

Development of the Ireland–Claisen Rearrangement of Allyl-2-alkoxyacetate Bearing an Allylic Amine and the Transformation to 3-Hydroxy-4-hydroxymethylpyrrolidine

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An unprecedented Ireland–Claisen rearrangement of allylglycolate bearing an allylic amine was realized, affording the corresponding product with excellent diastereoselectivity and in high yield. The product was analyzed by X-ray crystallography, and demonstrated the transformation to the 4-hydroxymethylpyrrolidin-3-ol.

Keywords: Ireland–Claisen rearrangement; Synthesis; Allylglycolate; Allylic amine.

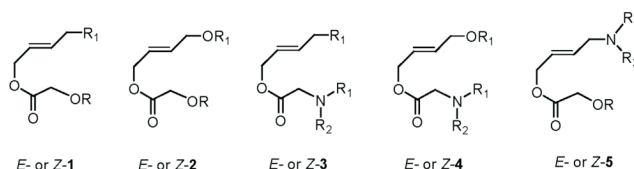
INTRODUCTION

For nearly a century, the Claisen Rearrangement¹ has played a pivotal role in organic synthesis, and a variety of modifications have been discovered for different purposes. Among the variant rearrangements, the Ireland–Claisen rearrangement² is one of the most interesting protocols, efficiently providing γ,δ -unsaturated carbonyl compounds with high stereoselectivity.³ Recently, the observation of Claisen rearrangements in biological systems has further attracted scientists' interests,⁴ resulting in the development of the organocatalysis Claisen rearrangements.⁵ Traditionally, the Ireland–Claisen rearrangements of the corresponding allylglycolate (*e.g.*, **1–4**) have received much attention,⁶ and the reactions of the variants bearing oxygen or nitrogen at different positions of the starting substrates have been reported (Scheme I).⁷ Nevertheless, the Ireland–Claisen rearrangement of an allylglycolate bearing an allylic amine (*e.g.*, **5**),⁸ as depicted in Scheme I, has received little attention. It would be interesting to explore the reaction of such substrates, to reveal the stereoselectivity of the transformation, and to identify synthetic applications. Herein we report such observations and an application toward the synthesis of 4-hydroxymethylpyrrolidin-3-ol, an important intermediate in the synthesis of nucleotides or sodium channel blockers.⁹

RESULTS AND DISCUSSION

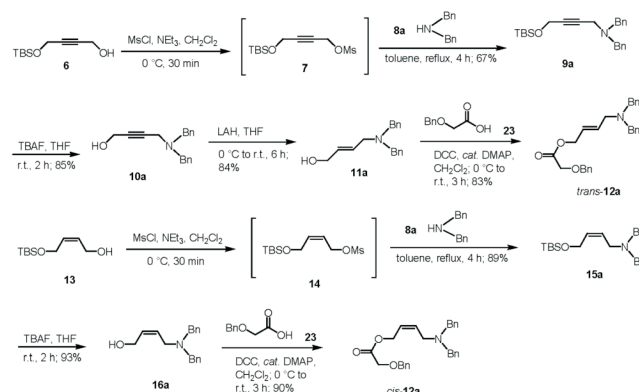
Initially, the *trans*-glycolate **12a** was prepared through a sequence of reactions starting from 1-(*tert*-butyldimethylsilyloxy)but-2-yn-4-ol (**6**)¹⁰ by mesylation and amination (MsCl, Et₃N, CH₂Cl₂, 0 °C, 30 min; Bn₂NH, toluene, reflux, 4 h; 89%) to give the propargyl amine **9a**. Depro-

Scheme I Selected allylglycolate variants for the Ireland–Claisen rearrangement



tection of **9a** followed by reduction afforded the allylic amine **11a**, which was reacted with 2-(benzyloxy)acetic acid to provide the *trans*-glycolate **12a** (Scheme II). On the other hand, the *cis*-glycolate **12a**¹¹ was prepared starting from the allylether **13** by a similar reaction sequence (Scheme II).

Scheme II Preparation of the allylglycolate **12a**

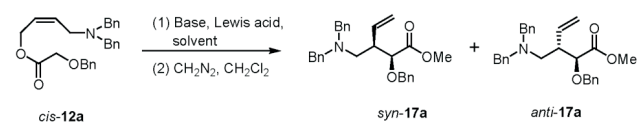


At the outset of the study, the rearrangement of *cis*-glycolate **12a** was carried out using various Lewis acids, Lewis bases, and solvents, see Table 1. The series of screenings revealed that the best yield was obtained by the

Dedicated to the memory of Professor Yung-Son Hon (1955–2011).

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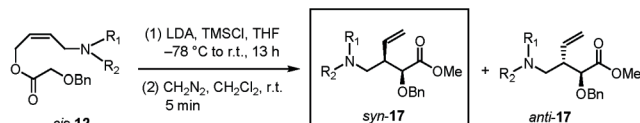
Table 1. Reaction condition screening for the rearrangement



Entry	Base	Solvent	Lewis acid	Time (h)	Yield ^b (%)
1	LDA	THF	TMSCl	13	90
2	LiHMDS	THF	TMSCl	13	82
3	NaHMDS	THF	TMSCl	13	54
4	KHMDS	THF	TMSCl	12	73
5	LDA	toluene	TMSCl	12	69
6	LDA	Ether	TMSCl	12	79
7	LDA	THF	TBSCl	13	76
8	LDA	THF	TMSOTf	12	65
9	NEt ₃	CH ₂ Cl ₂	TMSOTf	14	76 ^c

^b Isolated yields; unless otherwise noted, dr (*cis:trans*) >95:5. ^c dr (*cis:trans*) = 75:25.

reaction conditions involving LDA–TMSCl in THF at ambient temperature for 13 h, followed by in situ treatment of diazomethane to give a 90% yield of the *trans*-**17** as the only observable diastereomer (Table 1, entries 1). Replacement of LDA by LiHMDS, NaHMDS, or KHMDS under the same reaction conditions afforded a lower yield of the products, 54–82%, (Table 1, entries 2–4). The reaction under LDA–TMSCl condition in other solvents, *e.g.*, toluene, Et₂O, provided lower yields, 69–79% (Table 1, entries 5–6). Replacement of TMSCl by other acids, such as TBSCl or TMSOTf, also gave lower yields, (Table 1, en-

Table 2. Scope of the rearrangement of *cis*-glycolate **12**


Entry	Product	Yield (%) ^a	dr ^b
1	17a R ₁ = R ₂ = Bn	90	>95:5
2	17b R ₁ = allyl, R ₂ = Bn	81	>95:5
3	17c R ₁ = Ph, R ₂ = Me	82	16:1
4	17d R ₁ = <i>c</i> -C ₆ H ₁₁ , R ₂ = Bn	63	>95:5
5	17e R ₁ = R ₂ = <i>i</i> -C ₃ H ₇	71	>95:5
6	17f R ₁ = R ₂ = pyrrolidine	79	>95:5
7	17g R ₁ = R ₂ = piperidine	57	>95:5
8	17h R ₁ = Me, R ₂ = Bn	90	>95:5
9	17i R ₁ = allyl, R ₂ = <i>c</i> -C ₆ H ₁₁	58	>95:5
10	17j R ₁ = PMB, R ₂ = Bn	67	>95:5
11	17k R ₁ = R ₂ = N ₃	60	4:1

^a Isolated yield. ^b Determined by ¹H NMR of the crude products.

tries 7–8). It is noteworthy that the reaction with Et₃N–TMSOTf in CH₂Cl₂ yielded a 3:1 ratio of the *cis/trans* mixture of **17** (Table 1, entry 9). The structure of the product *syn*-**17** was unambiguously confirmed based on the X-ray analysis (Figure 1).

With the optimized conditions in hand, a series of *cis*-glycolates **12** was prepared according the aforementioned protocol and subjected to the rearrangement conditions. The results are summarized in Table 2, and, for most cases, the reaction gave *syn*-**17** in good yields (57–90%) with excellent diastereoselectivities, except for two examples. For the case of **17c**, the diastereomeric ratio dropped to 16:1, then decreased to 4:1, for the case of **17k** in which the two substituents on the nitrogen were azide, Table 2, entries 3 and 11.

The reaction conditions of the Ireland–Claisen rearrangement were alternatively applied to the *trans*-glycolate **12**, prepared according the aforementioned protocol, followed by esterification, to afford the *anti*-**17** as the major isomers (Table 3). For most cases, the *anti*-isomers were the only observable products (dr: >95:5). In a few cases, the diastereomeric ratio dropped to 3:1 or 7:1 (Table 3, entries 2, 3, 8, and 11).

