

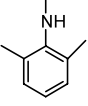

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Table 1. Enantioselective phase-transfer-catalysed alkylation of **3a–b** promoted by **1c**

Entry	R (Substrate)	Base ^a	Solvent	Temp. (°C)	Time (hrs.)	ee ^b (%)	Yield ^c (%)
1		50% aq. KOH	CH ₂ Cl ₂	0	24	13	4b , 58
2	3b	50% aq. KOH	Toluene: CHCl ₃ (7 : 3)	0	35	16	4b , 54
3	3b	CsOH.H ₂ O	CH ₂ Cl ₂	-50	28	20	4b , 52
4		CsOH.H ₂ O	CH ₂ Cl ₂	0	14	97	4a , 78
5	3a	CsOH.H ₂ O	CH ₂ Cl ₂	-50	23	99	4a , 83

^a12 equivalent.^bDetermined using chiral HPLC.^cYields of isolated product.

of steps to the final drug **2b**, glycine amide **3b** is a more stable synthon than the *tert*-butyl ester **3a**. Also, being a sterically hindered amide it can ward off unwanted *N*-alkylation. This was easily prepared¹⁶ from glycine and 2,6-dimethylaniline in 70% yield followed by benzophenone imine¹⁷ protection according to the reported procedure.¹⁸ When we subjected glycine amide **3b** to alkylation with 1-chloro-4-iodobutane, using CsOH.H₂O as a base with the catalyst **1c** at -50°C, we got the alkylated product **4b** in only 20% ee (entry 3, Table 1), as detected by chiral HPLC (Scheme 1). We repeated the reaction using various conditions, but the enantioselectivity of the alkylation did not improve significantly (entry 1–3, Table 1).

In order to ascertain the possible reasons for the low enantioselectivities, we repeated the reaction using benzyl bromide as the alkylating agent but using sterically more demanding groups at the amide nitrogen such as *iso*-propyl and *tert*-butyl groups, than the planer 2,6-dimethylbenzene. The enantioselectivity of the alkylation improved to give **6c** and **6d** in 42% and 53% ee, respectively (entries 2 and 3, Table 2). At this point we decided to see if *N,N*-disubstituted glycine amides would boost the stereoselectivity. Repeating the alkylation on *N,N*-disubstituted imine glycine amides **3e–h** under similar conditions as given in Scheme 2, certainly showed much higher enantioselectivity (ee 67–80%, entries 4–9, Table 2). It should be noted however, that *N,N*-dicyclo imine glycine amide did not improve the enantioselectivity (ee 75%, entry 9, Table 2) as compared to *N,N*-

diethyl imine glycine amide (ee 80%, entry 7, Table 2). This significant increase in enantioselectivity in the alkylation of *N,N*-disubstituted imine glycine amides prompted us to carry out modelling studies with various imine glycine amides and the catalysts.

Published X-ray data of *N*-anthracenylmethyl substituted quaternary ammonium salt of cinchona alkaloids³ show that they have a single accessible binding site for the enolates. Our modelling studies, performed using Sybyl 6.8¹⁹ along similar lines as done previously,²⁰ showed two possible ion-pair arrangements between the catalyst and the enolate. One ion-pair formed between the catalyst and enolate of imine glycine *tert*-butyl ester and that of the imine *N,N*-disubstituted glycine amides must be mainly existing in the most favorable arrangement as shown in Figure 1, which would give rise to the observed (*S*)-alkylated product, but when we consider the mono *N*-substituted glycine amide, like *N-tert*-butyl amide **3d**, there is a huge fall in stereoselectivity when compared to that of *O-tert*-butyl ester **3a** (53 and 95%, respectively) even though both carry the same sterically bulky *tert*-butyl group (entries 10 and 3, Table 2).

Our modelling studies show that there is a possibility of hydrogen bonding interaction between the amide NH of the enolate of **3d** and the quinoline nitrogen atom when the enolate is placed in the orientation as in Figure 2, which gives rise to the product having opposite stereochemistry. Though this orientation is less favourable due to the steric crowding, as the *tert*-butyl

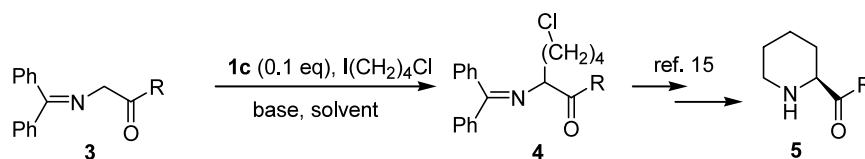
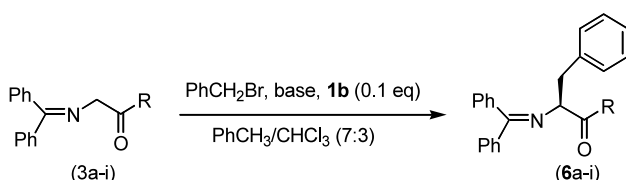
**Scheme 1.** Alkylation with 1-chloro-4-iodobutane.

