High Selectivity from Configurational Match/Mismatch in **Carbon-Hydrogen Insertion Reactions of Steroidal Diazoacetates** Catalyzed by Chiral Dirhodium(II) Carboxamidates

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Diazo decomposition of steroidal diazoacetates, where the point of attachment is the 3-position of the steroid A-ring, catalyzed by chiral dirhodium(II) carboxamidates results in products from carbon-hydrogen insertion in high yield and selectivities. Use of S-configured catalysts shows a distinctive preference for insertion into the 3-position to form β -lactone products. The *R*-configured catalysts direct insertion preferentially to the equatorial C-H bond at the 2-position. Substituents or functional groups at the 5/6-position prevent C-H insertion from taking place at the 4-position. Even in the best case with the 5/6-positions fully saturated, however, insertion into the 3-position remains competitive with insertion into the 4-position. Corresponding 3-substituted phenyldiazoacetates give only β -lactone products, and selectivity here is highest with chiral dirhodium(II) prolinate catalysts. A model is presented to explain these results. Overall, this methodology is versatile for functionalization of the steroid A-ring at positions 2 and 3.

We previously reported exceptional diastereoselection and regioselection in chiral catalyst controlled intramolecular carbon-hydrogen insertion reactions of cyclohexyl diazoacetates.^{1,2} Using enantiomerically pure 2substituted cyclohexyl diazoacetates and a series of chiral dirhodium(II) carboxamidates, virtually complete diastereocontrol was achieved when there was a match between substrate and catalyst configurations (e.g., Scheme 1).¹ When there was a significant mismatch, mixtures of products were obtained. Similar selectivities were obtained for reactions of conformationally restricted 4-substituted cyclohexyl diazoacetates;^{2,3} there are no reported examples using 3-substituted cyclohexyl diazoacetates.

Carbon-hydrogen insertion normally takes place to form five-membered ring products,⁴⁻⁷ but there is an increasing number of examples of four-membered ring formation, especially when insertion can occur adjacent to a heteroatom, especially oxygen and nitrogen.^{4,8-10} Furthermore, reactivity for insertion into C-H bonds

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Scheme 1 Rh₂(5S-MEPY) CH₂Cl₂ Me 94% Rh₂(4R-MPPIM)₄ N₂ Me (1S,2R)-1 CH₂Cl₂ 98% C 2

follows the order tertiary > secondary >> primary,¹¹ but conformational influences can and do override electronic preferences.¹² Insertion into an equatorial C-H bond is favored over insertion into an axial C-H bond,^{1,2} although an exception has been noted.13

Although there has continued to be a flurry of activity in metal-catalyzed C-H insertion reactions,⁴ and high selectivity continues to be achieved,¹⁴ there have been few reports of the influence of remote substituents on selectivity.⁴⁻⁷ Steroidal systems are especially useful

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Table 1. Products from Catalytic Diazo Decomposition of Cholesteryl Diazoacetate (8)^a

		rel yield, %	
catalyst	product yield, ^b %	9	10
$Rh_2(5R-MEPY)_4$	81	94	6
$Rh_2(4R-MEOX)_4$	81	89	11
$Rh_2(5S-MEPY)_4$	74	33	67
$Rh_2(4S-MEOX)_4$	80	10	90
$Rh_2(4S-MEAZ)_4$	69	27	73
$Rh_2(4S-IBAZ)_4$	31 ^c	32	68
$Rh_2(OAc)_4$	$<5^{c}$		

^a Reactions were performed in refluxing CH₂Cl₂ using 1.0 mol % of catalyst. b Weight yield after chromatographic separation of catalyst. ^c Major product was that from O-H insertion.

substrates for such investigations because of their conformational rigidity, but they have previously received only a minimum of attention with diazo decomposition reactions.⁴ We wish to report that exceptional diastereoselectivity and regiocontrol can be achieved in chiral dirhodium(II) carboxamidate catalyzed reactions of steroidal-3-diazoacetates and that there exists a surprising effective competition between γ - and β -lactone formation that is dependent on the configurational match between substrate and catalyst.

Results and Discussion

Cholest-5-en-3 β -yl diazoacetate (8) was selected for the initial survey. Prepared from cholesterol by diketene condensation, diazo transfer, and deacetylation in 68% overall yield, this diazoacetate was subjected to standard conditions for diazo decomposition¹ with the series of chiral dirhodium(II) carboxamidate catalysts 4-7.15-17



Two products were obtained (eq 1) with γ -lactone 9 favored by the *R*-configured catalysts and β -lactone **10** dominant with the S-configured catalysts (Table 1). A product from C-H insertion into the steroidal 4-position could not be confirmed and, if present, was only a trace constituent. The reason for the absence of insertion into the 4-position is not obvious. Significantly, the use of the achiral catalyst Rh₂(OAc)₄ resulted in a low yield of insertion products when reactions were performed under the same conditions. The γ -lactone product (9) stereochemistry was ascertained by NMR analyses.



High product yields and high selectivities (\geq 9:1) can be achieved with Rh₂(MEPY)₄ and Rh₂(MEOX)₄ catalysts. Lower selectivities and lower product yields occur in reactions catalyzed by Rh₂(IBAZ)₄ and Rh₂(MPPIM)₄.¹⁸ Use of chiral bis-oxazoline ligands **11** for copper(I) complexes¹⁹ from Cu(MeCN)₄PF₆ allowed C-H insertion to take place, but selectivities were low (9/10 = 72:28 with S-11 and 37:63 with R-11).



