Synthesis and Antitumor Activity of Lipid A Analogs Having a Phosphonooxyethyl Group with α - or β -Configuration at Position 1

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Three novel lipid A analogs, which have an α - or β -glycosidically bound phosphonooxyethyl group instead of the α -glycosyl phosphate group of natural lipid A, were synthesized. The first analog (2) had an α -phosphonooxyethyl group on the identical acylated disaccharide 4'-phosphate structure found in natural lipid A (from *Escherichia coli*) and hence differed from the latter only in the nature of the acidic group at position 1. The second one (3) had tetradecanoyl groups in place of the two (R)-3-hydroxytetradecanoyl groups bound to the 2- and 3-hydroxyl function of 2, retaining the α -phosphonooxyethyl group. The structure of the third analog (4) was the same as that of 3 except that the phosphonooxyethyl group of the former was β -oriented.

Compounds 2 and 3 exhibited potent activity against Meth A at the same level as natural lipid A, whereas 4 showed less activity. This fact revealed that the glycosidic phosphate is not a prerequisite for the antitumor activity of lipopolysaccharide. It can be replaced with a phosphonooxyethyl group without any loss of activity provided that the α -anomeric configuration at C-1 is retained. The replacement of the hydroxytetradecanoyl groups with tetradecanoyl groups does not change the activity either.

Keywords lipid A analogs; phosphonooxyethyl group; phosphate group; antitumor activity; Meth A fibrosarcoma; configuration; (R)-3-hydroxytetradecanoic acid

It is well known that lipopolysaccharide (LPS) has various potent biological activities including antitumor activity, induction of interferon, mitogenic activity for B-lymphocytes, and lethal toxicity. Most of these so-called endotoxic activities have been attributed to the lipophilic moiety of the LPS molecule called lipid A. The fundamental chemical structure of lipid A isolated from many *Enterobacteriae* was elucidated to have a backbone of the 1,4′-bisphosphorylated β (1→6) disaccharide of D-glucosamine. The complete chemical structure of *Escherichia coli* (*E. coli*) lipid A was deduced as 1 shown in the figure below and this structure was unequivocally confirmed by total synthesis. The synthetic *E. coli* lipid A showed biological activity identical with its natural counterpart. The synthetic is the synthesis of the synthesis of the synthesis.

Thereafter, many structural analogs of natural lipid A have been chemically synthesized⁶⁾ and tested for biological activities.⁷⁾ The results have given important information concerning the structure–activity relationship. For example, the phosphate groups seemed to play a significant role in the expression of the activity because the synthetic compound which lacks either of the two phosphates was found to have lower activity than the original bisphos-

$$(HO)_{2}OPO \xrightarrow{OR^{3}} HO \xrightarrow{OR^{1}} OPO(OH)_{2}$$

$$1$$

$$OH$$

$$R^{1}: CH_{3}(CH_{2})_{10}CHCH_{2}CO$$

$$R^{2}: CH_{3}(CH_{2})_{10}OCO$$

$$CH_{3}(CH_{2})_{10}CHCH_{2}CO$$

$$R^{3}: CH_{3}(CH_{2})_{12}OCO$$

СН₃(СН₂)₁₀СНСН₂СО

phorylated compound.

In the course of our synthetic efforts toward new antitumor compounds, we became interested in replacing the glycosyl phosphate moiety of lipid A. Though removal of even one of the phosphates of lipid A lowered its antitumor potency as described above, we anticipated that the substitution of them with other polar groups could not necessarily lead to similar loss of the activity. Of the two phosphate groups in the lipid A molecule, one at the glycosyl hydroxyl group is difficult to introduce synthetically and is chemically unstable. Hence, the presence of this phosphoric ester makes the synthesis itself and the purification of the product considerably difficult. We expected that substitution of this particular phosphate with a stable polar (acidic) group not only makes the synthetic strategy very flexible but also opens wide possibilities to create novel antitumor active compounds, which could be less toxic and have potential clinical application, based on the endotoxic lipid A.

According to this consideration we synthesized lipid A analogs with a glycosidically bound phosphonooxyethyl group. We expected that the acidity and the solubility of a phosphonooxyethyl glycoside resemble those of the corresponding glycosyl phosphate so that this type of substitution would not cause a serious change of the physical character of the molecules. In addition, the phosphoric ester in a phosphonooxyethyl glycoside was assumed to be more stable than a glycosyl phosphate since the phosphoric acid in the former structure is bound not directly at a glycosyl but at an alcoholic hydroxyl group of ethylene glycol.

We described in this paper a synthesis of three analogs 2, 3, 4 of natural lipid A. All three compounds are phosphonooxyethyl glycosides of a $\beta(1\rightarrow6)$ -linked glucosamine disaccharide which is phosphorylated at position 4' and acylated at positions 2, 2', 3, 3'. Compound 2 has the

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a: $R^4 = (R)$ -3-benzyloxytetradecanoyl

b: R⁴= tetradecanoyl

R⁵= tetradecanoyl

Troc = 2,2,2-trichloroethoxycarbonyl

i) HO(CH₂)₂OH, HCl; ii) Ac₂O, Py; iii) Zn/AcOH; iv) R⁴OH–DCC or R⁴Cl; v) NaOCH₃; vi) (CH₃)₂C(OCH₃)₂, H⁺; vii) (PhO)₂POCl, DMAP; viii) 90% AcOH

Chart 1

identical composition and distribution of acyl groups as in natural $E.\ coli$ lipid A. The phosphonooxyethyl group of 2 is in α -configuration like the α -phosphate in lipid A as well. Both 3 and 4 have again the identical acylation pattern as 2 except that the two (R)-3-hydroxytetradecanoyl groups on positions 2 and 3 are replaced with a tetradecanoyl group. The anomeric configurations are α in 3 but β in 4. The two compounds are hence stereo-isomers of each other with regards to the anomeric phosphonooxyethyl groups.

Chemistry The synthesis was performed in principle in a similar way to that reported for lipid A.⁸⁾ A distinct difference is that the phosphonooxyethyl group could be already introduced at the monosaccharide stage because of the chemical stability of this function. In case of lipid A synthesis, the glycosyl phosphate had to be introduced at the final stage just before the deprotection.

A key intermediate 14, which served as a glycosyl acceptor to form a disaccharide 29 with α -phosphonooxyethyl group, was prepared starting from N-(2,2,2-trichloroethoxycarbonyl)-D-glucosamine (5),^{4b)} as shown in Chart 1.

On reaction of 5 with excess ethylene glycol in the presence of hydrogen chloride, the predominant formation of α -glycoside 6 was recognized on thin layer chromatography (TLC), but it was not isolated because of the presence of excess ethylene glycol. Thus, treatment of the reaction mixture with acetic anhydride in pyridine in order to facilitate the removal of ethylene glycol yielded the peracetylated α -glycoside 7 in a 70% yield from 5. After cleavage of the 2,2,2-trichloroethoxycarbonyl (Troc) group of 7 with Zn dust in acetic acid, a (R)-3-benzyloxytetradecanoyl group was introduced to the amino group by the

active-ester method to give the N-benzyloxytetradecanoyl derivative 9a. Or alternatively, a tetradecanoyl group was introduced at the same position by the acid chloride method to afford 9b. Thereafter, all acetoxy protecting groups of 9a and 9b were removed, and the 4- and 6-hydroxyl groups were protected by the isopropylidene formation. The position of the isopropylidene group was confirmed from the nuclear magnetic resonance (NMR) spectrum of 15, obtained by acetylation of 11b. The primary hydroxyl group of 11 was phosphorylated by treatment with diphenylphosphorochloridate to give 12. The 3-hydroxyl groups of 12a and 12b were then acylated with (R)-3-benzyloxytetradecanoic acid and dicyclohexylcarbodiimide (DCC) or with tetradecanoyl chloride in the presence of 4-dimethylaminopyridine (DMAP) respectively. Hydrolysis of the isopropylidene group with 90% acetic acid afforded the glucosamine components 14a and 14b to be used as the glycosyl acceptors.