Table 2. Enantioselective phase-transfer-catalysed benzylation of **3a–j** promoted by **1b**

Entry	R (Substrate)	Base ^a	Temp. (°C)	Time (hrs.)	ee ^b (%) (config.) ^c	Yield ^d (%)
1	3b	50% aq. KOH	0	8	16 (<i>S</i>)	6b , 65
2	3c	50% aq. KOH	0	10	42 (<i>S</i>)	6c , 80
3	3d	50% aq. KOH	0	10	53 (<i>S</i>)	6d , 84
4	3e	50% aq. KOH	0	25	67 (<i>S</i>)	6e , 82
5	3f	50% aq. KOH	0	24	66 (<i>S</i>)	6f , 76
6	3g	50% aq. KOH	0	28	75 (<i>S</i>)	6g , 78
7 ^a	3g	CsOH.H ₂ O	-20	4	80 (<i>S</i>)	6g , 84
8 ^a	3h	CsOH.H ₂ O	0	5	70 (<i>S</i>)	6h , 55
9 ^a	3h	CsOH.H ₂ O	-20	7	75 (<i>S</i>)	6h , 60
10	3a	50% aq. KOH	0	4	95 (<i>S</i>)	6a , 85

^aReaction done using toluene : CH₂Cl₂ (7:3) as solvent.^a12 equivalent.^bDetermined using chiral HPLC.^cDetermined after hydrolyzing the products and comparing the optical rotation with L-phenylalanine.^dYields of isolated product.**Scheme 2.** Alkylation with benzyl bromide.

group is placed between quinoline ring and vinyl substituent, and also due to increased charge separation (loosening of ion-pair), it still seems to form due to the added advantage of hydrogen bonding. This explains the observed fall in the stereoselectivity.

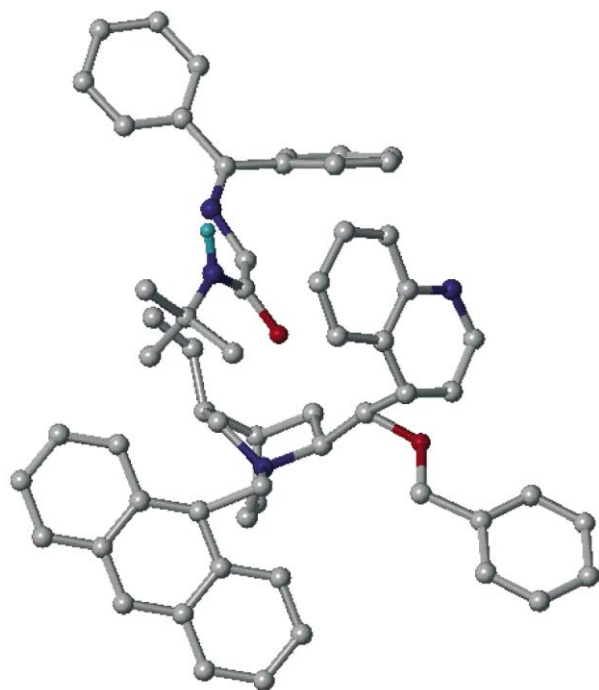
The above explanation also seems to fit in with the difference in the levels of diastereoselectivities observed by Marouka⁶ while benzylating the dipeptide Gly-L-Phe. High levels of diastereoselectivities were achieved using chiral spiro ammonium catalysts, where there was

no possibility of hydrogen bonding, whereas catalyst **1c** gave poor induction.

All the secondary amides showed low enantioselectivity with **3b** being the least. This is in agreement with the work done earlier on various esters by the O'Donnell's group,¹ where they observed that phenyl esters were sterically less demanding due to their planar structure than *tert*-butyl esters and hence showed worse asymmetric induction.