With the dihydrogen analogue of **8**, 5α -cholestan- 3β yl diazoacetate (12a), a strikingly different outcome occurred as a result of competition for C-H insertion at the 4-position. Three products were isolated in excellent vields (eq 2), and the results from reactions with a series of dirhodium(II) carboxamidate catalysts are reported in Table 2. That the stereochemistry of the γ -lactone products was trans in 13a and 15a was confirmed by NMR through 2D HSQC and 1D NOE experiments.



The *R*-configured ligands on dirhodium(II) do favor attack at the equatorial C-H bond at the 2-position, and the formation of 15a also occurs through attack at an equatorial C-H bond. This is consistent with prior

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 Table 2.
 Products from Catalytic Diazo Decomposition of Cholestanyl Diazoacetate (12a)^a

		rel yield, %		
catalyst	product yield, ^b %	13a	14a	15a
Rh ₂ (5 <i>R</i> -MEPY) ₄	89	72	20	8
$Rh_2(4R-MEOX)_4$	86	60	33	7
$Rh_2(4R-MPPIM)_4$	86	87	8	5
$Rh_2(5S-MEPY)_4$	87	6	62	32
$Rh_2(4S-MEOX)_4$	91	4	70	26
$Rh_2(4S-MPPIM)_4$	76	16	66	20
$Rh_2(4S-MEAZ)_4$	85	6	39	55
$Rh_2(4S-BNAZ)_4$	80	7	44	49

^{*a*} Reactions were performed in refluxing CH₂Cl₂ using 1.0 mol % of catalyst. ^{*b*} Weight yield after chromatographic separation of catalyst.

investigations.^{1,4} However, the competition between β -lactone (**14a**) and γ -lactone (**15a**) formation with the *S*configured catalysts is surprising and probably results from constraints placed on the configuration of the reacting metal carbene in its approach to the equatorial C–H bond at the 4-position. These constraints could be from substitution at the 5-position or from the angular methyl group at the 10-position. In any case, the fact that one or both of these remote substituents profoundly influences regiocontrol is an indication of the strength of these catalyst interactions. Virtually identical results were obtained with 5α -androstan-17-on- 3β -yl diazoacetate (**12b**), and these data are reported in Table 3; however, note that Rh₂(OAc)₄ offers virtually no selectivity for insertion.

Diazo decomposition of the 6-keto analogue of cholesteryl diazoacetate (**16**) for which the stereoelectronic influence of the 6-keto group was expected to inhibit C–H insertion β to this functional group²⁰ gave results that greatly favored formation of the β -lactone product (eq 3).



Results from catalytic reactions with representative chiral dirhodium carboxamidates are reported in Table 4. In only one other case, that of the diazo decomposition of nonracemic 2-octyl diazoacetate,¹ have we seen such a high tendency to undergo β -lactone formation.

We have recently reported that intramolecular C–H insertion of phenyldiazoacetates exhibited preferential formation of β -lactones with secondary alkyl esters (eq 4).¹⁰ Enanticontrol approached 50% ee, but diastereo-



 Table 3. Products from Catalytic Diazo Decomposition of Androstan-17-on-3-yl Diazoacetate (12b)^a

		rel yield, %		
catalyst	product yield, ^b %	13b	14b	15b
Rh ₂ (5 <i>R</i> -MEPY) ₄	84	73	23	4
$Rh_2(4R-MEOX)_4$	89	60	37	3
$Rh_2(5S-MEPY)_4$	83	3	65	32
Rh ₂ (4.S-MEOX) ₄	86	3	87	19
$Rh_2(4S-MEAZ)_4$	76	1	42	57
Rh ₂ (OAc) ₄	62	22	32	46

 a Reactions were performed in refluxing CH_2Cl_2 using 1.0 mol % of catalyst. b Weight yield after chromatographic separation of catalyst.

 Table 4. Products from Catalytic Diazo Decomposition of 16^a

		rel yield, %	
catalyst	product yield, ^b %	17	18
Rh ₂ (5 <i>R</i> -MEPY) ₄	85	61	39
$Rh_2(4R-MEOX)_4$	88	43	57
$Rh_2(5.S-MEPY)_4$	76	8	92
Rh ₂ (4S-MEOX) ₄	90	3	97

^{*a*} Reactions were performed in refluxing CH₂Cl₂ using 1.0 mol % of catalyst. ^{*b*} Weight yield after chromatographic separation of catalyst.

Table 5.	Products from Catalytic Diazo Decomposition
	of Cholesteryl Phenyldiazoacetate 21 ^a

		rel yield, %	
catalyst	product yield, ^b %	22	23
$Rh_2(OAc)_4$	69	22	78
$Rh_2(4R-MEAZ)_4$	69	16	84
Rh ₂ (4.S-MEAZ) ₄	66	38	62
$Rh_2(S-DOSP)_4$	58	16	84
$Rh_2(S-DOSP)_4^c$	69	10	90

^{*a*} Unless specified otherwise, reactions were performed in refluxing CH_2Cl_2 using 1.0 mol % of catalyst. ^{*b*} Weight yield after chromatographic separation of catalyst. ^{*c*} Reaction performed in refluxing pentane.

control was low. To ascertain the applicability of phenyldiazoacetates in the steroidal series we prepared the 3β -cholesteryl ester. Prior investigations had established that dirhodium(II) carboxylates and the reactive dirhodium(II) azetidinone-carboxylate catalysts (7) were active toward phenyldiazoacetate decomposition, but other carboxamidate catalysts (**4**–**6**) were not.¹⁶ Diazo decomposition resulted in the formation of two products, both β -lactones, in good yield (eq 5).



