



Pergamon

# Synthesis of novel discodermolide analogues with modified hydrogen-bonding donor/acceptor sites

Ian Paterson\* and Oscar Delgado

University Chemical Laboratory, Lensfield Road, Cambridge CB2 1EW, UK

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**Abstract**—A series of novel structural analogues of the potent microtubule-stabilizing anticancer agent discodermolide were synthesised, with modifications in the C16–C20 region to create new oxygenated H-bonding donor/acceptor sites for tubulin binding. By starting from an advanced C9–C24 intermediate, fully synthetic discodermolide analogues, incorporating either an additional hydroxyl group **3**, an oxetane **4** or a cyclic carbonate **5**, were obtained in 10 or 11 steps by using a versatile aldol construction of the C6–C7 bond.

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As a structurally unique antimetabolic agent, the marine-derived polyketide discodermolide (**1**, Fig. 1) is a promising candidate for clinical development in cancer chemotherapy.<sup>1–4</sup> Sharing a similar microtubule-stabilizing mechanism of action to Taxol/paclitaxel (**2**),<sup>3a–d</sup> discodermolide inhibits the proliferation of numerous cancer cell lines, including those that are Taxol-resistant, and shows synergy with Taxol.<sup>3c</sup> Moreover, in hollow fibre and xenograft mouse models, discodermolide induces significant growth inhibition of human tumours *in vivo*.<sup>4</sup> Its potential as a new chemotherapeutic agent for the treatment of solid tumours, including

drug-resistant breast and ovarian cancer, has recently led to discodermolide entering Phase I clinical trials. Due to the low isolation yield from the deep-sea sponge source,<sup>2</sup> total synthesis presently offers the only viable means of drug supply. This situation has stimulated considerable interest in developing a practical synthetic route for producing discodermolide,<sup>1</sup> thus also enabling access to novel analogues for biological evaluation. Recently, studies focussed on exploring structure–activity relationships for discodermolide, with the aim of defining a pharmacophore model, have been reported by several groups.<sup>4,5</sup>

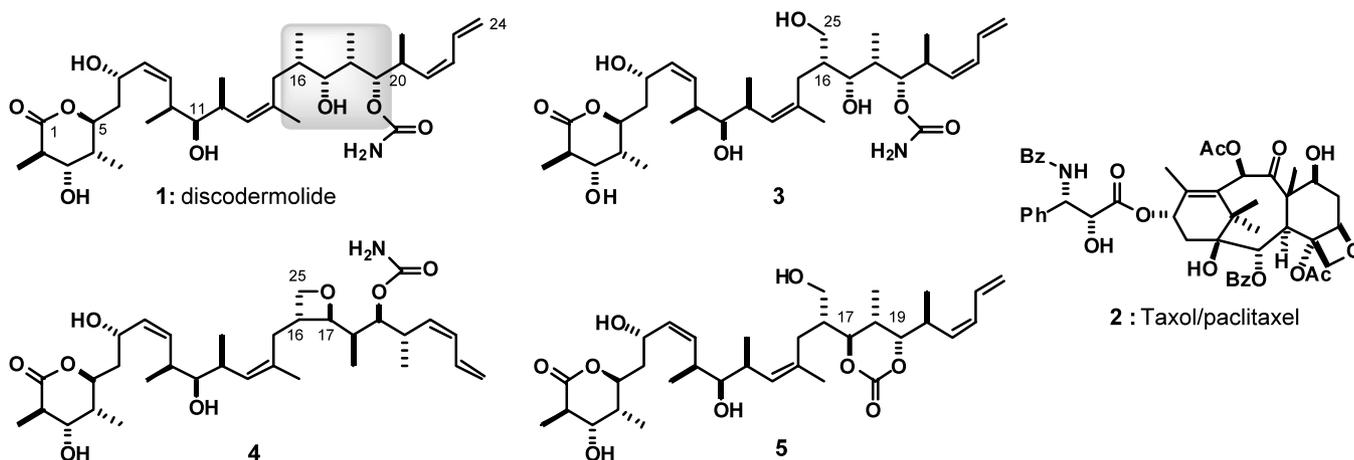
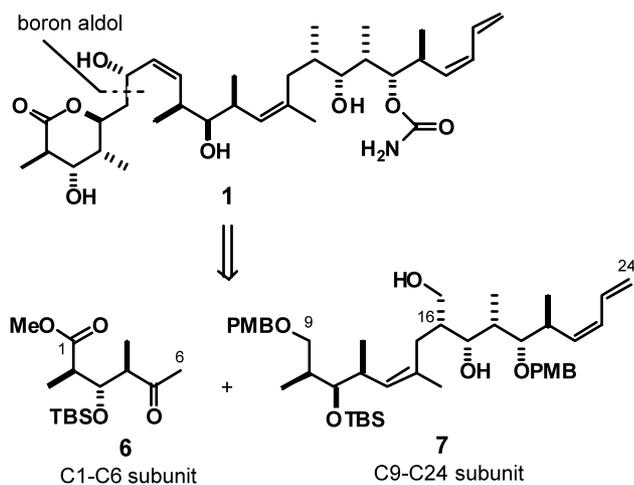


Figure 1.