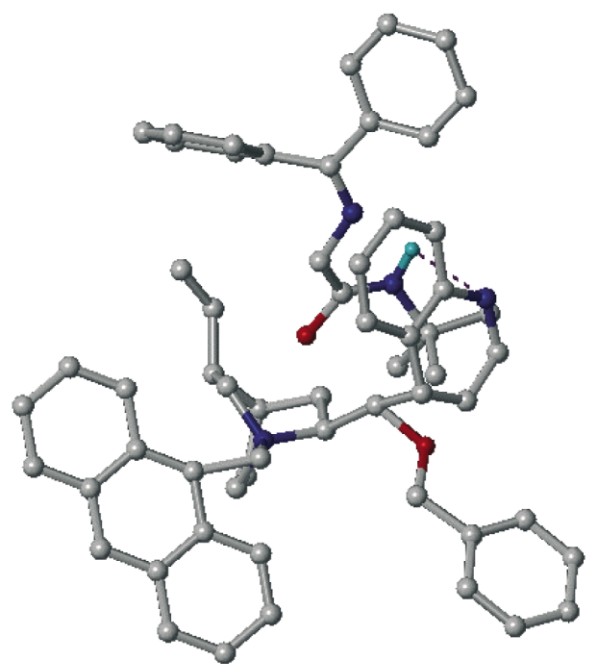
3. Conclusions

We have shown for the first time that achiral benzophenone imine glycinamides can also be used as suitable substrates for asymmetric alkylations using cinchona derived catalysts under PTC conditions. We have also tried to explain the poorer enantioselectivity of mono-substituted glycinamides as compared to *N,N*-disubstituted benzophenone imine glycinamides. On reduction, these compounds are able to yield useful chiral vicinal diamines; work in this direction is in progress.



● = Carbon, ● = Oxygen, ● = Nitrogen ● = Hydrogen.

Figure 1. Ion-pair between the catalyst¹⁹ and the (*E*)-enolate of **3d** in the conformation which will give rise to the (*S*)-product.



● = Carbon, ● = Oxygen, ● = Nitrogen ● = Hydrogen.

Figure 2. Ion-pair between the catalyst¹⁹ and the (*E*)-enolate of **3d** in the conformation, which will give rise to the (*R*)-product (Hydrogen bonding shown in dotted line)

4. Experimental

4.1. General

Melting points are uncorrected. ¹H and ¹³C spectra were recorded at 300 MHz Bruker Advance spectrometer with chemical shifts in ppm and tetramethylsilane as the internal standard. Infra-red absorption spectra were recorded on a Nicolet Impact 410 spectrometer, the frequencies in the IR spectra are indicated in cm⁻¹. Mass spectra data were recorded on a Finnigan-MAT LCMS spectrometer with electron spray ionization and atmospheric pressure ionization modes. Elemental analyses were recorded on a Elementa Vario EL. HPLC was performed on a Shimadzu SPD-10A using chiral phase column (DIACEL Chiralcel, OD-H), with the conditions outlined in the relevant experiment. TLC was performed on plates pre-coated (0.25 mm) with silica gel 60, Merck F-254. The plates were visualized by the use of a combination of UV (254 nm) and iodine. Column chromatography was carried out with silica gel Merck 60 (80–230 mesh). Molecular modelling studies were done using Sybyl 6.8 installed on an Octane 2 SG workstation.

4.2. General procedure for the preparation of benzophenone imine glycinamides

Benzophenone imine glycinamides were synthesized from the appropriate glycinamide hydrochloride¹⁶ and diphenylmethyl imine¹⁷ according to the reported procedure.¹⁸

4.2.1. *N*¹-(2,6-Dimethylphenyl)-*N*²-(diphenylmethylene)-glycinamide. Compound **3b** was obtained from *N*¹-(2,6-dimethylphenyl)-2-aminoacetamide hydrochloride¹⁶ in 82% yield; mp 118–119°C; ν_{max} (KBr) 3330, 3053, 2914, 2881, 1684, 1627, 1499, 1440, 1286, 776, 699; ¹H NMR δ (CDCl₃, 300 MHz) 2.30 (s, 6H), 4.16 (s, 2H), 7.11 (s, 3H), 7.20 (d, *J*=7.5 Hz, 2H), 7.34–7.52 (m, 6H), 7.67 (d, *J*=7.4 Hz, 2H), 8.94 (s, 1H); ¹³C NMR δ (CDCl₃, 300 MHz) 18.4, 56.6, 127.0, 127.0, 128.0, 128.1, 128.2, 128.8, 128.9, 130.7, 133.8, 135.0, 135.8, 138.4, 168.8, 170.2; MS (*m/z*): 344 (24%), 343 (100%). Anal. calcd for C₂₃H₂₂N₂O: C, 80.67; H, 6.48; N, 8.18 Found: C, 80.59; H, 6.50; N, 8.15.

