

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 16 (2006) 1605-1609

Acetylenic TACE inhibitors. Part 3: Thiomorpholine sulfonamide hydroxamates

J. I. Levin,^{a,*} J. M. Chen,^a L. M. Laakso,^a M. Du,^a J. Schmid,^a W. Xu,^c T. Cummons,^b J. Xu,^a G. Jin,^a D. Barone^d and J. S. Skotnicki^a

^aWyeth Research, 401 N. Middletown Rd., Pearl River, NY 10965, USA ^bWyeth Research, PO Box CN-8000, Princeton, NJ 08543, USA ^cWyeth Research, 200 Cambridge Park Drive, Cambridge, MA 02140, USA ^dAmgen, Seattle, WA 98101, USA

Received 16 September 2005; revised 5 December 2005; accepted 7 December 2005 Available online 19 January 2006

Abstract—A series of thiomorpholine sulfonamide hydroxamate TACE inhibitors, all bearing propargylic ether P1' groups, was explored. In particular, compound **5h** has excellent in vitro potency against isolated TACE enzyme and in cells, oral activity in a model of TNF- α production and a collagen-induced arthritis model, was selected as a clinical candidate for the treatment of RA. © 2005 Elsevier Ltd. All rights reserved.

The commercial success of biologics, including Enbrel[®] and Remicade[®], that modulate levels of both membrane-bound 26 kDa TNF and the soluble 17 kDa form of this pro-inflammatory cytokine has led to an intensive search for orally active small molecules that might be similarly effective in the treatment of rheumatoid arthritis (RA).¹

One attractive approach for affecting TNF levels is the inhibition of TNF- α converting enzyme (TACE/ADAM-17), the metalloprotease responsible for the shedding of membrane-bound TNF, thereby reducing circulating levels of soluble TNF.² Among the compounds directed at this mechanism of action are selective inhibitors of TACE, including the clinical candidate BMS-561392 (1, Fig. 1),^{3a,b} and those that inhibit enzymes from the related family of matrix metalloproteinases (MMPs) in addition to TACE, like PKF242-484 (2, Fig. 1)^{3c} and clinical candidate Ro 32-7315 (3, Fig. 1).^{3d} The issue of whether broad spectrum inhibition of TACE and MMPs, or more selective inhibition of TACE, would be optimal remains to be settled by RA clinical trials. Many broad spectrum MMP



Figure 1. Literature TACE inhibitors.

inhibitors have displayed dose-limiting toxicity in oncology and osteoarthritis clinical trials,⁴ but the inhibition of MMPs that are overexpressed in synovial tissue and degrade cartilage in RA joints may provide a more effective therapeutic.⁵

We, and others, have previously disclosed several series of sulfonamide hydroxamic acid TACE inhibitors bearing novel propargylic P1' groups, exemplified by TMI-1 (4, Fig. 1).⁶ Compound 4 is an excellent

Keywords: TACE inhibitor; Thiomorpholine; Sulfonamide: hydroxamate; Rheumatoid arthritis.

^{*}Corresponding author. Tel.: +1 845 602 3053; fax: +1 845 602 5561; e-mail: levinji@wyeth.com

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter @ 2005 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2005.12.020

inhibitor of TACE and many MMPs, with good potency in human whole blood and potent oral activity in an animal model of RA.^{6b} Unfortunately, poor solubility and poor bioavailability in rat and dog precluded the development of compound **4** as a clinical candidate. We now report our efforts to optimize the potency, physical properties, and bioavailability of the thiomorpholine sulfonamides by varying the P1' terminus of these inhibitors and appending substituents on the thiomorpholine ring (**5a**–**y**, Table 1), resulting in a clinical candidate for the treatment of RA.

Initial attempts at increasing the potency and selectivity of **4** focused on the size of the alkynyl ether P1' moiety and the position of the carbon–carbon triple bond (Scheme 1), which had previously been optimized on an anthranilate-derived scaffold.^{6a} Sulfonylation of thiomorpholine methyl or *t*-butyl ester **6**⁷ with in situ silylated 4-hydroxybenzenesulfonyl chloride provided phenol **7** after work-up with methanol.⁸ The phenol next underwent Mitsunobu alkylation with the selected alcohol, followed by cleavage of the ester and conversion of the resulting carboxylate into the desired hydroxamic acids **5a–5g**.