\* Corresponding author.

In parallel with our efforts to develop a highly practical synthesis of discodermolide,<sup>1a,6</sup> we have previously prepared a range of epimeric and truncated analogues.<sup>5a</sup> As an extension to this work, we now report the total synthesis of the more elaborate discodermolide analogues **3**, **4** and **5**, where these feature novel oxygenated hydrogen-bonding donor/acceptor sites within the C16–C20 region (as indicated by the shading in **1**), potentially enhancing interactions with the binding site in  $\beta$ -tubulin. Notably, the cyclic carbonate **5** arises from an unexpected acid-mediated cyclisation of the oxetane analogue **4**.

In the SAR studies on discodermolide reported originally by Schreiber and co-workers,<sup>5b</sup> the natural (*S*)-configuration at the C16 methyl-bearing stereocentre proved to be essential for retaining biological activity. To extend these studies further into the C16–C20 region, we envisaged adapting our total synthesis of (+)-discodermolide (Scheme 1) that employs the methyl ketone **6** and 1,3-diol **7** as C1–C6 and C9–C24 subunits, respectively.<sup>6a,b</sup> In this work, the required methyl substitution at C16 was secured by performing a controlled deoxygenation on **7**. This approach presents a convenient opportunity for analogue chemistry if the oxygenation associated with the C16 position is retained, enabling the modification of the hydrogen bonding



Scheme 1.

donor/acceptor sites in this region of the linear polyketide backbone.

To this end, the preparation of the hydroxymethyl analogue **3** (with one additional H-bond donor/acceptor site) and the oxetane-containing analogue **4** (with one H-bond donor site removed) was initially undertaken. In the latter case, the introduction of the Lewis and Brønsted basic oxetane also introduces a conformational lock, preventing rotation about the C16–C17 bond, as well as conferring some superficial resemblance to the D-ring oxetane in paclitaxel (**2**). Otherwise, these localised structural changes were not expected to lead to any severe perturbation of the conformational preferences over the rest of the molecule. In this regard, modelling studies (MacroModel 7.2)<sup>7</sup> indicated that both the pentaol **3** and oxetane **4** largely mimicked the conformational preferences of discodermolide (**1**),<sup>8</sup> favouring a similar low energy U-shaped arrangement (Fig. 2).

The strategy adopted for the preparation of discodermolide analogues **3** and **4** relied on a late-stage aldol coupling between the methyl ketone **6** and the appropriate (*Z*)-enal containing the modified C7–C24 region. As shown in Scheme 2, the synthesis of the first analogue **3**, having a hydroxymethyl substituent at C16, started out from the 1,3-diol **7**.<sup>6</sup> *Bis*-silylation of **7** with TBSOTf/2,6-lutidine, followed by oxidative cleavage of both PMB ethers with DDQ, gave the diol **8** (90%). Selective oxidation of the primary alcohol using TEMPO/PhI(OAc)<sub>2</sub><sup>9</sup> and a Still–Gennari HWE olefination<sup>10</sup> of the resulting aldehyde generated the desired (*Z*)-enoate **9** exclusively in 84% yield. Following installation of the carbamate moiety at the C19 hydroxyl in **9** under standard conditions (Cl<sub>3</sub>CC(O)NCO; K<sub>2</sub>CO<sub>3</sub>, MeOH),<sup>11</sup> DIBAL-H reduction of the methyl ester to provide the corresponding alcohol and subsequent Dess–Martin oxidation afforded (*Z*)-enal **10** (96%).

