC derivative of 2,5-dimethoxyaniline (**34**) was oxidised using PhI(OAc)₂^{19,23-25} to give monoacetal **35** in 78% yield (Scheme 3). Several epoxidation methods were investigated for the conversion of **35** into monoepoxide **36**. On a small scale (0.4 mmol) sodium perborate²⁶ gave the best results (36% + 45% recovery of starting material) but the efficiency diminished as the reaction was scaled up. On a larger scale the preferred procedure involved the use of

(30)

Novel manumycin analogues





 H_2O_2/K_2CO_3 in aqueous THF (methoxy conjugate addition adducts were observed when methanol was employed as solvent).

In order to obtain the key amine intermediate 27 from 36 selective removal of the BOC group in the presence of the methyl acetal was required. This was efficiently achieved using boron trifluoride-diethyl ether complex and activated molecular sieves in dichloromethane at room temperature. This novel and mild procedure, which was developed for this transformation, has also proved to be more widely applicable.²⁷ To our delight, amine **27** proved to be a stable, crystalline solid (mp 156–157°C) which could be stored at -20 °C for several weeks without noticeable decomposition. It is a vinylogous amide and therefore, not surprisingly, proved to be rather unreactive to standard acylation conditions. Acetylation was achieved in good yield (79%; 84% based on recovered starting material) with excess acetic anhydride and DMAP in THF but the transformation did not give reproducible yields, and did not proceed at all when acetyl chloride was employed. The requirement for an excess of an acid anhydride seemed to limit the value of this route for the preparation of manumycin analogues and therefore other acylating conditions were explored (Scheme 4). It was eventually found that acylation to give amides 28 could be carried out using a number of acid chlorides 37^{28a} providing lithium tert-butoxide in THF was employed as the base.^{28b} Reduction of ketones 28 was achieved with sodium borohydride and then acetal hydrolysis carried out using tosic acid and pyridinium *p*-toluenesulfonate (PPTS)¹⁹ to give separable mixtures of the syn- and anti-hydroxy epoxides 29. Compound syn-29a is the anti-tumour agent LL-C10037 α 17 and its NMR characteristics were entirely consistent with published data.^{14,19} This new route²² to (\pm) -LL-C10037 α is only 7 steps long and, although unoptimised, proceeds in ca. 10% overall yield. It also reduces the number of protection-deprotection steps compared to the published procedure (9 steps, ca. 7% overall yield).¹⁹ Amides 29b and **29c** are analogues of alisamycin (8) and asukamycin (7), respectively, lacking the lower side chains.





We also extended this chemistry to produce the manumycin quinone analogues **30** lacking the C-4 polyene chain (Scheme 4). Thus PDC oxidation of alcohols **29** produced the known epoxyquinones **30a**^{14b,29} and **30c**^{5a} in fair to excellent yields.

(b) Alternative Routes to Epoxyquinones 30

The route to epoxyquinones **30** shown in Scheme 4 is rather long and other routes were investigated as shown in Schemes 5 and $6^{30,31}$ Thus, we first attempted to prepare amine **39** which, on acylation, would lead directly to a range of quinones. The direct hydrolysis of **36** to give **38** or **39** was not successful but an efficient indirect route to prepare **38** was developed (Scheme 5). Once again, the use of boron trifluoride–diethyl ether²⁷ gave efficient removal of the BOC group, amine **39** being obtained as a crystalline material (mp 133–134 °C dec.) which proved to

be stable when stored at -20 °C. Unfortunately, amine **39** was very unreactive towards acylation using a range of conditions (including *tert*-BuOLi, THF). It was eventually found that acylation could be accomplished using acid chlorides in the presence of bis(trimethylsilyl)trifluoromethylacetamide (BSTFA) as a chloride scavenger.³² Even with these optimum conditions, however, the acylation yields were satisfactory only with saturated acid chlorides. With benzoyl chloride, the yield was reduced and no acylation at all was observed with polyunsaturated systems such as **37c**.



Scheme 5

The routes to epoxyquinones shown in Scheme 5 is still lengthy and is limited to more reactive acid chlorides. We therefore developed the third route shown in Scheme 6. In this procedure 2,5-dimethoxyaniline (34) is acylated in the first step and then hypervalent iodine oxidation followed by acetal removal gives the amidoquinones 40. Monoepoxidation of dienones 40 was attempted using hydrogen peroxide and tert-butyl hydroperoxide with a range of bases (Na₂CO₃, NaOH, triton B, triethylamine, pyridine etc.) but it was eventually discovered that the best procedure utilised tert-butyl hydroperoxide with DBU as base.³³ Epoxides **30** were found to be very sensitive towards basic conditions and thus, for optimum yields, it was important to minimise the reaction time. This was not a problem given the susceptibility of 40 towards epoxidation: reaction times of 30 seconds gave yields of 67–81%. For these high yields of epoxides 30 it was essential to remove the DBU and excess peroxide immediately after the completion of the reaction. This was achieved by washing the reaction mixture with saturated aqueous iron(II) sulfate in place of the metasulfite procedure³³ normally employed.

(c) Organometallic Addition Reactions

As shown in Scheme 2, the proposed synthetic route to the manumycin natural products involved elaboration of ke-



Scheme 6

tones 28 by organometallic addition to C-4. This approach appeared to be fraught with potential problems, particularly concerning the regio and stereoselectivity of the addition reaction. The likely low reactivity of the C-4 carbonyl group, which is a vinylogous imide, and which would presumably be further deactivated by deprotonation of the amide substituent, was a particular concern. In view of this, we first carried out the model studies as shown in Scheme 7. To our delight, reaction of amides 28b and 28c with phenyllithium in THF, gave good yields of adducts 41 as single diastereoisomers with no other byproducts observed. The syn-stereochemistry of compounds 41 was assigned on steric grounds and by analogy with the reduction reactions shown in Scheme 5 where the syn-isomer predominates.^{34a} In view of the importance of this stereochemical assignment, however, we felt that more concrete evidence was required. Unfortunately, high field NMR spectroscopy did not provide an unambiguous answer and we therefore sought a crystalline adduct. Suit-





able crystals were obtained from adduct **42** produced by phenyllithium addition to the *N*-BOC analogue **36**, and X-ray crystallography^{34b} confirmed the *syn*-hydroxy epoxide relationship [which is present in alisamycin (**8**) and other members of the manumycin family].

(d) Construction of the Manumycin Lower Side Chain

As mentioned earlier (Scheme 2) we planned to elaborate the lower manumycin side chain via a palladium-catalysed Stille coupling procedure.^{21,35,36} Model studies, shown in Scheme 8, were carried out to establish the viability of this approach with cyclohexane carbinol vinylstannane 43^{37} and bromodienes $44^{21,38}$ as substrates. A range of palladium catalysts were investigated, initially with ester 44a, and although adduct 45a was isolated it was discovered that dimerisation of the vinylstannane occurred to a significant extent to produce 46 unless the reaction solvent was rigorously degassed.³⁹



Using 5% PdCl2(MeCN)₂ under these conditions, **44a** gave trienyl ester **45a** in 82% yield. Acid **44b** also underwent efficient coupling, although the first formed product (tentatively assumed to be the stannyl ester⁴⁰) had to be treated with fluoride and then hydrolysed to provide adduct **45b**. In this case, the optimum procedure utilised the palladium(0) catalyst generated from PdCl₂(PPh₃)₂ and DIBAL-H using Negishi's procedure.³⁶ We then attempted the direct introduction of the fully functionalised lower side chain unit **33** (previously described by us²¹), and found that coupling again proceeded in good yield using PdCl₂(MeCN)₂ providing that the solvent was efficiently deoxygenated. Unfortunately, using these conditions, adduct **47** was contaminated by minor impurities. We dis-



covered that these impurities could be avoided by use of the Negishi catalyst³⁶ with adduct **47** being obtained in an excellent 83% yield. The *E*,*E*,*E*-stereochemistry of adducts **45** and **47** were established by high field NMR studies.

(e) Initial Approach to Alisamycin

We were now in a position to attempt the first synthesis of a member of the manumycin family of antibiotics. Alisamycin (8) was chosen to illustrate the methodology. Alisamycin^{6–9} is a potent antibiotic (MIC 1.6 mg/mL against *staphylococcus aureus*) which also possesses weak antifungal and cytotoxic activity.⁶ Its structure was established by Chatterjee et al.⁷ using a combination of mass spectrometry, IR, UV and NMR spectroscopy, and the absolute configuration was revealed as 4*R*, 5*S*, 6*R* using the exciton chirality method together with CD studies of degradation products.⁸ The enantiomer **9** also occurs naturally.⁹ The first synthetic approach to racemic alisamycin is shown in Scheme 9.⁴¹



Thus, treatment of protected quinone **28b** with *E*-2-tributylstannylethenyllithium (**31**), generated from the corresponding bis-stannane,²⁰ introduced an embryonic C-4 side chain giving **48** in 55% yield (79% based on recovered starting material). Stille coupling of vinylstannyl reagent **48** with bromodienamide **33**²¹ proceeded efficiently using PdCl₂(PPh₃)₂/DIBAL-H in DMF/THF at room temperature to give **49** in 72% yield. All that was required to complete the synthesis of alisamycin was to deprotect the acetal functionality of **49**. Unfortunately, using a number of procedures, ^{31,42,43} either little reaction occurred or, under more forcing conditions, decomposition was observed. Similar problems were encountered when attempting to deprotect the phenyl analogue **41b** (Scheme 10): the only product that could be isolated was ketone **50** resulting from enamide hydrolysis (with lithium perchlorate in MeCN⁴³ **50** was formed in almost quantitative yield).



(f) Successful Synthesis of Alisamycin and Nisamycin via Organometallic Additions to Ouinone **30b**

We therefore turned to a more direct (and even more speculative) synthetic approach (Scheme 11). In this procedure, organometallic addition was carried out using the unprotected epoxyquinone **30b**. Treatment with *E*-2tributylstannylethenyllithium (**31**) gave two major products **51** and **52** (46%, 1:1.3), resulting from attack at each of the ring carbonyl groups. The stereochemical assignments were again made on the basis of steric considerations and, in the case of **52**, this was confirmed by conversion into alisamycin and nisamycin.