Table 1. In vitro potency of cyclic sulfonamide hydroxamic acids



Compound	R ¹	R ²	TACE ^a	THP ^b	MMP-l ^a	MMP-13 ^a
4 (TMI-1)	CH ₂ CCCH ₃	Н	8	94/87	7	3
5a	CH ₂ CCH	Н	19	91/76	1	1
5b	(CH ₂) ₂ CCH	Н	32	64/36	2	1
5c	(CH ₂) ₃ CCH	Н	40	38/16	11	8
5d	CH ₂ CHCHCH ₃	Н	35	80/50	7	2
5e	$(CH_2)_3CH_3$	Н	42	38/11	7	9
5f	CH ₂ CC(CH ₂) ₃ CH ₃	Н	23	82/48	29	7
5g	CH ₂ CCCCH	Н	42	88/62	6	11
5h	CH ₂ CCCH ₂ OH	Н	20	95/89	33	8
5i	CH ₂ CC(CH ₂) ₂ OH	Н	62	57/25	35	14
5j	CH ₂ CC(CH ₂) ₃ OH	Н	40	51/19	159	86
5k	CH ₂ CC(CH ₂) ₄ OH	Н	53	77/52	101	17
51	CH ₂ CCCH ₂ NHBoc	Н	74	79/53	26	4
5m	CH ₂ CC(CH ₂) ₂ NHBoc	Н	112	37/14	300	7
5n	CH ₂ CC(CH ₂) ₃ NHBoc	Н	153	27/5	42	2
50	CH ₂ CC(CH ₂) ₄ NHBoc	Н	199	70/40	87	4
5p	CH ₂ CCCH ₂ NH ₂	Н	67	40/10	97	8
5q	CH ₂ CC(CH ₂) ₂ NH ₂	Н	138	4/0	1052	27
5r	CH ₂ CC(CH ₂) ₃ NH ₂	Н	28	22/0	226	9
5s	CH ₂ CC(CH ₂) ₄ NH ₂	Н	40	37/13	92	7
5t	CH ₂ CCCH ₃	CH ₂ CO ₂ H	20	89/73	17	7
5u	CH ₂ CCCH ₃	CH ₂ CONH ₂	42	91/76	35	29
5v	CH ₂ CCCH ₃	CH ₂ NH ₂	57	94/78	21	24
5w	CH ₂ CCCH ₃	CH ₂ NHSO ₂ CH ₃	11	91/81	19	14
5x	CH ₂ CCCH ₃	$(CH_2)_2OH$	15	90/74	21	23
5у	CH ₂ CCCH ₃	(CH ₂) ₂ NH(CH ₂ CH ₃) ₂	143	87/67	33	48

^a IC₅₀ (nM).

 $^{b}\%$ Inhibition at 3/1 $\mu M.$



Scheme 1. Reagents: (i) a—4-HOPhSO₂Cl, BTSA; b—MeOH; (ii) R¹OH, PPh₃, DEAD; (iii) HCl (g) or LiI; (iv) a—(COCl)₂, DMF; b—NH₂OH.

In an effort to achieve additional interactions between the protein and the inhibitor, we also examined alcohol, amine, and carbamate derivatives of the butynyl P1' moiety that could potentially contact the glutamate and valine residues in the TACE S3' subsite. The alcohols were prepared as shown in Scheme 2. The one carbon tether was prepared by direct Mitsunobu alkylation of 7 with 2-butyn-1,4-diol. Acetylation of the resulting alcohol gave 8. Ester cleavage of 8 with TFA, and conversion of the unmasked carboxylic acid to the hydroxamic acid, provided 5h. The two-, three-, and four-carbon tethers were prepared starting from THP-protected 3butyn-1-ol, 4-pentyn-1-ol, and 5-hexyn-1-ol. These alkynes were metallated with *n*BuLi and the resulting anion



Scheme 2. Reagents: (i) $HOCH_2CCCH_2OH$, PPh_3 , DEAD; (ii) Ac_2O ; (iii) HCl (g); (iv) $a-(COCl)_2$, DMF; $b-NH_2OH$; (v) $HOCH_2CC(CH_2)_nOTHP$, PPh_3 , DEAD; (vi) LiI, EtOAc; (vii) $a-(COCl)_2$, DMF or EDC, HOBT; $b-NH_2OH$; (viii) PPTS.

was quenched with formaldehyde. Mitsunobu alkylation of 7b then gave 9, followed by ester cleavage and hydroxamate formation, and subsequent removal of the THP moiety to give 5i-5k.