The influence of the catalyst on diastereoselectivity (Table 5) was substantial with the greatest selectivity

achieved with Davies' $Rh_2(S$ -DOSP)₄ (24) in pentane.²¹ Once again, remote substituents control diastereoselectivity to a substantial degree.²²



24: Ar = p-CH₃(CH₂)₁₁C₆H₄

What is obvious from the data presented here is that steric factors play a significant role in determining product selectivity. Access to the 4-position for C-H insertion is possible only with the least sterically demanding azetidinone-ligated dirhodium(II) catalysts (Tables 2 and 3). However, insertion into a C-H bond at the 2-position occurs with high selectivity using sterically more robust dirhodium(II) carboxamidate catalysts. In each case, insertion takes place into an equatorial C-H bond.

The formation of β -lactone product is favored by S-configured catalysts. This process occurs in synthetically meaningful yields even though there is significant ring strain introduced by the process. The driving force here is, in part, insertion into an oxygen-activated tertiary C-H bond, but steric factors must also contribute.

One way of understanding the selectivities that have been achieved is through the use of models for the reactant metal carbene.²³ Preference for insertion at the 2-position over insertion at the 4-position may be understood by comparison of 25 with 26. We have previously documented the preference for the conformation in which the carbonyl group is anti to the dirhodium(II) core.^{24,25}



In 25, a slight rotation of the steroid brings the C–H bond at the 3-position into close contact with the carbene carbon. In 26, the rotation required to bring the C-H bond at the 2- or 4-positions into contact with the carbene carbon also twists the bulk of the steroid away from the ligated metal (envision motion of the methyl group at C-10 clockwise or counterclockwise). That placement of a double bond between carbons 5 and 6 completely inhibits insertion into the C-H bond at C-4 is not obvious even with these models. However, competition between insertion at the 3-position and the 4-position (Tables 2

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and 3) is consistent with 26 elaborated to describe the stereochemistry of the ligands on rhodium.

Experimental Section

General Methods. ¹H NMR (300, 500, or 600 MHz) and ¹³C NMR (75, 125, or 150 MHz) were recorded from solutions in CDCl₃, and chemical shifts are reported in parts per million (ppm, δ) downfield from internal Me₄Si (TMS). Mass spectra were obtained using electron ionization on a quadrapole instrument. Infrared spectra were recorded by an FT-IR instrument either as a thin film on sodium chloride plates or as solutions as indicated, and absorptions are reported in wavenumbers (cm⁻¹). Melting points are uncorrected. Elemental analysis was performed by Atlantic Microlab, Inc. Dichloromethane was distilled from calcium hydride prior to use in catalytic reactions. Tetrahydrofuran was distilled from sodium and benzophenone. Methanesulfonyl azide was prepared from methanesulfonyl chloride and sodium azide and was not distilled.²⁶ Synthetic reagents were purified by distillation or crystallization prior to use. The preparation of Rh₂(5*R*-MEPY)₄ and Rh₂(5S-MEPY)4,24 Rh₂(4R-MEOX)4 and Rh₂(4S-MEOX)4,27 $Rh_2(4S-MPPIM)_{4,1}^{15}$ and $Rh_2(4S-MEAZ)_{4,1}^{16}$ $Rh_2(4S-IBAZ)_{4,2}^{28}$ and $Rh_2(4S-BNAZ)_{4,2}^{28}$ by acetate displacement from $Rh_2(OAc)_{4}$ have been reported.

Cholest-5-en-3 β **-yl diazoacetate (8)** was prepared by a two-pot modification of the standard three-step procedure.¹² To a stirred solution of cholesterol (3.00 g, 7.75 mmol) and triethylamine (98 µL, 0.70 mmol) in dry THF (30 mL) at 0 °C was added diketene (0.975 g, 11.6 mmol) via syringe over 5 min. The resulting yellow solution was stirred for 1 h at 0 °C and then warmed to room temperature. After 24 h, the mixture was again cooled to 0 °C, at which time triethylamine (1.57 g, 15.5 mmol) and methanesulfonyl azide (1.88 g, 15.5 mmol) were added via syringe. The reaction mixture was allowed to warm slowly to room temperature. After 24 h, the reaction was diluted with water (25 mL), and the diazoacetoacetate was isolated by extraction with ethyl acetate (3 \times 20 mL). The organic layers were combined, washed with water (3 \times 20 mL) and brine (50 mL), dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. Flash column chromatography on silica gel (15% ethyl acetate in hexanes) afforded the diazoacetoacetate in 87% overall yield (3.35 g, 6.75 mmol) as a yellow oil. Cholest-5-en-3 β -yl diazoacetoacetate: ¹H NMR (300 MHz, CDCl₃) δ 5.40 (d, J = 5.1 Hz, 1H), 4.81–4.68 (m, 1H), 2.47 (s, 3H), 2.42-2.34 (comp, 2H), 2.06-0.95 (comp, 29H), 1.03 (s, 3H), 0.92 (d, J = 6.3 Hz, 3H), 0.87 (d, J = 6.6Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.68 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 190.2, 160.8, 139.0, 123.1, 76.4, 75.4, 56.6, 56.1, 49.9, 42.2, 39.6, 39.4, 38.2, 36.8, 36.5, 36.1, 35.7, 31.8, 31.7, 28.2, 28.1, 28.0, 27.9, 24.2, 23.8, 22.8, 22.5, 21.0, 19.2, 18.6, 11.8.