4.2.2. *N*²-(Diphenylmethylene)-*N*¹-isopropylglycinamide. Compound **3c** was obtained from *N*¹-isopropyl-2-aminoacetamide hydrochloride¹⁶ in 87% yield; mp 88–89°C; ν_{max} (KBr) 3345, 3054, 2964, 2871, 1646, 1528, 1314, 1286, 1173, 779, 698; ¹H NMR δ (CDCl₃, 300 MHz) 1.25 (d, *J*=6.54 Hz, 6H), 3.93 (s, 2H), 4.17 (m, 1H), 7.13 (d, *J*=7.54 Hz, 2H), 7.2–7.4 (m, 5H), 7.62 (d, *J*=7.02 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 22.7, 40.7, 56.5, 127.0, 128.1, 128.2, 128.7, 128.8, 130.5, 135.9, 138.6, 169.6, 169.9; MS (*m/z*): 282 (22%), 281 (100%). Anal. calcd for C₁₈H₂₀N₂O: C, 77.11; H, 7.19; N, 9.99 Found: C, 76.98; H, 7.28; N, 10.03.

4.2.3. *N*¹-(*tert*-Butyl)-*N*²-(diphenylmethylene)-glycinamide. Compound **3d** was obtained from *N*¹-

(*tert*-butyl)-2-aminoacetamide hydrochloride¹⁶ in 85% yield; mp 108–109°C; ν_{\max} (KBr) 3374, 3080, 2966, 1672, 1623, 1526, 1449, 1286, 1223, 777, 699; ¹H NMR δ (CDCl₃, 300 MHz) 1.44 (s, 9H), 3.85 (s, 2H), 7.13 (d, J =7.39 Hz, 2H), 7.14–7.46 (m, 6H), 7.6 (d, J =7.64 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 28.7, 50.5, 56.9, 127.1, 128.1, 128.6, 128.8, 130.5, 135.9, 138.6, 169.4, 169.5; MS (m/z): 296 (20%), 295 (100%). Anal. calcd for C₂₆H₂₈N₂O: C, 81.21; H, 7.34; N, 7.29. Found: C, 80.83; H, 7.41; N, 7.24.

4.2.4. *N*-(Diphenylmethylene)-2-oxo-2-pyrrolidin-1-ylethanamine. Compound **3e** was obtained from 2-oxo-2-pyrrolidin-1-ylethylamine hydrochloride¹⁶ in 92% yield; mp 94–95°C; ν_{\max} (KBr): 2977, 2876, 1627, 1449, 1425, 1324, 793, 709; ¹H NMR δ (CDCl₃, 300 MHz) 1.79–1.97 (m, 4H), 3.48 (q, J =6.61 Hz, 4H), 4.18 (s, 2H), 7.23 (d, J =6.3 Hz, 2H), 7.29–7.46 (m, 6H), 7.64 (d, J =7.79 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 24.0, 26.0, 45.8, 46.4, 57.3, 127.7, 127.9, 128.4, 128.5, 130.1, 135.9, 139.3, 168.4, 170.8; MS (m/z): 294 (18%), 293 (100%). Anal. calcd for C₁₉H₂₀N₂O: C, 78.05; H, 6.89; N, 9.58. Found: C, 77.96; H, 6.95; N, 9.53.

4.2.5. *N*-(Diphenylmethylene)-2-oxo-2-piperidin-1-ylethanamine. Compound **3f** was obtained from 2-amino-1-piperidino-1-ethanone hydrochloride¹⁶ in 86% yield; mp 73–74°C; ν_{\max} (KBr) 3488, 2941, 2856, 1625, 1441, 1253, 1230, 1021, 707; ¹H NMR δ (CDCl₃, 300 MHz) 1.57–1.60 (m, 6H), 3.27–3.57 (m, 4H), 4.23 (s, 2H), 7.18–7.49 (m, 8H), 7.63 (d, J =8.05 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 24.4, 25.4, 26.1, 26.4, 42.9, 45.0, 46.8, 56.9, 127.7, 127.9, 128.1, 128.4, 128.5, 128.5, 129.9, 130.1, 132.3, 135.9, 139.3, 168.1, 170.2; MS (m/z): 294 (18%), 293 (100%). Anal. calcd for C₂₀H₂₂N₂O: C, 78.40; H, 7.24; N, 9.08. Found: C, 77.80; H, 7.32; N, 8.99.