Enolisation of methyl ketone **6** with (+)-Ipc<sub>2</sub>BCl/Et<sub>3</sub>N in Et<sub>2</sub>O,<sup>6a,b,12</sup> followed by addition to aldehyde **10**, proceeded in 65% yield with 8:1 *dr* in favour of the desired (*7S*)-configured adduct **11**. In this complex aldol coupling, the chiral boron reagent is needed to overturn the strong inherent facial bias of the aldehyde

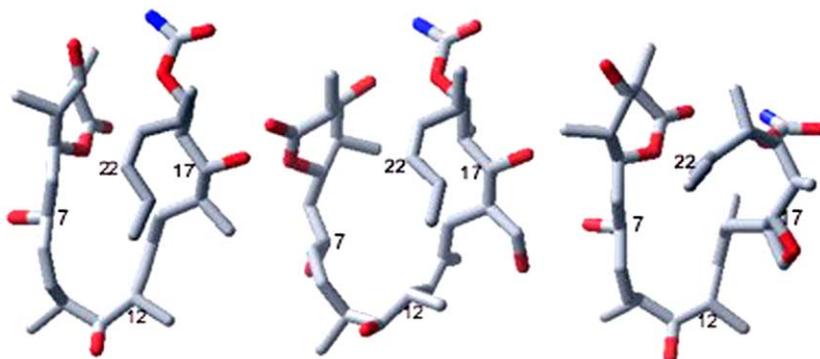
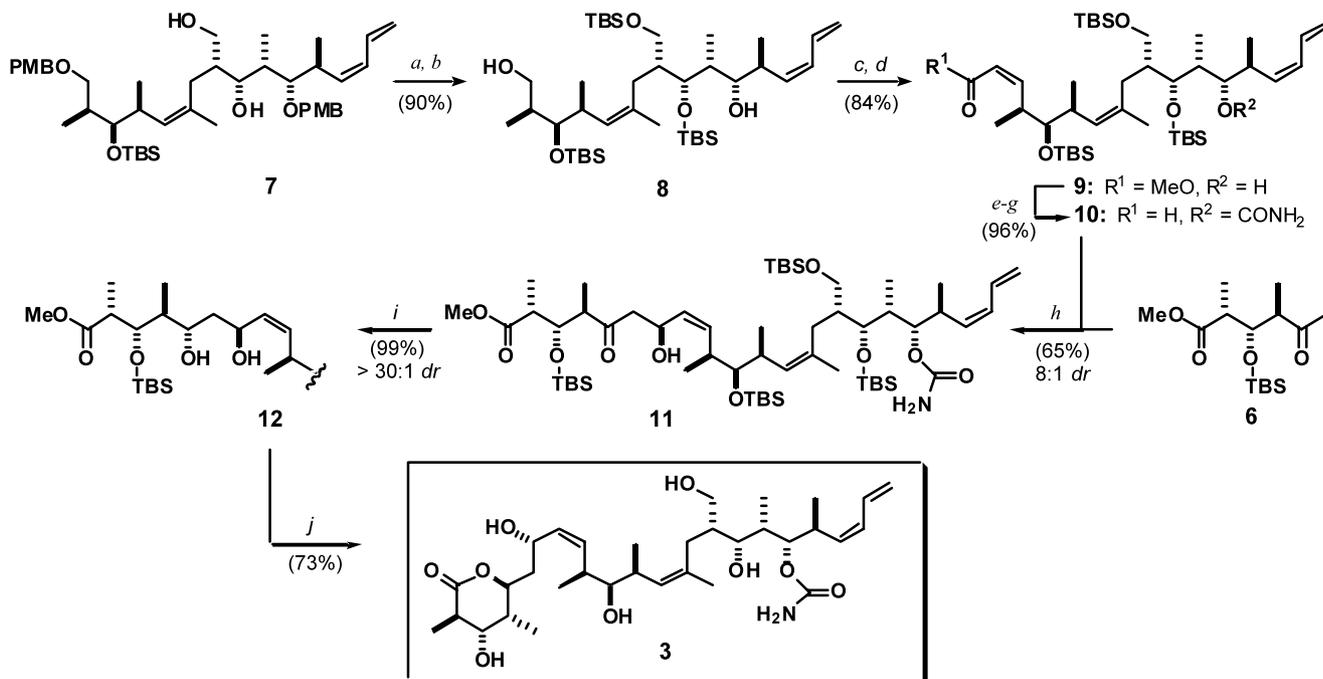


Figure 2. Energy minimised conformations of **1**, **3** and **4**.<sup>7,8</sup>



**Scheme 2.** Reagents and conditions: (a) TBSOTf, 2,6-lutidine, CH<sub>2</sub>Cl<sub>2</sub>, -78 to 20°C, 3 h; (b) DDQ, CH<sub>2</sub>Cl<sub>2</sub>/pH 7 buffer, 0°C, 2 h; (c) cat. TEMPO, PhI(OAc)<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 4 h; (d) (CF<sub>3</sub>CH<sub>2</sub>O)<sub>2</sub>P(O)CH<sub>2</sub>CO<sub>2</sub>Me, K<sub>2</sub>CO<sub>3</sub>, 18-C-6, PhMe, -20 to 0°C, 3 h; (e) Cl<sub>3</sub>CC(O)NCO, CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 30 min; K<sub>2</sub>CO<sub>3</sub>, MeOH, 20°C, 2 h; (f) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, -78°C, 4 h; (g) Dess–Martin periodinane, CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 30 min; (h) **6**, (+)-Ipc<sub>2</sub>BCl, Et<sub>3</sub>N, Et<sub>2</sub>O, 0°C, 1.5 h; **10**, -78 to -20°C, 16 h; (i) Me<sub>4</sub>NBH(OAc)<sub>3</sub>, MeCN/AcOH, -20°C, 1 h; (j) 3 M HCl, MeOH, 20°C, 72 h.