The corresponding key intermediate 26 with a β -phosphonooxyethyl group, was prepared starting from D-glucosamine β -peracetate 16, as shown in Chart 2.

The oxazoline derivative 17, which can be readily derived from 16, was treated with ethylene glycol monobenzyl ether⁹⁾ in the presence of p-toluenesulfonic acid (p-TsOH) to give the β -glycoside 18 though in a rather low yield. The N-acetyl group of 18 was cleaved by Meerwein's reagent followed by acid hydrolysis. The free amino group of 19 was acylated with tetradecanoyl chloride to give 20. Thereafter, all acetyl groups of 20 were removed and the 4- and 6-hydroxyl functions were protected by the isopropylidene group. The remaining 3-hydroxyl group of the 4,6-isopropylidene derivative 22 was again acylated

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i) BF₃Et₂O; ii) HO(CH₂)₂OBn, p-TsOH; iii) Et₃OBF₄, HCl; iv) R⁵Cl, Et₃N; v) NaOCH₃; vi) (CH₃)₂C(OCH₃)₂, p-TsOH; vii) R⁵Cl, Py, DMAP; viii) H₂, Pd/C; ix) (PhO)₂POCl, Py, DMAP; x) 90% AcOH

Chart 2

c: R^4 = tetradecanoyl, R^5 = 2-(diphenylphosphonooxy)ethoxy, R^6 =H

Chart 3

with tetradecanoyl chloride. The benzyl group of the acylation product 23, was cleaved by hydrogenolysis in acetic acid in water-free conditions to give 24 in an 80% yield, leaving the isopropylidene group intact. This reaction did not proceed when ethanol or tetrahydrofuran (THF) was used as a solvent. The hydroxyl group of 24 was then phosphorylated with diphenylphosphorochloridate to give 25, and hydrolysis with 90% acetic acid afforded the

glucosamine component 26.

The other glucosamine component 28 used as the glycosyl donor was obtained by the treatment of 27^{4b} with HBr-acetic acid. Coupling of the two glucosamine components and the subsequent conversion to 2, 3 and 4 were performed as shown in Chart 3.

Reaction of the bromide (28) with 14a, b or with 26 as an acceptor proceeded in the presence of mercuric cyanide

Table I. Antitumor Activity of Lipid A Analogs against Meth A Fibrosarcoma

Compound number	Dose (μg/mouse)	$T/C (\%)^{a)}$	Mortality ^{b)}	Complete regression ^{c)}
2	100 × 3	14 ^{e)}	1/8	1/7
3	100×3	$12^{e)}$	0/8	2/8
4	100×3	41^{d}	0/8	0/8
1	100×3	$10^{e)}$	0/8	1/8
Control		100	0/8	0/8

a) (Mean tumor weight in tested group/that in control group) \times 100. b) Number of mice died from toxicity/ number of mice tested. c) Number of tumor free-mice/number of mice survived on 21th day. d) p < 0.01. e) p < 0.001 vs. control (Student's test).

to give the desired $\beta(1\rightarrow 6)$ disaccharide **29a**, **b** and **c**, respectively. The Troc group of **29** was cleaved by treatment with Zn dust in acetic acid, and the resultant free amino group was acylated with (*R*)-3-dodecanoyloxytetradecanoic acid by the 1-hydroxybenzotriazole (HOBt) active ester method¹⁰⁾ to give **31**. The benzyl and phenyl groups were removed in two steps by hydrogenolysis with palladium and platinum catalysts, respectively. The crude products **2**, **3** and **4** were purified by preparative TLC (CHCl₃-MeOH- H_2O , 6:4:0.5—0.7) followed by desalting with Dowex 50 (H⁺ type), and the free forms of **2**, **3** and **4** were lyophilized from 0.1% aqueous triethylamine solution to give the respective triethylamine salts.

Antitumor Activity The antitumor activity of synthetic compounds was tested in BALB/c mice. Briefly, a group 8 male mice were inoculated intradermally with Meth A syngenic fibrosarcoma cells (2×10^5) . The triethylamine salt of each of the compounds was dissolved in 5% (v/v) aqueous glucose containing 0.1% triethylamine solution. The resulting solution was then administered to the mice at doses of $100 \,\mu\text{g/mouse}$ through the tail vein on the 7th, 12th and 17th days after the implantation. The percentage antitumor effect on the growth of the Meth A was determined by dividing the average tumor weight of the test group on the 21st day by the average tumor weight of the control group and multiplying the quotient by 100. Table I shows the results.

Discussion

The three synthetic compounds, **2**, **3** and **4**, having a phosphonooxyethyl group at position 1 showed statistically significant antitumor activity in comparison with the untreated control. Among them, **2** and **3**, whose phosphonooxyethyl glycosides occupied the α -configuration, exhibited high activity of the same level as that of *E. colitype* lipid A. By contrast, compound **4** with a β -phosphonooxyethyl group was significantly less active.

We can conclude from this result that the presence of the α -glycosyl phosphate is not essential for the antitumor activity of lipid A. Its substitution with α -phosphonooxyethyl glycoside proved to cause no loss of the activity. The α -orientation of the polar group at position 1 is, however, assumed to be important. Change in the anomeric configuration on this substitution resulted in a significant reduction of the activity as observed in the case of 4, though this compound was still definitely active. It should also be noted that no significant change in activity was

observed by replacement of the 3-hydroxytetradecanoyl groups with tetradecanoyl groups.

This work provided a new possibility in synthesization of a wide variety of antitumor active novel structural analogs of lipid A by replacing not only the acyl groups as already mentioned but also the phosphate function(s). Many of the stable polar groups might also serve as candidates for the substituents. Though our preliminary observation suggested that the present phosphonooxyethyl derivatives could unfortunately still be toxic, we expect that production of non-toxic lipid A analogs would be possible through synthetic approaches in this line.

Experimental

All melting points are uncorrected. Proton nuclear magnetic resonance ($^1\mathrm{H-NMR}$) spectra were determined on a Varian XL-200 spectrometer (200 MHz) in chloroform-d solutions unless otherwise noted. The chemical shifts are given in δ values with tetramethylsilane (TMS) as the internal standard. Optical rotations were measured with a Horiba SEDA-200 polarimeter at 25 °C. Mass spectra (MS) were obtained on a JMS-HX 110 or JMX-300 instrument. Precoated Silica gel 150 A PLK5F plates (1.0 mm thickness; Whatman) were used for preparative TLC. Organic solutions were dried over sodium sulfate.