Adducts **51** and **52** were separated by chromatography and individually coupled to bromodienamide **33** using $PdCl_2(PPh_3)_2/DIBAL-H$ as before. The use of **52** produced alisamycin **8** in 64% yield, the structure being confirmed by comparison of IR, MS, ¹H and ¹³C NMR data with authentic spectra.⁴⁴ The NMR data for the epoxycyclohexenone nucleus was particularly informative [found $\delta_{\rm H}$: H-3, 7.41 (d, J = 2.5 Hz), H-5, 3.72 (dd, J = 3.5, 2.5 Hz), H-6, 3.67 (d, J = 3.5 Hz); $\delta_{\rm C}$: C-3, 126.0, C-5, 57.4, C-6, 53.0; authentic⁷ $\delta_{\rm H}$: H-3, 7.40 (d, J = 2.6 Hz), H-5, 3.70 (dd, J = 3.6, 2.6 Hz), H-6, 3.65 (d, J = 3.6 Hz); $\delta_{\rm C}$: C-3, 126.3, C-5, 57.4, C-6, 52.9]. It should be noted that although this produces racemic **8**, this is in fact a mixture of two natural products, alisamycin and *ent*-alisamycin. The novel regioisomer **53** of alisamycin was also prepared by homologation of **51** in 75% yield.

This methodology was also employed to prepare nisamycin (15), and its methyl ester 54, as shown in Scheme 12. Thus, vinylstannane 52, used to prepare alisamycin, was coupled with bromodiene 44a to produce the methyl ester of nisamycin 54 in 82% yield. All attempts to convert ester 54 into acid 15 were unsuccessful, however. Acid 15 was therefore prepared, albeit in moderate yield, by the direct coupling reaction between vinylstannane 52 and bromodiene 44b. This reaction again produces the stannyl ester derivative 54 ($R = SnBu_3$) which is converted into the acid by treatment with KF followed by neutralisation with phosphate buffer.⁴⁰ Adduct **15** is (\pm) -nisamycin, the only member of the manumycin family to lack the terminal C₅N unit. This is the first synthesis of nisamycin, an antibiotic which also possesses antifungal and cytotoxic properties, and which has proved useful for the preparation of manumycin analogues containing novel amide linkages.¹² Synthetic nisamycin 15 was fully characterised and gave NMR data fully consistent with those published [δ_{H} : 7.41 (d, 1 H, J = 2.5 Hz, H-3), 3.72 (dd, 1 H, J = 4.0, 2.5 Hz, H-5), 3.66 (d, 1 H, J = 4.0 Hz, H-6); $\delta_{\rm C}$: 126.3 (C-3), 57.4 (C-5), 52.9 (C-6). Lit.^{12a} values, $\delta_{\rm H}$: 7.41





(d, 1 H, J = 2.1 Hz), 3.70 (dd, 1 H, J = 4.0, 2.1 Hz), 3.63 (d, 1 H, J = 4.0 Hz); $\delta_{\rm C}$: 126.9, 57.3, 52.8].

(g) Summary and Future Work

We have devised new chemistry for the preparation of a range of manumycin analogues of increasing complexity and utilised these procedures to prepare the natural products LL-C10037 α , alisamycin and nisamycin, the last two for the first time. The novel organometallic methodology for Southern side-chain introduction and elaboration is applicable to all of the manumycin family. We are currently optimising the synthetic route to the nucleus in enantiomerically pure form and will then apply this methodology to other members of this group of natural products.⁴⁵

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NMR spectra were recorded on Jeol GX-270 or Bruker AMX 500 instruments. TMS or CDCl₃/CHCl₃ was used as the internal standard. Carbon spectra were verified using DEPT experiments. Mps were recorded on an Electrothermal IA9100 digital melting point apparatus and are uncorrected. IR spectra were recorded on an ATI Mattson Genesis FTIR spectrometer. Low resolution electron impact (EI) mass spectra were recorded on a Kratos MS 25 spectrometer. Chemical ionisation (CI), fast atom bombardment (FAB) and HRMS were recorded on a Micromass Autospec spectrometer. Elemental analyses were carried out at the University of East Anglia, Norwich. Chromatography is medium pressure flash column chromatography and was performed using ICN silica gel (32–63) or Matrex silica gel 60 (70–200) using the eluant specified. Preparative TLC was carried out using pre-prepared plates (Merck silica gel 60 F-254, 5715). PE is petroleum ether (bp 40–60 °C). Where necessary Et_2O and THF were distilled from Na/benzophenone ketyl, and CH_2Cl_2 from CaH, immediately before use. H_2O is distilled water. Except where specified, all reagents were purchased from commercial sources and were used without further purification. All synthetic compounds are racemic.

3-tert-Butoxycarbonylamino-4,4-dimethoxycyclohexa-2,5-dien-1-one (35):

To a stirred solution of *N*-*tert*-butylcarbonyl-2,5-dimethoxyaniline⁴⁶ (10.0 g, 0.041 mol) in anhyd MeOH (100 mL) at 0°C under N₂ was added PhI(OAc)₂ (15.9 g, 0.049 mol) portionwise over 1 h. The mixture was stirred at 0°C for 6 h before being diluted with CH₂Cl₂ (200 mL)/H₂O (100 mL) and then neutralised using aq NaHCO₃. The organic layers were separated and washed with H₂O (60 mL) and brine (60 mL), dried (MgSO₄) and the solvent evaporated under reduced pressure to give a brown oil. Column chromatography (Et₂O/PE, 3:7 and then 1:1) gave **35** as an off-white solid, yield: 8.35 g (78%); *R*_f 0.30 (Et₂O/PE, 1:1); mp 130–131°C.

IR (CCl₄): v = 3415, 2986, 1743, 1672, 1502, 1232, 1153 cm⁻¹. ¹H NMR (270 MHz, CDCl₃): $\delta = 7.01$ (d, 1 H, J = 1.9 Hz, H-2), 6.85 (br s, 1 H, NH), 6.53 (d, 1 H, J = 10.2 Hz, H-5), 6.41 (dd, 1 H, $J \approx$ 10.2, 1.9 Hz, H-6), 3.27 (s, 6 H, 2 × OMe), 1.51 (s, 9 H, ^tBu). ¹³C NMR (67.5 MHz, CDCl₃): $\delta = 185.3$, 151.3, 148.2, 138.0, 133.1, 111.0, 94.1, 82.1, 51.3, 27.9.

MS (CI): m/z (%) = 270 (MH⁺, 100).

HRMS (CI): Calc for $C_{13}H_{20}NO_5$: 270.1342. Found: 270.1338. Anal: Calc C, 57.98; H, 7.11; N, 5.20. Found: C, 58.21; H, 7.13; N, 5.07.

3-*tert*-Butoxycarbonylamino-4,4-dimethoxy-5,6-epoxycyclohex-2-en-1-one (36):

To a stirred solution of dienone **35** (2.7 g, 10.03 mmol) and H_2O_2 (29% soln., 34 mL, 301.2 mmol) in THF (50 mL) was added aq K_2CO_3 (0.8 *M*, 12.5 mL, 10.0 mmol) dropwise at 0 °C. After stirring at r.t. for 6 d, the mixture was diluted with EtOAc (50 mL) and brine (10 mL) added. The aqueous layer was then extracted with EtOAc (6 × 50 mL). The combined organic layers were dried (MgSO₄) and the solvent evaporated under reduced pressure to give a brown oil. Column chromatography (CH₂Cl₂/EtOAc, 95:5) gave the title compound **36** as a white solid; yield: 1.4 g (49%); R_f 0.40 (CH₂Cl₂/EtOAc, 95:5); mp 144–145 °C.

IR (CCl₄): $v = 3416, 2980, 1747, 1679, 1630, 1500, 1153 \text{ cm}^{-1}$.

¹H NMR (270 MHz, CDCl₃): δ = 7.05 (br s, 1 H, NH), 6.77 (d, 1 H, J = 2.2 Hz, H-2), 3.82 (d, 1 H, J = 4.1 Hz, H-5), 3.65 (s, 3 H, OMe), 3.51 (dd, 1 H, J = 4.1, 2.2 Hz, H-6), 3.31 (s, 3 H, OMe), 1.50 (s, 9 H, ¹Bu).

¹³C NMR (67.5 MHz, CDCl₃): δ = 192.3, 151.1, 146.6, 106.1, 95.5, 82.5, 52.1, 51.5, 51.4, 50.6, 28.1.

MS (CI): m/z (%) = 286 (MH⁺, 100).

HRMS (CI): Calc for C₁₃H₂₀NO₆: 286.1291. Found: 286.1290.

Anal: Calc C, 54.73; H, 6.71; N, 4.91. Found: C, 54.87; H, 6.78; N, 4.81.

Unreacted starting material **35** (350 mg, 13%) was also recovered from this reaction.

3-Amino-4,4-dimethoxy-5,6-epoxycyclohex-2-en-1-one (27):

To a stirred solution of BOC compound **36** (291 mg, 1.02 mmol) and powdered activated 4 Å molecular sieves (625 mg) in anhyd CH₂Cl₂ (15 mL) was added BF₃·Et₂O (0.19 mL, 1.53 mmol) in CH₂Cl₂ (5 mL) dropwise at 0 °C over 30 min. The reaction was left to stir at r.t. for 20 h. On completion, the solvent was evaporated under reduced pressure to give a brown oil. Column chromatography (EtOAc/ MeOH, 95:5) gave **27** as a white solid; yield: 136 mg (72%); $R_{\rm f}$ 0.30 (EtOAc); mp 156–157 °C.

IR (CDCl₃): $v = 3401, 2944, 2854, 1621, 1255 \text{ cm}^{-1}$.

¹H NMR [270 MHz, (CD₃)₂CO] : δ = 6.42 (br s, 2 H, NH₂), 5.00 (d, 1 H, *J* = 2.0 Hz, H-2), 3.84 (d, 1 H, *J* = 4.0 Hz, H-5), 3.56 (s, 3 H, OMe), 3.35 (s, 3 H, OMe), 3.24 (dd, 1 H, *J* = 4.0, 2.0 Hz, H-6). ¹³C NMR (67.5 MHz, CDCl₃): δ = 190.6, 156.9, 96.3, 95.7, 52.8, 51.3, 51.7, 50.2.

