



## Symmetrical approach of spiro-pyrazolidinediones as acetyl-CoA carboxylase inhibitors

Makoto Kamata\*, Tohru Yamashita, Asato Kina, Michiko Tawada, Satoshi Endo, Atsushi Mizukami, Masako Sasaki, Akiyoshi Tani, Yoshihide Nakano, Yuuki Watanabe, Naoki Furuyama, Miyuki Funami, Nobuyuki Amano, Kohji Fukatsu

Pharmaceutical Research Division, Takeda Pharmaceutical Company, Ltd, 2-26-1, Muraoka-higashi, Fujisawa, Kanagawa 251-8555, Japan

### ARTICLE INFO

#### Article history:

Received 10 April 2012

Revised 8 May 2012

Accepted 15 May 2012

Available online 19 May 2012

#### Keywords:

Acetyl-CoA carboxylase

ACC

Spiro-pyrazolidinedione

Spiro-pyrazolo[1,2-*a*]pyridazine

Obesity

Fatty acid oxidation

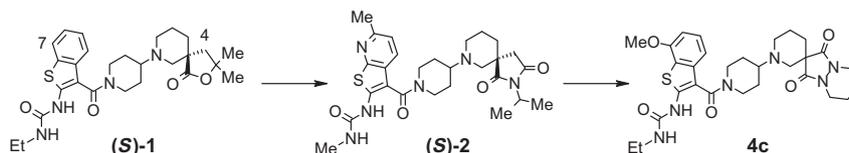
### ABSTRACT

Spiro-pyrazolidinedione derivatives without quaternary chiral center were discovered by structure-based drug design and characterized as potent acetyl-CoA carboxylase (ACC) inhibitors. The high metabolic stability of the spiro-pyrazolo[1,2-*a*]pyridazine scaffold and enhancement of the activity by incorporation of a 7-methoxy group on the benzothiophene core successfully led to the identification of compound **4c** as an orally bioavailable and highly potent ACC inhibitor. Oral administration of **4c** significantly decreased the values of the respiratory quotient in rats, indicating the stimulation of fatty acid oxidation.

© 2012 Elsevier Ltd. All rights reserved.

Acetyl-CoA carboxylase (ACC) catalyzes the biotin-dependent carboxylation of acetyl-CoA to form malonyl-CoA,<sup>1</sup> which is a known substrate for fatty acid synthase in de novo lipogenesis as well as an allosteric inhibitor of carnitine palmitoyltransferase 1 (CPT-1).<sup>2</sup> ACC inhibitors, which could reduce fatty acid synthesis and accelerate fatty acid oxidation by lowering malonyl-CoA levels,

between the log*D* value and metabolic stability.<sup>5</sup> This time we made an attempt to adopt a symmetrical approach for spiro structures as one of the solutions. Herein, we describe the synthesis and structure-activity relationship (SAR) of spiro-pyrazolidinediones derivatives and the excellent metabolic stability of the spiro-pyrazolo[1,2-*a*]pyridazine scaffold in human hepatic microsomes.

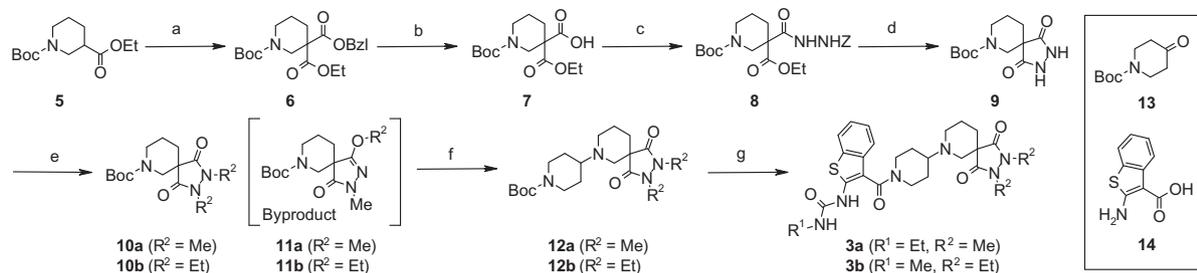


are widely known to be promising therapeutic targets for the treatment of obesity.<sup>3</sup> We have previously reported on spiro series as novel ACC inhibitors.<sup>4,5</sup> In particular, spiro-lactone (**S**)-**1** containing a ureidobenzothiophenyl group was designed by structure-based drug design, and its potent inhibitory activity has been demonstrated.<sup>4</sup> Moreover, we successfully identified highly potent and orally bioavailable inhibitor (**S**)-**2** through optimization of the benzothiophene and spiro-piperidine groups by using the relationship