4.2.6. *N*²-(Diphenylmethylene)-*N*¹,*N*¹-diethylglycinamide. Compound **3g** was obtained from *N,N*-diethyl-2-aminoacetamide hydrochloride¹⁶ in 90% yield; mp 104–105°C; ν_{\max} (KBr) 3054, 2975, 2927, 288, 1649, 1480, 1429, 1259, 1144, 773, 701; ¹H NMR δ (CDCl₃, 300 MHz) 1.26 (t, J =7.06 Hz, 6H), 3.37 (m, 4H), 4.23 (s, 2H), 7.20–7.23 (m, 2H), 7.26–7.47 (m, 6H), 7.64–7.66 (m, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 12.8, 14.1, 40.0, 41.7, 56.4, 127.6, 127.7, 128.3, 128.4, 129.9, 135.8, 139.2, 168.7, 170.4; MS (m/z): 296 (20%), 295 (100%). Anal. calcd for C₁₉H₂₂N₂O: C, 77.52; H, 7.53; N, 9.52. Found: C, 77.34; H, 7.58; N, 9.49.

4.2.7. *N*²-(Diphenylmethylene)-*N*¹,*N*¹-dicyclohexylglycinamide. Compound **3h** was obtained from *N,N*-dicyclohexyl-2-aminoacetamide hydrochloride¹⁶ in 80% yield; mp 116–117°C; ν_{\max} (KBr) 3052, 2928, 2850, 1624, 1443, 1313, 1182, 783, 700; ¹H NMR δ (CDCl₃, 300 MHz) 1.05–1.32 (m, 6H), 1.46–1.58 (m, 6H), 1.65–1.76 (m, 7H), 2.49 (bs, 1H), 2.91 (bs, 1H), 3.68 (m, 1H), 4.19 (s, 2H), 7.21 (dd, J =1.42 Hz, J =5.67 Hz, 2H), 7.30–7.48 (m, 6H), 7.64 (d, J =6.76 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 25.2, 25.8, 26.5, 29.7, 31.2, 55.9, 57.7, 59.1, 127.8, 127.8, 128.3, 128.5, 130.0, 135.9, 139.4, 168.8, 169.9; MS (m/z): 404 (24%), 403 (100%).

Anal. calcd for C₂₇H₃₄N₂O: C, 80.56; H, 8.51; N, 6.96. Found: C, 80.39; H, 8.64; N, 6.90.

4.3. General procedure for the alkylation of benzophenone imine glycinate and *tert*-butyl *N*-(diphenylmethylene)glycinate

Alkyl halide (8.4 mmol) was added to a mixture of the appropriate benzophenone imine glycinate **3b–h** or *tert*-butyl *N*-(diphenylmethylene)glycinate¹⁸ (**3a**, 1.6 mmol) and *N*-anthracenylmethyl cinchonidium bromide²¹ (0.16 mmol) in toluene/chloroform (7:3) 10 mL. The reaction mixture was cooled to the appropriate temp, base (16.94 mmol) was added and the resulting mixture stirred vigorously until the starting material had been consumed (4–28 h). The suspension was diluted with diethyl ether (35 mL), washed with water (2×10 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure to give crude products **4a–b**, **6a–h**. Purification of the residue by silica chromatography afforded the desired products in 58–85% yield. The enantioselectivity was determined by chiral HPLC analysis (DIACEL Chiralcel OD-H, hexane/*iso*-propanol, flow rate 0.5 mL, min^{−1}, 20°C, λ 254 nm). The absolute configuration was determined by comparison of the specific rotation of the hydrolyzed product(s) with L-phenylalanine.

4.3.1. *tert*-Butyl-*N*-(diphenylmethylene)-L-phenylalaninate. Compound **6a** was obtained from **3a** in 85% yield; pale yellow oil; ¹H NMR δ (CDCl₃, 300 MHz) 1.41 (s, 9H), 3.22–3.09 (m, 2H), 4.08 (dd, J =4.98 Hz, J =9.08 Hz, 1H), 6.53–7.62 (m, 15H); *R*_t HPLC (99.5:0.5, hexane/*iso*propanol, 8.1 min (*S*-isomer) 17.5 min (*R*-isomer)).