Finally, we explored the possibility of extending the product to 4-(hydroxymethyl)pyrrolidin-3-ol (Scheme III). The *syn*-**17** was converted to the *anti*-lactam **18** by deprotection of the allyl group and spontaneous lactamization (Pd–C, MeOH–HOAc, 100 °C, 3d), followed oxidative

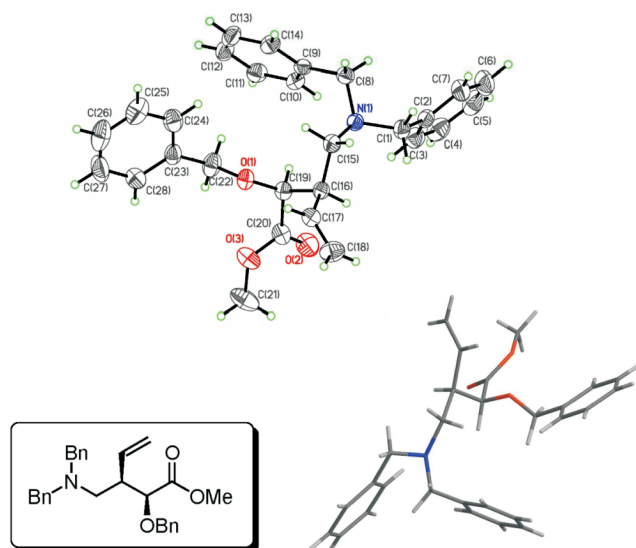
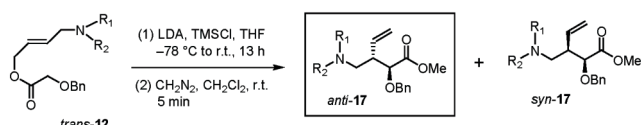


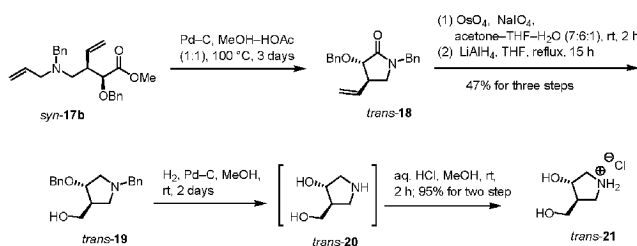
Fig. 1. Stereo plots of the X-ray crystal structures of *syn*-**17a**: C, gray; N, blue; O, red, and the ORTEP diagram.

Table 3. Scope of the rearrangement of *trans*-glycolate **12**


Entry	Product	Yield (%)	dr
1	17a R ₁ = R ₂ = Bn	82	>95:5
2	17b R ₁ = allyl, R ₂ = Bn	76	7:1
3	17c R ₁ = Ph, R ₂ = Me	72	3:1
4	17d R ₁ = <i>c</i> -C ₆ H ₁₁ , R ₂ = Bn	63	>95:5
5	17e R ₁ = R ₂ = <i>i</i> -C ₃ H ₇	54	>95:5
6	17f R ₁ = R ₂ = pyrrolidine	58	>95:5
7	17g R ₁ = R ₂ = piperidine	82	>95:5
8	17h R ₁ = Me, R ₂ = Bn	86	8:1
9	17i R ₁ = allyl, R ₂ = <i>c</i> -C ₆ H ₁₁	62	>95:5
10	17j R ₁ = PMB, R ₂ = Bn	61	>95:5
11	17k R ₁ = R ₂ = N ₃	61	7:1

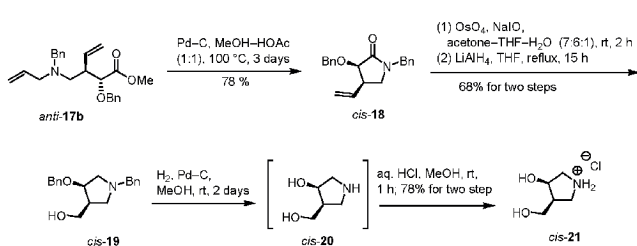
cleavage (OsO₄, NaIO₄, acetone–THF–H₂O (7:6:1) and reduction with LiAlH₄ to give 47% yield of **19** in three consecutive steps without purification of the intermediates (Scheme III). Deprotection of **19** (Pd–C, H₂, MeOH) afforded the pyrrolidine **20**, which was further treated with an aqueous HCl solution to give 4-(hydroxymethyl)pyrrolidin-3-ol.¹²

Scheme III



The same reaction sequence was alternatively applied in the *anti*-**17b** to give the *cis*-4-(hydroxymethyl)pyrrolidin-3-ol (Scheme IV).

Scheme IV



In summary, an unprecedented Ireland–Claisen rearrangement of an allylglycolate bearing an allylic amine was realized, affording the corresponding methyl 2-alkoxy-3-aminomethylpent-4-enoate with excellent yields and diastereoselectivity. The structure of the product was confirmed unambiguously by X-ray analysis of the appropriate product. Transformations of the products to the *cis*- and *trans*-4-hydroxymethylpyrrolidin-3-ol were achieved.

EXPERIMENTAL

Reactions were normally carried out under nitrogen atmosphere in glassware. Merck silica gel 60 (particle size 0.04–0.063 mm) was employed for flash chromatography. ¹H NMR spectra were obtained in CDCl₃ unless otherwise noted at 400 MHz (Bruker DPX-400) or 500 MHz (Varian-Unity INOVA-500). ¹³C NMR spectra were obtained at 100 MHz or 125 MHz. The melting point was recorded on a melting point apparatus (MPA100 – Automated melting point system, Stanford Research Systems, Inc.) and is uncorrected.

Typical experimental procedure for the preparation of *trans*-glycolate **12a**

To a solution of **11a** (492 mg, 1.84 mmol), N,N-dicyclohexylcarbodiimide (493 mg, 2.39 mmol) and N,N-4-dimethylaminopyridine (22 mg, 0.18 mmol) in CH₂Cl₂ (18 mL) was added benzyloxyacetic acid (398 mg, 2.39 mmol) at 0 °C. The solution was stirred at ambient temperature for 3 h until the completion of reaction, as monitored by TLC. The solution was filtered to remove the precipitate, and the filtrate was concentrated *in vacuo* to give the crude product. The residue was purified by flash column chromatography with 10% EtOAc–hexane (*R_f* = 0.57 in 20% EtOAc–hexane) to afford product *trans*-glycolate **12a** as a yellow oil (625 mg, 83% yield). Selected spectroscopic data: IR (CH₂Cl₂): 3027, 2924, 2797, 1753, 1454, 1193, 1125, 970, 736, 698 cm^{−1}; ¹H NMR (CDCl₃, 400 MHz): δ 7.36–7.23 (m, 15 H), 5.90–5.83 (m, 1 H), 5.79–5.72 (m, 1 H), 4.64 (d, *J* = 6.4 Hz, 2 H), 4.62 (s, 2 H), 4.10 (s, 2 H), 3.56 (s, 4 H), 3.07 (d, *J* = 5.6 Hz, 2 H); ¹³C NMR (CDCl₃, 100 MHz): δ 170.0 (C), 139.2 (two C), 137.1 (C), 133.4 (CH), 128.8 (four CH), 128.5 (two CH), 128.2 (four CH), 128.04 (two CH), 127.99 (CH), 126.9 (two CH), 126.3 (CH), 73.3 (CH₂), 67.2 (CH₂), 65.0 (CH₂), 57.9 (two CH₂), 54.7 (CH₂); MS (*m/z*, relative intensity): 415 (M⁺, 75), 338 (7), 324 (5), 250 (14), 249 (20), 210 (8), 158 (11), 106 (6), 92 (11), 91 (100), 65 (6); exact mass calculate for C₂₇H₂₉NO₃ (M⁺): 415.2147; found 415.2155.

Typical experimental procedure for the preparation of *cis*-glycolate **12a**

To a solution of **16a** (1.24 g, 4.66 mmol), N,N-dicyclohexylcarbodiimide (1.25 g, 6.05 mmol) and N,N-4-dimethylaminopyridine (56 mg, 0.46 mmol) in CH₂Cl₂ (46 mL) was added benzyloxyacetic acid (1.00 g, 6.05 mmol) at 0 °C. The solution was stirred at ambient temperature for 3 h until the completion of reaction, as monitored by TLC. The solution was filtered to remove the precipitate, and the filtrate was concentrated *in vacuo* to give the crude product. The residue was purified by flash column chromatography with 10% EtOAc-hexane (*R_f* = 0.77 in 25% EtOAc-hexane) to afford product *cis*-glycolate **12a** as a yellow oil (1.74 g, 90% yield). Selected spectroscopic data: IR (CH₂Cl₂): 3028, 2922, 1750, 1455, 1193, 1126, 737, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.36–7.20 (m, 15 H), 5.88–5.76 (m, 1 H), 5.69–5.63 (m, 1 H), 4.64 (d, *J* = 6.8 Hz, 2 H), 4.60 (s, 2 H), 4.06 (s, 2 H), 3.55 (s, 4 H), 3.10 (d, *J* = 6.4 Hz, 2 H); ¹³C NMR (CDCl₃, 100 MHz): δ 170.0 (C), 139.2 (two C), 137.0 (C), 132.8 (CH), 128.7 (four CH), 128.4 (two CH), 128.2 (four CH), 128.0 (two CH), 127.9 (CH), 126.9 (two CH), 125.7 (CH), 73.3 (CH₂), 67.1 (CH₂), 60.6 (CH₂), 58.1 (two CH₂), 50.0 (CH₂); MS (*m/z*, relative intensity): 415 (M⁺, 32), 250 (14), 249 (43), 197 (8), 196 (5), 158 (27), 106 (21), 92 (11), 91 (100), 65 (5); exact mass calculate for C₂₇H₂₉NO₃ (M⁺): 415.2147; found 415.2153.