2-Acetoxyethyl 3,4,6-Tri-O-acetyl-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)-α-D-glucopyranoside (7) To ethylene glycol (5.0 ml, 88 mmol) and 5 M HCl dioxane solution (0.5 ml) was added 5 (5.00 g, 14 mmol), and the mixture was stirred at 90 °C for 4h. After cooling with ice-water, pyridine (75 ml) and acetic anhydride (30.6 g, 300 mmol) were added and the mixture was stirred under cooling for 20 min. Stirring was then continued at room temperature for an additional 16h. The reaction mixture was poured into 350 ml of ice-water and stirred. The precipitate was collected by filtration and was washed with water. The resulting solid was dissolved in CHCl3, and the solution was washed with 1 m HCl, then with saturated aqueous NaCl, and dried. After evaporation of the solvent, the residue was recrystallized from ethanol to give 7 (5.64 g, 71%) as colorless prisms. mp 138—140 °C. $[\alpha]_D$ +74.0° $(c=1.2, CHCl_3)$. Anal. Calcd for C₁₉H₂₆Cl₃NO₁₂: C, 40.26; H, 4.62; N, 2.47. Found: C, 40.48; H, 4.75; N, 2.51. IR (KBr): 3340, 1755, 1540 cm⁻¹. NMR δ : 2.02 (3H, s, OAc), 2.04 (3H, s, OAc), 2.12 (6H, s, OAc × 2), 3.7—4.3 (8H, m), 4.67 and 4.85 (each 1H, AB type d, $J=12\,\text{Hz}$, CH_2CCl_3), 4.97 (1H, t, J=4 Hz, H-1), 5.16 (1H, t, J=10 Hz, H-3 or 4) 5.31 (1H, t, J=10 Hz, H-3 or 4). MS m/z: 568 $\lceil (M+2)^+ \rceil$.

2-Acetoxyethyl 3,4,6-Tri-O-acetyl-2-[(R)-3-benzyloxytetradecanoylamino]-2-deoxy-α-D-glucopyranoside (9a) To a solution of 7 (3.00 g, 5.29 mmol) in AcOH (40 ml) was added Zn dust (3.00 g), and the mixture was vigorously stirred for 1 h at room temperature. The insoluble materials were filtered off, and the filtrate was concentrated *in vacuo*. The resulting oily product was dissolved in CHCl₃, and the solution was washed with 5% aqueous NaHCO₃, and dried. The solvent was distilled off at reduced pressure to give an oil.

Separately, DCC (1.32 g, 6.40 mmol) was added to a solution of (R)-3-benzyloxytetradecanoic acid (1.95 g, 5.82 mmol) and HOBt (0.98 g, 6.40 mmol) in THF (30 ml) with ice-cooling, and the mixture was stirred for 3 h at room temperature. The precipitate was filtered off to give an active ester solution. The active ester solution was added to the oily amine prepared above, and N-methylmorpholine (0.7 ml, 64 mmol) was added to the solution with ice cooling. The mixture was stirred for 3 h at room temperature. After evaporation of the solvent, the residue was purified by silica gel column chromatography and eluted with CHCl₃-acetone (25:1–12:1) to give **9a** (3.70 g, 99%) as an oil. NMR δ : 0.87 (3H, t, J=6 Hz), 1.26 (18H, s, CH₂), 2.00 (3H, s, OAc), 2.04 (3H, s, OAc), 2.08 (6H, s, OAc), 2.18 (2H, COCH₂), 4.49 and 4.59 (each 1H, AB type d, J=12 Hz, OCH₂C₆H₅), 4.76 (1H, d, J=4 Hz, H-1), 5.10 (2H, m), 6.30 (1H, d, J=10 Hz, NH), 7.36 (5H, s, arom. H).

Acetoxyethyl 3,4,6-Tri-O-acetyl-2-deoxy-2-tetradecanoylamino- α -D-glucopyranoside (9b) To a solution of 7 (4.96 g, 8.75 mmol) in AcOH (60 ml) was added Zn dust (7.00 g), and the mixture was vigorously stirred for 1 h at room temperature. The insoluble materials were removed by filtration, and the filtrate was concentrated *in vacuo*. The residue was dissolved in dioxane, dioxane containing dry HCl was added to the solution, and the solvent was removed by evaporation. The resulting oily product was dissolved in CH₂Cl₂ (70 ml), and N-methylmorpholine

(2.88 ml, 26.2 mmol) and tetradecanoyl chloride (3.24 g, 13.1 mmol) were added to the solution with ice-cooling. After stirring for 1 h, MeOH (10 ml) was added to the reaction mixture. After stirring for 10 min, the reaction mixture was diluted with CHCl₃, washed with 1 m HCl, then with saturated aqueous NaCl, and dried. After evaporation of the solvent, the residue was purified by silica gel column chromatography (benzene–EtOAc, 9:1—1:1) to give **9b** (4.77 g, 91%) as a colorless oil. NMR δ : 0.88 (3H, t), 1.26 (20H, s, CH₂), 2.02 (3H, s, OAc), 2.05 (3H, s, OAc), 2.13 (6H, s, OAc), 2.18 (2H, COCH₂), 3.7—4.5 (8H, m), 4.91 (1H, d, J=4 Hz, H-1), 5.15 (1H, t, J=10 Hz, H-3 or 4), 5.27 (1H, t, J=10 Hz, H-3 or 4), 5.78 (1H, d, J=10 Hz, NH). MS m/z: 601 (M⁺), 602 [(M+H)⁺].

2-Hydroxyethyl 2-[(R)-3-Benzyloxytetradecanoylamino]-2-deoxy-α-D**glucopyranoside (10a)** To a solution of **9a** (3.68 g, 5.20 mmol) in dry MeOH (30 ml), a MeOH solution (3 ml) containing freshly prepared MeONa (ca. 2.8 mmol) was added, and the mixture was stirred for 20 min at room temperature. The solution was neutralized with Amberlist-15, and the resin was filtered off. The filtrate was concentrated at reduced pressure. The residue was washed with Et₂O to give **10a** (2.49 g, 89%) as a pale brown powder, which was recrystallized from EtOH–H₂O. mp 125—127 °C. [α]_D +73.3° (c=0.9, MeOH). *Anal*. Calcd for C₂₉H₄₉NO₈: C, 64.54; H, 9.15; N, 2.60. Found: C, 64.33; H, 8.86; N, 2.59. IR (KBr): 3310, 1645, 1555 cm⁻¹. NMR δ: 0.90 (3H, t, J=7 Hz), 1.30 (18H, s, CH₂), 2.43 (1H, dd, J=6, 14 Hz, NCOCH₂), 2.58 (1H, dd, J=8, 14 Hz, COCH₂), 4.58 (2H, OCH₂C₆H₅), 4.73 (1H, d, J=4 Hz, H-1).

2-Hydroxyethyl 2-Deoxy-2-tetradecanoylamino-α-D-glucopyranoside (10b) Compound **9b** (4.77 g, 7.93 mmol) was treated with MeONa (*ca.* 9 mmol) in the same manner as described for **10a** to give **10b** (3.02 g, 88%) as a white solid. Recrystallization from EtOH–H₂O gave a purified product. mp 158—160 °C. [α]_D +82.1° (c=0.8, THF–H₂O, 4:1). *Anal.* Calcd for C₂₂H₄₃NO₇: C, 60.94; H, 10.00; N, 3.23. Found: C, 61.09; H, 9.95; N, 3.25. IR (KBr): 3300 (br), 1645, 1555 cm⁻¹. NMR δ: 0.90 (3H, t), 1.28 (20H, s, CH₂), 2.25 (2H, t, J=7 Hz, COCH₂), 4.79 (1H, d, J=4 Hz, H-1). MS m/z: 434 [(M+H)⁺].