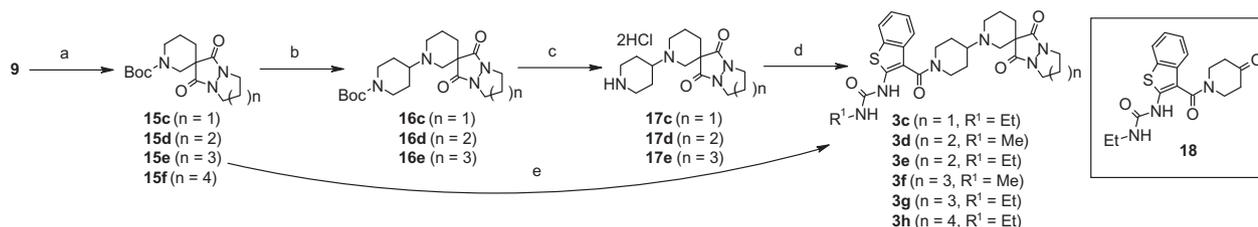
Spiro-pyrazolidinediones **3a**, **b** were synthesized following the route illustrated in Scheme 1. Carboxylation of ester **5** with benzyl chloroformate gave diester **6**. After the benzyl group of **6** was deprotected under reductive conditions, amide **8** was obtained by condensation with **7** and *Z*-hydrazine. Removal of the *Z* group in **8** by catalytic hydrogenation followed by cyclization under basic and heat conditions afforded spiro-pyrazolidinedione **9**. Dialkylation of **9** afforded *N,N*-dialkyl derivatives **10a**, **b** with moderate yield, which were easily separated from byproduct *N,O*-dialkyl compounds **11a**, **b** by using silica gel column chromatography. Deprotection of the Boc groups in **10a**, **b** followed by reductive

\* Corresponding author. Tel.: +81 466 32 1130; fax: +81 466 29 4451.

E-mail address: [Kamata\\_Makoto@takeda.co.jp](mailto:Kamata_Makoto@takeda.co.jp) (M. Kamata).



**Scheme 1.** Reagents and conditions: (a) 1.1 M LHMDS, ZCl, cyclopentylmethyl ether,  $-78^{\circ}\text{C}$ , quant.; (b)  $\text{H}_2/\text{Pd-C}$ , THF, 90%; (c)  $\text{ZNNH}_2$ , WSC, HOBT, DMF, quant.; (d) (i)  $\text{H}_2/\text{Pd-C}$ , THF, 90%; (ii) pyridine,  $100^{\circ}\text{C}$ , 91%; (e) MeI or EtI, NaH, DMF, 53% (**10a**), 50% (**10b**); (f) (i) 4 M HCl–EtOAc, 94% ( $\text{R}^2 = \text{Me}$ ), quant. ( $\text{R}^2 = \text{Et}$ ); (ii) **13**,  $\text{NaBH}(\text{OAc})_3$ , THF–AcOH, 30% (**12a**), 55% (**12b**); (g) (i) 4 M HCl–EtOAc, 92% ( $\text{R}^2 = \text{Me}$ ), quant. ( $\text{R}^2 = \text{Et}$ ); (ii) **14**, WSC, HOBT,  $\text{Et}_3\text{N}$ , DMF, 72% ( $\text{R}^2 = \text{Me}$ ), 85% ( $\text{R}^2 = \text{Et}$ ); (iii)  $\text{R}^1\text{NCO}$ , pyridine,  $80^{\circ}\text{C}$ , 77% (**3a**), 66% (**3b**).