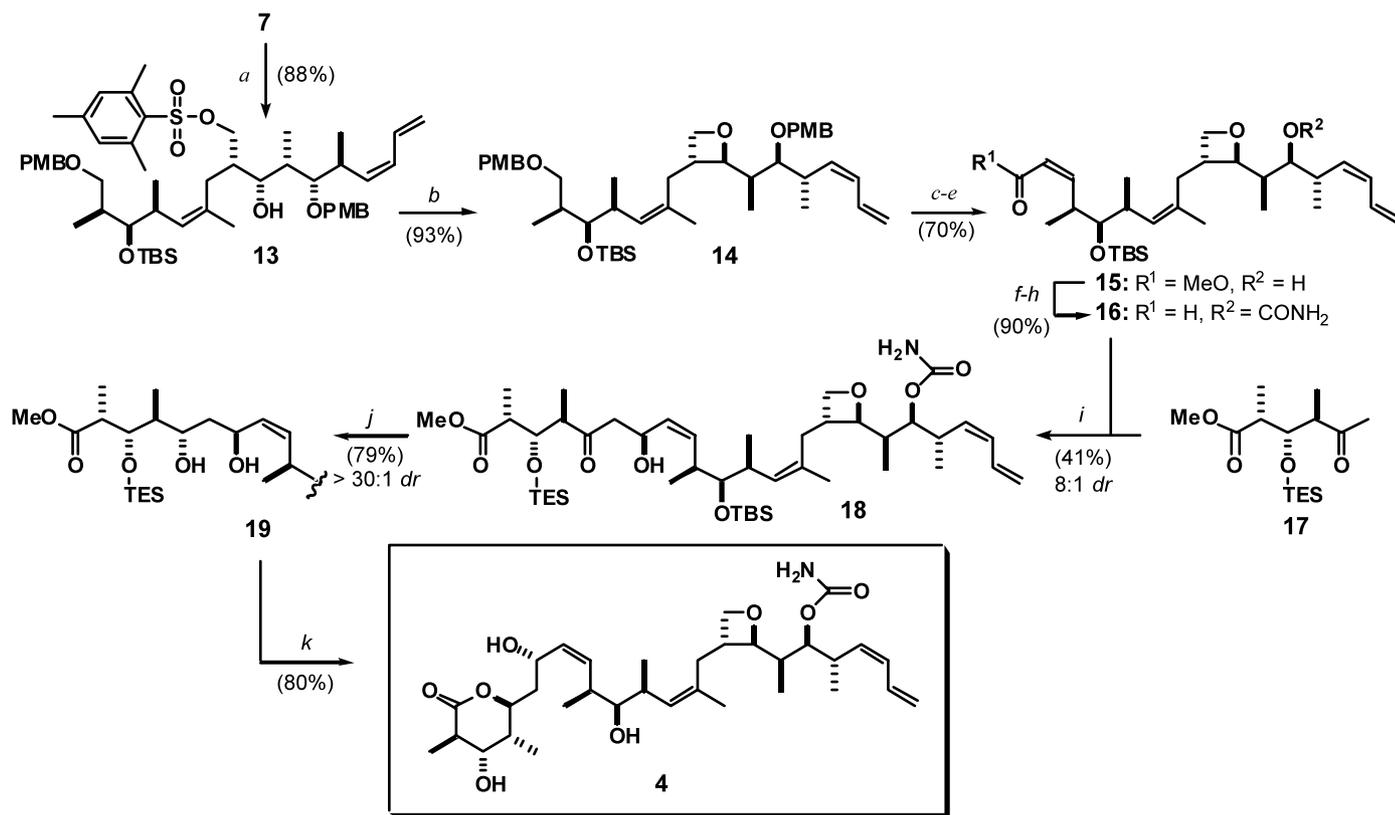
component. Subsequent hydroxyl-directed 1,3-*anti* reduction<sup>13</sup> of the β-hydroxy ketone **11** with Me<sub>4</sub>NBH(OAc)<sub>3</sub> then provided diol **12** smoothly (99%). Final treatment of this diol **12** with 3 M HCl in MeOH (72 h) caused deprotection of the four TBS ethers with concomitant lactonisation to afford the desired C16-hydroxymethyl analogue **3** (73%).<sup>14</sup> Overall, this synthesis of pentaol **3** proceeded in 10 steps and 30% yield from diol **7** (Scheme 2).