Typical experimental procedure for the Ireland-Claisen rearrangement of glycolate **12a**, preparation of *syn*-**17a**

To a solution of *cis*-glycolate **12a** (110 mg, 0.26 mmol) in dry THF (4 mL) was added a solution of lithium diisopropylamide in THF (2.0 M in THF, 0.26 mL, 0.52 mmol) at -78 °C. The solution was stirred for 15 h, followed by the addition of chloro trimethylsilane (TMSCl, 0.10 mL, 0.78 mmol). The solution was gradually warm up to room temperature and stirred for 13 h, followed by the addition of saturated aqueous NH₄Cl (3 mL). The solution was extracted with EtOAc (5 mL × 3), dried over MgSO₄, and concentrated *in vacuo* to give the crude product as yellow oil. The residue was dissolved in CH₂Cl₂, followed by the slow addition of a solution of diazomethane in ether, monitored by TLC. The solution was concentrated *in vacuo* to give the crude product. The residue was purified by flash column chromatography with 10% EtOAc-hexane (*R_f* = 0.68 in 20% EtOAc-hexane) to afford product *syn*-**17a** as a yellow oil (104 mg, 90% yield).

Spectroscopic data for *syn*-**17a**: IR (CH₂Cl₂): 3028, 2923, 2851, 1749, 1455, 1266, 1204, 1136, 737, 699 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.33–7.13 (m, 15 H), 5.79–5.71 (m, 1 H), 5.08–4.99 (m, 2 H), 4.54 (d, *J* = 11.2 Hz, 1 H), 4.29 (d, *J* = 2.0 Hz, 1 H), 3.99 (d, *J* = 11.2 Hz, 1 H), 3.69 (s, 3 H), 3.63 (d, *J* = 13.6 Hz, 2 H), 3.45 (d, *J* = 13.6 Hz, 2 H), 2.81–2.73 (m, 2 H) 2.47–2.40 (m, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 173.0 (C), 139.2 (two C), 137.8 (C), 135.9 (CH), 129.1 (four CH), 128.2 (four CH), 128.1 (two CH), 127.8 (two CH), 127.4 (CH), 126.9 (two CH), 117.3 (CH₂), 78.7 (CH), 72.5 (CH₂), 58.5 (two CH₂), 55.0 (CH₂), 51.6 (CH₃), 46.3 (CH); MS (*m/z*, relative intensity): 429 (M⁺, 49), 211 (17), 210 (100), 181 (4), 92 (8), 91 (87), 65 (5); exact mass calculate for C₂₈H₃₁NO₃ (M⁺): 429.2304; found 429.2298.

For *anti*-**17a**: 91 mg, 82% yield; *R_f* = 0.74 (Hexane/EtOAc = 4 : 1), yellow oil.

Spectroscopic data for *anti*-**17a**: IR (CH₂Cl₂): 3027, 2923, 2851, 2797, 1748, 1494, 1453, 1201, 1128, 916, 745, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.32–7.18 (m, 15 H), 5.68–5.59 (m, 1 H), 5.12–5.08 (m, 2 H), 4.64 (d, *J* = 12.0 Hz, 1 H), 4.32 (d, *J* = 12.0 Hz, 1 H), 3.91 (d, *J* = 5.6 Hz, 1 H), 3.64 (d, *J* = 13.6 Hz, 2 H), 3.60 (s, 3 H), 3.40 (d, *J* = 13.6 Hz, 2 H), 2.86–2.81 (m, 1 H) 2.67 (dd, *J* = 12.8, 5.6 Hz, 1 H), 2.54 (dd, *J* = 12.8, 8.8 Hz, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 171.9 (C), 139.3 (two C), 137.40 (CH), 137.39 (C), 129.0 (four CH), 128.2 (two CH), 128.0 (four CH), 127.8 (two CH), 127.7 (CH), 126.7 (two CH), 117.1 (CH₂), 80.4 (CH), 72.5 (CH₂), 58.3 (two CH₂), 54.0 (CH₂), 51.4 (CH₃), 45.9 (CH); MS (*m/z*, relative intensity): 429 (M⁺, 29), 211 (10), 210 (100), 181 (4), 92 (7), 91 (79), 65 (4); exact mass calculate for C₂₈H₃₁NO₃ (M⁺): 429.2304; found 429.2296.

For *syn*-**17b**: 516 mg, 81% yield; *R_f* = 0.63 (Hexane/EtOAc = 5 : 1); yellow oil.

Spectroscopic data for *syn*-**17b**: IR (CH₂Cl₂): 3065, 3028, 2949, 2925, 2803, 1752, 1455, 1260, 1202, 1136, 918, 740, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.30–7.21 (m, 10 H), 5.85–5.74 (m, 2 H), 5.15–5.00 (m, 4 H), 4.69 (d, *J* = 11.2 Hz, 1 H), 4.37 (d, *J* = 2.0 Hz, 1 H), 4.26 (d, *J* = 11.2 Hz, 1 H), 3.71 (s, 3 H), 3.68 (d, *J* = 13.6 Hz, 1 H), 3.46 (d, *J* = 13.6 Hz, 1 H), 3.10 (dd, *J* = 14.0, 6.0 Hz, 1 H), 2.96 (dd, *J* = 14.0, 6.8 Hz, 1 H), 2.79–2.73 (m, 2 H), 2.41 (dd, *J* = 17.2, 10.4 Hz, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 173.2 (C), 139.3 (C), 137.8 (C), 135.8 (CH), 135.4 (CH), 129.0 (two CH), 128.2 (four CH), 127.9 (two CH), 127.6 (CH), 126.9 (CH), 117.6 (CH₂), 117.4 (CH₂), 78.5 (CH),

72.8 (CH₂), 58.3 (CH₂), 56.8 (CH₂), 54.7 (CH₂), 51.6 (CH₃), 46.1 (CH); MS (*m/z*, relative intensity): 379 (M⁺, 49), 161 (11), 160 (100), 92 (5), 91 (73); exact mass calculate for C₂₄H₂₉NO₃ (M⁺): 379.2147; found 379.2138.

For *anti*-**17b**: 78 mg, 76% yield; R_f = 0.69 (Hexane/EtOAc = 4 : 1); yellow oil.

Spectroscopic data for *anti*-**17b**: IR (CH₂Cl₂): 3065, 3028, 2948, 2838, 2801, 1750, 1454, 1258, 1202, 1132, 917, 739, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.33–7.21 (m, 10 H), 5.87–5.69 (m, 2 H), 5.14–5.05 (m, 4 H), 4.72 (d, *J* = 12.0 Hz, 1 H), 4.37 (d, *J* = 12.0 Hz, 1 H), 3.98 (d, *J* = 4.4 Hz, 1 H), 3.67 (s, 3 H), 3.61 (d, *J* = 13.6 Hz, 1 H), 3.50 (d, *J* = 13.6 Hz, 1 H), 3.07–2.96 (m, 2 H), 2.87–2.79 (m, 1 H), 2.72 (dd, *J* = 12.8, 6.8 Hz, 1 H), 2.50 (dd, *J* = 12.8, 7.6 Hz, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 172.1 (C), 139.2 (C), 137.6 (C), 137.5 (CH), 135.6 (CH), 129.1 (two CH), 128.3 (two CH), 128.0 (two CH), 127.8 (two CH), 127.7 (CH), 126.7 (CH), 117.3 (CH₂), 116.9 (CH₂), 80.1 (CH), 72.6 (CH₂), 58.2 (CH₂), 56.7 (CH₂), 53.7 (CH₂), 51.5 (CH₃), 45.8 (CH); MS (*m/z*, relative intensity): 379 (M⁺, 63), 161 (11), 160 (93), 92 (8), 91 (100), 65 (5); exact mass calculate for C₂₄H₂₉NO₃ (M⁺): 379.2147; found 379.2144.

For *syn*-**17c**: 97 mg, 82% yield; R_f = 0.68 (Hexane/EtOAc = 5 : 1); yellow oil.

Spectroscopic data for *syn*-**17c**: IR (CH₂Cl₂): 2950, 2870, 1752, 1599, 1507, 1203, 1133, 748, 694 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.39–7.30 (m, 5 H), 7.23–7.17 (m, 2 H), 6.74–6.64 (m, 3 H), 5.95–5.85 (m, 1 H), 5.13–5.07 (m, 2 H), 4.75 (d, *J* = 11.2 Hz, 1 H), 4.25 (d, *J* = 11.2 Hz, 1 H), 4.10 (d, *J* = 2.8 Hz, 1 H), 3.70 (s, 3 H), 3.61 (dd, *J* = 14.8, 8.8 Hz, 1 H), 3.26 (dd, *J* = 14.8, 6.0 Hz, 1 H), 2.99–2.93 (m, 1 H), 2.89 (s, 3 H); ¹³C NMR (CDCl₃, 100 MHz): δ 172.3 (C), 149.3 (C), 137.6 (C), 134.7 (CH), 129.1 (two CH), 128.3 (two CH), 127.9 (two CH), 127.8 (CH), 118.4 (CH₂), 116.4 (CH), 112.3 (two CH), 78.3 (CH), 72.4 (CH₂), 54.2 (CH₂), 51.7 (CH₃), 46.3 (CH), 38.9 (CH₃); MS (*m/z*, relative intensity): 339 (M⁺, 100), 121 (8), 120 (100), 91 (6); exact mass calculate for C₂₁H₂₅NO₃ (M⁺): 339.1834; found 339.1840.

For *anti*-**17c**: 73 mg, 72% yield; R_f = 0.74 (Hexane/EtOAc = 4 : 1); yellow oil.