MS (CI): m/z (%) = 186 (MH⁺, 100).

HRMS (CI): Calc for C₈H₁₂NO₄: 186.0766. Found: 186.0769.

Preparation of Amides (28); General Procedure:

A solution of lithium *tert*-butoxide (1.0 M solution in THF, 1.1 equiv.) was added dropwise over 15 min to a stirred solution of amine **27** (1 equiv.) in anhyd THF at 0 °C under N₂.After stirring at 0 °C for 30 min, a solution of the acid chloride **37**^{28a} (1.2 equiv.) in THF was added dropwise over 1 h. The reaction was allowed to warm and then stirred at r.t. until acylation was complete. The THF was evaporated under reduced pressure and EtOAc and H₂O were added. The two layers were separated and the aqueous layer extracted with EtOAc. The combined organic layers were washed with brine, dried (MgSO₄) and the solvent evaporated under reduced pressure to give a dark oily residue. Column chromatography (EtOAc/PE, 1:1) gave the desired compound, usually as an off-white solid which could be recrystallised from CH₂Cl₂/PE.

3-Acetamido-4,4-dimethoxy-5,6-epoxycyclohex-2-en-1-one (**28a**): White solid; yield: 65%; R_f 0.45 (EtOAc); mp 107–108 °C.

IR (CCl₄): v = 3415, 2956, 2927, 2854, 1722, 1680, 1498, 1223, 1120, 1064 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 7.66 (br s, 1 H, NH), 7.15 (d, 1 H, J = 2.0 Hz, H-2), 3.82 (d, 1 H, J = 4.3 Hz, H-5), 3.66 (s, 3 H, OMe), 3.53 (dd, 1 H, J = 4.3, 2.0 Hz, H-6), 3.30 (s, 3 H, OMe), 2.19 (s, 3 H, Me).

¹³C NMR (67.5 MHz, CD₃COCD₃): δ = 193.7, 170.9, 147.7, 108.3, 96.3, 52.6, 52.3, 51.4, 50.4, 24.9.

MS (CI): m/z (%) = 245 (MNH₄⁺, 35), 228 (MH⁺, 100). HRMS (CI): Calc for C₁₀H₁₄NO₅: 228.0872. Found: 228.0873.

3-(5-Cyclohexylpenta-2E,4E-dienamido)-4,4-dimethoxy-5,6-epoxy-cyclohex-2-en-1-one (**28b**):

White solid; yield: 73%; $R_f 0.37$ (EtOAc/PE, 1:1); mp 135–136°C. IR (CH₂Cl₂): $\nu = 3406$, 3051, 2983, 2854, 1701, 1674, 1633, 1548, 1500, 1340, 1261, 1120 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 7.71 (s, 1 H, NH), 7.28 (dd, 1 H, J = 14.8, 9.7 Hz, H-3'), 7.22 (d, 1 H, J = 1.9 Hz, H-2), 6.19–6.06 (m, 2 H, H-4' and H-5'), 5.88 (d, 1 H, J = 14.8 Hz, H-2'), 3.82 (d, 1 H, J = 4.1 Hz, H-5), 3.64 (s, 3 H, OMe), 3.51 (dd, 1 H, J = 4.1, 1.9 Hz, H-6), 3.28 (s, 3 H, OMe), 2.09 (m, 1 H, cy-H), 1.70–1.03 (m, 10 H, cy). ¹³C NMR (67.5 MHz, CDCl₃): δ = 192.7, 164.9, 151.6, 145.2, 125.3, 120.7, 145.6, 108.6, 95.4, 52.0, 51.4, 51.2, 50.6, 41.1, 32.1, 25.9, 25.4.

MS (CI): m/z (%) = 348 (MH⁺, 96), 316 (100).

HRMS: Calc for C₁₉H₂₆NO₅: 348.1811. Found: 348.1812.

3-(7-Cyclohexylhepta-2E,4E,6E-trienamido)-4,4-dimethoxy-5,6-epoxycyclohex-2-en-1-one (28c):

White solid; yield: 67%; R_f 0.35 (EtOAc/PE, 1:1); mp 151–153 °C.

IR (CH₂Cl₂): $\nu = 3392$, 3051, 2987, 1694, 1668, 1606, 1550, 1498, 1265, 1157, 1120 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 7.81 (s, 1 H, NH), 7.36 (dd, 1 H, J = 14.9, 11.6 Hz, H-3'), 7.27 (d, 1 H, J = 2.0 Hz, H-2), 6.60 (dd, 1 H, J = 14.9, 10.6 Hz, H-5'), 6.24 (dd, 1 H, J = 11.6, 14.9 Hz, H-4'), 6.13 (dd, 1 H, J = 10.6, 15.2 Hz, H-6'), 5.97 (d, 1 H, J = 14.9 Hz, H-2'), 5.95 (dd, 1 H, J = 15.2, 6.8 Hz, H-7'), 3.86 (d, 1 H, J = 4.0 Hz, H-5), 3.67 (s, 3 H, OMe), 3.54 (dd, 1 H, J = 4.0, 2.0 Hz, H-6), 3.32 (s, 3 H, OMe), 2.10–2.05 (m, 1 H, cy-H), 1.77–1.06 (m, 10 H, cy).

¹³C NMR (67.5 MHz, CDCl₃): δ = 192.9, 165.0, 147.1, 144.7, 143.0, 127.3, 127.3, 121.3, 145.8, 108.6, 95.4, 52.1, 51.5, 51.3, 50,61, 41.1, 32.4, 26.0, 25.8.

MS (CI): m/z (%) = 374 (MH⁺, 53), 342 (100).

HRMS (CI): Calc for C₂₁H₂₈NO₅: 374.1968. Found: 374.1964.

Preparation of SecondaryAlcohols (29); General Procedure:

To a stirred solution of amide 28 in MeOH was added NaBH₄ (1 equiv.) at 0° C under N₂. After stirring for ca. 1 h at 0° C, the reaction was warmed to r.t. at which point H2O was added followed by sufficient sat. NH₄Cl to give a neutral solution. The MeOH was evaporated under reduced pressure and the residue extracted with EtOAc. The organic layer was dried (Na₂SO₄) and the solvent evaporated under reduced pressure to give a quantative yield of product as a mixture of the two diastereomers. The crude material was dissolved in acetone/H₂O (8:1) and *p*-toluenesulphonic acid (0.15 equiv.) and pyridinium *p*-toluenesulphonate (0.9 equiv.) added at 0°C under N₂. The mixture was stirred at r.t. until the reaction was complete (3.5-7 d). Volatiles were then removed under reduced pressure and H₂O added. Extraction with EtOAc, drying of the combined organic layers (Na₂SO₄) and removal of the solvent under reduced pressure gave the crude product. Column chromatography or preparative TLC (EtOAc/PE) gave the syn and anti-diastereomers as white solids.

2-(Acetamido)-5,6-epoxy-4-hydroxy-cyclohex-2-en-1-one (29a):

Anti-isomer [(\pm)-5-epi-LL-C10037 α] as colourless needles; yield: 12%; $R_{\rm f}$ 0.25 (EtOAc/DCM, 1:1); mp 148–149°C (from CH₂Cl₂/ Et₂O) (lit.¹⁹ mp 154°C dec.).

IR (CHCl₃): v = 3687, 3595, 3392, 3039, 1682, 1516, 1375, 1026 cm⁻¹.

¹H NMR (270 MHz, $CDCl_3$): $\delta = 7.70$ (br s, 1H, NH), 7.61 (dd, 1H, J = 5.3, 2.4 Hz, H-3), 4.90 (dddd, 1H, J = 7.0, 5.3, 1.3, 1.2 Hz, H-4), 3.85 (ddd, 1H, J = 3.5, 2.4, 1.3 Hz, H-5), 3.62 (dd, 1H, J = 3.5, 1.2 Hz, H-6), 2.51 (d, 1H, J = 7.0 Hz, OH), 2.14 (3H, Me).

¹³C NMR (67.5 MHz, CDCl₃): δ = 188.7, 169.6, 130.3, 122.9, 63.1, 57.3, 52.2, 24.7.

MS (CI): m/z (%) = 201 (MNH₄⁺, 47), 184 (MH⁺, 100).

HRMS (CI): Calc for C₈H₁₀NO₄: 184.0610. Found: 184.0614.

Syn-isomer [(±)-LL-C10037 α **17**] as a colourless powder; yield: 37%; $R_{\rm f}$ 0.20 (EtOAc/CH₂Cl₂, 1:1); mp 168–170 °C dec. (from acetone/Et₂O) (lit.¹⁹ mp 167 °C dec.).

IR (CHCl₃): v = 3687, 3597, 3394, 3033, 1684, 1516, 1373, 1032 cm⁻¹.

¹H NMR (270 MHz, $CDCl_3$): $\delta = 7.56$ (br s, 1H, NH), 7.44 (dd, 1H, J = 3.0, 2.2 Hz, H-3), 4.85 (ddd, 1H, J = 10.5, 3.2, 3.0 Hz, H-4), 3.88 (ddd, 1H, J = 3.9, 3.2, 2.2 Hz, H-5), 3.60 (d, 1H, J = 3.9 Hz, H-6), 2.25 (d, 1H, J = 10.5 Hz, OH), 2.14 (3H, Me).

¹³C NMR (67.5 MHz, CDCl₃): δ = 188.4, 169.2, 128.4, 124.5, 64.5, 54.1, 52.6, 24.6.

MS (CI): m/z (%) = 201 (MNH₄⁺, 100), 184 (MH⁺, 34).

HRMS (CI): Calc for C₈H₁₀NO₄: 184.0610. Found: 184.0611.

2-(5-Cyclohexylpenta-2E,4E-dienamido)-5,6-epoxy-4-hydroxycyclohex-2-en-1-one (**29b**):

Anti isomer as a white solid; yield: 19%; R_f 0.36 (EtOAc/PE, 1:2); mp 184–185 °C.

IR (CH₂Cl₂): $\nu = 3944$, 3691, 3062, 2989, 1676, 1632, 1550, 1514, 1422, 1265, 1155, 1024 cm⁻¹.