Our synthetic plan for the more challenging discodermolide analogue **4** relied upon the early introduction of the oxetane moiety (Scheme 3). Thus, selective sulfonation of 1,3-diol **7** with 2,6-mesitylenesulfonyl chloride and Et<sub>3</sub>N gave **13** in 88% yield. In the course of our discodermolide synthesis,<sup>6b</sup> treatment of **13** with Super-Hydride<sup>®</sup>, with a view to reductively displacing the sulfonate group, unexpectedly led to the formation of oxetane **14**. This fortuitous reaction proved less useful on a preparative scale, where a complex mixture of products was obtained. After some optimisation, we found that generation of the potassium alkoxide of **13** with KO<sup>*t*</sup>-Bu in THF led to clean cyclisation to afford **14** in 93% yield.

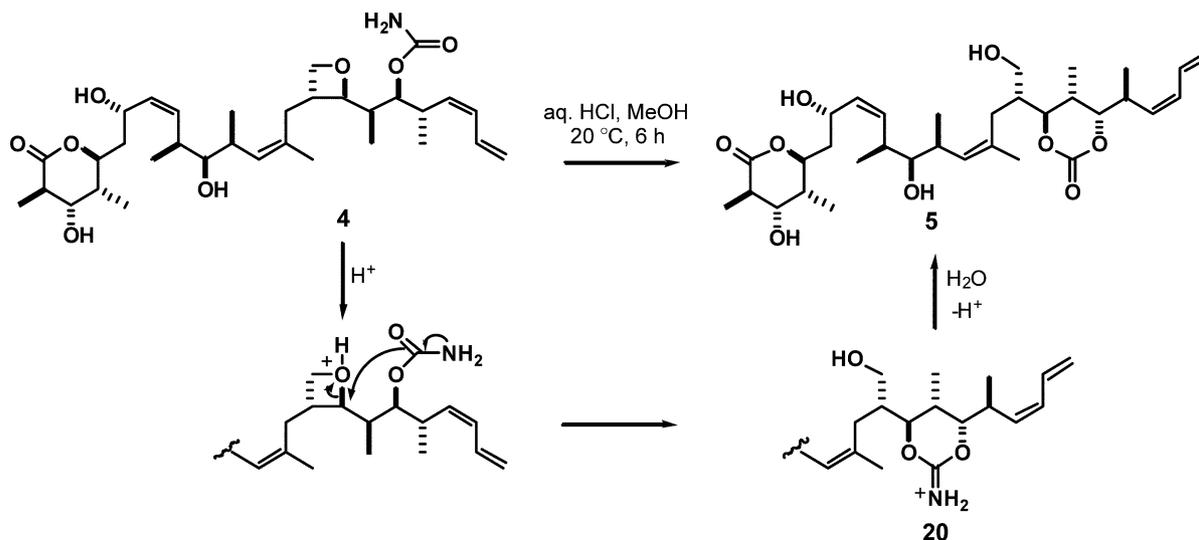
Removal of the PMB ethers in **14**, selective oxidation and a (*Z*)-selective HWE reaction then provided ester **15**. Following introduction of the carbamate and oxidation state adjustment, the aldehyde **16** was obtained in readiness for aldol coupling with the C1–C6 subunit. In this case, however, the presence of the acid-sensitive oxetane functionality necessitated a reassessment of the

protecting group strategy. Thus, the TBS ether at the C3 hydroxyl in methyl ketone **6** was first replaced with the more labile TES ether, as in **17**, in order to avoid undesired oxetane opening in the final desilylation step.<sup>15</sup> Under our standard conditions, the aldol coupling between **16** and **17** proceeded in 41% yield with 8:1 *dr* in favour of the (*7S*)-adduct **18**. Following controlled 1,3-*anti* reduction to give **19**, brief treatment with 3M HCl in MeOH (1 h) then provided the target oxetane analogue **4** (62%).<sup>14</sup> Altogether, this sequence proceeded in 11 steps and 13% yield from 1,3-diol **7**.