Spectroscopic data for *anti*-**17c**: IR (CH₂Cl₂): 2948, 2871, 1747, 1599, 1507, 1346, 1204, 1132, 993, 749, 695 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.39–7.29 (m, 5 H), 7.22–7.18 (m, 2 H), 6.69–6.66 (m, 3 H), 5.85–5.76 (m, 1 H), 5.15–5.08 (m, 2 H), 4.83 (d, *J* = 11.6 Hz, 1 H), 4.40 (d, *J* = 11.6 Hz, 1 H), 3.93 (d, *J* = 4.8 Hz, 1 H), 3.60 (dd, *J* = 14.4,

7.6 Hz, 1 H), 3.49 (s, 3 H), 3.28 (dd, *J* = 14.4, 7.2 Hz, 1 H), 3.17–3.12 (m, 1 H), 2.86 (s, 3 H); ¹³C NMR (CDCl₃, 100 MHz): δ 171.4 (C), 149.1 (C), 137.5 (C), 136.4 (CH), 128.9 (two CH), 128.3 (two CH), 127.9 (two CH), 127.8 (CH), 117.8 (CH₂), 116.1 (CH), 112.2 (two CH), 79.3 (CH), 72.8 (CH₂), 53.6 (CH₂), 51.2 (CH₃), 45.7 (CH), 39.8 (CH₃); MS (*m/z*, relative intensity): 339 (M⁺, 100), 121 (9), 120 (100), 105 (7), 91 (11), 77 (5); exact mass calculate for C₂₁H₂₅NO₃ (M⁺): 339.1834; found 339.1831.

For *syn*-**17d**: 98 mg, 63% yield; R_f = 0.50 (Hexane/EtOAc = 8 : 1); yellow oil.

Spectroscopic data for *syn*-**17d**: IR (CH₂Cl₂): 2927, 2853, 1751, 1454, 1248, 1203, 1137, 915, 838, 737, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.35–7.20 (m, 10 H), 5.82–5.73 (m, 1 H), 5.04 (dd, *J* = 10.4, 2 Hz, 1H), 4.96 (dd, *J* = 17.6, 2 Hz, 1H), 4.68 (d, *J* = 11.2 Hz, 1 H), 4.40 (d, *J* = 2.0 Hz, 1 H), 4.19 (d, *J* = 11.2 Hz, 1 H), 3.67 (s, 3 H), 3.64 (s, 2 H), 2.80 (dd, *J* = 12.8, 9.6 Hz, 1 H), 2.54–2.49 (m, 1 H), 2.45 (dd, *J* = 12.8, 5.2 Hz, 1 H), 2.41–2.17 (m, 1 H), 1.87–1.80 (m, 1 H), 1.76–1.64 (m, 3 H), 1.58–0.98 (m, 6 H); ¹³C NMR (CDCl₃, 100 MHz): δ 173.4 (C), 141.3 (C), 138.0 (C), 136.0 (CH), 128.6 (two CH), 128.2 (two CH), 128.1 (two CH), 127.8 (two CH), 127.5 (CH), 126.5 (CH), 117.1 (CH₂), 78.4 (CH), 72.6 (CH₂), 59.6 (CH), 55.2 (CH₂), 52.0 (CH₂), 51.5 (CH₃), 47.2 (CH), 29.8 (CH₂), 27.7 (CH₂), 26.3 (CH₂), 26.2 (CH₂), 26.1 (CH₂); MS (*m/z*, relative intensity): 421 (M⁺, 44), 203 (13), 202 (100), 120 (11), 92 (4), 91 (54), 55 (5); exact mass calculate for C₂₇H₃₅NO₃ (M⁺): 421.2617; found 421.2624.

For *anti*-**17d**: 79 mg, 63% yield; R_f = 0.49 (Hexane/EtOAc = 9 : 1); yellow oil.

Spectroscopic data for *anti*-**17d**: IR (CH₂Cl₂): 2927, 2853, 1748, 1452, 1265, 1203, 1131, 736, 699 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.33–7.19 (m, 10 H), 5.71–5.62 (m, 1 H), 5.06–5.01 (m, 2 H), 4.67 (d, *J* = 11.6 Hz, 1 H), 4.34 (d, *J* = 11.6 Hz, 1 H), 3.99 (d, *J* = 4.8 Hz, 1 H), 3.66 (s, 3 H), 3.66–3.55 (m, 2 H), 2.74 (dd, *J* = 12.4, 5.2 Hz, 1 H), 2.66–2.59 (m, 1 H), 2.55 (dd, *J* = 12.4, 8.0 Hz, 1 H), 2.41–2.35 (m, 1 H), 1.78–1.71 (m, 4 H), 1.24–0.96 (m, 6 H); ¹³C NMR (CDCl₃, 100 MHz): δ 172.2 (C), 141.3 (C), 137.58 (C), 137.56 (CH), 128.5 (two CH), 128.2 (two CH), 127.92 (two CH), 127.89 (two CH), 127.7 (CH), 126.4 (CH), 116.8 (CH₂), 80.2 (CH), 72.6 (CH₂), 59.2 (CH), 54.9 (CH₂), 51.4 (CH₃), 50.9 (CH₂), 46.9 (CH), 29.2 (CH₂), 28.0 (CH₂), 26.4 (CH₂), 26.21 (CH₂), 26.15 (CH₂); MS (*m/z*, relative intensity): 421 (M⁺, 27), 203 (15), 202 (100), 120 (15), 92 (6), 91 (83), 55 (8); exact mass calculate for

$C_{27}H_{35}NO_3$ (M^+): 421.2617; found 421.2613.

For *syn-17e*: 124 mg, 71% yield; $R_f = 0.44$ (Hexane/EtOAc = 4 : 1); yellow oil.

Spectroscopic data for *syn-17e*: IR (CH_2Cl_2): 2965, 1751, 1455, 1382, 1254, 1207, 1174, 1133, 916, 843, 736, 698, 668, 595 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.39–7.27 (m, 5 H), 5.88–5.79 (m, 1 H), 5.07–4.99 (m, 2 H), 4.74 (d, $J = 11.2$ Hz, 1 H), 4.42 (d, $J = 1.6$ Hz, 1 H), 4.35 (d, $J = 11.2$ Hz, 1 H), 3.73 (s, 3 H), 3.03–2.96 (m, 2 H), 2.68–2.58 (m, 2 H), 2.51–2.45 (m, 1 H), 1.00–0.96 (m, 12 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 173.8 (C), 138.1 (C), 136.2 (CH), 128.2 (two CH), 127.7 (two CH), 127.5 (CH), 117.0 (CH₂), 78.1 (CH), 72.4 (CH₂), 51.5 (CH₃), 47.8 (two CH), 47.5 (CH), 46.4 (CH₂), 21.7 (two CH₃), 20.1 (two CH₃); MS (m/z , relative intensity): 333 (M^+ , 26), 304 (4), 115 (7), 114 (100), 91 (18), 72 (13); exact mass calculate for $C_{20}H_{31}NO_3$ (M^+): 333.2304; found 333.2309.

For *anti-17e*: 71 mg, 54% yield; $R_f = 0.50$ (Hexane/EtOAc = 1 : 1); yellow oil.

Spectroscopic data for *anti-17e*: IR (CH_2Cl_2): 2964, 2928, 2869, 1752, 1455, 1386, 1361, 1253, 1206, 1118, 915, 735, 698 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.37–7.28 (m, 5 H), 5.75–5.66 (m, 1 H), 5.09–5.04 (m, 2 H), 4.75 (d, $J = 12.0$ Hz, 1 H), 4.40 (d, $J = 12.0$ Hz, 1 H), 4.03 (d, $J = 4.4$ Hz, 1 H), 3.72 (s, 3 H), 2.98–2.91 (m, 2 H), 2.74–2.64 (m, 1 H), 2.44 (dd, $J = 12.8$, 7.2 Hz, 1 H), 0.93 (d, $J = 6.4$ Hz, 12 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 172.4 (C), 137.75 (C), 137.73 (CH), 128.3 (two CH), 128.0 (two CH), 127.7 (CH), 116.6 (CH₂), 80.0 (CH), 72.7 (CH₂), 51.4 (CH₃), 47.6 (two CH), 47.2 (CH), 45.5 (CH₂), 20.8 (two CH₃), 20.7 (two CH₃); MS (m/z , relative intensity): 333 (M^+ , 5), 115 (8), 114 (100), 91 (22), 72 (15), 70 (4); exact mass calculate for $C_{20}H_{31}NO_3$ (M^+): 333.2304; found 333.2304.

For *syn-17f*: 90 mg, 79% yield; $R_f = 0.61$ (CH_2Cl_2 /MeOH = 10 : 1); yellow oil.

Spectroscopic data for *syn-17f*: IR (CH_2Cl_2): 2951, 2929, 2788, 1752, 1455, 1278, 1202, 1132, 916 737, 698 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.37–7.28 (m, 5 H), 5.86–5.77 (m, 1 H), 5.10–5.04 (m, 2 H), 4.77 (d, $J = 11.6$ Hz, 1 H), 4.40 (d, $J = 11.6$ Hz, 1 H), 4.28 (d, $J = 2.8$ Hz, 1 H), 3.72 (s, 3 H), 2.76–2.69 (m, 2 H), 2.54–2.51 (m, 2 H), 2.38–2.32 (m, 3 H), 1.75–1.68 (m, 4 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 173.0 (C), 137.9 (C), 136.1 (CH), 128.2 (two CH), 128.0 (two CH), 127.7 (CH), 117.3 (CH₂), 79.0 (CH), 72.9 (CH₂), 57.0 (CH₂), 54.0 (two CH₂), 51.6 (CH₃), 46.8 (CH), 23.6 (two CH₂); MS (m/z , relative intensity): 303

(M^+ , 15), 91 (9), 85 (4), 84 (100); exact mass calculate for $C_{18}H_{25}NO_3$ (M^+): 303.1834; found 303.1839.