Acetyl cleavage of cholest-5-en- 3β -yl diazoacetoacetate was performed by the addition of LiOH monohydrate (0.979 g, 23.3 mmol) to a biphasic solution of the diazoacetoacetate (3.35 g, 6.75 mmol) in THF (15 mL) and distilled water (10 mL). The dark brown solution was stirred at room temperature until acetyl cleavage was complete. The reaction mixture was extracted with ethyl acetate (3 \times 20 mL). The organic layers were combined and washed with water (3 \times 15 mL), brine (3 × 15 mL), dried over anhydrous MgSO₄ filtered and concentrated under reduced pressure. Flash column chromatography on silica gel (3% ethyl acetate in hexanes) afforded cholest-5en- 3β -yl diazoacetate (8) as a pale yellow solid in 68% yield (1.93 g, 5.27 mmol): mp 138–139 °C; $[\alpha]^{26}_{D} = -24.5$ (c 2.79, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 5.38 (d, J = 5.1 Hz, 1H), 4.74-4.68 (m, 1H), 4.70 (bs, 1H), 2.38-2.28 (comp, 2H), 2.02-

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1.94 (comp, 2H), 1.89–1.79 (comp, 3H), 1.63–0.90 (comp, 21H), 1.01 (s, 3H), 0.91 (d, J = 6.6 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.67 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 166.3, 139.5, 122.7, 74.6, 56.7, 56.1, 50.0, 46.3, 42.3, 39.7, 39.5, 38.3, 36.9, 36.6, 36.2, 35.8, 31.9, 31.8, 28.2, 28.0, 27.9, 24.3, 23.8, 22.8, 22.6, 21.0, 19.3, 18.7, 11.9; IR (film) 2120 (C=N₂), 1698 (C=O), 1404, 1196 cm⁻¹. Anal. Calcd for C₂₉H₄₆N₂O₂: C, 76.6; H, 10.20; N, 6.16. Found: C, 76.67; H, 10.28; N, 6.06.

Diazo Decomposition of Cholest-5-en-3 β -yl Diazoacetate. To a stirred solution of Rh₂(4*R*-MEOX)₄ (1.9 mg, 0.0011 mmol) in dry CH₂Cl₂ (5 mL) heated at reflux under a nitrogen atmosphere was added cholesteryl diazoacetate 8 (50.0 mg, 0.111 mmol) in dry CH₂Cl₂ (3 mL) over 5 h via syringe pump. The reaction mixture was heated at reflux for an additional half hour. After being cooled to room temperature, the reaction solution was filtered through a short silica plug, which was subsequently washed with 10 mL of 30% ethyl acetate in hexanes. The solvent was removed under reduced pressure to afford 34 mg of product (72% yield) consisting of γ -lactone 9 and β -lactone **10**. The ratio of products in the crude reaction mixture was determined by ¹H NMR spectroscopy. Column chromatography on silica gel (3% ethyl acetate in hexanes) afforded 30 mg of 2-(3 β -hydroxy-5-cholesten-2 α -yl)acetic acid lactone (9) (63% yield) and 3 mg of 3-(3 β -hydroxy-5-cholesten- 3α -yl)acetic acid lactone (10) (6% yield). The Rh₂(4S-MEOX)₄catalyzed reaction of diazoacetate 8 afforded 4 mg of 2-(3 β hydroxy-5-cholesten- 2α -yl)acetic acid lactone (9) and 30 mg of 3-(3β -hydroxy-5-cholesten- 3α -yl)acetic acid lactone (10) (65% yield). 4-(3β -Hydroxy-5-cholesten-4 α -yl)acetic acid lactone, the product from Č-H insertion at the 4-position, did not form to an appreciable amount in any diazo decomposition reaction.

2-(**3** β -Hydroxy-5-cholesten-2 α -yl)acetic acid lactone (9): mp 137–138 °C; [α]²²_D = -11.4 (*c* 1.75, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 5.49 (d, *J* = 5.2 Hz, 1H), 3.69 (ddd, *J* = 12.2, 10.3, 4.7 Hz, 1H), 2.58 (dd, *J* = 12.2, 4.4 Hz, 1H), 2.53 (dd, *J* = 15.2, 5.2 Hz, 1H, lactone α -H_{eq}), 2.57–2.49 (comp, 1H), 2.19 (t, *J* = 15.2, 1H, lactone α -H_{ax}), 2.34–2.13 (comp, 1H), 2.07–1.98 (comp, 3H), 1.88–1.80 (comp, 1H), 1.61–0.70 (comp, 20H), 1.07 (s, 3H), 0.91 (d, *J* = 6.5 Hz, 3H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.86 (d, *J* = 6.6 Hz, 3H), 0.68 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 176.6, 138.1, 125.6, 84.9, 56.6, 56.1, 50.1, 42.3, 40.9, 40.4, 39.6, 39.5, 38.6, 37.7, 36.1, 35.9, 35.7, 31.8, 31.6, 28.2, 28.0, 24.2, 23.8, 22.8, 22.5, 21.0, 20.6, 18.7, 11.8; IR (CDCl₃) 1787 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₉H₄₇O₂ 427.3576, found 427.3589 (M⁺¹).