Fortuitously, a further structurally novel discodermolide analogue was isolated from the preceding acid-mediated deprotection step to form the oxetane analogue **4**. Initially, the presence of a minor byproduct in the reaction mixture was detected and, following its isolation by normal-phase HPLC purification, NMR analysis suggested opening of the oxetane ring and loss of the carbamate moiety. On a preparative scale, treatment of oxetane **4** with aq. HCl/MeOH for 6 h led to clean conversion into the same byproduct (79%), where the structure was assigned as the cyclic carbonate **5** (Scheme 4). Mechanistically, **5** presumably arises by acid-catalysed regioselective opening of the oxetane at the C17 position by the carbonyl oxygen of the adjacent carbamate group, followed by hydrolysis of the resulting imino carbonate intermediate **20**. Detailed NMR analysis of **5** indicated that this intramolecular oxetane opening reaction proceeded with inversion of configuration at C17.<sup>16</sup>



**Scheme 3.** Reagents and conditions: (a) 2,4,6-Me<sub>3</sub>(C<sub>6</sub>H<sub>2</sub>)SO<sub>2</sub>Cl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 20 h; (b) *t*-BuOK, THF, 0°C, 30 min; (c) DDQ, CH<sub>2</sub>Cl<sub>2</sub>/pH7 buffer, 0°C, 2 h; (d) cat. TEMPO, PhI(OAc)<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 4 h; (e) (CF<sub>3</sub>CH<sub>2</sub>O)<sub>2</sub>P(O)CH<sub>2</sub>CO<sub>2</sub>Me, K<sub>2</sub>CO<sub>3</sub>, 18-C-6, PhMe, -20 to 0°C, 3 h; (f) Cl<sub>3</sub>CC(O)NCO, CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 30 min; K<sub>2</sub>CO<sub>3</sub>, MeOH, 20°C, 2 h; (g) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, -78°C, 4 h; (h) Dess–Martin periodinane, CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 30 min; (i) **17**, (+)-Ipc<sub>2</sub>BCl, Et<sub>3</sub>N, Et<sub>2</sub>O, 0°C, 1.5 h; **16**, -78 to -20°C, 16 h; (j) Me<sub>4</sub>NBH(OAc)<sub>3</sub>, MeCN/AcOH, -20°C, 1 h; (k) 3 M HCl, MeOH, 20°C, 1 h.



**Scheme 4.**

In summary, we have synthesised three novel analogues **3**, **4** and **5** by adapting our versatile aldol-based route to discodermolide.<sup>6</sup> Biological evaluation of these compounds regarding their cell growth inhibitory activities

and tubulin polymerization properties may provide useful information in helping to define the preferred binding mode of discodermolide to  $\beta$ -tubulin, and these studies will be reported in due course.

### Acknowledgements

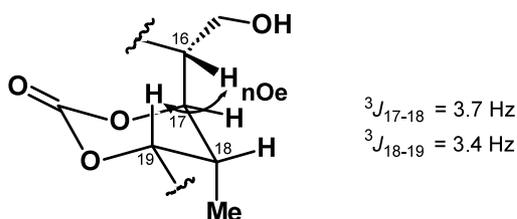
We thank the EC (Network HPRN-CT-200-0018), Gobierno de Canarias, Cambridge European Trust and Novartis Pharma AG for support, and Professor J. Vilarrasa and Oriol Pineda (University of Barcelona) for helpful discussions over the modelling studies.