For *anti-17f*: 70 mg, 58% yield; $R_f = 0.55$ (CH_2Cl_2 /MeOH = 10 : 1); yellow oil.

Spectroscopic data for *anti-17f*: IR (CH_2Cl_2): 2951, 2925, 2788, 1750, 1455, 1258, 1201, 1132, 916, 736, 698 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.36–7.28 (m, 5 H), 5.84–5.75 (m, 1 H), 5.16–5.08 (m, 2 H), 4.75 (d, $J = 11.6$ Hz, 1 H), 4.39 (d, $J = 11.6$ Hz, 1 H), 3.96 (d, $J = 4.8$ Hz, 1 H), 3.71 (s, 3 H), 2.84–2.77 (m, 1 H), 2.73–2.70 (m, 1 H), 2.58–2.42 (m, 5 H), 1.71–1.66 (m, 4 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 172.2 (C), 137.6 (C), 137.4 (CH), 128.3 (two CH), 127.9 (two CH), 127.7 (CH), 117.0 (CH₂), 80.2 (CH), 72.8 (CH₂), 56.2 (CH₂), 54.2 (two CH₂), 51.5 (CH₃), 46.5 (CH), 23.5 (two CH₂); MS (m/z , relative intensity): 303 (M^+ , 10), 91 (18), 85 (6), 84 (100), 55 (6); exact mass calculate for $C_{18}H_{25}NO_3$ (M^+): 303.1834; found 303.1836.

For *syn-17g*: 65 mg, 57% yield; $R_f = 0.21$ (Hexane/EtOAc = 5 : 1); yellow oil.

Spectroscopic data for *syn-17g*: IR (CH_2Cl_2): 2930, 2852, 1752, 1600, 1455, 1267, 1197, 1132, 750, 698 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.37–7.27 (m, 5 H), 5.83–5.74 (m, 1 H), 5.07–5.01 (m, 2 H), 4.76 (d, $J = 11.6$ Hz, 1 H), 4.41 (d, $J = 11.6$ Hz, 1 H), 4.35 (d, $J = 3.2$ Hz, 1 H), 3.72 (s, 3 H), 2.80–2.73 (m, 1 H), 2.52 (dd, $J = 12.4$, 10.4 Hz, 1 H), 2.45–2.43 (m, 2 H), 2.20–2.13 (m, 3 H), 1.53–1.47 (m, 4 H), 1.42–1.39 (m, 2 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 173.4 (C), 138.0 (C), 136.0 (CH), 128.2 (two CH), 128.1 (two CH), 127.6 (CH), 117.3 (CH₂), 78.7 (CH), 73.1 (CH₂), 59.5 (CH₂), 54.7 (two CH₂), 51.6 (CH₃), 45.0 (CH), 26.2 (two CH₂), 24.5 (CH₂); MS (m/z , relative intensity): 317 (M^+ , 46), 99 (6), 98 (100), 91 (8); exact mass calculate for $C_{19}H_{27}NO_3$ (M^+): 317.1991; found 317.1981.

For *anti-17g*: 93 mg, 82% yield; $R_f = 0.26$ (Hexane/EtOAc = 2 : 1); yellow oil.

Spectroscopic data for *anti-17g*: IR (CH_2Cl_2): 2931, 2853, 1752, 1455, 1260, 1207, 1121, 1027, 916, 736, 698 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.36–7.28 (m, 5 H), 5.79–5.73 (m, 1 H), 5.12–5.05 (m, 2 H), 4.78 (d, $J = 11.6$ Hz, 1 H), 4.39 (d, $J = 11.6$ Hz, 1 H), 3.97 (d, $J = 4.4$ Hz, 1 H), 3.74 (s, 3 H), 2.88–2.81 (m, 1 H), 2.59 (dd, $J = 12.8$, 8.4 Hz, 1 H), 2.34–2.21 (m, 5 H), 1.49–1.45 (m, 4 H), 1.39–1.35 (m, 2 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 172.3 (C), 137.82 (C), 137.75 (CH), 128.2 (two CH), 127.9 (two CH), 127.7 (CH), 116.5 (CH₂), 79.9 (CH), 72.8 (CH₂), 58.8 (CH₂), 54.8 (two CH₂), 51.5 (CH₃), 44.8 (CH), 25.9 (two CH₂), 24.4 (CH₂); MS (m/z , relative intensity): 317 (M^+ , 80), 302

(30), 138 (80), 137 (84), 124 (13), 122 (16), 98 (53), 91 (100), 86 (10), 85 (65), 84 (45), 83 (10), 65 (11), 55 (26); exact mass calculate for $C_{19}H_{27}NO_3$ (M^+): 317.1991; found 317.1999.

For *syn*-**17h**: 104 mg, 90% yield; R_f = 0.26 (Hexane/EtOAc = 2 : 1); yellow oil.

Spectroscopic data for *syn*-**17h**: IR (CH_2Cl_2): 2949, 2845, 2790, 1751, 1455, 1266, 1202, 1137, 1025, 917, 740, 698 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.31–7.21 (m, 10 H), 5.85–5.76 (m, 1 H), 5.11–5.04 (m, 2 H), 4.74 (d, J = 11.2 Hz, 1 H), 4.39 (d, J = 2.8 Hz, 1 H), 4.34 (d, J = 11.2 Hz, 1 H), 3.72 (s, 3 H), 3.52 (d, J = 12.8 Hz, 1 H), 3.42 (d, J = 12.8 Hz, 1 H), 2.83–2.77 (m, 1 H), 2.70 (dd, J = 12.0, 10.0 Hz, 1 H), 2.29 (dd, J = 12.0, 5.6 Hz, 1 H), 2.14 (s, 3 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 173.1 (C), 137.8 (two C), 135.7 (CH), 129.0 (two CH), 128.22 (two CH), 128.15 (two CH), 128.0 (two CH), 127.7 (CH), 126.9 (CH), 117.5 (CH_2), 78.5 (CH), 73.0 (CH_2), 62.5 (CH_2), 58.4 (CH_2), 51.6 (CH_3), 45.8 (CH), 42.2 (CH_3); MS (m/z , relative intensity): 353 (M^+ , 7), 135 (9), 134 (100), 92 (4), 91 (64), 65 (3); exact mass calculate for $C_{22}H_{27}NO_3$ (M^+): 353.1991; found 353.1998.

For *anti*-**17h**: 94 mg, 86% yield; R_f = 0.55 (Hexane/EtOAc = 3 : 1); yellow oil.

Spectroscopic data for *anti*-**17h**: IR (CH_2Cl_2): 2949, 2924, 2844, 2791, 1749, 1454, 1265, 1206, 1135, 1025, 917, 737, 699 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.34–7.22 (m, 10 H), 5.82–5.72 (m, 1 H), 5.16–5.10 (m, 2 H), 4.76 (d, J = 11.6 Hz, 1 H), 4.39 (d, J = 11.6 Hz, 1 H), 4.00 (d, J = 4.4 Hz, 1 H), 3.68 (s, 3 H), 3.57 (d, J = 13.2 Hz, 1 H), 3.36 (d, J = 13.2 Hz, 1 H), 2.91–2.84 (m, 1 H), 2.67 (dd, J = 12.8, 7.2 Hz, 1 H), 2.41 (dd, J = 12.8, 7.6 Hz, 1 H), 2.11 (s, 3 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 172.2 (C), 138.9 (C), 137.6 (C), 137.4 (CH), 129.2 (two CH), 128.3 (two CH), 128.0 (two CH), 127.9 (two CH), 127.7 (CH), 126.8 (CH), 116.9 (CH_2), 79.9 (CH), 72.7 (CH_2), 62.5 (CH_2), 57.6 (CH_2), 51.5 (CH_3), 45.5 (CH), 42.2 (CH_3); MS (m/z , relative intensity): 353 (M^+ , 84), 135 (9), 134 (100), 92 (6), 91 (65), 65 (4); exact mass calculate for $C_{22}H_{27}NO_3$ (M^+): 353.1991; found 353.1997.

For *syn*-**17i**: 78 mg, 58% yield; R_f = 0.46 (Hexane/EtOAc = 8 : 1); yellow oil.