2-Hydroxyethyl 2-[(R)-3-Benzyloxytetradecanoylamino]-2-deoxy-4,6-O-isopropylidene-α-D-**glucopyranoside (11a)** To a solution of **10a** (1.20 g, 2.22 mmol) in dimethylformamide (DMF) (5 ml) were added p-TsOH·H₂O (38 mg) and 2,2-dimethoxypropane (0.69 g, 6.66 mmol), and the mixture was stirred for 5 h at room temperature. After neutralization with 5% aqueous NaHCO₃, the solvent was removed by distillation at reduced pressure. The residue was dissolved in EtOAc, washed with H₂O, then with saturated aqueous NaCl, and dried. After the evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃– MeOH, 19:1) to give **11a** (0.98 g, 76%) as a colorless viscous oil. [α]_D +31.4° (c=0.9, CHCl₃). NMR δ: 0.89 (3H, t, J=6 Hz), 1.28 (18H, s, CH₂), 1.45 (3H, s, CH₃), 1.56 (3H, s, CH₃), 2.50 (2H, dd, COCH₂), 4.20 (1H, m, H-2), 4.51 and 4.63 (each 1H, AB type d, J=12 Hz, CH₂C₆H₅), 4.70 (1H, d, J=4 Hz, H-1), 6.94 (1H, d, J=8 Hz, NH), 7.40 (5H, s, arom. H). MS m/z: 580 (M⁺), 546 [(M-16)⁺].

2-Hydroxyethyl 2-Deoxy-4,6-*O*-isopropylidene-2-tetradecanoylamino-α-D-glucopyranoside (11b) As described for 11a, compound 10b (0.87 g, 2.0 mmol) was reacted with dimethoxypropane to give 11b (0.78 g, 82%) as a colorless viscous oil. NMR δ: 0.88 (3H, t), 1.26 (20H, s, CH₂), 1.46 (3H, s, CH₃), 1.56 (3H, s, CH₃), 2.16 (2H, t, J=7 Hz), 3.6—3.9 (9H, m), 4.10 (1H, m), 4.89 (1H, d, J=4 Hz, H-1). MS m/z: 458 [(M-15) $^+$].

2-Acetoxyethyl 3-O-Acetyl-2-deoxy-4,6-O-isopropylidene-2-tetradecanovlamino-α-D-glucopyranoside (15) To a solution of 11b (0.12 g, 0.25 mmol) in $\mathrm{CH_2Cl_2}$ (7 ml) were added pyridine (1.5 ml) and acetic anhydride (0.21 ml, 2.10 mmol), and the mixture was stirred for 21 h at room temperature. The reaction mixture was diluted with CHCl₃, and the solution was washed with 1 M HCl, then with saturated aqueous NaCl, and dried. After evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃-acetone, 19:1) to give 15 (0.14 g, quant.) into a waxy solid. Recrystallization from MeOH-H₂O gave the purified product. mp 58-60 °C. Anal. Calcd for C29H51NO9: C, 62.45; H, 9.22; N, 2.51. Found: C, 62.06; H, 8.91; N, 2.50. IR (KBr): 3300, 1745, 1650, 1550 cm⁻¹. NMR δ : 0.88 (3H, t, J=6 Hz), 1.26 (20H, s, CH₂), 1.38 (3H, s, CH₃), 1.48 (3H, s, CH₃), 2.06 (3H, s, OAc), 2.11 (3H, s, OAc), 2.16 (2H, t, J=7 Hz, COCH₂), 3.6—3.9 (6H, m, H-4, 5, 6 and OCH_2CH_2), 4.30 (3H, m, H-2 and CH_2OAc), 4.83 (1H, d, J=4 Hz, H-1), 5.18 (1H, m, H-3), 5.85 (1H, d, J = 10 Hz, NH).

2-(Diphenylphosphonooxy)ethyl 2-[(R)-3-Benzyloxytetradecanoylamino]-2-deoxy-4,6-O-isopropylidene- α -D-glucopyranoside (12a) To a solution of 11a (0.83 g, 1.43 mmol), pyridine (0.17 ml, 2.15 mmol) and DMAP (0.26 g, 2.15 mmol) in CH₂Cl₂ (8 ml) was added diphenylphosphoro-

chloridate (0.42 g, 1.57 mmol) with ice cooling. After the mixture was stirred for 4h at room temperature, MeOH (3 ml) was added. After stirring for a while, the solvent was removed by evaporation. The residue was purified by silica gel column chromatography (CHCl₃–acetone, 19:1) to give **12a** (0.98 g, 84%) as a colorless oil. NMR δ : 0.88 (3H, t, J=7 Hz), 1.25 (18H, s, CH₂), 1.46 (3H, s, CH₃), 1.53 (3H, s, CH₃), 2.47 (2H, d, J=6 Hz, COCH₂), 4.2 (3H, m, H-2 and CH₂OP), 4.49 and 4.57 (each 1H, AB type d, J=12 Hz, CH₂C₆H₅), 4.64 (1H, d, J=4 Hz, H-1), 7.2—7.4 (15H, m, arom, H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-4,6-*O***-isopropylidene-2-tetradecanoylamino-** α -**D-glucopyranoside (12b)** In the same manner as described for **12a**, compound **11b** (0.77 g, 1.63 mmol) was reacted with diphenylphosphorochloridate to give **12b** (0.81 g, 71%) as a colorless viscous oil. NMR δ : 0.88 (3H, t), 1.24 (20H, s, CH₂), 1.46 (3H, s, CH₃), 1.56 (3H, s, CH₃), 2.17 (2H, t, J=8 Hz), 3.6—4.0 (m), 4.2 (1H, m, H-2), 4.45 (2H, m, CH₂OP), 4.79 (1H, d, J=4 Hz), 7.2—7.4 (10H, m, arom. H). MS m/z: 706 (M⁺).