**Scheme 2.** Reagents and conditions: (a)  $\text{Br}(\text{CH}_2)_n\text{Br}$ , NaH, DMF, 48% (**15c**), 70% (**15d**), 70% (**15e**), 14% (**15f**); (b) (i) 4 M HCl–EtOAc, 87% ( $n = 1$ ), 87% ( $n = 2$ ); (ii) **13**,  $\text{NaBH}(\text{OAc})_3$ , THF or  $\text{CH}_2\text{Cl}_2$ , 30% (**16a**), 55% (**16b**), 71% (2 steps, **16c**); (c) 4 M HCl–EtOAc, 92% ( $n = 1$ ), quant. ( $n = 2$ ), 90% ( $n = 3$ ); (d) (i) **14**, WSC, HOBT,  $\text{Et}_3\text{N}$ , DMF, 65% ( $n = 1$ ), 32% ( $n = 2$ ); (ii)  $\text{R}^1\text{NCO}$ , pyridine,  $70^{\circ}\text{C}$ , 71% (**3c**), 34% (**3d**), 83% (**3e**), 29% (2 steps, **3f**); (e) (i) 4 M HCl–EtOAc; (ii) **18**,  $\text{NaBH}(\text{OAc})_3$ ,  $\text{Et}_3\text{N}$ ,  $\text{CH}_2\text{Cl}_2$ , 51% (2 steps, **3g**), 37% (2 steps, **3h**).

alkylation with Boc-4-piperidone (**13**) provided compounds **12a, b**, which were converted to desired compounds **3a, b** by deprotection of the Boc groups, acylation with 2-amino-3-benzothiophenecarboxylic acid (**14**), and formation of an ureido group using alkyl isocyanate.

Other spiro-pyrazolidinediones (**3c–f**) were also easily synthesized (Scheme 2), in a manner similar to the one shown in Scheme 1. Only **3g, h** were directly prepared by reductive alkylation of deprotected **15e, f** with 1-ethyl-3-(3-[(4-oxopiperidin-1-yl)carbonyl]-1-benzothiophen-2-yl)urea (**18**).<sup>6</sup>

Synthesis of **4a–c** with 7-methoxybenzothiophenyl derivatives is shown in Scheme 3. Starting from commercially available thiophene **19**,  $\alpha$ -keto bromide **20** was prepared according to the known protocol.<sup>7</sup> Successive aromatization and alkylation of phenol **21** afforded **22**, which was hydrolyzed to give carboxylic acid **23**. Compound **23** was converted to **4a–c** by condensation with **17d** and subsequent formation of an ureido group.

The spiro compounds prepared in this study were evaluated for their inhibitory activities toward human ACC1/2 by using the malachite green method.<sup>8</sup> The results of ACC inhibition, human metabolic stability,<sup>9</sup> and lipophilicity ( $\log D$ ) are summarized in Tables 1–3. Because the 50% inhibitory concentration ( $\text{IC}_{50}$ ) ratios of ACC2 and ACC1 inhibitory activities were nearly the same, we primarily discuss the SAR for ACC2 inhibition in the present report.

As previously reported,<sup>4</sup> the co-crystal data of human ACC2 and (*S*)-**1** revealed that the carbonyl group of the spiro-lactone ring interacts with the backbone of Gly2162. In fact, the chiral resolu-

**Table 1**

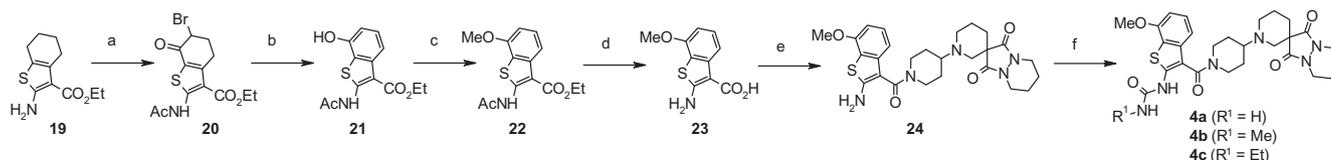
Acetyl-CoA carboxylase (ACC) inhibitory activity, human metabolic stability (hMS), and  $\log D$  values of spiro-pyrazolidinediones **3a–h**