The Boc carbamates **5**I–**50** and the primary amine derivatives **5**p–**5**s were prepared from the protected alcohol intermediates **8** and **9** according to Scheme 3. Thus, after deprotection of **8** and **9** the alcohols were converted into the propargylic bromides, followed by displacement with sodium azide, reduction of the resulting azide to the primary amine, and Boc protection to give **10**. The ester moiety was next converted into the desired hydroxamate in two steps to give carbamates **5**I–**50**. Removal of the Boc group then afforded the primary amines **5**p–**5**s as TFA or HCl salts.

Finally, analysis of computer models of compound 4 bound to the active site of TACE indicated that the 6-position (see \mathbb{R}^2 in 5, Table 1) of the thiomorpholine is solvent exposed, and that substituents affecting the physical properties of the molecule might be placed there with minimal loss of enzyme inhibitory activity. A similar strategy has been used on a series of pipecolic



Scheme 3. Reagents: (i) MeOH, K_2CO_3 (for 8), PPTS (for 9) (ii) PPh₃, CBr₄; (iii) NaN₃; (iv) Bu₃P, THF, H₂O; (v) Boc₂O; (vi) LiI; (vii) a—HOBt, EDC; b—NH₂OH; (viii) HCl (g) or TFA.



Scheme 4. Reagents: (i) a—HCl (g); b—SOCl₂, MeOH; (ii) 4butynyloxybenzene sulfonyl chloride, TEA; (iii) BrCH₂CH=CHCO₂*t*-Bu, K₂CO₃; (iv) Bu₃P, THF, H₂O.

acid-based TACE inhibitors.⁹ The preparation of these compounds, **5t–5y**, from D-penicillamine disulfide (**11**) utilizing a key diastereoselective intra-molecular Michael reaction to construct the thiomorpholine ring of **13** from **12**, has been reported elsewhere and is summarized briefly in Scheme $4.^{10}$

All of the sulfonamide hydroxamic acids were tested in vitro¹¹ for their ability to inhibit MMP-1, MMP-13, and TACE.¹² Activity against MMP-1 was assessed since the inhibition of this enzyme had been postulated to contribute to musculoskeletal side effects seen for MMP inhibitors.⁴ The inhibition of MMP-13 was measured since this may help prevent the degradation of cartilage in RA.⁵ The ability of compounds to inhibit TNF production in LPS-stimulated THP-1 cells was also assessed, although IC₅₀s were determined for only the most active analogs.¹³ The in vitro potencies for the cyclic sulfonamide hydroxamic acid analogs bearing a variety of P1' groups are shown in Table 1.

As for the analogs of anthranilate-based TACE inhibitors,^{6a} the shorter propargyl ether P1' analog **5a** is substantially more active than butynyl ether derivative 4 against MMP-1, while slightly less active against TACE. Increasing the length of the linker between the alkyne and the P1' ether oxygen (5b-5c) progressively decreased MMP-1 and TACE activity, and activity against LPSstimulated TNF production in THP-1 cells. The transolefin analog 5d is 4-fold less potent than 4 against cell-free TACE and also slightly less active than 4 in THP-1 cells. This trend continues with the fully saturated butyl ether P1' derivative 5e, which has dramatically less cellular activity than 4 and 5d, though it is essentially equipotent to 5d against cell-free TACE. Lengthening of the butynyl ether to the heptynyl ether 5f or the rigid divne 5g results in a slight loss of activity against both cell-free TACE and in cells. That analogs 5f and 5g do not increase selectivity for TACE over MMP-1, though the P1' groups of both can clearly extend past the arginine side chain that normally forms the base of the shallow MMP-1 S1' pocket, confirms that this residue is mobile and can move to accommodate larger P1' substituents on the ligand.¹⁴ All of compounds **5a–5g** remain potent inhibitors of MMP-13.