4.3.2. *tert*-Butyl-6-chloro-*N*-(diphenylmethylene)-L-norleucinate. Compound **4a** was obtained from **3a** in 83% yield; pale yellow oil; ¹H NMR δ (CDCl₃, 300 MHz) 1.30–1.44 (m, 11H), 1.71 (m, 2H), 1.80 (q, J =7.61 Hz, 2H), 3.49 (t, J =6.66 Hz, 2H), 3.92 (t, J =6.46 Hz, 1H), 7.16–7.65 (m, 9H), 7.80 (d, J =7.17 Hz, 1H); *R*_t HPLC (99.5:0.5, hexane/*iso*propanol, 12.0 min (*S*-isomer) 14.9 min (*R*-isomer)).

4.3.3. 6-Chloro-*N*¹-(2,6-dimethylphenyl)-*N*²-(diphenylmethylene)-L-norleucinamide. Compound **4b** was obtained from **3b** in 58% yield; colourless oil; $[\alpha]_D^{25}$ = −5.8 (*c* 0.5, CH₂Cl₂), ee 20%; ν_{\max} (film) 3347, 3062, 2926, 2249, 1715, 1494, 1315, 908, 733; ¹H NMR δ (CDCl₃, 300 MHz) 1.55–1.78 (m, 6H), 2.18 (s, 6H), 3.31 (m, 1H), 3.46 (t, J =6.43 Hz), 4.93 (s, 1H), 7.06–7.46 (m, 13H), 8.53 (s, 1H); ¹³C NMR δ (CDCl₃, 300 MHz) 18.5, 23.1, 32.2, 34.7, 44.6, 66.0, 127.5, 127.7, 128.1, 128.2, 128.5, 128.8, 128.9, 130.7, 133.7, 135.6, 136.7, 138.3, 169.8, 171.4; MS (m/z): 434 (24%), 433 (100%). Anal. calcd for C₂₇H₂₉ClN₂O: C, 74.90; H, 6.75; N, 6.47. Found: C, 74.78; H, 6.83; N, 6.43; The enantiomeric excess was determined after reducing the imine to amine; *R*_t HPLC (hexane/*iso*propanol, 15.2 min (*S*-isomer), 19.8 min (*R*-isomer)).

4.3.4. *N*-(2,6-Dimethylphenyl)-*N*-(diphenylmethylene)-*L*-phenylalaninamide. Compound **6b** was obtained from **3b** in 65% yield; Colourless oil; $[\alpha]_D^{25} = -4.3$ (*c* 1, CH₂Cl₂), ee 16%; ν_{\max} (film) 3340, 3057, 3025, 2920, 2853, 1686, 1621, 1492, 1283, 771, 697; ¹H NMR δ (CDCl₃, 300 MHz) 2.20 (s, 6H), 3.18 (dd, *J*=3.57 Hz, *J*=9.45 Hz, 1H), 3.33 (dd, *J*=2.98 Hz, *J*=10.07 Hz, 1H), 4.33 (dd, *J*=3.07 Hz, *J*=6.31 Hz, 1H), 6.46 (d, *J*=6.98 Hz, 2H), 7.03–7.44 (m, 14H), 7.63 (d, *J*=7.03 Hz, 2H), 8.26 (s, 1H); ¹³C NMR δ (CDCl₃, 300 MHz) 18.4, 41.6, 68.0, 126.3, 127.1, 127.2, 128.1, 128.3, 128.5, 130.2, 130.6, 133.7, 135.0, 135.3, 137.7, 138.8, 170.2, 170.9; MS (*m/z*): 434 (38%), 433 (100%). Anal. calcd for C₃₀H₂₈N₂O: C, 83.30; H, 6.52; N, 8.18. Found: C, 83.20; H, 6.56; N, 8.13. *R*_t HPLC (98:2 hexane/*iso*-propanol, 25.62 min (*S*-isomer) 53.44 min (*R*-isomer)).