	Substituents		$\text{IC}_{50}$ (nM)		hMS <sup>a</sup>	$\log D^b$
	$\text{R}^1$	$\text{R}^2$	ACC1	ACC2		
<b>3a</b>	Et	Me	120	20	12	2.03
<b>3b</b>	Me	Et	41	8.1	120	2.33
<b>3c</b>	Et	$-(\text{CH}_2)_3-$	170	33	25	1.93
<b>3d</b>	Me	$-(\text{CH}_2)_4-$	200	36	12	2.14
<b>3e</b>	Et	$-(\text{CH}_2)_4-$	130	23	18	2.45
<b>3f</b>	Me	$-(\text{CH}_2)_5-$	48	8.7	71	2.14
<b>3g</b>	Et	$-(\text{CH}_2)_5-$	43	7.5	100	2.48
<b>3h</b>	Et	$-(\text{CH}_2)_6-$	22	5.1	200	2.73

<sup>a</sup> In vitro metabolic clearance in human hepatic microsomes ( $\mu\text{L}/\text{min}/\text{mg}$  protein).

<sup>b</sup>  $\log D$  values were obtained using high-performance liquid chromatography (HPLC) at pH 7.  $\text{IC}_{50}$  = 50% inhibitory concentration.

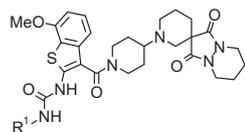
tion of **1** demonstrated that (*S*)-**1** ( $\text{ACC}_2$ :  $\text{IC}_{50} = 5.4$  nM) was more potent than (*R*)-**1** ( $\text{ACC}_2$ :  $\text{IC}_{50} = 1,600$  nM). Moreover, a wide space filled with water at the periphery of the 4-position (methylene) in spiro-lactone (*S*)-**1** was also observed (Fig. 1). The result suggests



**Scheme 3.** Reagents and conditions: (a) (i)  $\text{Ac}_2\text{O}$ , AcOH; (ii)  $\text{Ce}(\text{SO}_4)_2$ , AcOH,  $\text{CHCl}_3$ , 45%; (iii)  $\text{Br}_2$ ,  $\text{CHCl}_3$ , reflux, 89%; (b)  $\text{Li}_2\text{CO}_3$ , DMF,  $120^{\circ}\text{C}$ , 41%; (c) MeI,  $\text{K}_2\text{CO}_3$ , DMF, 59%; (d) 8 M NaOH, EtOH, reflux, 68%; (e) **17d**, WSC, HOBT,  $\text{Et}_3\text{N}$ , DMF, 81%; (f)  $\text{Cl}_3\text{CCONCO}$ , THF then 7 M  $\text{NH}_3$  in MeOH, 77% (**4a**) or  $\text{R}^1\text{NCO}$ , pyridine,  $70^{\circ}\text{C}$ , 85% (**4b**), 51% (**4c**).

**Table 2**

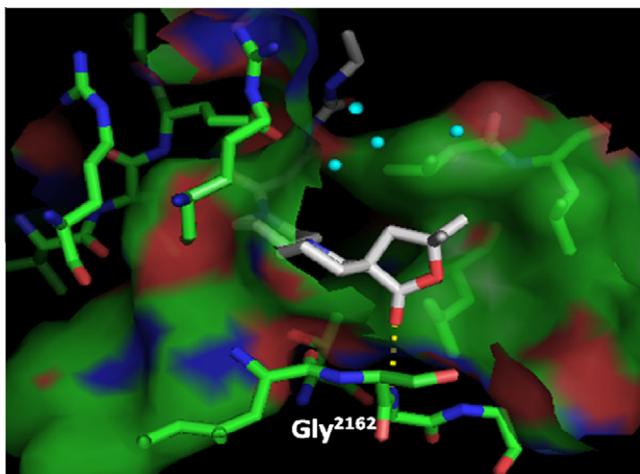
Acetyl-CoA carboxylase (ACC) inhibitory activity, human metabolic stability (hMS), and log*D* values of spiro-pyrazolo[1,2-*a*]pyridazines **4a–c**



	R <sup>1</sup>	IC <sub>50</sub> (nM)		hMS <sup>a</sup>	log <i>D</i> <sup>b</sup>
		ACC1	ACC2		
<b>4a</b>	H	22	5.9	15	1.76
<b>4b</b>	Me	25	6.1	6	2.08
<b>4c</b>	Et	21	4.9	35	2.33

<sup>a</sup> In vitro metabolic clearance in human hepatic microsomes (μL/min/mg protein).