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- All new compounds gave spectroscopic data in agreement with the structures indicated. Analogue **3** had <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 6.61 (1H, ddd, *J* = 16.8, 10.8, 10.4 Hz, H<sub>23</sub>), 6.04 (1H, app t, *J* = 11.0 Hz, H<sub>22</sub>), 5.53 (1H, dd, *J* = 11.0, 8.0 Hz, H<sub>8</sub>), 5.41 (1H, dd, *J* = 10.7, 10.5 Hz, H<sub>9</sub>), 5.34 (1H, dd, *J* = 10.4, 10.3 Hz, H<sub>21</sub>), 5.24 (2H, m, H<sub>13</sub>, H<sub>24A</sub>), 5.13 (1H, d, *J* = 10.3 Hz, H<sub>24B</sub>), 4.74 (1H, ddd, *J* = 10.3, 8.0, 3.4 Hz, H<sub>7</sub>), 4.70–4.58 (4H, m, H<sub>5</sub>, H<sub>19</sub>, CONH<sub>2</sub>), 3.82 (1H, br d, *J* = 11.0 Hz, H<sub>25A</sub>), 3.73 (1H, dd, *J* = 3.8, 3.8 Hz, H<sub>3</sub>), 3.58–3.54 (2H, m, H<sub>17</sub>, H<sub>25B</sub>), 3.20 (1H, dd, *J* = 6.8, 4.3 Hz, H<sub>11</sub>), 3.00 (1H, ddq, *J* = 10.1, 6.8, 6.8 Hz, H<sub>20</sub>), 2.78 (1H, ddq, *J* = 9.7, 6.7, 6.7 Hz, H<sub>10</sub>), 2.66 (1H, qd, *J* = 7.1, 4.4 Hz, H<sub>2</sub>), 2.63–2.56 (1H, m, H<sub>12</sub>), 2.14–1.87 (8H, m, H<sub>4</sub>, H<sub>6A</sub>, H<sub>15A</sub>, H<sub>15B</sub>, H<sub>18</sub>, 3×OH), 1.78–1.61 (5H, m obs, H<sub>6B</sub>, 14-Me, OH), 1.33 (3H, d, *J* = 7.3 Hz, 2-Me), 1.08 (3H, d, *J* = 7.1 Hz, 4-Me), 1.05–0.99 (9H, d, 10-Me, 18-Me, 20-Me), 0.97 (3H, d, *J* = 7.0 Hz, 12-Me); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 173.8, 157.3, 134.5, 133.3, 132.9, 132.5, 132.0, 130.6, 130.1, 118.2, 79.5, 79.0, 73.2, 64.3, 43.1, 40.9, 39.1, 37.7, 36.2, 35.7, 35.3, 35.1, 31.2, 31.1, 30.9, 29.7, 23.3, 18.3, 17.5, 15.6, 15.3, 12.6, 8.4; HRMS (ES<sup>+</sup>) calcd for C<sub>33</sub>H<sub>55</sub>NO<sub>9</sub>Na [M+Na]<sup>+</sup> 615.3775, found 615.3765. Analogue **4** had <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 6.62 (1H, ddd, *J* = 16.9, 10.8, 10.5 Hz, H<sub>23</sub>), 6.03 (1H, app t, *J* = 11.0 Hz, H<sub>22</sub>), 5.53 (1H, dd, *J* = 11.1, 7.7 Hz, H<sub>8</sub>), 5.44 (1H, dd, *J* = 10.7, 10.3 Hz, H<sub>9</sub>), 5.29 (1H, dd, *J* = 10.6, 10.4 Hz, H<sub>21</sub>), 5.21 (1H, dd, *J* = 16.8, 1.2 Hz, H<sub>24A</sub>), 5.15–5.09 (2H, m, H<sub>13</sub>, H<sub>24B</sub>), 4.73 (1H, ddd, *J* = 8.3, 8.3, 3.2 Hz, H<sub>7</sub>), 4.65–4.59 (2H, m, H<sub>5</sub>, H<sub>19</sub>), 4.58 (2H, br s, CONH<sub>2</sub>), 4.53 (1H, dd, *J* = 7.9, 6.1 Hz, H<sub>25A</sub>), 4.25–4.21 (2H, m, H<sub>17</sub>, H<sub>25B</sub>), 3.74 (1H, dd, *J* = 4.1, 3.9 Hz, H<sub>3</sub>), 3.19 (1H, dd, *J* = 6.6, 5.2 Hz, H<sub>11</sub>), 2.98 (1H, ddq, *J* = 9.0, 6.9, 6.9 Hz, H<sub>20</sub>), 2.81–2.65 (3H, m, H<sub>2</sub>, H<sub>10</sub>, H<sub>16</sub>), 2.63 (1H, ddq, *J* = 10.1, 6.9, 6.9 Hz, H<sub>12</sub>), 2.46 (1H, dd, *J* = 13.7, 10.6 Hz, H<sub>15A</sub>), 2.36 (1H, dd, *J* = 13.8, 5.0 Hz, H<sub>15B</sub>), 2.22–2.00 (4H, m, H<sub>4</sub>, 3×OH), 1.86 (1H, ddq, *J* = 13.5, 6.8, 3.4 Hz, H<sub>18</sub>), 1.73 (1H, ddd, *J* = 14.4, 9.4, 2.4 Hz, H<sub>6A</sub>), 1.72 (1H, ddd, *J* = 14.2, 10.7, 3.3 Hz, H<sub>6B</sub>), 1.62 (3H, s, 14-Me), 1.33 (3H, d, *J* = 7.2 Hz, 2-Me), 1.09 (3H, d, *J* = 6.9 Hz, 18-Me), 1.05 (3H, d, *J* = 6.9 Hz, 10-Me), 1.04–1.00 (6H, m, 4-Me, 20-Me), 0.97 (3H, d, *J* = 6.8 Hz, 12-Me); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 173.7, 156.6,

134.0, 133.8, 132.9, 132.2 (2C), 130.2, 130.1, 117.9, 89.5, 79.0, 77.2, 75.6, 73.7, 73.2, 64.7, 43.1, 41.0, 40.5, 37.7, 36.1, 35.7, 35.6, 35.0, 34.7, 23.4, 18.4, 17.4, 15.6, 15.3, 12.5, 8.3; HRMS (ES+) calcd for  $C_{33}H_{53}NO_8Na$   $[M+Na]^+$  614.3669, found 614.3671.