2-(Diphenylphosphonooxy)ethyl 3-O-[(R)-3-Benzyloxytetradecanoyl]-2-[(R)-3-benzyloxytetradecanoylamino]-2-deoxy-α-D-glucopyranoside (14a) To a solution of 12a (0.96 g, 1.18 mmol) and (R)-3-benzyloxytetradecanoic acid (0.59 g, 1.78 mmol) in CH₂Cl₂ (10 ml) were added DMAP (30 mg, 0.24 mmol) and DCC (0.39 g, 1.90 mmol) with ice cooling. After the mixture was stirred for 18 h at room temperature, the insoluble materials were removed by filtration. The filtrate was washed with 0.2 m HCl, then with saturated aqueous NaCl, and dried. After the evaporation of the solvent, 90% AcOH (30 ml) was added to the residue, followed by stirring for 30 min at 90 °C. The solvent was distilled off, and the resulting residue was purified by silica gel column chromatography (CHCl₃-MeOH, 19:1) to give 14a (1.23 g, 96%). $[\alpha]_D$ +31.4° (c=1.2, CHCl₃). NMR δ : 0.88 (6H, t, J = 7 Hz), 1.28 (s, CH_2), 2.34 (2H, d, J = 6 Hz, $NCOCH_2$), 2.6 (2H, dd, COCH₂), 4.2—4.3 (3H, m, H-2 and CH₂OP), 4.47 and 4.55 (each 1H, AB type d, J = 12 Hz, $CH_2C_6H_5$, 4.56 (2H, s, $CH_2C_6H_5$), 4.71 (1H, d, J=4 Hz, H-1), 5.13 (1H, m, H-3), 6.83 (1H, d, J=10 Hz, NH), 7.2—7.4 (20H, m, arom. H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-3-O-tetradecanoyl-2-tetradecanoylamino-α-D-glucopyranoside (14b) To a solution of 12b (0.81 g, 1.15 mmol) in CH₂Cl₂ (10 ml) were added pyridine (0.45 ml, 5.75 mmol), DMAP (50 mg) and tetradecanoyl chloride (0.45 g, 1.84 mmol) with ice cooling. The mixture was stirred for 30 min at 0 °C and then for 30 min at room temperature. After the addition of MeOH (2 ml) and stirring for a few minutes, the mixture was diluted with CHCl₃. The solution was washed with 1 M HCl, then with saturated aqueous NaCl, and dried. After evaporation of the solvent, 90% AcOH (30 ml) was added to the residue. The mixture was stirred for 40 min at 90 °C, and then the solvent was distilled off. The resulting residue was purified by silica gel column chromatography (CHCl₃-MeOH, 19:1) to give 14b (0.92 g, 92%) as a semi-solid. $[\alpha]_D + 40.8^\circ$ (c = 1.1, CHCl₃). NMR δ : 0.90 (6H, t), 1.24 (s, CH_2), 2.07 (2H, t, J=8 Hz, $COCH_2$), 2.34 (2H, t, J=8 Hz, $COCH_2$), 3.7—4.0 (6H, m), 4.4 (3H, m, H-2 and CH₂OP), 4.84 (1H, d, J=4 Hz, H-1), 5.09 (1H, m, H-3), 6.24 (1H, d, J=9 Hz, NH), 7.2—7.4 (10H, m, arom. H). MS m/z: 876 (M+).

2-Benzyloxyethyl 2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy-β-D-glucopyranoside (18) To a suspension of CaSO₄ (8.2 g) in CH₂Cl₂ (90 ml) were added BF₃·Et₂O (3.79 ml, 30 mmol) and 16 (7.78 g, 20 mmol) at room temperature. After the reaction mixture was stirred for 2.5 h, an additional BF₃·Et₂O (1.26 ml, 10 mmol) was added and the mixture was stirred for 16.5 h at room temperature. The mixture was poured into 250 ml of 5.2% aqueous NaHCO3 and extracted with CHCl3. The extract was washed with saturated aqueous NaCl and dried. The solvent was evaporated in vacuo to give a viscous oil. To a solution of the resultant oil and ethylene glycol monobenzylether (6.09 g, 40 mmol) in dry CHCl₃ (90 ml) was added p-TsOH H₂O (190 mg, 1.00 mmol). This mixture was refluxed for 6.5 h and washed with 5% aqueous NaHCO3, then with saturated aqueous NaCl, and dried. After evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃-acetone, 19:1-7:1). The resulting residue was washed with Et₂O to give 18 (3.60 g, 37%). Recrystallization from MeOH-H₂O gave a purified product of 18. mp 106-107 °C. $[\alpha]_D - 17.0$ ° (c = 1.2, CHCl₃). Anal. Calcd for $C_{23}H_{31}NO_{10}$: C, 57.37; H, 6.49; N, 2.91. Found: C, 57.32; H, 6.36; N, 2.95. IR (KBr): 3250, 1750, 1650, 1575 cm⁻¹. NMR δ : 1.86 (3H, s, OAc), 2.04 (6H, s, $OAc \times 2$), 2.09 (3H, s, OAc), 3.6—4.3 (8H, m), 4.58 (2H, s, $CH_2C_6H_5$), 4.78 (1H, d, J=8 Hz, H-1), 5.11 (1H, t, J=10 Hz, H-3), 5.28 (1H, t, J = 10 Hz, H-4), 5.56 (1H, d, J = 10 Hz, NH), 7.38 (5H, s, arom. H). MS

2-Benzyloxyethyl 3,4,6-Tri-O-acetyl-2-deoxy-2-tetradecanoylamino-β-

D-glucopyranoside (20) To a solution of **18** (1.50 g, 3.1 mmol) in CH₂Cl₂ (30 ml) were added K₂CO₃ (1.55 g, 11.2 mmol) and freshly prepared Et₃O·BF₄ at room temperature. After stirring for 17.5 h, the mixture was washed with H2O, then with saturated aqueous NaCl, and dried. Evaporation of the solvent gave an oil. This oil was dissolved in THF (30 ml), and 1 m HCl (3.2 ml) was added to the solution with ice cooling. The solution was stirred for 40 min at room temperature and concentrated in vacuo to give 19 (1.56 g) as white powder. As described for 9a, compound 19 was reacted with tetradecanoyl chloride (0.92 g, 3.7 mmol) to give 20 (1.11 g, 55%) as a white powder. Recrystallization from benzene-hexane gave a purified product of 20. mp 101-102 °C. $[\alpha]_D$ -5.4° (c=0.9,CHCl₃). Anal. Calcd for C₃₅H₅₅NO₁₀: C, 64.69; H, 8.53; N, 2.16. Found: C, 64.69; H, 8.33; N, 2.27. NMR δ : 0.88 (3H, t, J = 6 Hz, CH₃), 1.26 (20H, s, CH₂), 2.02, 2.04 and 2.09 (11H, 3s, OAc×3 and COCH₂), 3.6—4.1 (6H, m), 4.15 (1H, dd, J=12, 2Hz, H-6), 4.82 (1H, dd, J=12, 5 Hz, H-6), 4.58 (2H, s, $\underline{CH}_2C_6H_5$), 4.82 (1H, d, J=8 Hz, H-1), 5.12 (1H, t, J = 10 Hz, H-3), 5.31 (1H, t, J = 10 Hz, H-4), 5.45 (1H, d, J = 8 Hz, H-4)NH), 7.38 (5H, s, arom. H). MS m/z: 649 (M⁺).

2-Benzyloxyethyl 2-Deoxy-2-tetradecanoylamino-β-D-glucopyranoside (21) In a manner similar to that for 10a, compound 20 (1.05 g, 1.6 mmol) was treated with MeONa (ca. 2.2 mmol) in MeOH (20 ml) to give 21 (0.74 g, 87%) as a white powder. Recrystallization from EtOH–H₂O gave a purified product of 21. mp 155—156 °C. [α]_D –18.1° (c=0.5, MeOH). *Anal.* Calcd for C₂₉H₄₉NO₇: C, 66.51; H, 9.43; N, 2.67. Found: C, 66.63; H, 9.43; N, 2.75. IR (KBr): 3451, 3390, 1665, 1535 cm⁻¹. NMR δ: 0.88 (3H, t, J=6 Hz), 1.28 (20H, s, CH₂), 2.04 (2H, t, J=7 Hz), 3.5—4.1 (m), 4.55 (1H, d, J=8 Hz, H-1), 4.62 (2H, s, $\underline{\text{CH}}_2\text{C}_6\text{H}_5$), 7.40 (5H, s, arom. H). MS m/z: 525 [(M+H)⁺].