<sup>b</sup> Log*D* values were obtained by using high-performance liquid chromatography (HPLC) at pH 7. IC<sub>50</sub> = 50% inhibitory concentration.



**Figure 1.** Detailed crystal structure of the spiro-lactone core in (*S*)-**1** with the carboxyl terminal (CT) domain of human acetyl-CoA carboxylase 2 (ACC2).

that incorporation of a polar group, for example, an oxo group at the 4-position in the spiro-lactone, would be favorable for ACC2 inhibition and prompted us to design a symmetrical spiro-pyrazolidinedione scaffold. The results of the in vitro activities of the spiro-pyrazolidinedione derivatives are summarized in Table 1. All synthesized compounds showed activities in the order of 10<sup>−8</sup> to 10<sup>−9</sup> M. As introduction of a more lipophilic and bulky substituent on the imidazolone or lactam core potentiated the activity,<sup>5</sup> compounds **3b** and **3f–h** displayed potencies of nanomolar order.

In addition, we could identify a high metabolic property of pyrazolidinediones. Examination of substituents on the spiro-5-

**Table 3**

Pharmacokinetic profile of **4c**<sup>a</sup>

	i.v. (0.1 mg/kg)			p.o. (1 mg/kg)			
	CL <sub>total</sub> <sup>b</sup> (L/h/kg)	V <sub>ss</sub> <sup>c</sup> (L/kg)	MRT <sup>d</sup> (h)	C <sub>max</sub> <sup>e</sup> (ng/mL)	T <sub>max</sub> <sup>f</sup> (h)	AUC <sup>g</sup> (ngh/mL)	F <sup>h</sup> (%)
<b>4c</b>	0.78	0.97	1.3	71	0.25	180	14

<sup>a</sup> Rat cassette dosing (*n* = 3); nonfasted CrI: CD(SD)(IGS) rats.

<sup>b</sup> Total clearance.

<sup>c</sup> Volume of distribution at steady state.

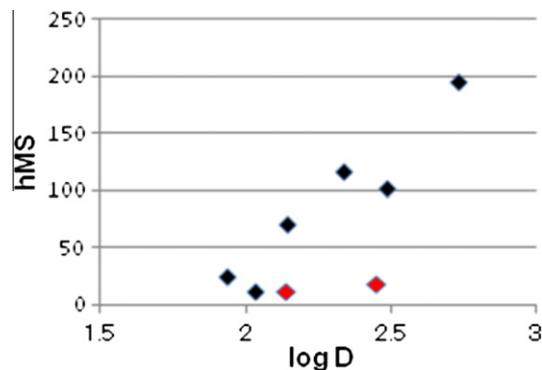
<sup>d</sup> Mean residence time.

<sup>e</sup> Maximal plasma concentration.

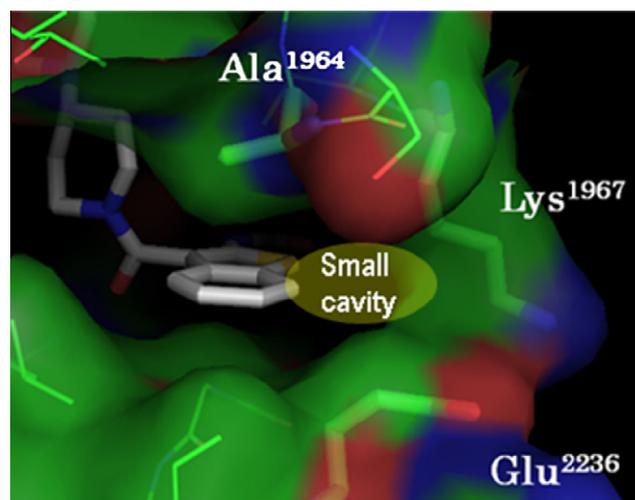
<sup>f</sup> Time of maximal concentration.

<sup>g</sup> Area under the plasma concentration versus time curve.

<sup>h</sup> Bioavailability.