Spectroscopic data for *syn*-**17i**: IR (CH_2Cl_2): 2928, 2853, 1752, 1453, 1258, 1201, 1135, 1051, 995, 916, 735, 697 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.39–7.27 (m, 5 H), 5.84–5.75 (m, 2 H), 5.15–4.99 (m, 4 H), 4.75 (d, J = 11.2 Hz, 1 H), 4.41 (d, J = 2.4 Hz, 1 H), 4.35 (d, J = 11.2 Hz, 1

H), 3.72 (s, 3 H), 3.13 (d, J = 6.4 Hz, 2 H), 2.74–2.61 (m, 2 H), 2.45–2.37 (m, 2 H), 1.76–1.56 (m, 6 H), 1.25–0.99 (m, 4 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 173.5 (C), 138.2 (CH), 138.0 (C), 136.0 (CH), 128.2 (two CH), 127.8 (two CH), 127.5 (CH), 117.1 (CH_2), 115.7 (CH_2), 78.5 (CH), 72.7 (CH_2), 60.3 (CH), 54.0 (CH_2), 51.61 (CH_2), 51.55 (CH), 47.1 (CH), 30.0 (CH_2), 28.3 (CH_2), 26.3 (CH_2), 26.21 (CH_2), 26.17 (CH_2); MS (m/z , relative intensity): 371 (M^+ , 57), 153 (9), 152 (100), 91 (12), 70 (16), 55 (5); exact mass calculate for $C_{23}H_{33}NO_3$ (M^+): 371.2460; found 371.2465.

For *anti*-**17i**: 82 mg, 62% yield; R_f = 0.27 (Hexane/EtOAc = 9 : 1); yellow oil.

Spectroscopic data for *anti*-**17i**: IR (CH_2Cl_2): 2927, 2854, 1751, 1453, 1257, 1202, 1131, 1027, 995, 915, 735, 698 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.40–7.28 (m, 5 H), 5.81–5.68 (m, 2 H), 5.15–4.94 (m, 4 H), 4.76 (d, J = 11.6 Hz, 1 H), 4.39 (d, J = 11.6 Hz, 1 H), 3.99 (d, J = 3.6 Hz, 1 H), 3.71 (s, 3 H), 3.15–3.01 (m, 2 H), 2.75–2.69 (m, 2 H), 2.47–2.40 (m, 2 H), 1.74–1.64 (m, 5 H), 1.26–1.11 (m, 5 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 172.3 (C), 138.1 (CH), 137.8 (C), 137.7 (CH), 128.2 (two CH), 127.9 (two CH), 127.6 (CH), 116.6 (CH_2), 115.6 (CH_2), 79.9 (CH), 72.7 (CH_2), 59.9 (CH), 53.9 (CH_2), 51.4 (CH_3), 50.5 (CH_2), 46.6 (CH), 28.87 (CH_2), 28.85 (CH_2), 26.4 (CH_2), 26.24 (CH_2), 26.22 (CH_2); MS (m/z , relative intensity): 371 (M^+ , 25), 153 (12), 152 (100), 91 (15), 70 (25), 55 (9); exact mass calculate for $C_{23}H_{33}NO_3$ (M^+): 371.2460; found 371.2453.

For *syn*-**17j**: 104 mg, 67% yield; 0.54 (Hexane/EtOAc = 5 : 1); mp. 56–58 °C; yellow solid.

Spectroscopic data for *syn*-**17j**: IR (CH_2Cl_2): 2926, 2834, 1751, 1611, 1509, 1455, 1249, 1203, 1136, 1030, 741, 698 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz): δ 7.31–7.13 (m, 12 H), 6.81 (d, J = 8.0 Hz, 2 H), 5.79–5.71 (m, 1 H), 5.08–5.04 (m, 2 H), 4.54 (d, J = 11.2 Hz, 1 H), 4.30 (d, J = 1.6 Hz, 1 H), 3.98 (d, J = 11.2 Hz, 1 H), 3.73 (s, 3 H), 3.69 (s, 3 H), 3.64 (d, J = 13.6 Hz, 1 H), 3.58 (d, J = 13.2 Hz, 1 H), 3.42 (d, J = 13.6 Hz, 1 H), 3.36 (d, J = 13.2 Hz, 1 H), 2.82–2.74 (m, 2 H), 2.45–2.37 (m, 1 H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 173.3 (C), 158.8 (C), 139.6 (C), 138.0 (C), 136.1 (CH), 131.4 (C), 130.5 (two CH), 129.3 (two CH), 128.4 (two CH), 128.3 (two CH), 127.9 (two CH), 127.6 (CH), 127.1 (CH), 117.5 (CH_2), 113.8 (two CH), 78.8 (CH), 72.7 (CH_2), 58.5 (CH_2), 57.9 (CH_2), 55.4 (CH_3), 54.9 (CH_2), 51.9 (CH_3), 46.5 (CH); MS (m/z , relative intensity): 459 (M^+ , 47), 241 (7), 240 (43), 122 (7), 121 (100), 91 (15); exact mass calculate for $C_{29}H_{33}NO_4$ (M^+): 459.2410; found 459.2415.

For *anti*-**17j**: 70 mg, 61% yield; 0.36 (Hexane/EtOAc = 8 : 1); yellow oil.

Spectroscopic data for *anti*-**17j**: IR (CH₂Cl₂): 2923, 1748, 1540, 1508, 1456, 1247, 739, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.32–7.14 (m, 12 H), 6.81 (d, *J* = 8.4 Hz, 2 H), 5.67–5.58 (m, 1 H), 5.16–5.04 (m, 2 H), 4.65 (d, *J* = 11.6 Hz, 1 H), 4.32 (d, *J* = 11.6 Hz, 1 H), 3.91 (d, *J* = 5.6 Hz, 1 H), 3.78 (s, 3 H), 3.65 (d, *J* = 6.4 Hz, 1 H), 3.62–3.59 (m, 4 H), 3.38 (d, *J* = 13.6 Hz, 1 H), 3.34 (d, *J* = 13.6 Hz, 1 H), 2.87–2.80 (m, 1 H), 2.66 (dd, *J* = 12.8, 5.6 Hz, 1 H), 2.52 (dd, *J* = 12.8, 8.8 Hz, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 172.0 (C), 158.5 (C), 139.6 (C), 137.50 (CH), 137.46 (C), 131.3 (C), 130.2 (two CH), 129.0 (two CH), 128.3 (two CH), 128.0 (two CH), 127.8 (two CH), 127.7 (CH), 126.7 (CH), 117.1 (CH₂), 113.4 (two CH), 80.5 (CH), 72.5 (CH₂), 58.2 (CH₂), 57.6 (CH₂), 55.2 (CH₃), 53.9 (CH₂), 51.5 (CH₃), 45.9 (CH); MS (*m/z*, relative intensity): 459 (M⁺, 45), 241 (7), 240 (39), 122 (4), 121 (100), 91 (13); exact mass calculate for C₂₉H₃₃NO₄ (M⁺): 459.2410; found 459.2415.

For *syn*-**17k**: 71 mg, 60% yield; 0.58 (Hexane/EtOAc = 4 : 1); yellow oil.

Spectroscopic data for *syn*-**17k**: IR (CH₂Cl₂): 2924, 2869, 2100, 1751, 1455, 1203, 1127, 738, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.36–7.31 (m, 5 H), 5.82–5.73 (m, 1 H), 5.19–5.14 (m, 2 H), 4.80 (d, *J* = 11.6 Hz, 1 H), 4.39 (d, *J* = 11.6 Hz, 1 H), 4.17 (d, *J* = 3.2 Hz, 1 H), 3.75 (s, 3 H), 3.44 (dd, *J* = 12.0, 8.4 Hz, 1 H), 3.35 (dd, *J* = 12.0, 6.4 Hz, 1 H), 2.77–2.75 (m, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 171.7 (C), 137.2 (C), 133.3 (CH), 128.4 (two CH), 128.2 (two CH), 128.0 (CH), 119.5 (CH₂), 77.6 (CH), 72.9 (CH₂), 52.0 (CH₂), 51.9 (CH₃), 47.1 (CH); MS (*m/z*, relative intensity): 275 (M⁺, 3), 149 (6), 113 (14), 105 (5), 92 (9), 91 (100), 83 (4), 82 (4), 81 (5), 71 (5), 69 (4), 65 (8), 57 (7), 55 (6); exact mass calculate for C₁₄H₁₇N₃O₃ (M⁺): 275.1270; found 275.1275.

For *anti*-**17k**: 72 mg, 61% yield; 0.55 (Hexane/EtOAc = 4 : 1); yellow oil.

Spectroscopic data for *anti*-**17k**: IR (CH₂Cl₂): 2924, 2869, 2100, 1749, 1455, 1266, 1200, 1119, 739, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.36–7.30 (m, 5 H), 5.78–5.69 (m, 1 H), 5.21–5.17 (m, 2 H), 4.70 (d, *J* = 11.2 Hz, 1 H), 4.41 (d, *J* = 11.2 Hz, 1 H), 3.96 (d, *J* = 6.8 Hz, 1 H), 3.73 (s, 3 H), 3.49 (d, *J* = 5.6 Hz, 2 H), 2.80–2.74 (m, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 171.7 (C), 137.0 (C), 134.5 (CH), 128.4 (two CH), 128.14 (two CH), 128.07 (CH),

119.1 (CH₂), 78.8 (CH), 72.9 (CH₂), 51.8 (CH₃), 51.6 (CH₂), 47.0 (CH); MS (*m/z*, relative intensity): 275 (M⁺, 12), 113 (11), 112 (4), 92 (8), 91 (100), 82 (4), 65 (7); exact mass calculate for C₁₄H₁₇N₃O₃ (M⁺): 275.1270; found 275.1276.