¹H NMR (500 MHz, CD₃COCD₃): δ = 8.32 (s, 1 H, NH), 7.66 (dd, 1 H, *J* = 4.4, 2.3 Hz, H-3), 7.20 (dd, 1 H, *J* = 14.9, 10.6 Hz, H-3,), 6.33 (d, 1 H, *J* = 14.9 Hz, H-2'), 6.21 (dd, 1 H, *J* = 10.6, 15.3 Hz, H-4'), 6.13 (dd, 1 H, *J* = 15.3, 6.8 Hz, H-5'), 4.80 (m, 1 H, H-4), 3.83 (ddd, 1 H, *J* = 3.7, 2.3, 0.9 Hz, H-5), 3.58 (dd, 1 H, *J* = 3.7, 0.9 Hz, H-6), 2.83 (br s, 1 H, OH), 2.10–2.04 (m, 1 H, cy-H) and 1.76–1.12 (m, 10 H, cy).

¹³C NMR (67.5 MHz, CD₃COCD₃): δ = 190.4, 166.1, 150.0, 143.7, 127.2, 125.1, 131.8, 123.8, 63.7, 58.7, 53.6, 42.1, 33.3, 26.9, 26.7. MS (CI): m/z (%) = 304 (MH⁺, 100).

HRMS (CI): Calc for C₁₇H₂₂NO₄: 304.1549. Found: 304.1553.

Syn isomer as a white solid; yield: 67%; mp 177–178°C; $R_{\rm f}$ 0.22 (EtOAc/PE, 1:2).

IR (CH₂Cl₂): v = 3328, 2924, 1652, 1548, 1371, 1252, 1108, 1066, 1005 cm⁻¹.

¹H NMR (500 MHz, CD₃COCD₃): δ = 8.23 (s, 1 H, NH), 7.49 (dd, 1 H, *J* = 2.8, 2.7 Hz, H-3), 7.19 (dd, 1 H, *J* = 14.9, 10.7 Hz, H-3'), 6.31 (d, 1 H, *J* = 14.9 Hz, H-2'), 6.20 (dd, 1 H, *J* = 15.3, 10.7 Hz, H-4'), 6.12 (dd, 1 H, *J* = 15.3, 6.8 Hz, H-5'), 4.93 (ddd, 1 H, *J* = 7.6, 2.9, 2.8 Hz, H-4), 3.85 (ddd, 1 H, *J* = 4.1, 2.9, 2.7 Hz, H-5), 3.51 (d, 1 H, *J* = 4.1 Hz, H-6), 2.81 (m, 1 H, OH), 2.14–2.06 (m, 1 H, cy-H), 1.76–1.11 (m, 10 H, cy).

¹³C NMR (67.5 MHz, CD₃COCD₃): δ = 190.4, 165.8, 149.9, 143.4, 127.6, 127.2, 129.8, 123.8, 65.3, 55.1, 53.3, 42.1, 33.3, 26.9, 26.7.

MS (Cl): m/z (%) = 304 (MH⁺, 100).

HRMS (CI): Calc for $C_{17}H_{22}NO_4$: 304.1549. Found: 304.1553.

Anal: Calc C, 67.31; H, 6.98; N, 4.62. Found: 66.90; H, 6.84; N, 4.59.

2-(7-Cyclohexylhepta-2E,4E,6E-trienamido)-5,6-epoxy-4-hydroxy-cyclohex-2-en-1-one (**29c**):

Anti isomer as a white solid; yield: 16%; $R_{\rm f}$ 0.39 (EtOAc/PE, 1:2); mp 180–182 °C.

IR (CH₂Cl₂): v = 3944, 3693, 3062, 2985, 1673, 1606, 1423, 1273, 1155, 987 cm⁻¹.

¹H NMR (500 MHz, CD₃COCD₃): δ = 8.33 (s, 1 H, NH), 7.67 (dd, 1 H, *J* = 3.9, 2.4 Hz, H-3), 7.26 (dd, 1 H, *J* = 14.9, 11.3 Hz, H-3'), 6.63 (dd, 1 H, *J* = 14.9, 10.7 Hz, H-5'), 6.38 (d, 1 H, *J* = 14.9 Hz, H-2'), 6.32 (dd, 1 H, *J* = 14.9, 11.3 Hz, H-4'), 6.18 (dd, 1 H, *J* = 15.3, 10.7 Hz, H-6'), 5.92 (dd, *J* = 15.3, 7.1 Hz, H-7'), 4.80 (m, 1 H, H-4), 3.83 (br dd, 1 H, *J* = 3.7, 2.4 Hz, H-5), 3.58 (d, 1 H, *J* = 3.7 Hz, H-6), 2.80–2.77 (m, 1 H₂, OH), 2.11–2.06 (m, 1 H, cy-H) and 1.75–1.11 (m, 10 H, cy).

¹³C NMR (125 MHz, CD₃COCD₃): δ = 190.1, 165.2, 145.9, 142.8, 141.8, 129.3, 128.6, 124.4, 129.1, 124.1, 63.5, 58.4, 53.2, 41.8, 33.2, 26.7, 26.5.

MS (CI): m/z (%) = 330 (MH⁺, 100).

HRMS (CI): Calc for C₁₉H₂₄NO₄: 330.1705. Found: 330.1697.

Syn isomer as a white solid; yield: 64%; mp 175–176°C; $R_{\rm f}$ 0.22 (EtOAc/PE, 1:2).

IR (CH₂Cl₂): v = 3944, 3693, 3052, 2987, 1674, 1608, 1516, 1423, 1263, 1157, 987 cm⁻¹.

¹ H NMR (500 MHz, CD₃COCD₃): δ = 8.23 (s, 1 H, NH), 7.49 (dd, 1 H, *J* = 2.9, 2.7 Hz, H-3), 7.25 (dd, 1 H, *J* = 14.9, 11.4 Hz, H-3'), 6.63 (dd, 1 H, *J* = 14.9, 10.7 Hz, H-5'), 6.35 (d, 1 H, *J* = 14.9 Hz, H-2'), 6.32 (dd, 1 H, *J* = 14.9, 11.4 Hz, H-4'), 6.18 (dd, 1 H, *J* = 15.3, 10.7 Hz, H-6'), 5.91 (dd, 1 H, *J* = 15.3, 7.0 Hz, H-7'), 4.92 (ddd, 1 H, *J* = 7.5, 3.0, 2.9 Hz, H-4), 3.85 (ddd, 1 H, *J* = 4.1, 3.0, 2.7 Hz, H-5), 3.50 (d, 1 H, *J* = 4.1 Hz, H-6), 2.79 (m, 1 H, OH), 2.07 (m, 1 H, cy-H), 1.75–1.11 (m, 10 H, cy).

¹³C NMR (67.5 MHz, CD₃ COCD₃): δ = 190.2, 165.7, 145.9, 142.8, 141.7, 129.26, 128.7, 127.4, 129.6, 124.3, 65.2, 54.9, 53.2, 41.9, 33.3, 26.8, 26.6.

MS (CI): m/z (%) = 330 (MH⁺, 100).

HRMS (CI): Calc for $C_{19}H_{24}NO_4$: 330.1705. Found: 330.1697.

Oxidation of Secondary Alcohols 29; General Procedure:

To a stirred solution of alcohol **29** (1 equiv.) in CH_2Cl_2 under N_2 at 0°C was added PDC (1.5–2.0 equiv.) in one portion. After stirring at r.t. overnight (athough the reactions were often finished earlier), the inorganics were removed by filtration and the solvent evaporated under reduced pressure. The resulting brown oil was purified by column chromatography (EtOAc/PE).

2-Acetamido-5,6-epoxy-1,4-benzoquinone (30a):

As a pale yellow solid from **29a** (*syn*); yield: 87%; R_f 0.60 (EtOAc/DCM, 1:1); mp (from EtOAc/PE) 169–170 °C (dec.).

IR (CCl₄): v = 3386, 1728, 1699, 1685, 1610, 1502, 1315, 1213, 758 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 7.88 (br s, 1 H, NH), 7.52 (d, 1 H, J = 2.3 Hz, H-3), 3.92 (d, 1 H, J = 3.6 Hz, H-6), 3.84 (dd, 1 H, J = 3.6, 2.3 Hz, H-5), 2.23 (s, 3 H, Me).

¹³C NMR (67.5 MHz, CDCl₃): δ = 191.1, 188.0, 169.3, 138.5, 115.4, 53.8, 52.4, 24.9.

MS (CI): m/z (%) = 199 (MNH₄⁺, 18), 182 (MH⁺, 80).

HRMS (CI): Calc for C₈H₈NO₄: 182.0453. Found: 182.0456.

The ¹H NMR data was entirely consistent with published^{14b} values. See also preparation by two other procedures described later.

2-(7-Cyclohexylhept-2E,4E,6E-trienamido)-5,6-epoxy-1,4-benzoquinone (**30c**):

As a dark yellow oil from **29c** (*anti*); yield: 48%; $R_f 0.72$ (EtOAc/PE, 1:1).

IR (CHCl₃): v = 3373, 3028, 2929, 2854, 1693, 1678, 1606, 1504, 1317, 1261, 1174, 1117, 1088, 1012, 723 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 7.85 (br s, 1 H, NH), 7.63 (d, 1 H, J = 2.4 Hz, H-3), 7.39 (dd, 1 H, J = 14.9, 11.4 Hz, H-3'), 6.63 (dd, 1 H, J = 15.0, 10.3 Hz, H-5'), 6.31–6.09 (m, 2 H, H-4' and H-6'), 5.95 (d, 1 H, J = 14.9, H-2'), 6.01–5.94 (m, 1 H, H-7'), 3.93 (d, 1 H, J = 3.6 Hz, H-6), 3.85 (dd, 1 H, J = 2.4, 3.6 Hz, H-5), 1.80–1.05 (m, 11 H, cy-H and cy). ¹³C NMR (67.5 MHz, CDCl₃): δ = 191.0, 188.2, 164.9, 147.6, 145.6, 143.7, 130.9, 128.8, 127.2, 120.4, 115.2, 53.9, 52.5, 41.1, 32.4, 260.

MS (EI): m/z (%) =327 (M⁺, 13). **MS** (E): m/z (%) =327 (M⁺, 13). **MS** (E): m/z (%) =327 (M⁺, 13).