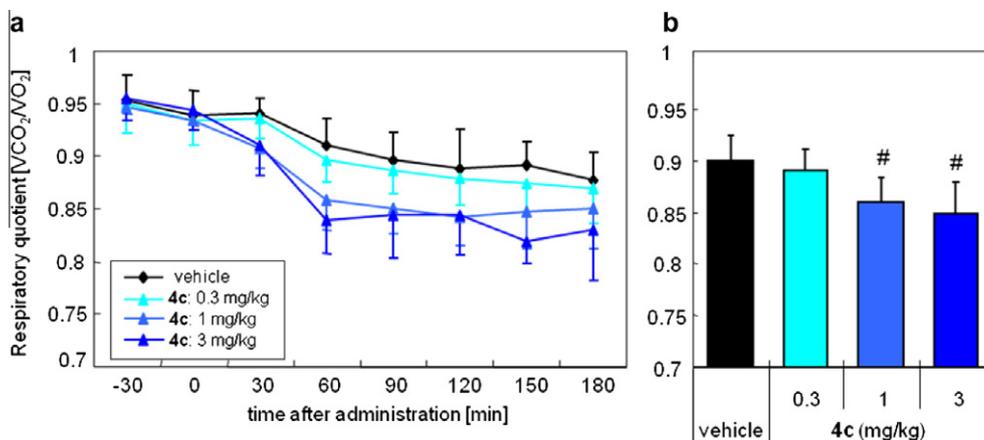
**Figure 2.** Relationships between log*D* values and metabolic stability of spiro-pyrazolidinediones **3a–h**. Red diamonds correspond to pyrazolo[1,2-*a*]pyridazine derivatives **3d** and **3e** and black diamonds correspond to the compounds. hMS: human metabolic stability.



**Figure 3.** Detailed crystal structure of the benzothiophene part in (*S*)-**1** with the carboxyl terminal (CT) domain of human acetyl-CoA carboxylase 2 (ACC2).

membered ring suggested that the log*D* values inversely correlated with metabolic stability in human microsomes except for those of **3d**, **e** (Fig. 2). Interestingly, pyrazolo[1,2-*a*]pyridazine derivatives (red diamonds: **3d**, **e**) exhibited higher metabolic stability, though the log*D* values of **3d**, **e** were the same as those of **3b** and **3f–h**. Therefore, we selected the pyrazolo[1,2-*a*]pyridazine core and modified the benzothiophene moiety to enhance the activity.

The results of the spiro-compounds possessing the pyrazolo[1,2-*a*]pyridazine moiety are summarized in Table 2. As the X-ray co-crystal data of ACC2 with (*S*)-**1** also suggest the presence of a small cavity formed by Glu2236, Lys1967, and Ala1964 at the 7-position of the benzothiophene ring (Fig. 3), introduction of a 7-methoxy group on the ring was examined as to whether it potentiates the



**Figure 4.** Decrease in the respiratory quotient (RQ) values by **4c**. Wistar fatty rats ( $n = 6$ , male) were administered with either vehicle or compound **4c** (0.3, 1, 3 mg/kg). (a) Times course of RQ. (b) Average RQ values at 3 h after administration. Values are presented as means  $\pm$  SEM.  $^{\#}p < 0.025$  (Williams test).

ACC2 inhibitory activity. In fact, the 7-methoxybenzothiophene derivatives **4a–c** exhibited potencies of nanomolar order. Moreover, all compounds retained excellent metabolic stability independent of their  $\log D$  values. Among these compounds, **4c** with the highest ACC inhibitory activity had a favorable pharmacokinetic profile in rat cassette dosing<sup>10</sup> as described in Table 3.

The selected compound **4c** also exhibited potent rat ACC inhibitory activity (rACC1:  $IC_{50} = 28$  nM, rACC2:  $IC_{50} = 13$  nM) and was tested at doses of 0.3, 1, and 3 mg/kg in Wistar fatty rats to confirm increased fatty acid oxidation by lowering the respiratory quotient (RQ). The compound **4c**-induced decrease in RQ values was dose-dependent and reached a significant level at a dose of 1 mg/kg, indicating stimulation of fatty acid oxidation (Fig. 4).<sup>11</sup>

In conclusion, we designed and synthesized various spiro-pyrazolidinedione derivatives without an asymmetric center. The compounds displayed high ACC inhibitory activities and their  $\log D$  values inversely correlated with their metabolic stability in human microsomes. Only pyrazolo[1,2-*a*]pyridazines **3d** and **3e** showed excellent metabolic stability independent of their  $\log D$  values. Moreover, X-ray co-crystal data showed that the introduction of a methoxy group at the 7-position of the benzothiophene core increased the activity. We identified orally bioavailable and highly potent ACC inhibitor **4c**, which stimulated fatty acid oxidation in rats, by combining the pyrazolo[1,2-*a*]pyridazine scaffold and the 7-methoxybenzothiophene moiety.