The alcohols 5h-5k were all less active than the butynyl analog 4 in the TACE FRET assay, with the shortest analog, 5h, being the most potent. TACE activity does not fall off substantially with increasing length, but activity in THP cells decreases dramatically in going from **5h** (89% inhibition at $1 \mu M$, IC₅₀ = 140 nM) to the pentynyl and hexynyl alcohols 5i and 5j (<25% inhibition at 1 µM). Interestingly, the heptynyl alcohol 5k recovers substantial cell activity (52% inhibition at 1 μ M), relative to 5i-j. The same general trend is seen for carbamates 51-50. Despite their unimpressive activity against TACE enzyme, at least 80-fold less potent than 4, both 51 and 50 display modest cellular activity $(\geq 40\%)$ at 1 µM, while 5m and 5n are only weakly active even at $3 \mu M$. The amines **5p** and **5q** are essentially equivalent in activity to their corresponding carbamates, 51 and 5m, against TACE, but are far less potent in the THP cellular assay. Furthermore, the hexynyl and heptynyl amines, 5r and 5s, are both approximately 5fold more active in the TACE FRET assay than the analogous carbamates, 5n and 5o, but are less active in cells. Although all of the amines are only weakly active in cells, the butynyl and heptynyl analogs are again the most potent, as in the alcohol and carbamate series. All of the alcohols, carbamates, and amines, 5h-5s, maintain excellent activity against MMP-13, with little or no selectivity for TACE over MMP-1.

An X-ray crystal structure has been obtained for 5p bound to the active site of TACE (Fig. 2).¹⁵ It is interesting to note that the amine moiety of 5p is within hydrogen bonding distance to both a valine residue and a glutamate residue in the S3' pocket of TACE, yet it is less active against the enzyme than 4, which makes neither of these contacts.

The in vitro activity of the 6-substituted thiomorpholines **5t–5y** demonstrates that a variety of functionality



Figure 2. X-ray structure of compound 5p bound to TACE.

is tolerated at this position, as was expected. Thus, the carboxylic acid 5t is only 2-fold less active than 4 against cell-free TACE and slightly less active than 4 in THP cells at $1 \mu M$. The corresponding primary amide 5u and the primary amine 5v are at least 5-fold less active than 4 in the FRET assay, but still sub-micromolar in cells. Sulfonamide 5w and alcohol 5x are the most potent of the 6-substituted thiomorpholines in the TACE FRET assay, similar in potency to 4, and both are sub-micromolar inhibitors in THP cells. Diethylamine 5y, like the other analog bearing a basic amine at the thiomorpholine 6-position (5v), loses significant enzyme activity in the FRET assay, 18-fold compared to 4, but is still reasonably potent in THP cells, affording 67% inhibition of LPS-stimulated TNF production at $1 \mu M$. We have previously noted that discrepancies between the level of enzyme activity in the TACE FRET assay and in THP cells are frequently not attributable to cell permeability and protein binding issues.¹⁶

The in vivo activity of the compounds in Table 1 with the greatest activity in the THP assay was initially measured by their ability to inhibit the LPS-stimulated production of TNF- α after oral dosing in a mouse.¹³ Of compounds 5a-5y, several compounds equaled or exceeded the level of activity of 4 in this model at a 50 mg/kg po dose. When dosed orally at 50 mg/kg, 5h, 5v, 5x, and 5y each provide \geq 95% inhibition of TNF- α levels one hour after administration of LPS. The most potent analog in this model is propargylic alcohol derivative 5h with an ED₅₀ of 5 mg/kg po. Analogs 5h and 5v were also examined in a mouse prophylactic collagen-induced arthritis (CIA) efficacy model,¹³ using an LPS boost, and both gave statistically significant ($p \leq 0.05$) reductions in clinical severity scores at 10 mg/kg bid po. In fact, in this model a 10 mg/kg po dose of 5h provides a > 50% reduction in clinical severity score $(p \leq 0.05, n = 15)$, comparable to a 30 µg qd dose of Enbrel[®]. Also, in human whole blood 5h inhibits LPS-stimulated TNF production with an IC_{50} of 300 nM. Analysis of in vitro stability of 5h in liver microsomes from rat, dog, monkey, and human shows diminished levels of metabolites resulting from loss of the P1' moiety, as compared to 4. In addition, 5h is permeable in Caco-2 cells (Mean $P_{\rm app} = 2.2 \times 10^{-6}$ cm/s), is not highly protein bound (~50%), and has vastly improved solubility in simulated gastric and intestinal fluids (>1.5 mg/ mL) as compared to 4 (<0.08 mg/mL). Perhaps as a result of these properties, bioavailability of 5h at 10 mg/kg (mouse: 24%, rat: 8%, and dog: 36%) is in general better than that of **4** (mouse: 71%, rat: 5%, and dog: 0%), particularly in dogs.