3-(3 β -Hydroxy-5-cholesten-3 α -yl)acetic acid lactone (10): mp 119–121 °C; [α]²⁶_D = -24.0 (c 0.35, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 5.42 (d, J = 5.1 Hz, 1H), 3.03 (d, J = 16.2 Hz, 1H, lactone α -H), 2.95 (d, J = 16.2 Hz, 1H, lactone α -H), 2.24–1.73 (comp, 8H), 1.61–0.97 (comp, 20H), 1.05 (s, 3H), 0.91 (d, J = 6.3 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.68 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 168.3, 138.5, 124.2, 79.2, 56.6, 56.1, 50.0, 46.2, 42.7, 42.3, 39.6, 39.5, 36.7, 36.5, 36.2, 35.7, 32.0, 31.9, 31.8, 28.2, 28.0, 24.2, 23.8, 22.8, 22.5, 21.1, 19.2, 18.7, 11.8; IR (CDCl₃) 1818 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₉H₄₇O₂ 427.3576, found 427.3581 (M⁺¹).

5 α -**Cholestan-3** β -**yl diazoacetate (12a)** was prepared by the standard method. 12 To a stirred solution of $5\alpha\mbox{-cholestan-}$ $3\beta\text{-ol}$ (3.00 g, 7.72 mmol) and triethylamine (11 $\mu\text{L},~0.077$ mmol) in dry THF (35 mL) at 0 °C was added diketene (0.973 g, 11.6 mmol) via syringe over 5 min. The resulting yellow solution was stirred for 1 h at 0 °C and then warmed to room temperature. After 24 h, the mixture was again cooled to 0 °C, at which time triethylamine (1.17 g, 11.6 mmol) and methanesulfonyl azide (1.40 g, 11.6 mmol) were added via syringe. The reaction mixture was allowed to warm slowly to room temperature. After 24 h, the reaction was diluted with water (25 mL), and the diazoacetoacetate was isolated by extraction with ethyl acetate (3 \times 75 mL). The organic layers were combined, washed with 1 M NaOH (25 mL), water (3 imes50 mL), and brine (50 mL), dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. Flash column chromatography on silica gel (15% ethyl acetate in hexanes) afforded the diazoacetoacetate in 87% overall yield (3.37 g, 6.72 mmol) as a yellow oil. 5 α -Cholestan-3 β -yl diazoacetoacetate: ¹H NMR (300 MHz, CDCl₃) δ 4.90–4.77 (m, 1H), 2.47 (s, 3H), 2.04–0.95 (comp, 30H), 0.90 (d, J = 6.6 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.84 (s, 3H), 0.74–0.60 (comp, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 190.4, 161.0, 75.3, 56.4, 56.2, 54.1, 44.7, 42.5, 39.9, 39.5, 36.7, 36.1, 35.8, 35.4, 34.2, 31.9, 28.6, 28.3, 28.2, 28.0, 27.6, 24.2, 23.8, 22.8, 22.5, 21.2, 18.6, 12.2, 12.0.

Acetyl cleavage of 5 α -cholestan-3 β -yl diazoacetoacetate was performed as previously described by the addition of LiOH monohydrate (0.421 g, 10.0 mmol) to a biphasic solution of the diazoacetoacetate (3.55 g, 6.68 mmol) in THF (25 mL) and distilled water (25 mL). Flash column chromatography on silica gel (5% ethyl acetate in hexanes) afforded 5α -cholestan- 3β -yl diazoacetate (12a) in 88% yield (2.67 g, 5.89 mmol) as a pale yellow solid: mp 141–144 °C; $[\alpha]^{34}_{D} = +15.4$ (c 1.22, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 4.83–4.70 (m, 1H), 4.69 (bs, 1H), 2.05-0.92 (comp, 30H), 0.90 (d, J = 6.6 Hz, 3H), 0.86(d, J = 6.6 Hz, 6H), 0.82 (s, 3H), 0.74–0.55 (comp, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 166.5, 74.4, 56.4, 56.2, 54.2, 46.3, 44.6, 42.6, 39.9, 39.5, 36.7, 36.1, 35.8, 35.5, 35.4, 34.2, 32.0, 28.6, 28.2, 28.0, 27.7, 24.2, 23.8, 22.8, 22.5, 21.2, 18.6, 12.2, 12.0; IR (film) 2117 (C=N₂), 1672 (C=O) cm⁻¹. Anal. Calcd for C₂₉H₄₈N₂O₂: C, 76.27; H, 10.59; N, 6.13. Found: C, 76.34; H, 10.73; N, 6.01.