HRMS (EI): Calc for $C_{19}H_{21}NO_4$: 327.1471. Found: 327.1480. The NMR data were consistent with published^{5a} values. See also preparation described later.

2-tert-Butoxycarbonylamino-5,6-epoxy-1,4-benzoquinone (38):

Following the general procedure for the preparation of secondary alcohols and then the general procedure for the oxidation of secondary alcohols, oxo acetal **36** gave dione **38** which was purified by chromatography (EtOAc/CH₂Cl₂, 1:9) to give **38** as a light yellow oil; yield: 55% from **36**; R_f 0.72 (EtOAc/PE, 1:1).

IR (CCl₄): v = 3390, 2983, 2935, 1747, 1703, 1680, 1612, 1502, 1317, 1236, 1215, 1151, 997 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 7.26 (br s, 1 H, NH), 7.16 (d, 1 H, J = 2.3 Hz, H-3), 3.89 (d, 1 H, J = 3.6 Hz, H-6), 3.81 (dd, 1 H, J = 3.6, 2.3 Hz, H-5), 1.50 (s, 9H, ¹Bu).

¹³C NMR (67.5 MHz, CDCl₃): δ = 190.6, 187.7, 150.9, 139.8, 113.1, 83.1, 53.8, 52.5, 28.0.

MS (Cl): m/z (%) = 257 (MNH₄⁺, 100), 240 (MH⁺, 57).

HRMS (CI): Calc for C₁₁H₁₄NO₅: 240.0872. Found: 240.0872.

2-Amino-5,6-epoxy-1,4-benzoquinone (39):

Urethane **38** (221 mg, 0.92 mmol) was dissolved in anhyd CH₂Cl₂ (10 mL) under N₂ at 0 °C and powdered, activated 4Å molecular sieves (800 mg) were added followed by dropwise addition of a solution of BF₃·Et₂O (0.17 mL, 1.1 mmol), the temperature was allowed to rise slowly to r.t. and the mixture was stirred at r.t. for 20 h. Direct purification by chromatography (EtOAc/PE, 1:1 to pure EtOAc) afforded a yellow powder; yield: 126 mg (98%); $R_{\rm f}$ 0.30 (EtOAc/PE, 1:1) mp 133–134 °C (dec.).

IR (DCM): v = 3685, 3504, 3392, 1710, 1655, 1620, 1404, 1344, 1010, 839 cm⁻¹.

¹ H NMR (270 MHz, $CDCl_3$): $\delta = 5.56$ (d, 1 H, J = 2.3 Hz, H-3), 4.89 (br s, 2 H, NH₂), 3.75 (d, 1 H, J = 3.6 Hz, H-6), 3.65 (dd, 1 H, J = 3.6, 2.3 Hz, H-5).

¹³C NMR (67.5 MHz, CDCl₃): δ = 189.6, 189.5, 147.1, 103.3, 54.3, 52.6.

MS (CI): m/z (%) = 157 (MNH₄⁺, 48), 140 (MH⁺, 100).

HRMS (Cl): Calc for C₆H₉N₂O₃: 157.0613. Found: 157.0619.

Preparation of Amides 30 by Acylation of 39; General Procedure:

Amine **39** (1 equiv.) was dissolved in anhyd CH_2Cl_2 and stirred under N_2 at 0 °C and BSTFA (5 equiv.) was added. Then, a solution

of the acid chloride (1.5 equiv.) in CH₂Cl₂ was added over 10 min. The resulting mixture was stirred at r.t. for up to 4 d until reaction was complete. Volatile material was removed under reduced pressure and the residue was purified by chromatography.

2-Acetamido-5,6-epoxy-1,4-benzoquinone (30a): Yield: 65% (+ 25% recovered 39): data as before.

2-Lauramido-5,6-epoxy-1,4-benzoquinone (30d):

As yellow needles; yield: 99%; Rf 0.40 (EtOAc/PE, 1:4); mp 108 °C (from CH₂Cl₂/PE).

IR (CCl₄): v = 3384, 2927, 2856, 1699, 1682, 1610, 1500, 1315, 1213, 1174 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 7.83 (br s, 1 H, NH), 7.55 (d, 1 H, J = 2.4 Hz, H-3), 3.92 (d, 1 H, J = 3.6 Hz, H-6), 3.84 (dd, 1 H, J = 3.6, 2.4 Hz, H-5), 2.44–2.32 (m, 2 H), 1.65–1.59 (m, 2 H), 1.26–1.18 (m, 16 H), 0.91–0.82 (m, 3 H).

¹³C NMR (67.5 MHz, CDCl₃): δ = 191.1, 188.2, 172.5, 138.5, 115.3, 53.8, 52.4, 37.9, 33.7, 31.9, 29.5, 29.4, 29.3, 29.2, 29.0, 24.9, 22.7, 14.1. MS (EI): m/z (%) = 321 (M⁺, 4), 43 (100).

HRMS (EI): Calc for C₁₈H₂₇NO₄: 321.1940. Found: 321.1939.

2-Benzamido-5,6-epoxy-1,4-benzoquinone (30e):

As yellow crystals; yield: 65%; Rf 0.25 (EtOAc/PE, 1:4); mp 111-112°C (from CH₂Cl₂/PE);

IR (CCl₄): v = 3390, 1694, 1682, 1601, 1510, 1490, 1350, 1311, 1238, 1215, 1172, 1063, 901, 881, 785, 704 cm^{-1}

¹H NMR (270 MHz, CDCl₃): $\delta = 8.66$ (br s, 1 H, NH), 7.86–7.83 (m, 2 H, ArH), 7.69 (d, 1 H, J = 2.3 Hz, H-3), 7.65–7.49 (m, 3 H, ArH), 3.98 (d, 1 H, J = 3.6 Hz, H-6), 3.87 (dd, 1 H, J = 3.6, 2.3 Hz, H-5). ¹³C NMR (67.5 MHz, CDCl₃): δ = 191.0, 188.2, 165.8, 138.8,

133.2, 132.8, 129.1, 127.3, 115.5, 53.8, 52.5. MS (EI): m/z (%) = 243 (M⁺, 1), 105 (100).

HRMS (EI): Calc for C₁₃H₉NO₄: 243.0532. Found: 243.0538.

Preparation of Amides 30 by Epoxidation of Quinones 40; General Procedure:

(a) Preparation of Quinones 40:

To a solution of 2,5-dimethoxyaniline (34) (1 equiv.), pyridine (1.1 equiv.) and a catalytic amount (ca. 5 mg/mmol amine) of DMAP in CH_2Cl_2 was added the corresponding acid chloride 37 (1.1 equiv.) in CH₂Cl₂ at 0°C under N₂. The resulting solution was then stirred at r.t. for 12 h. The mixture was then poured into H₂O, and extracted with EtOAc. The organic layers were combined and washed with 2 M HCl and sat. aq NaHCO₃, and dried (Na₂SO₄). After the removal of the solvent, the dark residue was purified by column chromatography to give the corresponding amide which was used directly in the next step.

A solution of the amide (1 equiv.) in anhyd MeOH was stirred at 0°C under N₂ and treated portionwise with iodobenzene diacetate (1.5 equiv.). The reaction was then stirred at 0 °C for 4 h. If starting material was still remaining (TLC), the reaction was warmed to r.t. for 1 h. On completion, the mixture was diluted with CH2Cl2 and H2O and then neutralized with aq NaHCO3. The reaction was extracted with CH₂Cl₂ and the organic layers combined, washed with H₂O and brine, and then treated with Amberlyst-15 (ca. 3.5 g/mmol) and several drops of H₂O. In the case of 40a the hydrolysis was carried out at r.t. for about 12 h, and in the case of **40b,f** the reaction was run at 35-40 °C for about 4 h and monitored by TLC. The Amberlyst-15 was then filtered off, the solvent evaporated and the residue was washed with a small amount of Et₂O to give the quinones 40 as dark yellow solids after filtration. The filtrates were concentrated and subjected to column chromatography to give additional product.

2-Acetamido-/,4-benzoquinone (40a):

As a dark solid; yield: 66%; mp 145-147°C (Lit:⁴⁷ 146-148°C with data in accord with the literature.47

2-(5-Cyclohexylpenta-2E,4E-dienamido)-1,4-benzoquinone (40b): As an orange solid; yield: 70%; R_f 0.33 (EtOAc/PE, 1:3); mp 164-165°C

IR (CHCl₃): v = 3369, 2929, 2854, 1666, 1631, 1595, 1500, 1342, 1321, 1178, 1115, 1001, 893 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 8.03 (s, 1 H, NH), 7.67 (d, 1 H, J = 2.3 Hz, H-3), 7.34 (dd, 1 H, J = 14.8, 10.2 Hz, H-3'), 6.81–6.72 (m, 2 H, H-4' and H-5'), 6.24–6.16 (m, 2 H, H-5 and H-6), 5.95 (d, 1 H, J =

14.8 Hz, H-2'), 2.20-2.02 (m, 1 H, cy-H), 1.78-1.12 (m, 10 H, cy). ¹³C NMR (67.5 MHz, CDCl₃): δ = 187.9, 182.8, 164.9, 152.1, 145.7,

125.4, 120.3, 138.5, 138.3, 133.1, 114.5, 41.2, 32.1, 25.9, 25.8.

MS (EI): m/z (%) = 285 (M⁺, 21), 202 (27), 163 (78).

HRMS (EI): Calc. for C₁₇H₁₉NO₃: 285.1365. Found: 285.1373.

The NMR data were consistent with published^{12c} values.

2-(E-Cinnamido)-1,4-benzoquinone (40f):

As a dark solid; yield: 65%, R_f 0.30 (PE/EtOAc, 2:1); mp 204-205.5 °C (dec.).

IR (nujol): v = 3326, 1693, 1663, 1645, 1625, 1597, 1500, 1345, 1140, 883, 721 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 8.19 (br s, 1 H, NH), 7.77 (d, 1 H, J = 15.5 Hz, H-3'), 7.73 (d, 1 H, J = 2.2 Hz, H-3), 7.57 (m, 2 H, ArH), 7.45-7.40 (m, 3 H, ArH), 6.84-6.74 (m, 2 H, H-5,6), 6.59 (d, 1 H, J = 15.5 Hz, H-2').