In summary, we have expanded a series of thiomorpholine sulfonamide hydroxamates bearing a propargylic P1' group as inhibitors of MMPs and TACE. One of these compounds, **5h**, has activity equivalent to that of compound **4** in an animal model of RA, but with improved solubility and PK properties, and has been advanced to clinical trials. The complete pharmacological profile of propargylic alcohol **5h** (TMI-005) will be reported in due course.¹⁷

Acknowledgments

We thank A. Sung, S. Skala, and R. Mulvey for technical support in the MMP and THP assays, and R. A. Black, R. Cowling, L. Killar, K. M. Mohler, and Y. Zhang for their insight and guidance during the course of this work. We also thank U. Jain and the PCC group for physical data, and C. Huselton and group for PK data.

References and notes

- (a) Newton, R. C.; Decicco, C. P. J. Med. Chem. 1999, 42, 2295; (b) Palladino, M. A.; Bahjat, F. R.; Theodorakis, E. A.; Moldawer, L. L. Nat. Rev. Drug Disc. 2003, 2, 736; (c) Skotnicki, J. S.; Levin, J. I. Ann. Rep. Med. Chem. 2003, 38, 153.
- (a) Newton, R. C.; Solomon, K. A.; Covington, M. B.; Decicco, C. P.; Haley, P. J.; Friedman, S. M.; Vaddi, K. *Ann. Rheum. Dis.* 2001, *60*, iii25; (b) Moss, M. L.; Bartsch, J. *Biochemistry* 2004, *43*, 7227.
- (a) Grootveld, M.; McDermott, M. F. Curr. Opin. Invest. Drugs 2003, 4, 598; (b) Duan, J. J.-W.; Lu, Z.; Xue, C.-B.; He, X.; Seng, J. L.; Roderick, J. J.; Wasserman, Z. R.; Liu, R.-Q.; Covington, M. B.; Magolda, R. L.; Newton, R. C.; Trzaskos, J. M.; Decicco, C. P. Bioorg. Med. Chem. Lett. 2003, 13, 2035; (c) Trifilieff, A.; Walker, C.; Keller, T.; Kottirsch, G.; Neumann, U. Br. J. Pharmacol. 2002, 135, 1655; (d) Beck, G.; Bottomley, G.; Bradshaw, D.; Brewster, M.; Broadhurst, M.; Devos, R.; Hill, C.; Johnson, W.; Kim, H.-J.; Kirtland, S.; Kneer, J.; Lad, N.; Mackenzie, R.; Martin, R.; Nixon, J.; Price, G.; Rodwell, A.; Rose, F.; Tang, J.-P.; Walter, D. S.; Wilson, K.; Worth, E. J. Pharmacol. Exp. Ther. 2002, 302, 390.
- For excellent discussions of MMP inhibition and related side effects, see: (a) Peterson, J. T. *Heart Fail. Rev.* 2004, 9, 63; (b) Renkiewicz, R.; Qiu, L.; Lesch, C.; Sun, X.; Devalaraja, R.; Cody, T.; Kaldjian, E.; Welgus, H.; Baragi, V. *Arthritis Rheum.* 2003, 48, 1742.
- (a) Yoshihara, Y.; Nakamura, H.; Obata, K.; Yamada, H.; Hayakawa, T.; Fujikawa, K.; Okada, Y. Ann. Rheum. Dis. 2000, 59, 455; (b) Konttinen, Y. T.; Ainola, M.; Valleala, H.; Ma, J.; Ida, H.; Mandelin, J.; Kinne, R. W.; Santavirta, S.; Sorsa, T.; Lopez-Otin, C.; Takagi, M. Ann. Rheum. Dis. 1999, 58, 691.
- (a) Chen, J. M.; Jin, G.; Sung, A.; Levin, J. I. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1195; (b) Levin, J. I.; Chen, J. M.; Laakso, L. M.; Du, M.; Du, X.; Venkatesan, A. M.; Sandanayaka, V.; Zask, A.; Xu, J.; Xu, W.; Zhang, Y.; Skotnicki, J. S. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 4345;