2-Benzyloxyethyl 2-Deoxy-4,6-*O*-isopropylidene-2-tetradecanoylamino-β-D-glucopyranoside (22) As described for 11, compound 21 (710 mg, 1.36 mmol) was reacted with dimethoxypropane (0.42 g, 4.1 mmol) in CH₂Cl₂ (30 ml) in the presence of p-TsOH·H₂O to give 22 (710 mg, 93%) as a waxy solid. NMR δ : 0.88 (3H, t, J=6 Hz), 1.26 (20H, s, CH₂), 1.46 (3H, s, CH₃), 1.56 (3H, s, CH₃), 2.02 (2H, t, J=8 Hz), 3.3 (1H, m), 3.5—4.1 (9H, m), 4.62 (1H, d, J=8 Hz, H-1), 4.62 (2H, s, CH₂C₆H₅), 7.38 (5H, m, arom. H). MS m/z: 565 [(M+H)⁺].

2-Benzyloxyethyl 2-Deoxy-4,6-*O***-isopropylidene-3-***O***-tetradecanoyl-2-tetradecanoylamino-**β**-D-glucopyranoside (23)** As described for **13b**, compound **22** (0.70 g, 1.24 mmol) was treated with tetradecanoyl chloride (0.46 g, 1.86 mmol) to give **23** (0.79 g, 82%). Recrystallization from hexane gave a purified product of **23** mp 88—89 °C. [α]_D -29.0° (c=0.8, CHCl₃). *Anal.* Calcd for C₄₆H₇₉NO₈: C, 71.37; H, 10.29; N, 1.81. Found: C, 71.32; H, 10.19; N, 1.82. IR (KBr): 1740, 1665, 1530 cm⁻¹. NMR δ: 0.89 (6H, t, J=7 Hz), 1.26 (40H, s, CH₂), 1.38 (3H, s, CH₃), 1.47 (3H, s, CH₃), 2.05 (2H, t, J=8 Hz), 2.32 (2H, m), 3.38 (1H, m), 3.6—4.2 (8H, m), 4.57 (2H, s, CH₂C₆H₅), 4.62 (1H, d, J=8 Hz, H-1), 5.08 (1H, t, J=10 Hz, H-3), 5.54 (1H, d, J=10 Hz, NH), 7.38 (5H, m, arom. H). MS m/z: 774 (M⁺).

2-Hydroxyethyl 2-Deoxy-4,6-O-isopropylidene-3-O-tetradecanoyl-2tetradecanoylamino-β-D-glucopyranoside (24) A suspension of 23 (745 mg, 0.96 mmol) in AcOH (50 ml) was hydrogenolyzed over 5% palladium carbon as a catalyst at room temperature for 15 h (atmospheric pressure). The catalyst was removed by filtration, and the filtrate was freeze-dried to give a powder. The catalyst on the filter was washed with CHCl₃. The freezed-dried powder was dissolved in the CHCl₃ washing and the resulting solution was washed with 5% aqueous NaHCO3, then with H₂O, and dried. After evaporation of the solvent, the residue was purified by short silica gel column chromatography (CHCl₃-acetone, 9:1) to give 24 (529 mg, 80%) as a white powder, which gradually softened on heating without showing a definite melting point. IR (KBr): 3320, 1740, 1660, 1535 cm⁻¹. NMR δ : 0.89 (6H, t, J = 7 Hz), 1.26 (40H, s, CH₂), 1.38 (3H, s, CH_3), 1.48 (3H, s, CH_3), 2.15 (2H, t, J=7 Hz), 2.33 (2H, m), 3.40 (1H, m), 3.7-4.2 (9H, m), 4.50 (1H, d, J=8 Hz, H-1), 5.08 (1H, t, J=10 Hz, H-3), 5.28 (1H, d, J = 8 Hz, NH). MS m/z: 684 (M⁺).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-3-*O*-tetradecanoyl-2-tetradecanoylamino-β-D-glucopyranoside (26) As described for **12a** and **14a**, compound **24** (515 mg, 0.75 mmol) was reacted with diphenylphosphorochloridate (304 mg, 1.13 mmol), and the resulting oily **25** was treated with 90% AcOH to give **26** (491 mg, 74%) as an oil. [α]_D -19.5° (c=0.9, CHCl₃). NMR δ: 0.88 (6H, t, J=7 Hz), 1.24 (40H, s, CH₂), 2.08 (2H, t, J=8 Hz), 2.36 (2H, t, J=8 Hz), 3.40 (1H, m), 3.6—4.0 (8H, m), 4.20 (2H, m, CH₂OP), 4.62 (1H, d, J=8 Hz, H-1), 5.02 (3H, t, J=10 Hz, H-3), 5.96 (1H, d, J=8 Hz, NH), 7.2—7.4 (10H, m, arom. H). MS m/z: 875 (M⁺).

2-(Diphenylphosphonooxy)ethyl 3-O-[(R)-3-Benzyloxytetradecanoyl]-2-[(R)-3-benzyloxytetradecanoylamino]-2-deoxy-6-O-[2-deoxy-4-O-di-

phenylphosphono-3-O-[(R)-3-tetradecanovloxytetradecanovl]-6-O-(2.2.2trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)- β -Dglucopyranosyl]-α-D-glucopyranoside (29a) To a solution of 27 (450 mg, 0.36 mmol) in CH₂Cl₂ (2 ml) was added 25% HBr-AcOH (6 ml), and the mixture was stirred for 1.5h at room temperature. After the reaction mixture was diluted with CHCl₃, the solution was washed successively with ice-water, 5% aqueous NaHCO3 and saturated aqueous NaCl, and dried. Evaporation of the solvent gave 28 as an oil. The oil and 14a (392 mg, 0.36 mmol) were dissolved in CH₂Cl₂ (5 ml). To the soltuion were added CaSO₄ (0.5 g) and mercuric cyanide (182 mg, 0.72 mmol), and the mixture was refluxed for 14h. The insoluble materials were removed by filtration through Celite 545, and the filtrate was washed with 5% aqueous potassium iodide, then with saturated aqueous NaCl, and dried. After evaporation of the solvent, the residue was purified by silica gel column chromatography (benzene-EtOAc, 9:1-2:1) to give 29a (554 mg, 67%) as a colorless oil. NMR δ : 0.89 (12H, t, J = 7 Hz), 1.28 (s, CH₂), 2.2—2.7 (8H, m, COCH₂ × 4), 5.66 (1H, m, H'-3), 7.2—7.4 (30H,

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)- β -D-glucopyranosyl]-3-O-tetradecanoyl-2-tetradecanoylamino- α -D-glucopyranoside (29b) As was described for 29a, compound 28, obtained from 27 (400 mg, 0.32 mmol), was reacted with 14b (276 mg, 0.32 mmol) to give 29b (537 mg, 81%) as an oil. NMR δ : 0.88 (12H, t), 1.24 (78H, s, CH₂), 2.07 (2H, t, J=8 Hz), 2.3 (6H, m), 4.63 and 4.76 (AB type d, J=12 Hz), 5.63 (1H, m), 7.2—7.4 (20H, m, arom. H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphospho-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)- β -D-glucopyranosyl]-3-O-tetradecanoyl-2-tetradecanoylamino- β -D-glucopyranoside (29c) As described for 29a, compound 28 (ca. 0.32 mmol) was treated with 26 (280 mg, 0.32 mmol) to give 29c (522 mg, 79%) as a colorless viscous oil. NMR δ : 0.89 (12H, t, J=7 Hz), 1.24 (78H, s, CH₂), 2.07 (2H, t), 2.22 (2H, t), 2.38 (4H, m), 5.23 (1H, m), 5.58 (1H, t, J=10 Hz), 7.2—7.4 (20H, m, arom. H).