Typical experimental procedure for the preparation of **18**

To a solution of *syn*-**17b** (548 mg, 1.45 mmol) and acetic acid (7.5 mL) in MeOH (7.5 mL) was added Pd/C (31 mg, 0.29 mmol) at room temperature. The solution was stirred under H₂ atmosphere (1 atm) for 3 days, followed by filtrating through Celite and concentrated *in vacuo* to give the crude product as brown oil. The residue was purified by flash column chromatography with 10% EtOAc-hexane (*R_f* = 0.46 in 20% EtOAc-hexane) to afford product *anti*-**18** as a yellow oil (383 mg, 86% yield).

Spectroscopic data for *anti*-**18**: IR (CH₂Cl₂): 2925, 2856, 2360, 2337, 1697, 1492, 1440, 1253, 1130, 1081, 919, 738, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.42–7.23 (m, 10 H), 5.77–5.69 (m, 1 H), 5.16–5.08 (m, 3 H), 4.87 (d, *J* = 12.0 Hz, 1 H), 4.51 (d, *J* = 14.4 Hz, 1 H), 4.40 (d, *J* = 14.4 Hz, 1 H), 3.98 (d, *J* = 8.0 Hz, 1 H), 3.34–3.28 (m, 1 H), 2.97–2.89 (m, 2 H); ¹³C NMR (CDCl₃, 100 MHz): δ 172.3 (C), 137.9 (C), 136.1 (CH), 135.9 (C), 128.8 (two CH), 128.3 (two CH), 128.2 (two CH), 128.1 (two CH), 127.8 (CH), 127.7 (CH), 117.6 (CH₂), 80.1 (CH), 72.3 (CH₂), 47.5 (CH₂), 46.7 (CH₂), 44.3 (CH); MS (*m/z*, relative intensity): 307 (M⁺, 15), 202 (5), 201 (34), 200 (6), 174 (4), 92 (9), 91 (100), 77 (4), 65 (12); exact mass calculate for C₂₀H₂₁NO₂ (M⁺): 307.1572; found 307.1575.

For *syn*-**18**: 325 mg, 78% yield; 0.38 (Hexane/EtOAc = 4 : 1); yellow oil.

Spectroscopic data for *syn*-**18**: IR (CH₂Cl₂): 2923, 2854, 1695, 1434, 1253, 1124, 738, 698 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.39–7.22 (m, 10 H), 6.00–5.91 (m, 1 H), 5.16–5.09 (m, 2 H), 4.91 (d, *J* = 12.0 Hz, 1 H), 4.78 (d, *J* = 12.0 Hz, 1 H), 4.49 (d, *J* = 14.8 Hz, 1 H), 4.45 (d, *J* = 14.8 Hz, 1 H), 4.06 (d, *J* = 6.4 Hz, 1 H), 3.28–3.18 (m, 2 H), 3.01–2.94 (m, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 171.9 (C), 137.8 (C), 136.0 (C), 133.8 (CH), 128.7 (two CH), 128.3 (two CH), 128.1 (two CH), 127.8 (two CH), 127.7 (CH), 127.6 (CH), 117.8 (CH₂), 77.8 (CH), 72.0 (CH₂), 49.3 (CH₂), 46.5 (CH₂), 41.9 (CH); MS (*m/z*, relative intensity): 307 (M⁺, 25), 202 (13), 201 (84), 200 (13), 174 (27), 110 (5), 92 (9), 91 (100), 65 (11); exact mass calculate for C₂₀H₂₁NO₂ (M⁺): 307.1572; found 307.1576.

Typical experimental procedure for the preparation of 19

To a solution of *anti*-**18** (264 mg, 0.84 mmol) in acetone (12.6 mL), H₂O (1.8 mL), and THF (10.8 mL) was added NaIO₄ (901 mg, 4.22 mmol) and a solution of OsO₄ in H₂O (0.4 M, 54 μ L). The solution was stirred at room temperature for 2 h until the completion of reaction, as monitored by TLC. To the reaction mixture was added saturated aqueous Na₂SO₃ solution (5 mL), and removed the organic solvent under vacuum. The solution was extracted with EtOAc (5 mL $\times$ 3), dried over MgSO₄, and concentrated *in vacuo* to give the crude aldehyde product as yellow oil. To the solution of this crude product in dry THF (24 mL) was added LiAlH₄ (352 mg, 9.24 mmol) at 0 °C. The solution was heated to reflux for 15 h, followed by the addition of saturated aqueous NaOH solution (0.5 mL). The solution was filtered to remove the precipitate, and the filtrate was dried over MgSO₄, concentrated *in vacuo* to give the crude product. The residue was purified by flash column chromatography with 20% EtOAc-hexane (*R_f* = 0.22 in 20% EtOAc-hexane) to afford product *anti*-**19** as a yellow oil (135 mg, 55% yield).

Spectroscopic data for *anti*-**19**: ¹H NMR (CDCl₃, 400 MHz): δ 7.35–7.23 (m, 10 H), 4.48 (s, 2 H), 4.08–4.04 (m, 1 H), 3.72 (dd, *J* = 10.0, 4.4 Hz, 1 H), 3.64 (dd, *J* = 10.0, 4.0 Hz, 1 H), 3.61 (s, 2 H), 3.16 (dd, *J* = 10.0, 6.8 Hz, 1 H), 2.75 (dd, *J* = 9.2, 6.8 Hz, 1 H), 2.67 (dd, *J* = 9.2, 2.8 Hz, 1 H), 2.40 (dd, *J* = 10.0, 4.4 Hz, 1 H), 2.32–2.30 (m, 1 H); ¹³C NMR (CDCl₃, 100 MHz): δ 138.23 (C), 138.16 (C), 128.7 (two CH), 128.40 (two CH), 128.36 (two CH), 127.68 (two CH), 127.65 (CH), 127.2 (CH), 81.3 (CH), 71.4 (CH₂), 66.1 (CH₂), 60.5 (CH₂), 60.0 (CH₂), 56.5 (CH₂), 46.3 (CH); IR (CH₂Cl₂): 3351, 3062, 3029, 2923, 2861, 2798, 1454, 1093, 1068, 738, 698 cm⁻¹; MS (*m/z*, relative intensity): 297 (M⁺, 26), 206 (5), 191 (10), 160 (6), 120 (28), 92 (9), 91 (100), 65 (5); exact mass calculate for C₁₉H₂₃NO₂ (M⁺): 297.1729; found 297.1723.

For *syn*-**19**: 214 mg, 68% yield; 0.29 (Hexane/EtOAc = 1 : 4); yellow oil.

Spectroscopic data for *syn*-**19**: IR (CH₂Cl₂): 3396, 3028, 2917, 2795, 1494, 1454, 1092, 739, 702, 666, 604 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 7.36–7.27 (m, 10 H), 4.57 (d, *J* = 12.0 Hz, 1 H), 4.42 (d, *J* = 12.0 Hz, 1 H), 4.29–4.24 (m, 1 H), 3.78–3.77 (m, 2 H), 3.64 (d, *J* = 12.8 Hz, 1 H), 3.60 (d, *J* = 12.8 Hz, 1 H), 3.03 (dd, *J* = 9.6, 6.4 Hz, 1 H), 2.82–2.77 (m, 1 H), 2.54–2.48 (m, 3 H); ¹³C NMR (CDCl₃, 100 MHz): δ 138.6 (C), 137.9 (C), 128.7 (two

CH), 128.5 (two CH), 128.3 (two CH), 127.8 (CH), 127.5 (two CH), 127.0 (CH), 79.7 (CH), 72.0 (CH₂), 61.7 (CH₂), 60.5 (CH₂), 59.5 (CH₂), 55.1 (CH₂), 43.0 (CH).

Typical experimental procedure for the preparation of 21

To a solution of *anti*-**19** (253 mg, 0.85 mmol) in MeOH (9 mL) was added Pd/C (18 mg, 0.17 mmol) at room temperature. The solution was stirred under H₂ atmosphere (1 atm) for 2 days, followed by filtrating through Celite and concentrated *in vacuo* to give the crude product as brown oil. To a solution of the residue in MeOH (8 mL) was slowly added a solution of aqueous HCl solution (12 M, 0.4 mL) at room temperature. The solution was stirred for 2 h and concentrated *in vacuo* to give *anti*-**21** as brown oil. (124 mg, 95% yield). Brown oil.

Spectroscopic data for *anti*-**21**: ¹H NMR (hydrochloride, D₂O, 400 MHz): δ 4.49–4.46 (m, 1 H), 3.73–3.63 (m, 3 H), 3.50 (dd, *J* = 12.4, 4.8 Hz, 1 H), 3.33 (d, *J* = 11.5 Hz, 1 H), 3.23 (dd, *J* = 12.4, 5.6 Hz, 1 H), 2.49–2.57 (m, 1 H); ¹³C NMR (hydrochloride, D₂O, 100 MHz): δ 71.9 (CH), 60.9 (CH₂), 52.1 (CH₂), 47.9 (CH), 46.6 (CH₂).

For *syn*-**21**: 86 mg, 78% yield; yellow oil.

Spectroscopic data for *syn*-**21**: ¹H NMR (DMSO-d₆, 400 MHz): δ 9.32 (br s, 1 H), 9.12 (br s, 1 H), 5.37 (br s, 1 H), 4.29–4.26 (m, 1 H), 3.62 (dd, *J* = 10.8, 7.2 Hz, 1 H), 3.45 (dd, *J* = 11.2, 7.2 Hz, 1 H), 3.29–3.18 (m, 2 H), 3.08–3.04 (m, 1 H), 2.91–2.85 (m, 1 H), 2.26–2.19 (m, 1 H); ¹³C NMR (DMSO-d₆, 100 MHz): δ 68.8 (CH), 58.2 (CH₂), 53.0 (CH₂), 46.1 (CH), 45.5 (CH₂).