Diazo Decomposition of 5α **-Cholestan-** 3β **-yl Diazoac**etate. To a stirred solution of Rh₂(5*R*-MEPY)₄ (1.0 mg, 0.0011 mmol) in dry CH₂Cl₂ (5 mL) heated at reflux under a nitrogen atmosphere was added diazoacetate 12a (50.0 mg, 0.110 mmol) in dry CH₂Cl₂ (3 mL) over 5 h via syringe pump. The reaction mixture was heated at reflux for an additional 0.5 h to complete the reaction. After being cooled to room temperature, the reaction mixture was filtered through a short silica plug, which was subsequently washed with 10 mL of 20% ethyl acetate in hexanes. The solvent was removed under reduced pressure to afford 45 mg of product (95% yield) consisting of γ -lactone **13a**, 2-(3 β -hydroxy-5 α -cholestan-2 α -yl)acetic acid lactone; β -lactone **14a**, 3-(3 β -hydroxy-5 α -cholestan-3 α -yl)acetic acid lactone; and γ -lactone **15a**, 4-(3 β -hydroxy-5 α -cholestan- 4α -yl)acetic acid lactone. The relative ratio of products in the crude reaction mixture was determined by ¹H NMR spectroscopy. Column chromatography on silica gel (3% ethyl acetate in hexanes) afforded 27 mg of a mixture of 2-(3β -hydroxy- 5α cholestan-2 α -yl)acetic acid lactone (13a) and 4-(3 β -hydroxy-5a-cholestan-4a-yl)acetic acid lactone (15a) (90:10 ratio respectively, 58% yield) and 6 mg of 3-(3 β -hydroxy-5 α -cholestan- 3α -yl)acetic acid lactone (14a) (13% yield). Using the same procedure, the Rh₂(5S-MEPY)₄ catalyzed reaction of diazoacetate 12a afforded 14 mg 13a and 15a (13:87 ratio respectively, 30% yield) and 19 mg of (14a) (41% yield). The $Rh_2(4S-MEOX)_4$ catalyzed reaction of diazoacetate 12a afforded 12 mg of 13a and 15a (9:91 ratio respectively, 26% yield) and 21 mg of (14a)-(45% yield).

2-(3 β -Hydroxy-5 α -cholestan-2 α -yl)acetic acid lactone (13a): mp 178–180 °C; [α]²⁸_D = +38.8 (*c* 1.96, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 3.78 (dt, *J* = 11.0, 4.0 Hz, 1H), 2.46 (dd, *J* = 14.1, 4.6 Hz, 1H, lactone α -H_{eq}), 2.17 (t, *J* = 14.1 Hz, 1H, lactone α -H_{ax}), 2.15–2.08 (comp, 1H), 1.98 (dt *J* = 12.8, 3.4 Hz, 1H), 1.92 (dd, *J* = 12.8, 2.9 Hz, 1H), 1.86 (dt, *J* = 12.0, 3.6 Hz, 1H), 1.84–1.78 (m, 1H), 1.69 (dq, *J* = 13.2, 3.5 Hz, 1H), 1.62–1.20 (comp, 14H), 1.19–0.83 (comp, 9H), 0.90 (d, *J* = 6.6 Hz, 3H), 0.89 (s, 3H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.86 (d, *J* = 6.6 Hz, 3H), 0.72 (dt, *J* = 6.6, 2.7 Hz, 1H), 0.66 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.8, 85.6, 56.3, 56.2, 54.3, 45.7, 42.6, 40.4, 40.1, 39.9, 39.5, 37.9, 36.2, 36.1, 35.7, 35.4, 32.8, 32.0, 28.6, 28.2, 28.0, 24.1, 23.8, 22.8, 22.5, 21.4, 18.6, 14.1, 12.1; IR (film) 1799 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₉H₄₉O₂ 429.3733, found 429.3731 (M⁺¹).

3-(3 β -Hydroxy-5 α -cholestan-3 α -yl)acetic acid lactone (14a): mp 136–138 °C; [α]³⁰_D = +14.0 (*c* 1.34, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 3.07 (s, 2H, lactone α -H), 2.05 (dt, J = 13.6, 4.2 Hz, 1H), 1.98 (dt. J = 12.7, 3.3 Hz, 1H), 1.93 (t, J = 13.0 Hz, 1H), 1.84 (dt, J = 12.9, 3.6 Hz, 1H), 1.84–1.76 (m, 1H), 1.74 (dq, J = 13.1, 3.1 Hz, 1H), 1.69, (dq, J = 13.2, 3.5 Hz, 1H), 1.69–1.44 (comp, 4H), 1.40–1.20 (comp, 8H), 1.18–0.80 (comp, 11H), 0.90 (d, J = 6.5 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.85 (s, 3H), 0.65 (s, 3H), 0.63 (ddd, J = 12.3, 10.7, 4.1 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 168.5, 79.3, 56.4, 56.2, 54.1, 47.6, 44.6, 42.6, 39.9, 39.5, 38.4, 36.3, 36.1, 35.8, 35.4, 35.3, 32.0, 31.9, 28.5, 28.2, 28.0, 24.2, 23.8, 22.8, 22.5, 21.3, 18.6, 12.1, 11.8; IR (film) 1836 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₉H₄₉O₂ 429.3733, found

429.3731 (M⁺¹). **4**-(**3**β-**Hydroxy-5α-cholestan-4α-yl)acetic acid lactone** (**15a**): mp 117–119 °C; [α]³⁰_D = +0.8 (c 0.57, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 3.77 (ddd, J = 11.8, 10.2, 4.2 Hz, 1H), 2.43 (dd, J = 15.6, 6.0 Hz, 1H, lactone α-H_{eq}), 2.09 (t, J = 15.6Hz, 1H, lactone α-H_{ax}), 2.22–1.37 (comp, 10H), 1.36–1.20 (comp, 8H), 1.25 (s, 3H), 1.19–0.92 (comp, 9H), 0.91–0.80 (comp, 2H), 0.89 (d, J = 5.6 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.71 (dt, J = 11.0, 4.2 Hz, 1H), 0.66 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 177.0, 85.9, 56.4, 56.2, 54.2, 49.0, 43.9, 42.5, 39.8, 39.5, 36.4, 36.1, 35.8, 35.1, 34.5, 31.5, 29.7, 28.2, 28.0, 26.2, 25.4, 24.2, 23.8, 22.8, 22.6, 21.2, 18.6, 13.5, 12.1; IR (film) 1789 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₉H₄₉O₂ 429.3733, found 429.3731 (M⁺¹).