#### Acknowledgments

The authors thank Clifford D. Mol, Hua Zou, Douglas R. Dougan, BiChing Sang, and Gyorgy Snell for acquisition of the X-ray co-crystal data of the carboxyl terminal (CT) domain of human ACC2 in

complex with (*S*)-**1**. The authors appreciate the helpful discussion with Dr. Yu Momose.

#### Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.bmcl.2012.05.062>.

#### References and notes

- (a) Munday, M. R.; Hemingway, C. J. *Adv. Enzyme Regul.* **1999**, *39*, 205; (b) Ruderman, N. B.; Saha, A. K.; Vavvas, D.; Witters, L. A. *Am. J. Physiol.* **1999**, *276*, E1; (c) Ruderman, N. B.; Saha, A. K.; Kraegen, E. W. *Endocrinology* **2003**, *144*, 5166.
- Ruderman, N.; Prentki, M. *Nat. Rev. Drug Disc.* **2004**, *3*, 340.
- Harwood, H. J. *Expert Opin. Ther. Targets* **2005**, *9*, 267.
- Yamashita, T.; Kamata, M.; Endo, S.; Yamamoto, M.; Kakegawa, K.; Watanabe, H.; Miwa, K.; Yamano, T.; Funata, M.; Sakamoto, J.; Tani, A.; Mol, C. D.; Zou, H.; Dougan, D. R.; Sang, B.; Snell, G.; Fukatsu, K. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 6314.
- Kamata, M.; Yamashita, T.; Kina, A.; Funata, M.; Mizukami, A.; Sasaki, M.; Tani, A.; Funami, M.; Amano, N.; Fukatsu, K. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 3642.
- Synthesis of 1-ethyl-3-[(4-oxopiperidin-1-yl)carbonyl]-1-benzothiophen-2-yl)urea **18** was performed by acylation of 4-piperidone with **14** followed by formation of ureido group using ethyl isocyanate.
- (a) Cauquil-Caubère, I.; Kamenka, J.-M. *Eur. J. Med. Chem.* **1998**, *33*, 867; (b) Pinna, G. A.; Curzu, M. M.; Cignarella, G.; Barlocco, D.; D'Amico, M.; Filippelli, A.; De Novellis, V.; Rossi, F. *Eur. J. Med. Chem.* **1994**, *29*, 447.
- Bernal, C.; Palacin, C.; Boronat, A.; Imperial, S. *Anal. Biochem.* **2005**, *337*, 55.
- Koike, T.; Takai, T.; Hoashi, Y.; Nakayama, M.; Kosugi, Y.; Nakashima, M.; Yoshikubo, S.; Hirai, K.; Uchikawa, O. *J. Med. Chem.* **2011**, *54*, 4207.
- Negoro, N.; Sasaki, S.; Ito, M.; Kitamura, S.; Tsujihata, Y.; Ito, R.; Suzuki, M.; Takeuchi, K.; Suzuki, N.; Miyazaki, J.; Santou, T.; Odani, T.; Kanzaki, N.; Funami, M.; Tanaka, T.; Yasuma, T.; Momose, Y. *J. Med. Chem.* **2012**, *55*, 1538.
- Harwood, H. J., Jr.; Petras, S. F.; Shelly, L. D.; Zaccaro, L. M.; Perry, D. A.; Makowski, M. R.; Hargrove, D. M.; Martin, K. A.; Tracey, W. R.; Chapman, J. G.; Magee, W. P.; Dalvie, D. K.; Soliman, V. F.; Martin, W. H.; Mularski, C. J.; Eisenbeis, S. A. *J. Biol. Chem.* **2003**, *278*, 37099.