¹³C NMR (67.5 MHz, CDCl₃): δ = 187.8, 182.7, 164.5, 144.9, 138.5, 138.3, 133.9, 133.1, 130.8, 129.0, 128.3, 119.2, 114.9.

MS (EI): m/z (%) = 253 (M⁺, 8), 131 (100).

HRMS (EI): Calc. for C₁₅H₁₁NO₃: 253.0739. Found: 253.0745.

(b) Epoxidation of Quinones 40:

To a solution of quinone 40 (0.2 mmol) in EtOAc (40 mL) in a 100 mL separating funnel was added tert-butyl hydroperoxide in nonane (5-6 M, 5 equiv.) containing DBU (1.0 equiv.). The mixture was shaken for 30 sec, washed with sat. aq iron(II) sulphate $(3 \times 30 \text{ mL})$, then brine $(2 \times 30 \text{ mL})$, and then dried (Na_2SO_4) . After evaporation of the solvent, the residue was purified by column chromatography (EtOAc/PE, 1:4) to give epoxyquinones 30:

2-Acetamido-5,6-epoxy-1,4-benzoquinone (30a): Yield: 67%: data as before.

2-(5-Cyclohexylpenta-2E,4E-dienamido)-5, 6-epoxy-1,4-benzoquinone (30b):

As an off-white solid; yield: 81%; Rf 0.55 (PE/EtOAc, 1:1); mp 159-160°C.

IR (CCl₄): v = 3381, 2929, 2854, 1695, 1680, 1633, 1605, 1502, 1350, 1313, 1213, 1172, 1115, 999 cm⁻¹.

¹H NMR (270 MHz, CDCl₃): δ = 7.90 (br s, 1 H, NH), 7.61 (d, 1 H, J = 2.3 Hz, H-3), 7.36–7.17 (m, 1 H, vinyl), 6.23–6.09 (m, 2 H, vinyl), 5.93 (d, 1 H, J = 15.8 Hz, H-2'), 3.93 (d, 1 H, J = 3.6 Hz, H-6), 3.83 (dd, 1 H, J = 2.3, 3.6 Hz, H-5), 2.17–2.00 (m, 1 H, cy-H), 1.77-1.68 (m, 5 H, cy), 1.32-1.10 (m, 5 H, cy).

¹³C NMR (67.5 MHz, CDCl₃): δ = 191.0, 188.1, 165.0, 152.4, 146.1, 139.0, 125.3, 120.0, 115.2, 53.8, 52.5, 41.2, 32.1, 25.9, 25.7. MS (EI): m/z (%)= 301 (M⁺, 10), 163 (100).

HRMS (EI): Calc. for C₁₇H₁₉NO₄: 301.1314. Found: 301.1323.

2-(E-Cinnamido)-5,6-epoxy-1,4-benzoquinone (30f):

Yield: 68% as a white solid; $R_{\rm f}$ 0.27 (PE/EtOAc, 2:1); mp 183– 185°C (dec.).

IR (nujol): v = 3349, 1672, 1627, 1610, 1505, 1173, 1140, 868, 665 cm^{-1}

¹H NMR (270 MHz, CDCl₃): $\delta = 8.00$ (br s, 1 H, NH), 7.76 (d, 1 H, *J* = 15.5 Hz, H-3'), 7.67 (d, 1 H, *J* = 2.4 Hz, H-3), 7.59–7.52 (m, 2 H, ArH), 7.44–7.38 (m, 3 H, ArH), 6.54 (d, 1 H, J = 15.5 Hz, H-2'), 3.95 (d, 1 H, J = 3.6 Hz, H-6), 3.86 (dd, 1 H, J = 2.4, 3.6 Hz, H-5).

¹³C NMR (67.5 MHz, CDCl₃): δ = 191.0, 188.2, 164.5, 145.3, 138.9, 133.75, 130.9, 129.0, 128.3, 118.8, 115.6, 53.9, 52.5. MS (CI): *m*/*z* (%) = 287 (MNH₄⁺, 35), 270 (MH⁺, 100). HRMS (CI): Calc. for C₁₅H₁₂NO₄: 270.0766. Found: 270.0770.

3-(7-Cyclohexylhepta-2*E*,4*E*,6*E*-trienamido)-4,4-dimethoxy-5,6epoxy-1-phenylcyclohex-2-en-1-ol (41c); Typical Procedure:

To a stirred solution of PhLi (1.8 M in THF, 0.26 mL, 0.47 mmol) in THF (4 mL) at -40 °C under N₂ was added ketone **28c** (25 mg, 0.067 mmol) in THF (4 mL) dropwise over 3 h. After stirring at -40 °C for 30 min, the reaction was warmed to r.t., quenched with sat. NH₄Cl (50 mL) and extracted with EtOAc (5 × 40 mL). The combined organic layers were washed with H₂O (40 mL) and brine (40 mL), dried (Na₂SO₄) and the solvent evaporated under reduced pressure to give a yellow oil. Purification by column chromatography (EtOAc/PE, 1:1) gave **41c** as a colourless oil; yield: 26 mg (86%); *R*_f 0.40 (EtOAc/PE, 1:1).

IR (CHCl₃): v = 3691, 3606, 3408, 3032, 2931, 2856, 1699, 1682, 1603, 1512, 1450, 1371, 1246, 1122, 1063, 780 cm⁻¹.

¹H NMR (270 MHz, CD₃COCD₃): δ = 8.11 (s, 1 H, NH), 7.67–7.64 (m, 2 H, ArH), 7.37–7.23 (m, 3 H, ArH), 7.17 (dd, 1 H, *J* = 14.9, 11.3 Hz, H-3'), 6.79 (1 H, d, *J* = 2.3 Hz, H-2), 6.58 (dd, 1 H, *J* = 15.2, 10.8 Hz, H-6'), 6.26 (d, 1 H, *J* = 14.9 Hz, H-2'), 6.26 (dd, 1 H, *J* = 15.2, 11.3 Hz, H-4'), 6.15 (dd, 1 H, *J* = 15.2, 10.8 Hz, H-5'), 5.88 (dd, 1 H, *J* = 15.2, 7.3 Hz, H-7'), 3.68 (d, 1 H, *J* = 4.3 Hz, H-5), 3.49 (s, 3 H, OMe), 3.32 (dd, 1 H, *J* = 4.3, 2.3 Hz, H-6), 3.34 (s, 3 H, OMe), 2.08 (s, 1 H, OH), 1.80–1.07 (m, 11 H, cy).

¹³C NMR (125 MHz, CD₃COCD₃): δ = 165.0, 145.6, 142.0, 141.3, 129.4, 129.0, 128.7, 128.2, 127.2, 125.1, 96.3, 73.0, 53.2, 51.0, 50.6, 50.0, 41.9, 33.4, 26.7, 26.6.

MS (EI): m/z (%) = 451 (M⁺, 14).

HRMS (EI): Calc for C₂₇H₃₃NO₅: 451.2359. Found: 451.2359.

3-(5-Cyclohexylpenta-2E,4E-dienamido)-4,4-dimethoxy-5,6-epoxy-1-phenylcyclohex-2-en-1-ol (**41b**):

In a similar manner, ketone **28b** was converted into **41b**, which was fully characterised, in 60% yield.

3-[(*tert*-Butoxycarbonyl)amido]-4,4-dimethoxy-5,6-epoxy-1-phenylcyclohex-2-en-1-ol (42):

To a stirred solution of PhLi (1.8 M in THF, 0.29 mL, 0.53 mmol) in THF (3 mL) at -40 °C under N₂ was added ketone **36** (50 mg, 0.18 mmol) in THF (2 mL), dropwise over 3 h. After stirring at -40 °C for 1 h, the reaction was warmed slowly to r.t., quenched with sat. NH₄Cl (30 mL) and extracted with EtOAc (3 × 50 mL). The combined organic layers were washed with H₂O (40 mL) and brine (40 mL), dried (MgSO₄) and the solvent evaporated under reduced pressure to give a pale yellow oil. Purification by column chromatography (EtOAc/PE, 1:3) gave **42** as a pale yellow solid; yield: 59 mg (93%); *R*_f 0.49 (Et₂O/PE, 1:1); mp 217–219 °C.

IR (CHCl₃): $\nu = 3577$, 3425, 2981, 1725, 1504, 1450, 1369, 1346, 1242, 1159, 1124, 1063, 1041, 991, 947 cm⁻¹.

¹H NMR (270 MHz, $CDCl_3$): $\delta = 7.19-6.81$ (m, 5 H, ArH), 6.18 (s, 1 H, NH), 5.89 (br s, 1 H, H-2), 3.28 (d, 1 H, J = 4.3 Hz, H-5), 3.16 (s, 3 H, OMe), 3.08 (dd, 1 H, J = 4.3, 2.4 Hz, H-6), 2.95 (s, 3 H, OMe), 2.16 (s, 1 H, OH), 1.03 (s, 9 H, *t*-Bu).

¹³C NMR (67.5 MHz, CDCl₃): *δ* = 152.4, 141.7, 128.7, 128.4, 127.9, 127.7, 125.9, 113.8, 95.0, 72.9, 57.5, 50.9, 53.2, 49.9, 28.2.

MS (CI): m/z (%) = 381 (MNH₄⁺, 7), 363 (15), 346 (100).

HRMS (CI): Calc for C₁₉H₂₉N₂O₆: 381.20262. Found: 381.2026.

Methyl 7-(1'-Hydroxycyclohexyl)hepta-2*E*,4*E*,6*E*-trienoate (45a): A solution of stannane 43^{37} (50 mg, 0.12 mmol) in anhyd, degassed DMF (0.4 mL) followed by a solution of bromodiene $44a^{21,38}$ (21 mg, 0.11 mmol) in anhyd, degassed DMF (0.4 mL) were added to a solution of PdCl₂(MeCN)₂ (1.3 mg, 0.005 mmol) in anhyd, degassed DMF (0.2 mL) at r.t. under N₂. The reaction was stirred overnight at

r.t., then the solvent was removed under reduced pressure and the crude product directly purified by column chromatography (PE/EtOAc, 3:2) to give ester **45a** as a colourless oil; yield: 21.3 mg (82%); R_f 0.50 (PE/EtOAc, 1:1).