Supporting Information Available

Spectra data for the new compounds.

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REFERENCES

- For recent reviews, see: (a) Castro, A. M. M. *Chem. Rev.* **2004**, *104*, 2939–3002. (b) Majumdar, K. C.; Alam, S.; Chattopadhyay, B. *Tetrahedron* **2008**, *64*, 597. (c) Gonda, J. *Angew. Chem. Int. Ed.* **2004**, 3516. (d) Majumdar, K. C.; Bhattacharyya, T.; Chattopadhyay, B.; Sinha, B. *Synthesis* **2009**, 2117.
- For reviews, see: (a) Chai, Y. C.; Hong, S.-P.; Lindsay, H. A.;

- McFarland, C.; McIntosh, M. C. *Tetrahedron* **2002**, *58*, 2905. For recent examples of the Ireland–Claisen Rearrangement in synthesis, see: (b) Tellam, J. P.; Kociok-Kohn, G.; Carbery, D. R. *Org. Lett.* **2008**, *10*, 5199. (c) Ylioja, P. M.; Mosley, A. D.; Charlot, C. E.; Carbery, D. R. *Tetrahedron Lett.* **2008**, *49*, 1111.
3. For a recent example, see: Tellam, J. P.; Kociok-Kohn, G.; Carbery, D. R. *Org. Lett.* **2008**, *10*, 5199.
4. For a recent example, see: Khanjin, N. A.; Snyder, J. P.; Menger, F. M. *J. Am. Chem. Soc.* **1999**, *121*, 11831.
5. Kirsten, M.; Rehbein, J.; Hiersemann, M.; Strassner, T. *J. Org. Chem.* **2007**, *72*, 4001.
6. For select examples, see: (a) Burke, S. D.; Zhao, Q. *J. Org. Chem.* **2000**, *65*, 1489–1500. (b) Hong, J. H.; Oh, C.-H.; Cho, J.-H. *Tetrahedron* **2003**, *59*, 6103–6108. (c) Marchart; Mulzer, J.; Enev, V. S. *Org. Lett.* **2007**, *9*, 813–816. (d) Jung, D. Y.; Kang, S.; Chang, S.; Kim, Y. H. *Synlett* **2006**, 86–90. (e) Mues, H.; Kazmaier, U. *Synthesis* **2001**, 3, 487–498. (f) Kazmaier, U.; Mues, H.; Krebs, A. *Chem. Eur. J.* **2002**, *8*, 1850–1855. (g) Mues, H.; Kazmaier, U. *Synthesis* **2001**, 487–498. (h) Francais, A.; Bedel, O.; Picoul, W.; Meddour, A.; Courtieu, J.; Haudrechy, A. *Tetrahedron: Asymmetry* **2005**, *16*, 1141–1156. (i) Morimoto, Y.; Takaishi, M.; Kinoshita, T.; Sakaguchi, K.; Shibata, K. *Chem. Commun.* **2002**, 42–43. (j) Wen, S.-J.; Yao, Z.-J. *Org. Lett.* **2004**, *6*, 2721–2724. (k) Izzo, I.; Avallone, E.; Corte, L. D.; Maulucci, N.; Riccardis, F. D. *Tetrahedron: Asymmetry* **2004**, *15*, 1181–1186.
7. (a) Mulzer, J.; Giester, G.; Gilbert, M. *Helv. Chim. Acta.* **2005**, *88*, 1560. (b) Burke, S. D.; Zhao, Q. *J. Org. Chem.* **2000**, *65*, 1489–1500. (c) Shin, K. J.; Moon, H. R.; George, C.; Marquez, V. E. *J. Org. Chem.* **2000**, *65*, 2172–2178. (d) Probst, N. P.; Haudrechy, A.; Plé, K. *J. Org. Chem.* **2008**, *73*, 4338–4341. (e) Hong, J. H.; Oh, C.-H.; Cho, J.-H. *Tetrahedron* **2003**, *59*, 6103–6108. (f) Français, Bedel, O.; Picoul, W.; Meddour, A.; Courtieu, J.; Haudrechy, A. *Tetrahedron: Asymmetry* **2005**, *16*, 1141–1155. (g) Picoul, W.; Urchegui, R.; Haudrechy, A.; Langlois, Y. *Tetrahedron Lett.* **1999**, *40*, 4797–4800. (h) Davies, S. G.; Garner, A. C.; Nicholson, R. L.; Osborne, J.; Roberts, P. M.; Savory, E. D.; Smith, A. D.; Thomson, J. E. *Org. Biomol. Chem.* **2009**, *7*, 2604–2611. (i) Izzo, I.; Avallone, E.; Corte, L. D.; Maulucci, N.; Riccardis, F. D. *Tetrahedron: Asymmetry* **2004**, *15*, 1181–1186. (j) Morimoto, Y.; Takaishi, M.; Kinoshita, T.; Sakaguchi, K.; Shibata, K. *Chem. Commun.* **2002**, 42–43. (k) Sakaguchi, K.; Yamamoto, M.; Kawamoto, T.; Yamada, T.; Shinada, T.; Shimamoto, K.; Ohfuné, Y. *Tetrahedron Lett.* **2004**, *45*, 5869–5872. (l) Kazmaier, U.; Paehler, S.; Endermann, R.; Haebich, D.; Kroll, H.-P.; Riedl, B. *Bioorg. Med. Chem.* **2002**, *10*, 3905–3914. (m) Pal, U.; Del Valle, J. R.; Ranatunga, S.; Ariyaratna, Y. *Org. Lett.* **2009**, *11*, 5298–5301. (n) Sharma, V.; Kelly, G. T.; Watanabe, C. M. H. *Org. Lett.* **2008**, *10*, 4815–4818.
8. For an only example, see: Hauske, J. R.; Julin, S. M. *Tetrahedron Lett.* **1993**, *34*, 4909–4912.
9. (a) Huang, H.; Greenberg, M. M. *J. Org. Chem.* **2008**, *73*, 2695–2703. (b) Marquez, V. E.; Wang, P.; Nicklaus, M. C.; Maier, M.; Manoharan, M.; Christman, J. K.; Banavali, N. K.; Mackerell, A. D. *Nucleosides Nucleotides* **2001**, *20*, 451–460. (c) Besada, P.; Gonzalez Moa, M. J.; Teran, C. *Synthesis* **2008**, 2363–2368. (d) Ok, D.; Li, C.; Abbadie, C.; Felix, J. P.; Fisher, M. H.; Garcia, M. L.; Kaczorowski, G. J.; Lyons, K. A.; Martin, W. J.; Priest, B. T.; Smith, M. M.; Brande, S. W.; Matthew, J. W.; William, H. P. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 1358–1361.
10. For preparation conditions, see: (a) Najdi, S. D.; Olmstead, M. M.; Schore, N. E. *J. Organometal. Chem.* **1992**, *431*, 335–358. (b) Toshima, H.; Aramaki, H.; Ichihara, A. *Tetrahedron Lett.* **1999**, *40*, 3587–3590. (c) MacMahon, S.; Fong, R.; Baran, P. S.; Safonov, I.; Wilson, S. R.; Schuster, D. I. *J. Org. Chem.* **2001**, *66*, 5449–5455. (d) Schering, A. G. US Patent 7,001,916, 2006. (e) Kawamura, T.; Inagaki, F.; Narita, S.; Takahashi, Y.; Hirata, S.; Kitagaki, S.; Mukai, C. *Chem. Eur. J.* **2010**, *16*, 5173–5183. (f) Pour, M.; Negishi, E.-I. *Tetrahedron Lett.* **1996**, *37*, 4679–4682.
11. For preparation, see: (a) Marshall, J. A.; Garofalo, A. W. *J. Org. Chem.* **1996**, *61*, 8732–8738. (b) Righi, G.; Pescatore, G.; Bonadies, F.; Bonini, C. *Tetrahedron* **2001**, *57*, 5649–5656. (c) Mauleon, P.; Krinsky, J. L.; Toste, F. D. *J. Am. Chem. Soc.* **2009**, *131*, 4513–4520. (d) Soulie, J.; Ta, C.; Lallemand, J.-Y. *Tetrahedron* **1992**, *48*, 443–452. (e) Mitchell, M.; Qiao, L.; Wong, C.-H. *Adv. Synth. Cat.* **2001**, *343*, 6–7. (f) Lohse-Fraefel, N.; Carreira, E. M. *Chem. Eur. J.* **2009**, *15*, 12065–12081. (g) Kumaraswamy, G.; Ramakrishna, D.; Santhakumar, K. *Tetrahedron: Asymmetry* **2010**, *21*, 544–548. (h) Sodeoka, M.; Yamada, H.; Shibasaki, M. *J. Am. Chem. Soc.* **1990**, *112*, 4906–4911. (i) Ding, P.; Miller, M. J.; Chen, Y.; Helquist, P.; Oliver, A. Jayne; W. O. *Org. Lett.* **2004**, *6*, 1805–1808.
12. The synthetic material was identical in all respects (^1H , ^{13}C NMR) to the literature data: Karlsson, S.; Hogberg, H. E. *Tetrahedron: Asymmetry* **2001**, *12*, 1977–1982.