2-(Diphenylphosphonooxy)ethyl 3-O-[(R)-3-Benzyloxytetradecanoyl]-2-[(R)-3-benzyloxytetradecanoylamino]-2-deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-2-[(R)-3-dodecanoyloxytetradecanoylamino]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]- β -D-glucopyranosyl]- α -D-glucopyranoside (31a) As described for 9a, compound 29a (520 mg, 0.32 mmol) was treated with Zn dust, and the resulting oil was condensed with the HOBt active ester of (R)-3-dodecanoyloxytetradecanoic acid (149 mg, 0.35 mmol) to give 31a (327 mg, 61%) as a colorless oil. NMR δ : 0.89 (18H, t, J= 7 Hz), 1.28 (108H, s, CH₂), 2.2—2.7 (12H, m, COCH₂ × 6), 5.58 (1H, m, H-3), 7.2—7.4 (30H, m, arom. H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-2-[(R)-3-dodecanoyloxytetradecanoylamino]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]- β -D-glucopyranosyl]-3-O-tetradecanoyl-2-tetradecanoylamino- α -D-glucopyranoside (31b) In the manner described for 9a, compound 29b (527 mg, 0.26 mmol) was treated with Zn powder, and the resulting oil was condensed with the HOBt active ester of (R)-3-dodecanoyloxytetradecanoic acid (166 mg, 0.39 mmol) to give 31b (286 mg, 53%) as an oil. [α]_D +13.3° (c=0.5, CHCl₃). NMR δ : 0.88 (18H, t), 1.24 (112H, s, CH₂), 2.03 (2H, m), 2.3 (10H, m), 5.57 (1H, m), 7.2—7.4 (20H, m, arom, H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-2-[(R)-3-dodecanoyloxytetradecanoylamino]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]- β -D-glucopyranosyl]-3-O-tetradecanoyl-2-tetradecanoylamino- β -D-glucopyranoside (31c) As in the case of 9a, compound 29c (509 mg, 0.25 mmol) was condensed with Zn dust, and the resulting oil was reacted with the HOBt active ester of (R)-3-dodecanoyloxytetradecanoic acid (160 mg, 0.37 mmol) to give 31c (286 mg, 55%) as a colorless viscous oil. [α]_D -16.5° (c=0.7, CHCl₃). NMR δ : 0.88 (18H, t, J=7 Hz), 1.24 (112H, s, CH₂), 2.05 (2H, m), 2.3 (10H, m), 5.56 (1H, t, J=10 Hz), 7.2—7.4 (20H, m, arom. H).

2-Phosphonooxyethyl 2-Deoxy-6-O-[2-deoxy-2-[(R)-3-dodecanoyloxytetradecanoylamino]-4-O-phosphono-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-β-D-glucopyranosyl]-3-O-[(R)-3-hydroxytetradecanoyl]-2-[(R)-3-hydroxytetradecanoylamino]-α-D-glucopyranoside (2) Compound 31a (314 mg, 0.13 mmol) was dissolved in AcOH (12 ml) and hydrogenolyzed in the presence of 5% palladium—carbon for 6h at room temperature (atmospheric pressure). The catalyst was removed by filtration and the filtrate was condensed by evaporation *in vacuo*. The residue was hydrogenolyzed over platinum dioxide (atmospheric pressure), and

the resulting powder was purified by preparative TLC using CHCl₃–MeOH–H₂O (6:4:0.7). After extraction with CHCl₃–MeOH–H₂O–Et₃N (6:4:1:0.02), the solvent was evaporated *in vacuo*. The residue was dissolved in CHCl₃–MeOH–H₂O (6:4:1), and the solution was desalted with Dowex 50 (H⁺ type). The desalted solution was concentrated, and the residue was lyophilized from 0.1% aqueous Et₃N to give the Et₃N salt of **2** (141 mg) as a white powder. The physical data given below are those of the free acid form of **2**. mp 144–147 °C (dec.). [α]_D +9.8° (c=0.5, CHCl₃–MeOH, 3:1). IR (KBr): 1740, 1660 cm⁻¹. NMR (CDCl₃–CD₃OD, 1:1) &: 0.90 (18H, t, J=6Hz), 1.30 (108H, s, CH₂), 2.3–2.7 (12H, m, COCH₂×6), 5.2 (4H, m, CHCO×4).

2-Phosphqnooxyethyl 2-Deoxy-6-O-[2-deoxy-2-[(R)-3-dodecanoyloxytetradecanoylamino]-4-O-phosphono-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-β-D-glucopyranosyl]-3-O-tetradecanoyl-2-tetradecanoylaminoα-D-glucopyranoside (3) Compound 31b (260 mg, 0.12 mmol) was dissolved in THF (15 ml) and hydrogenolyzed with the mixture of platinum dioxide (290 mg) and 5% palladium-carbon (140 mg) as a catalyst at room temperature for 38 h (atmospheric pressure). The catalyst was removed by filtration, and washed with CHCl₃-MeOH-H₂O (6:4:0.5). The filtrate and the washing solution were combined, and the solvent was evaporated at reduced pressure. The residue was purified by preparative TLC (CHCl₃-MeOH-H₂O, 6:4:0.5), and the resultant extracted solution, in the same manner as described for 2, was desalted with Dowex 50 (H+ type). A portion of the desalted solution was concentrated, and the residue was lyophilized from dioxane suspension to give 3 (20 mg) as a white powder. mp 163—167 °C (dec.). $[\alpha]_D$ +15.0° (c=0.6, CHCl₃-MeOH, 3:1). Anal. Calcd for C₉₆H₁₈₂N₂O₂₄P₂·2H₂O: C, 62.45; H, 10.15; N, 1.52. Found: C, 62.13; H, 9.87; N, 1.52. IR (KBr): 1735, 1660, 1565 cm⁻¹. NMR (CDCl₃-CD₃OD, 1:1) δ : 0.90 (18H, t, J=6 Hz), 1.30 (112H, s, CH₂), 2.19 (2H, m), 2.3—2.4 (8H, m), 2.65 (2H, m), 5.2 (m).

The residual desalted solution was adjusted with $CHCl_3$ – $MeOH-H_2O-Et_3N$ (6:4:1:0.02) to about pH 8 with ice-cooling. After evaporation of the solvent, the residue was lyophilized from 0.1% aqueous Et_3N to give 36 mg of Et_3N salt of 3 as a white powder.

2-Phosphonooxyethyl 2-Deoxy-6-O-[2-deoxy-2-[(R)-3-dodecanoyloxytetradecanoylamino]-4-O-phosphono-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]- β -D-glucopyranosyl]-3-O-tetradecanoyl-2-O-tetradecanoylamino- β -D-glucopyranoside (4) In the same manner as described for 2, compound 31c (260 mg, 1.23 mmol) was hydrogenolyzed in the presence of a catalyst, and the resulting powder was purified by preparative TLC (CHCl₃-MeOH-H₂O, 6:4:0.6) and desalted. The product was lyophilized from 0.1% aqueous Et₃N to give the Et₃N salt of 4 (115 mg) as a white powder. mp 145—150 °C (dec.). [α]_D –11.8° (c=0.5, CHCl₃-MeOH, Et₃N salt of 4). IR (KBr): 1730, 1635, 1560 cm⁻¹. NMR (CDCl₃-CD₃OD, 1:1) δ : 0.89 (18H, t), 1.26 (112H, s, CH₂), 2.06 (2H, m), 2.3—2.5 (8H, m), 2.66 (2H, m).