IR (neat): v = 3376, 2930, 2857, 1699, 1615, 1420, 1260, 1134, 1004 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.32 (dd, 1 H, *J* = 15.3, 11.3 Hz, H-3), 6.56 (dd, 1 H, *J* = 10.8, 15.0 Hz, H-5), 6.39 (dd, 1 H, *J* = 10.8, 15.0 Hz, H-6), 6.32 (dd, 1 H, *J* = 11.3, 15.0 Hz, H-4), 6.03 (d, 1 H, *J* = 15.3 Hz, H-2), 5.88 (d, 1 H, *J* = 15.0 Hz, H-7), 3.75 (s, 3 H, OMe), 1.75–1.25 (m, 10 H, cy).

¹³C NMR (125 MHz, CDCl₃): δ = 167.5, 146.0, 144.6, 140.6, 129.6, 126.7, 120.3, 71.7, 51.5, 37.8, 25.4, 21.9.

MS (EI): m/z (%) = 236 (M⁺, 65), 55 (100).

HRMS (EI): Calc for $C_{14}H_{20}O_3$: 236.1413. Found: 236.1410.

7-(1'-Hydroxycyclohexyl)hepta-2*E*,4*E*,6*E*-trienoic Acid (45b):

A solution of DIBAL-H (1.0 M in THF, 0.02 mL, 0.020 mmol) was added to a solution of PdCl₂(PPh₃)₂ (7.2 mg, 0.01 mmol) in THF (0.5 mL) at r.t. under N₂. After stirring for 10 min, a solution of stannane **43**³⁷ (85 mg, 0.21 mmol) in anhyd, degassed DMF (0.5 mL) followed by a solution of bromodiene **44b**^{21,38} (33.8 mg, 0.19 mmol) in anhyd, degassed DMF (0.5 mL) were added to the solution of the pre-reduced catalyst at r.t. The reaction was stirred overnight at r.t., then treated with acetone (0.2 mL) and 1 M aq KF (0.2 mL). After stirring for 1 h, the mixture was filtered, washed with a 0.1 M solution of KH₂PO₄ (3 mL), extracted with EtOAc (3 × 3 mL) and the combined organic layers were dried (MgSO₄) and the solvent was removed. The crude product was purified by column chromatography (PE/EtOAc, 1:1) to give the ester **45b** as a white solid; yield: 30.4 mg (72%); R_f 0.17 (PE/EtOAc, 1:1); mp 96–98 C°.

IR (film): v = 3406, 2933, 2856, 1685, 1615, 1419, 1268, 1134, 1006 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.39 (dd, 1 H, *J* = 15.0, 11.5 Hz, H-3), 6.60 (dd, 1 H, *J* = 15.0, 10.5 Hz, H-5), 6.40 (dd, 1 H, *J* = 15.0, 10.5 Hz, H-6), 6.35 (dd, 1 H, *J* = 15.0, 11.5 Hz, H-4), 6.06 (d, 1 H, *J* = 15.0 Hz, H-2), 5.88 (d, *J* = 15.0 Hz, 1 H, H-7), 1.80–1.25 (m, 10 H, cy). ¹³C NMR (125 MHz, CDCl₃): δ = 171.7, 146.6 (2C), 141.5, 129.3,

126.5, 119.6, 71.7, 37.6, 25.3, 21.8.

MS (CI): m/z (%) = 240 (MNH₄⁺, 10), 205 (100).

HRMS (CI): Calc for C₁₃H₂₂NO₃: 240.1600. Found: 240.1593.

N-(2"-Hydroxy-5"-oxocyclopent-1"-en-1"-yl)-7-(1'-hydroxycyclohexyl)hepta-2*E*,4*E*,6*E*-trienamide (47):

Bromodiene 33^{21} (25 mg, 0.09 mmol) and stannane 43^{37} (46 mg, 0.11 mmol) were coupled using the same procedure described later for alisamycin. Purification by column chromatography (CH₂Cl₂/MeOH, 19:1) gave 47 as a yellow solid; yield: 24.3 mg (83%); $R_{\rm f}$ 0.13 (CH₂Cl₂/MeOH, 19:1); mp 132–133 °C.

IR (CDCl₃): v = 3689, 3600, 3373, 3155, 2985, 2935, 1817, 1793, 1628, 1603, 1531, 1469, 1379, 1217, 1165, 1095 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 13.64$ (s, 1 H, enol-OH), 7.54 (s, 1 H, NH), 7.34 (dd, 1 H, J = 14.9, 11.4 Hz, H-3'), 6.61 (dd, 1 H, J = 14.5, 10.7 Hz, H-5'), 6.40 (dd, 1 H, J = 15.2, 10.7 Hz, H-6'), 6.32 (dd, 1 H, J = 14.5, 11.4 Hz, H-4'), 6.06 (d, 1 H, J = 15.2 Hz, H-7'), 5.99 (d, 1 H, J = 14.9 Hz, H-2'), 2.80–2.50 (m, 4 H, H-4" and H-5"), 1.69–1.22 (m, 10 H, cy).

¹³C NMR (67.5 MHz, CDCl₃): *δ* = 197.3, 173.8, 167.1, 146.8, 144.4, 141.7, 129.1, 126.6, 119.8, 115.0, 71.7, 32.1, 25.6, 25.3, 21.9.

MS (EI): m/z (%) = 317 (M⁺, 24), 299 (21), 188 (100).

HRMS (EI): Calc for C /zH₂₃NO₄: 317.1627. Found: 317.1629.

2-(5'-Cyclohexylpenta-2'*E*,4'*E*-dienamido)-4-(2"-*E*-tributylstannylethenyl)-4-hydroxy-5,6-epoxycyclohex-2-en-1-one (52) and Regioisomer (51):

BuLi (2.5 M in hexanes, 0.24 mL, 0.60 mmol) was added dropwise over 15 min to a stirred solution of *E*-1,2-bis(tributylstannyl)ethene²⁰

(483 mg, 0.80 mmol) in THF (6 mL) under N₂. The reaction was stirred at $-78 \,^{\circ}$ C for 30 min and then at 0 $^{\circ}$ C for 90 min. The stirred solution of **31** was then re-cooled to $-78 \,^{\circ}$ C and epoxyquinone **30b** (60 mg, 0.20 mmol) in THF (2 mL) was added dropwise, under N₂ over 30 min. After stirring at $-78 \,^{\circ}$ C for 1 h, the reaction was quenched at this temperature with sat. NH₄Cl (10 mL) and extracted with EtOAc (3 × 40 mL). The combined organic layers were washed with brine (25 mL), dried (MgSO₄) and the solvent evaporated under reduced pressure to give a yellow oil. Purification by column chromatography (EtOAc/PE, 3:7) gave **52** as a colourless oil; yield: 31 mg (26%); *R*_f 0.62 (EtOAc/PE, 3:7).

IR (film): $v = 3345, 2954, 2925, 2852, 1667, 1632, 1612, 1522, 1464, 1450, 1365, 1246, 997 cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 7.56 (br s, 1 H, NH), 7.40 (d, 1 H, J = 2.6 Hz, H-3), 7.23 (dd, 1 H, J = 10.0, 15.0 Hz, H-3'), 6.49 (d, 1 H, J = 19.3 Hz, H-7), 6.14–6.11 (m, 2 H, H-4' and H-5'), 5.98 (d, 1 H, J = 19.3 Hz, H-8), 5.85 (d, 1 H, J = 15.0 Hz, H-2'), 3.67–3.63 (m, 2 H, H-5 and H-6), 2.15–2.05 (m, 1 H, cy-H), 1.77–0.93 (m, 37 H, cy + 3 × Bu).

 $^{13}\mathrm{C}$ NMR (125 MHz, CDCl₃): δ = 189.1, 164.9, 150.4, 145.5, 143.8, 133.5, 128.0, 127.4, 125.5, 121.2, 73.0, 57.7, 52.9, 41.1, 32.2, 29.0, 27.8, 26.0, 25.8, 13.6, 9.6.

MS (CI): m/z (%) = 620 [MH⁺ (for ¹²⁰Sn), 7].

HRMS (CI): Calc for $C_{31}H_{50}NO_4^{116}Sn$: 616.2757. Found: 616.2767.

The regioisomeric adduct **51** was also obtained as a colourless oil; yield: 24 mg (20%); R_f 0.38 (EtOAc/PE, 1:4).

IR (NaCl): v = 3334, 2954, 2926, 2852, 1666, 1633, 1610, 1504, 1373, 1340, 1211, 998 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.37 (br s, 1 H, NH), 7.34–7.24 (m, 1 H, H-3'), 7.12 (d, 1 H, J = 2.0 Hz, H-2), 6.67 (d, 1 H, J = 19.5 Hz, H-7), 6.20–6.08 (m, 2 H, H-4' and H-5'), 5.86 (d, 1 H, J = 19.5 Hz, H-8), 5.80 (d, 1 H, J = 14.5 Hz, H-2'), 3.61–3.55 (m, 2 H, H-5 and H-6), 2.92 (br s, 1 H, OH), 2.17–2.05 (m, 1 H, cy-H, 1.80–1.05 (m, 28 H, cy + 9 × CH₂), 1.00–0.80 (m, 9 H, 3 × Me).

¹³C NMR (125 MHz, CDCl₃): δ = 193.0, 165.4, 151.6, 150.5, 145.2, 143.1, 135.0, 125.4, 120.6, 107.0, 74.2, 57.1, 54.4, 41.2, 32.2, 29.0, 27.1, 26.0, 25.8, 13.7, 9.7.

MS (Cl): m/z (%) = 620 [MH⁺(for ¹²⁰Sn), 0].

HRMS (CI): Calc for $C_{31}H_{50}NO_4^{-116}Sn$: 616.2757. Found: 616.2750.