5α-Androstan-17-one-3β-yl diazoacetate (12b) was prepared as described for 12a. To a stirred solution of 5α androstan-17-on- 3β -ol (3.00 g, 10.3 mmol) and triethylamine (15.7 mg, 0.155 mmol) in dry THF (50 mL) at 0 °C was added diketene (1.30 mg, 15.5 mmol). Triethylamine (1.57 mL, 15.5 mmol) and methanesulfonyl azide (1.88 g, 15.5 mmol) were added. Flash column chromatography on silica gel (25% ethyl acetate in hexanes) afforded the diazoacetoacetate in 88% overall yield (3.62 g, 9.03 mmol) as a yellow oil. $5\alpha\text{-}An\text{-}$ drostan-17-on-3 β -yl diazoacetoacetate: ¹H NMR (300 MHz, CDCl₃) δ 4.89–4.77 (m, 1H), 2.46 (s, 3H), 2.42 (dd, J= 18.9, 9.0 Hz, 1H), 2.05 (dd, J = 19.0, 9.0 Hz, 1H), 2.00-0.95 (comp, 19H), 0.88 (s, 3H), 0.86 (s, 3H), 0.75 (dt, J = 11.7, 3.6 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 220.4, 189.7, 160.5, 76.0, 74.6, 53.8, 50.9, 47.3, 44.3, 36.2, 35.4, 35.2, 34.6, 33.7, 31.1, 30.4, 27.9, 27.2, 21.4, 20.1, 13.4, 11.9.

Acetyl cleavage of 5α -androstan-17-on- 3β -diazoacetoacetate was performed as previously described by the addition of LiOH monohydrate (0.566 g, 13.5 mmol) to a biphasic solution of the diazoacetoacetate (3.60 g, 8.99 mmol) in THF (30 mL) and distilled water (30 mL). Flash column chromatography on silica gel (25% ethyl acetate in hexanes) afforded 5α-androstan-17-on- 3β -yl diazoacetate (12b) in 77% yield (2.51 g, 6.97 mmol) as a pale yellow solid: mp 163–165 °C; $[\alpha]^{34}_{D} = +16.6$ (c 2.53, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 4.82–4.70 (m, 1H), 4.69 (bs, 1H), 2.42 (dd, J = 19.2, 9.0 Hz, 1H), 2.03 (dd, J = 19.2, 9.0 Hz, 1H), 1.97-1.12 (comp, 17H), 1.05-0.89 (comp, 2H), 0.84 (s, 6H), 0.75 (dt, J = 11.4, 4.2 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) & 221.2, 166.4, 74.1, 54.2, 51.3, 47.7, 46.2, 44.6, 36.6, 35.8, 35.6, 35.0, 34.1, 31.5, 30.7, 28.2, 27.6, 21.7, 20.4, 13.8, 12.2; IR (film) 2142 (C=N₂), 1720 (C=O), 1657 (C=O) cm⁻¹. Anal. Calcd for C21H30N2O3: C, 70.36; H, 8.44; N, 7.81. Found: C, 70.48; H, 8.55; N, 7.61.

Diazo Decomposition of 5α-Androstan-17-on-3β-yl Dia**zoacetate.** To a stirred solution of $Rh_2(5R-MEPY)_4$ (2.4 mg, 0.0028 mmol) in dry CH₂Cl₂ (5 mL) heated at reflux under a nitrogen atmosphere was added diazoacetate 12b (100 mg, 0.278 mmol) in dry CH₂Cl₂ (3 mL) over 5 h via syringe pump. The reaction mixture was heated at reflux for an additional 0.5 h to complete the reaction. After being cooled to room temperature, the reaction mixture was filtered through a short silica plug, which was subsequently washed with 10 mL of 30% ethyl acetate in hexanes. The solvent was removed under reduced pressure to afford 84 mg of product (91% yield) consisting of γ -lactone **13b** [2-(3 β -hydroxy-5 α -androstan-17on-2 α -yl)acetic acid lactone], β -lactone **14b** [3-(3 β -hydroxy-5 α androstan-17-on-3 α -yl)acetic acid lactone], and γ -lactone 15b [4-(3β -hydroxy- 5α -androstan-17-on- 4α -yl)acetic acid lactone]. The relative ratio of products in the crude reaction mixture was determined by ¹Ĥ NMR spectroscopy. Column chromatography on silica gel (2% ethyl acetate in CH2Cl2) afforded 56 mg of a mixture of 13b and 15b (95:5 ratio respectively, 61% yield) and 14 mg of 14b (15% yield). Using the same procedure, the $Rh_2(4.S-MEAZ)_4$ -catalyzed reaction of diazoacetate **12b** afforded 35 mg of **13b** and **15b** (5:95 ratio respectively, 38% yield) and 26 mg of **14b** (38% yield). The $Rh_2(4.S-MEOX)_4$ -catalyzed reaction of diazoacetate **12b** afforded 18 mg of **13b** and **15b** (13:87 ratio respectively, 20% yield) and 42 mg of **14b** (46% yield).