15. On acid treatment of **19**, rapid deprotection of the two TBS groups and the TES group was accomplished in 80% yield in 1 h. In contrast, the TBS ether at C3 on the  $\delta$ -lactone of discodermolide requires prolonged treatment with acid (2–3 days) for complete removal and this was found to be incompatible with the oxetane group.
16. Analysis of the  $^1H$  NMR data of cyclic carbonate **5** enabled the C17 configuration to be determined unambiguously. Strong nOe contacts were observed between  $H_{19}$  and  $H_{16}$ , which is consistent with the preferred chair-like conformation depicted below. The small vicinal coupling constants,  $^3J_{17-18}=3.7$  Hz and  $^3J_{18-19}=3.4$  Hz, are also in good agreement with the proposed stereostructure.



Analogue **5** had  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta_H$  6.61 (1H, ddd,  $J=16.8, 11.0, 10.4$  Hz,  $H_{23}$ ), 6.13 (1H, app t,  $J=10.9$  Hz,  $H_{22}$ ), 5.60–5.53 (2H, m,  $H_8, H_9$ ), 5.38 (1H, dd,  $J=10.4, 10.3$  Hz,  $H_{21}$ ), 5.28 (1H, d,  $J=16.8$  Hz,  $H_{24A}$ ), 5.23–5.19 (2H, m,  $H_{13}, H_{24B}$ ), 4.70 (1H, dddd,  $J=6.8, 6.5, 3.4, 3.3$  Hz,  $H_7$ ), 4.66 (1H, ddd,  $J=11.4, 10.5, 2.1$  Hz,  $H_5$ ), 4.33 (1H, dd,  $J=7.8, 3.7$  Hz,  $H_{17}$ ), 4.30 (1H, dd,  $J=7.8, 3.4$  Hz,  $H_{19}$ ), 3.77 (1H, dd,  $J=4.6, 4.0$  Hz,  $H_3$ ), 3.66 (1H, dd,  $J=11.0, 3.4$  Hz,  $H_{25A}$ ), 3.55 (1H, dd,  $J=10.7, 6.0$  Hz,  $H_{25B}$ ), 3.24 (1H, app t,  $J=5.6$  Hz,  $H_{11}$ ), 3.00 (1H, ddq,  $J=9.2, 7.3, 7.3$  Hz,  $H_{20}$ ), 2.82–2.76 (1H, m,  $H_{10}$ ), 2.69 (1H, qd,  $J=7.5, 5.0$  Hz,  $H_2$ ), 2.58 (1H, ddq,  $J=7.2, 3.7, 3.5$  Hz,  $H_{18}$ ), 2.51 (1H, ddq,  $J=9.7, 6.4, 6.4$  Hz,  $H_{12}$ ), 2.33 (1H, dd,  $J=13.7, 11.4$  Hz,  $H_{15A}$ ), 2.20–2.12 (1H, m,  $H_{15B}$ ), 2.06–1.99 (2H, m,  $H_4, H_{16}$ ), 1.87 (1H, ddd,  $J=14.5, 9.2, 2.3$  Hz,  $Me_{6A}$ ), 1.80–1.56 (6H, m,  $4\times OH, H_{6B}, 14-Me$ ), 1.34 (3H, d,  $J=7.2$  Hz, 2-Me), 1.16 (3H, d,  $J=7.1$  Hz, 18-Me), 1.10 (3H, d,  $J=7.0$  Hz, 4-Me), 1.08–1.05 (6H, m, 10-Me, 20-Me), 1.01 (3H, d,  $J=6.7$  Hz, 12-Me);  $^{13}C$  NMR (125 MHz,  $CDCl_3$ )  $\delta_C$  173.9, 149.2, 134.1, 132.9, 132.1, 131.8, 131.7 (2C), 130.7, 118.9, 85.4, 82.3, 79.2, 77.0 (obs), 72.9, 64.2, 60.9, 42.8, 41.2, 40.8, 36.1, 35.9, 35.8, 35.3, 29.5, 29.1, 23.2, 18.9, 17.2, 15.9, 15.4, 12.5, 11.8; HRMS (ES+) calcd for  $C_{33}H_{52}O_9Na$   $[M+Na]^+$  615.3509, found 615.3516.