(±)-Alisamycin (8) and Regioisomer (53):

A solution of DIBAL-H (1.0 M in THF, 0.051 mL, 0.051 mmol) was added to a solution of PdCl₂(PPh₃)₂ (18 mg, 0.026 mmol) in THF (4 mL) at r.t. under N₂. After stirring at r.t. for 5 min, 0.1 mL of this solution was removed and used immediately in the following coupling reaction. Solutions of vinylstannane **52** (10 mg, 0.016 mmol) in anhyd, degassed DMF (0.2 mL) and bromodiene **33**²¹ (4.9 mg, 0.018 mmol) in anhyd, degassed DMF (0.2 mL) were added in rapid succession to a vigorously stirred solution of the pre-generated catalyst (*ca.* 0.65×10^{-3} mmol) in THF (0.1 mL) at r.t. under N₂. After stirring at r.t. for 12 h, the mixture was poured into H₂O (10 mL) and extracted with EtOAc (3×15 mL). The combined organic layers were dried (Na₂SO₄) and the solvent evaporated under reduced pressure to give a yellow residue. Preparative TLC (CH₂Cl₂/EtOAc, 4:6) gave **8** as a yellow waxy solid; yield: 5.4 mg (64%); *R*_f 0.22 (CH₂Cl₂/MeOH, 95:5); mp 186–188 °C [lit.⁷ > 250 °C for (–)**8**].

IR (CHCl₃): v = 3311, 2960, 2925, 2852, 1666, 1599, 1514, 1369, 1259, 1095, 1012 cm⁻¹.

¹³C NMR (125 MHz, CDCl₃): δ = 197.1, 188.4, 173.5, 165.2, 165.0, 150.7, 144.1, 143.5, 139.5, 136.0, 131.6 (2 C), 128.1, 125.9, 125.4, 121.4, 121.0, 114.8, 71.2, 57.4, 53.0, 41.1, 32.2, 32.1, 26.0, 25.8, 25.6.

MS (FAB): m/z (%) = 543 (MNa⁺, 37), 521 (MH⁺, 100).

HRMS (FAB): Calc for $C_{29}H_{33}N_2O_7$: 521.2288. Found: 521.2294. The NMR data were consistent with published^{7,44} values.

Following the same procedure on the same scale, vinylstannane (**51**) gave the regioisomer (**53**) as a pale yellow solid; yield: 6.3 mg (75%); $R_f 0.39$ (CH₂Cl₂/MeOH, 9:1); mp = 233–236 °C (dec.). [P. (CHC) : x = 3257 - 2924 - 2850 - 1653 - 1616 - 1605 - 1497 - 1375

IR (CHCl₃): v = 3257, 2924, 2850, 1653, 1616, 1605, 1497, 1375, 1209, 1009 cm⁻¹.

¹H NMR (500 MHz, CDCl₃) δ = 13.50 (s, 1 H, enol-OH), 7.53 (br s, 1 H, NH), 7.42 (br s, 1 H, NH), 7.33 (dd, 1 H, *J* = 15.0, 11.5 Hz, H-11), 7.24 (dd, 1 H, *J* = 15.0, 10.0 Hz, H-3'), 7.12 (d, 1 H, *J* = 2.0 Hz, H-2), 6.64 (dd, 1 H, *J* = 14.5, 11.0 Hz, H-8), 6.57 (dd, 1 H, *J* = 14.5, 11.0 Hz, H-9), 6.46 (dd, 1 H, *J* = 14.5, 11.5 Hz, H-10), 6.20–6.12 (m, 2 H, H-4' and H-5'), 6.08 (d, 1 H, *J* = 15.0 Hz, H-2), 3.66 (d, 1 H, *J* = 4.5, Hz, H-7), 5.81 (d, 1 H, *J* = 15.0 Hz, H-2'), 3.66 (d, 1 H, *J* = 4.5, Hz, H-5), 3.59 (dd, 1 H, *J* = 4.5, 2.0 Hz, H-6), 3.27 (br s, 1 H, OH), 2.65–2.45 (m, 4 H, H-4" and H-5"), 2.18–2.04 (m, 1 H, cy-H), 1.85–1.05 (m, 10 H, cy).

¹³C NMR (125 MHz, CDCl₃): δ = 197.4, 192.6, 174.1, 165.4, 165.2, 152.0, 150.1, 145.5, 143.1, 138.5, 134.2, 132.7, 132.6, 125.4, 122.2, 120.5, 114.9, 106.9, 72.6, 56.9, 54.3, 41.2, 32.2, 26.0, 25.8, 25.6. MS (FAB⁺): m/z (%) = 543 (MNa⁺, 23), 521 (MH⁺, 32).

HRMS (FAB⁺): Calc for $C_{29}H_{33}N_2O_7$: 521.2288. Found: 521.2299.

Nisamycin Methyl Ester (54):

A solution of DIBAL-H (1.0 M in THF, 0.03 mL, 0.030 mmol) was added to a solution of $PdCl_2(PPh_3)_2$ (10.0 mg, 0.014 mmol) in THF (2.0 mL) at r.t. under N₂ and stirred for 10 min. To a portion (0.1 mL) of the pre-reduced catalyst at r.t. under N₂ was added a solution of stannane **52** (12 mg, 0.019 mmol) in anhyd, degassed DMF (0.2 mL) followed by a solution of bromodiene **44a** (4.5 mg, 0.023 mmol) in anhyd, degassed DMF (0.2 mL). The reaction was stirred overnight at r.t., diluted with EtOAc (5 mL), washed with H₂O (3 × 3 mL) and the organic layer dried (MgSO₄) and the solvent removed. The crude product was purified using preparative TLC (PE/EtOAc, 3:2) to give the ester **54** as a pale yellow oil; yield: 6.8 mg (82%); R_f 0.46 (PE/EtOAc, 3:2).

IR (film): v = 3363, 2924, 2852, 1697, 1670, 1618, 1520, 1361, 1259, 1134, 1003 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.56 (br s, 1 H, NH), 7.41 (d, 1 H, J = 2.5 Hz, H-3), 7.30 (dd, 1 H, J = 15.0, 11.0 Hz, H-11), 7.24 (dd, 1 H, J = 15.0, 10.5 Hz, H-3'), 6.62–6.50 (m, 2 H, H-8 and H-9), 6.41 (dd, 1 H, J = 15.0, 11.0 Hz, H- 10), 6.20–6.10 (m, 2 H, H-4' and H-5'), 5.94 (d, 1 H, J = 15.0 Hz, H-2'), 5.85 (d, 1 H, J = 15.0 Hz, H-7), 5.84 (d, 1 H, J = 15.0 Hz, H-12), 3.76 (s, 3 H, OMe), 3.71 (dd, 1 H, J = 4.0, 2.5 Hz, H-5), 3.66 (d, 1 H, J = 4.0 Hz, H-6), 2.80 (br s, 1 H, OH), 2.18–2.08 (m, 1 H, H-6'), 1.80–1.05 (m, 10 H, cy).

¹³C NMR (125 MHz, CDCl₃): δ = 188.6, 167.3, 165.1, 150.8, 144.1, 143.8, 138.5, 135.5, 132.2, 131.7, 128.0, 126.3, 125.4, 121.9, 120.9, 71.1, 57.4, 52.9, 51.6, 41.1, 32.2, 26.0, 25.8.

MS (FAB⁺): m/z (%) = 462 (MNa⁺, 28), 176 (100).

HRMS (FAB⁺): Calc for $C_{25}H_{29}NNaO_6$: 462.1893. Found: 462.1900.

(±)-Nisamycin (15):

A solution of DlBAL-H (1.0 M in THF, 0.028 mL, 0.028 mmol) was added to a solution of $PdCl_2(PPh_3)_2$ (10 mg, 0.014 mmol) in THF (1 mL) at r.t. under N₂ and the solution stirred for 10 min. To this solution of the pre-reduced catalyst at r.t. was added a solution of

stannane **52** (10 mg, 0.016 mmol) in anhyd, degassed DMF (0.2 mL) followed by a solution of bromodiene **44b** (3.5 mg, 0.019 mmol) in anhyd, degassed DMF (0.2 mL). The reaction was stirred for 24 h at r.t., and then treated with acetone (0.2 mL) and aq KF (1 M, 0.2 mL). After stirring for a further 1 h, the mixture was filtered, washed with KH₂PO₄ (0.1 M, 1 mL), extracted with EtOAc (3 × 3 mL), the combined organic layers were dried (MgSO₄) and the solvent removed. The crude product was purified by preparative TLC (CH₂Cl₂/MeOH, 95:5) to give the acid **15** as a pale yeiiow solid; yield: 4.0 mg (58%); $R_{\rm f}$ 0.27 (CH₂Cl₂/MeOH, 9:1); mp 107–109 °C.

IR (CHCl₃): v = 3360, 2954, 2926, 2852, 1680, 1614, 1520, 1367, 1261, 1128, 1005 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.57 (br s, 1 H, NH), 7.41 (d, 1 H, J = 2.5 Hz, H-3), 7.36 (dd, 1 H, J = 15.0, 11.5 Hz, H-11), 7.24 (dd, 1 H, J = 15.0, 9.5 Hz, H-3'), 6.65–6.55 (m, 2 H, H-8 and H-9), 6.45 (dd, 1 H, J = 14.5, 11.5 Hz, H-10), 6.20–6.09 (m, 2 H, H-4' and H-5'), 5.94 (d, 1 H, J = 15.0 Hz, H-2'), 5.87 (d, 1 H, J = 14.5 Hz, H-7), 5.85 (d, 1 H, J = 15.0 Hz, H-12), 3.72 (dd, 1 H, J = 4.0, 2.5 Hz, H-5), 3.66 (d, 1 H, J = 4.0 Hz, H-6), 2.97 (br s, 1 H, OH), 2.17–2.08 (m, 1 H, H-6'), 1.90–1.05 (m, 10 H, cy).

¹³C NMR (125 MHz, CDCl₃): δ = 188.5, 170.6, 165.2, 150.9, 145.8, 144.2, 139.4, 136.1, 131.9, 131.5, 128.0, 126.3, 125.4, 121.1, 120.8, 71.2, 57.4, 52.9, 41.1, 32.2, 26.0, 25.8.

MS (FAB⁺): m/z (%) = 426 (MH⁺, 14), 93 (100).

HRMS (FAB⁺): Calc for $C_{24}H_{28}NO_6$: 426.1917. Found: 426.1920. The NMR data were consistent with published^{12a} values.

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