References

- C. Galanos, O. Luderitz, E. Th. Rietschel, and O. Westphal, "International Review of Biochemistry, Biochemistry of Lipids II," Vol. 14, ed. by T. W. Goodwin, University Park Press, Baltimore, 1977, Chapter 6; O. Westphal, O. Luderitz, C. Galanos, H. Mayer, and E. Th. Rietschel, Adv. Immunopharmacol., 3, 13 (1986).
- 2) O. Westphal and O. Luderitz, Angew. Chem., 66, 407 (1954).
- M. Imoto, S. Kusumoto, T. Shiba, H. Naoki, T. Iwashita, E. Th. Rietschel, H.-W. Wollenweber, C. Galanos, and O. Luderitz, *Tetrahedron Lett.*, 24, 4017 (1983); M. Imoto, S. Kusumoto, T. Shiba, E. Th. Rietschel, C. Galanos, and O. Luderitz, *ibid.*, 26, 907 (1985).
- a) M. Imoto, H. Yoshimura, S. Kusumoto, and T. Shiba, *Proc. Jpn. Acad.*, Ser B, 60, 285 (1984);
 b) M. Imoto, H. Yoshimura, N.

- Sakaguchi, S. Kusumoto, and T. Shiba, *Tetrahedron Lett.*, **26**, 1545 (1985); c) M. Imoto, H. Yoshimura, T. Shimamoto, N. Sakaguchi, S. Kusumoto, and T. Shiba, *Bull. Chem. Soc. Jpn.*, **60**, 2205 (1987).
- J. Y. Homma, M. Matsuura, S. Kanegasaki, Y. Kawakubo, Y. Kojima, N. Shibukawa, Y. Kumazawa, A. Yamamoto, K. Tanamoto, T. Yasuda, M. Imoto, H. Yoshimura, S. Kusumoto, and T. Shiba, J. Biochem. (Tokyo), 98, 395 (1985); S. Kotani, H. Takada, M. Tsujimoto, T. Ogawa, I. Takahashi, T. Ikeda, K. Otsuka, H. Shimauchi, N. Kasai, J. Mashimo, S. Nagao, A. Tanaka, K. Harada, K. Nagaki, H. Kitamura, T. Shiba, S. Kusumoto, M. Imoto, and H. Yoshimura, Infect. Immun., 49, 225 (1985); C. Galanos, O. Luderitz, E. Th. Rietschel, O. Westphal, H. Brade, L. Brade, M. Freudenberg, U. Schade, M. Imoto, H. Yoshimura, S. Kusumoto, and T. Shiba, Eur. J. Biochem., 148, 1 (1985).
- S. Kusumoto, H. Yoshimura, M. Imoto, T. Shimamoto, and T. Shiba, *Tetrahedron Lett.*, 26, 909 (1985); M. Imoto, H. Yoshimura, M. Yamamoto, T. Shimamoto, S. Kusumoto, and T. Shiba, *Bull. Chem. Soc. Jpn.*, 60, 2197 (1987).
- S. Kotani, H. Takada, I. Takahashi, T. Ogawa, M. Tsujimoto, H. Shimauchi, T. Ikeda, H. Okamura, T. Tamura, K. Harada, S. Tanaka, T. Shiba, S. Kusumoto, and T. Shimamoto, Infect. Immun., 54, 673 (1986); H, Loppnow, L. Brade, H. Brade, E. T. Rietschel, S. Kusumoto, T. Shiba, and H. D. Flad, Eur. J. Immun., 16, 1263 (1986); N. Kasai, S. Arata, J. Mashimo, K. Okuda, Y. Hihara, S. Kotani, A. Takada, T. Shiba, S. Kusumoto, M. Imoto, H. Yoshimura, and T. Shimamoto, Infect. Immun., 51, 43 (1986); S. Kanegasaki, K. Tanamoto, T. Yasuda, J. Honma, M. Matsuura, M. Nakatsuka, Y. Kumazawa, A. Yamamoto, T. Shiba, S. Kusumoto, M. Imoto, H. Yoshimura, and T. Shimamoto, J. Biochem. (Tokyo), 99, 1203 (1986); S. Kotani, H. Takada, I. Takahashi, M. Tsujimoto, T. Ogawa, T. Ikeda, K. Harada, H. Okamura, T. Tamura, and S. Tanaka, Infect. Immun., 52, 872 (1986); S. Ukei, J. Iida, T. Shiba, S. Kusumoto, and I. Azuma, Vaccine, 4, 21 (1986); E. T. Rietschel, L. Brade, U. Schade, C. Galanos, M. Frendenberg, O. Luderitz, S. Kusumoto, and T. Shiba, Eur. J. Biochem., 169, 27 (1987); I. Takahashi, S. Kotani, H. Takada, M. Tsujimoto, T. Ogawa, T. Shiba, S. Kusumoto, M. Yamamoto, A. Hasegawa, M. Kiso, M. Nishijima, F. Amano, Y. Akamatsu, K. Harada, S. Tanaka, H. Okamura, and T. Tamura, Infect. Immun., 65, 57 (1987); I. Takahashi, S. Kotani, H. Takada, T. Shiba, and S. Kusumoto, Blood Purification, 6, 188 (1988); H. Takada, S. Kotani, S. Tanaka, T. Ogawa, I. Takahashi, M. Tsujimoto, T. Komuro, T. Shiba, S. Kusumoto, N. Kusunose, A. Hasegawa, and M. Kiso, Eur. J. Biochem, 175, 573 (1988); S. Arata, J. Mashimo, N. Kasai, K. Okuda, T. Aihara, S. Kotani, H. Takada, T. Shiba, S. Kusumoto, T. Shimamoto, and N. Kusunose, FEBS Microbiol. Lett., 49, 479 (1988); T. Nakamura, F. Takunage, T. Morita, S. Iwanaga, S. Kusumoto, T. Shiba, T. Kobayashi, and K. Inoue, Eur. J. Biochem., 176, 89 (1988); M. Tsujimoto, S. Kotani, T. Shiba, S. Kusumoto, A. Hasegawa, M. Kiso, and Y. Ono, Adv. Biosci., 68, 151 (1988); A. B. Cady, S. Kotani, T. Shiba, S. Kusumoto, and J. M. Krueger, Infect. Immun., 57, 396 (1989); M. Tsujimoto, S. Kotani, T. Okunaga, T. Kubo, H. Takada, T. Kubo, T. Shiba, S. Kusumoto, T. Takahashi, Y. Goto, and F. Kinoshita, Vaccine, 7, 39 (1989).
- 8) M. Imoto, H. Yoshimura, T. Shimamoto, N. Sakaguchi, S. Kusumoto, and T. Shiba, Bull. Chem. Soc. Jpn., 60, 2205 (1987).
- 9) L. M. Haines and E. Singleston, J. Chem. Soc., Dalton Trans., 1981, 1972
- 10) W. Konig and R. Geiger, Chem. Ber., 103, 788 (1970).