4.3.5. *N*-Isopropyl-*N*-(diphenylmethylene)-*L*-phenylalaninamide. Compound **6c** was obtained from **3c** in 80% yield; mp 114–115°C; $[\alpha]_D^{25} = -41.4$ (*c* 1, CH₂Cl₂), ee 42%; ν_{\max} (KBr) 3380, 3058, 2970, 2919, 1674, 1625, 1502, 1444, 1283, 764, 697; ¹H NMR δ (CDCl₃, 300 MHz) 1.14 (dd, *J*=2.77 Hz, *J*=3.75 Hz, 6H), 2.99 (dd, *J*=4.20 Hz, *J*=8.88 Hz, 1H), 3.15 (dd, *J*=3.29 Hz, *J*=9.77 Hz, 1H), 4.11 (dd, *J*=4.68 Hz, *J*=5.46 Hz, 2H), 6.48 (d, *J*=6.82 Hz, 2H), 6.59 (d, *J*=7.81 Hz, 1H), 7.02–7.58 (m, 13H); ¹³C NMR δ (CDCl₃, 300 MHz) 22.3, 22.7, 22.8, 40.8, 41.5, 67.5, 126.2, 127.3, 128.0, 128.1, 128.3, 128.3, 128.5, 130.1, 130.4, 135.5, 137.8, 139.2, 169.6, 171.6; MS (*m/z*): 372 (26%), 371 (100%); Anal. calcd for C₂₅H₂₆N₂O: C, 81.05; H, 7.07; N, 7.56. Found: C, 80.95; H, 7.14; N, 7.49; *R*_t HPLC (95:5 hexane/*iso*propanol, 9.4 min (*S*-isomer), 13.9 min (*R*-isomer)).

4.3.6. *N*-(*tert*-Butyl)-*N*-(diphenylmethylene)-*L*-phenylalaninamide. Compound **6d** was obtained from **3d** in 84% yield; mp 124–125°C; $[\alpha]_D^{25} = -7.8$ (*c* 1, CH₂Cl₂), ee 53%; ν_{\max} (KBr) 3378, 3033, 2964, 2928, 2870, 1664, 1617, 1507, 1447, 1278, 1224, 755, 700; ¹H NMR δ (CDCl₃, 300 MHz) 1.35 (s, 9H), 2.99 (dd, *J*=4.22 Hz, *J*=8.83 Hz, 1H), 3.14 (dd, *J*=3.25 Hz, *J*=9.80 Hz, 1H), 4.03 (dd, *J*=3.32 Hz, *J*=5.42 Hz, 1H), 6.86 (d, *J*=6.86 Hz, 2H), 6.69 (s, 1H), 7.01–7.42 (m, 10H), 7.56 (d, *J*=6.87 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 28.7, 41.5, 50.6, 67.9, 126.1, 127.2, 127.9, 128.1, 128.2, 128.4, 130.1, 130.3, 135.4, 137.8, 139.1, 169.0, 171.6; MS (*m/z*): 386 (25%), 385 (100%). Anal. calcd for C₂₆H₂₈N₂O: C, 81.21; H, 7.34; N, 7.29. Found: C, 80.98; H, 7.48; N, 7.23. *R*_t HPLC (95:5 hexane/*iso*-propanol, 9.6 min (*S*-isomer), 20.1 min (*R*-isomer)).

4.3.7. (2*S*)-*N*-(Diphenylmethylene)-1-oxo-3-phenyl-1-pyrrolidin-1-ylpropan-2-amine. Compound **6e** was obtained from **3e** in 82% yield; mp 126–127°C; $[\alpha]_D^{25} = -55.9$ (*c* 1, CH₂Cl₂), ee 67%; ν_{\max} (KBr) 2974, 2869, 1637, 1437, 1286, 773, 701; ¹H NMR δ (CDCl₃, 300 MHz) 1.54–1.64 (m, 4H), 2.72 (t, *J*=5.72 Hz, 2H), 3.21 (dd, *J*=7.11 Hz, *J*=5.82 Hz, 1H), 3.29–3.44 (m, 3H), 4.33 (t, *J*=6.99 Hz, 1H), 6.87 (d, *J*=3.58 Hz, 2H), 7.07–7.38 (m, 11H), 7.65 (d, *J*=7.34 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 23.7, 25.9, 40.9, 45.7, 45.9, 67.2, 126.2, 127.3, 127.8, 128.0, 128.3, 128.3, 128.7, 129.6, 130.1, 137.0, 138.1, 139.1, 169.2, 170.3; MS

(*m/z*): 384 (20%), 383 (100%). Anal. calcd for C₂₆H₂₆N₂O: C, 81.64; H, 6.85; N, 7.32. Found: C, 81.48; H, 6.93; N, 7.29. *R*_t HPLC (85:15, hexane/*iso*-propanol, 19.0 min (*S*-isomer), 21.9 min (*R*-isomer)).