(c) Tsukida, T.; Moriyama, H.; Inoue, Y.; Kondo, H.; Yoshino, K.; Nishimura, S. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 1569; (d) Moriyama, H.; Tsukida, T.; Inoue, Y.; Yokota, K.; Yoshino, K.; Kondo, H.; Miura, N.; Nishimura, S.-I. *J. Med. Chem.* **2004**, *47*, 1930.

- Bender, S.L.; Melnick, M. J. U.S. Patent 5,753,653, 1998; Chem. Abstr. 1998, 129, 16140.
- Levin, J. I.; Du, M. T.; Park, K. Synthetic Commun. 2004, 34, 2773.
- Letavic, M. A.; Axt, M. Z.; Barberia, J. T.; Carty, T. J.; Danley, D. E.; Geoghegan, K. F.; Halim, N. S.; Hoth, L. R.; Kamath, A. V.; Laird, E. R.; Lopresti-Morrow, L. L.; McClure, K. F.; Mitchell, P. G.; Natarajan, V.; Noe, M. C.; Pandit, J.; Reeves, L.; Schulte, G. K.; Snow, S. L.; Sweeney, F. J.; Tan, D. H.; Yu, C. H. *Bioorg. Med. Chem. Lett.* 2002, *12*, 1387.
- Laakso, L. M.; Levin, J. I.; Du, M.; Jin, G.; Cowling, R.; Skala, S.; Cummons, T.; Xu, J.; Barone, D.; Skotnicki, J. S. *Abstracts of Papers*, 226th National Meeting of the American Chemical Society, New York, NY, Sept 7–11, 2003; MEDI 116. Manuscript in preparation.
- 11. (a) Weingarten, H.; Feder, J. Anal. Biochem. **1985**, 147, 437(b) Inhibitor concentrations were run in triplicate. MMP IC_{50} determinations were calculated from a 4-parameter logistic fit of the data within a single experiment.
- Jin, G.; Huang, X.; Black, R.; Wolfson, M.; Rauch, C.; McGregor, H.; Ellestad, G. A.; Cowling, R. Anal. Biochem. 2002, 302, 269.
- For a description of all cell and in vivo assays, see: Zhang, Y.; Xu, J.; Levin, J.; Hegen, M.; Li, G.; Robertshaw, H.; Brennan, F.; Cummons, T.; Clarke, D.; Vansell, N.; Nickerson-Nutter, C.; Barone, D.; Mohler, K.; Black, R.; Skotnicki, J.; Gibbons, J.; Feldman, M.; Frost, P.; Larsen, G.; Lin, L.-L. J. Pharmacol. Exp. Ther. 2004, 309, 348.
- Lovejoy, B.; Welch, A. R.; Carr, S.; Luong, C.; Broka, C.; Hendricks, R. T.; Campbell, J. A.; Walker, K. A. M.; Martin, R.; Van Wart, H.; Browner, M. F. *Nat. Struct. Biol.* 1999, *6*, 217.
- 15. PDB deposition number: 2A8H.
- 16. Levin, J. I.; Du, M. T. Drug Design Discov. 2003, 18, 123.
- (a) Zhang, Y.; Xu, J.; Udata, C.; McDevitt, J.; Cummons, T.; Zeng, C.; Sun, L.; Zhu, Y.; Gibbons, J.; Skotnicki, J.; Lin, L.-L.; Levin, J. *Scientific Abstracts*, Annual European Congress of Rheumatology, Vienna, Austria, June 8–11, 2005; Abstract 1812.; (b) Hegen, M.; Zhang, Y.; Levin, J.; Xu, J.; Cummons, T.; Harding, K.; Sheppard, B. J.; Leach, M. W.; Lin, L.-L.; Gibbons, J.; Skotnicki, J.; Nickerson-Nutter, C. L. *Scientific Abstracts*, Annual European Congress of Rheumatology, Vienna, Austria, June 8–11, 2005; Abstract 157.