4.3.8. (2*S*)-*N*-(Diphenylmethylene)-1-oxo-3-phenyl-1-piperidin-1-ylpropan-2-amine. Compound **6f** was obtained from **3f** in 76% yield; mp 120–121°C; $[\alpha]_D^{25} = -48.7$ (*c* 1, CH₂Cl₂), ee 66%; ν_{\max} (KBr) 2925, 2858, 1624, 1496, 1443, 1219, 998, 694, 775; ¹H NMR δ (CDCl₃, 300 MHz) 1.10 (bs, 1H), 1.25 (bs, 1H), 1.50 (m, 4H), 3.09–3.18 (m, 2H), 3.28–3.34 (m, 2H), 3.51 (m, 2H), 4.51 (dd, *J*=1.44 Hz, *J*=6.17 Hz, 1H), 6.80 (m, 2H), 7.06–7.67 (m, 13H); ¹³C NMR δ (CDCl₃, 300 MHz) 24.5, 25.6, 26.2, 40.9, 41.9, 43.4, 46.2, 47.2, 50.4, 66.3, 126.3, 127.5, 127.7, 127.9, 128.2, 128.3, 128.4, 128.7, 129.4, 129.7, 130.1, 136.5, 138.3, 139.3, 169.0, 169.9; MS (*m/z*): 398 (10%), 397 (100%); Anal. calcd for C₂₇H₂₈N₂O: C, 81.78; H, 7.12; N, 7.06. Found: C, 81.69; H, 7.31; N, 6.99. *R*_t HPLC (90:10 hexane/*iso*-propanol, 12.3 min (*S*-isomer), 14.3 min (*R*-isomer)).

4.3.9. *N,N*-Diethyl-*N*-(diphenylmethylene)-*L*-phenylalaninamide. Compound **6g** was obtained from **3g** in 84% yield; mp 111–112°C; $[\alpha]_D^{25} = -51.8$ (*c* 1, CH₂Cl₂), ee 80%; ν_{\max} (KBr) 3058, 3022, 2970, 2930, 1648, 1611, 1446, 1260, 1078, 792, 701; ¹H NMR δ (CDCl₃, 300 MHz) 0.72 (t, *J*=7.04 Hz, 3H), 1.04 (t, *J*=7.04 Hz, 3H), 2.92–3.11 (m, 3H), 3.24–3.38 (m, 3H), 4.42 (t, *J*=6.72 Hz, 1H), 6.84 (d, *J*=4.29 Hz, 2H), 7.05–7.39 (m, 11H), 7.64 (d, *J*=6.91 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 12.8, 14.2, 40.5, 41.1, 66.0, 126.2, 127.4, 127.8, 128.1, 128.2, 128.3, 128.4, 128.7, 129.6, 130.1, 136.5, 138.2, 139.3, 168.8, 170.6; MS (*m/z*): 386 (20%), 385 (100%). Anal. calcd for C₂₆H₂₈N₂O: C, 81.21; H, 7.34; N, 7.29. Found: C, 81.12; 7.58; N, 7.24. *R*_t HPLC (95:5, hexane/*iso*propanol, 21.6 min (*S*-isomer), 27.5 min (*R*-isomer)).

4.3.10. *N,N*-Dicyclohexyl-*N*-(diphenylmethylene)-*L*-phenylalaninamide. Compound **6h** was obtained from **3h** in 60% yield; Pale yellow oil; $[\alpha]_D^{25} = -25.2$ (*c* 0.5, MeOH), ee 75%; ν_{\max} (film) 3048, 3022, 2930, 1625, 1446, 1319, 1260, 1098, 772, 701; ¹H NMR δ (CDCl₃, 300 MHz) 1.03–1.29 (m, 6H), 1.42–1.53 (m, 6H), 1.62–1.70 (m, 7H), 2.40 (bs, 1H), 2.91–3.0 (m, 2H), 3.24 (dd, *J*=3.25 Hz, *J*=9.87 Hz, 1H), 3.59 (m, 1H), 4.29 (t, *J*=6.92 Hz, 1H), 6.82 (d, *J*=3.55 Hz, 2H), 7.09–7.48 (m, 11H), 7.65 (d, *J*=6.76 Hz, 2H); ¹³C NMR δ (CDCl₃, 300 MHz) 25.3, 25.7, 26.8, 29.9, 31.2, 55.2, 59.3, 67.2, 126.2, 127.3, 127.8, 128.0, 128.3, 128.4, 128.7, 129.5, 130.1, 135.9, 138.7, 169.2, 170.2; MS (*m/z*): 495.3 (10%), 494 (25%), 493 (100%); Anal. calcd for C₃₄H₄₀N₂O: C, 82.88; H, 8.18; N, 5.69. Found: C, 82.74; H, 8.29; N, 5.63. *R*_t HPLC (98:2, hexane/*iso*-propanol, 8.1 min (*S*-isomer), 17.5 (*R*-isomer)).

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