2-(3 β -Hydroxy-5 α -androstan-17-on-2 α -yl)acetic acid lactone (13b): mp 208–210 °C; $[\alpha]^{29}_{D} = +86.8$ (*c* 1.74, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 3.79 (dt, J = 10.9, 3.9 Hz, 1H), 2.47 (dd, J = 14.7, 4.6 Hz, 1H, lactone α -H_{eq}), 2.42 (d, J = 9.4 Hz, 1H), 2.18 (t, J = 14.7 Hz, 1H, lactone α -H_{ax}), 2.25–1.76 (comp, 7H), 1.72–1.15 (comp, 11H), 1.01 (t, J = 12.2 Hz, 1H), 0.94 (s, 3H), 0.87 (s, 3H), 0.90–0.75 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 220.9, 176.6, 85.3, 54.4, 51.2, 47.7, 45.7, 40.4, 40.1, 38.0, 36.1, 35.8, 34.9, 32.7, 31.4, 30.8, 28.2, 21.7, 20.6, 14.1, 13.8; IR (film) 1777 (C=O), 1732 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₁H₃₁O₃ 331.2273, found 331.2278 (M⁺¹).

3-(3 β -Hydroxy-5 α -androstan-17-on-3 α -yl)acetic acid lactone (14b): mp 145–147 °C; $[\alpha]^{30}_{D} = +72.4$ (*c* 1.59, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 3.08 (s, 2H, lactone α -H), 2.46 (dd, J = 19.2, 8.9 Hz, 1H), 2.16–1.72 (comp, 7H), 1.71–1.43 (comp, 4H), 1.42–1.18 (comp, 5H), 0.89 (s, 3H), 0.87 (s, 3H), 1.17–0.79 (comp, 4H), 0.72 (dt, J = 12.0, 8.1 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 221.0, 168.3, 79.0, 54.1, 51.3, 47.7, 47.6, 44.6, 38.3, 36.2, 35.6, 34.9, 31.9, 31.4, 30.7, 29.7, 28.2, 21.8, 20.5, 13.8, 11.8; IR (film) 1824 (C=O), 1738 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₁H₃₁O₃ 331.2273, found 331.2277 (M⁺¹).

4-(3β-Hydroxy-5α-androstan-17-on-4α-yl)acetic acid lactone (15b): mp 136–138 °C; $[α]^{30}_{D} = +31.3$ (*c* 0.98, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 3.79 (ddd, *J* = 11.8, 10.2, 4.2 Hz, 1H), 2.47 (dd, *J* = 16.0, 6.5 Hz, 1H, lactone α-H_{eq}), 2.53–2.44 (comp, 1H), 2.16 (t, *J* = 15.8 Hz, 1H, lactone α-H_{ax}), 2.17–0.90 (comp, 19H), 0.93 (s, 3H), 0.87 (s, 3H), 0.81 (dt, *J* = 12.0, 4.2, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 220.9, 176.7, 85.7, 54.2, 51.3, 48.9, 47.7, 43.8, 37.4, 36.3, 35.7, 34.6, 34.4, 31.4, 30.3, 26.2, 25.1, 21.7, 20.5, 13.8, 13.6; IR (film) 1786 (C=O), 1734 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₁H₃₁O₃ 331.2273, found 331.2269 (M⁺¹).

5α-**Cholestan-6-on-3**β-**yl Diazoacetate (16).** To a stirred solution of 5 α -cholestan-6-on-3 β -ol (1.00 g, 2.48 mmol) and triethylamine (2.5 mg, 0.25 mmol) in dry THF (10 mL) at 0 °C was added diketene (0.313 g, 3.72 mmol). Triethylamine (0.377 g, 3.72 mmol) and methanesulfonyl azide (0.451 g, 3.72 mmol) were added. Flash column chromatography on silica gel (25% ethyl acetate in hexanes) afforded diazoacetoacetate in 59% overall yield (0.755 g, 1.47 mmol) as a yellow solid. 5α -**Cholestan-6-on-3** β **-yl diazoacetatoacetate:** ¹H NMR (300 MHz, CDCl₃) δ 4.89–4.76 (m, 1H), 2.47 (s, 3H), 2.39–2.25 (comp, 2H), 2.10-0.96 (comp, 27H), 0.91 (d, J = 6.3 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.79 (s, 3H),0.67 (s, 3H); $^{13}\mathrm{C}$ NMR (75 MHz, CDCl_3) δ 209.9, 189.8, 160.6, 74.2, 56.4, 56.1, 55.8, 53.4, 46.3, 42.7, 42.6, 40.6, 39.2, 39.1, 37.6, 36.0, 35.8, 35.4, 28.0, 27.8, 27.7, 26.8, 26.1, 23.7, 23.5, 22.6, 22.3, 21.2, 18.4, 12.8, 11.7.

Acetyl cleavage of 5α -cholestan-6-on- 3β -yl diazoacetoacetate was performed as previously described by the addition of LiOH monohydrate (0.141 g, 2.93 mmol) to a biphasic solution of the diazoacetoacetate (0.752 g, 1.47 mmol) in THF (8 mL) and distilled water (8 mL). Flash column chromatography on silica gel, (25% ethyl acetate in hexanes) afforded 5α -cholestan-6on- 3β -yl diazoacetate (**16**) in 80% yield (0.546 g, 1.17 mmol) as a pale crystalline yellow solid: mp 166–168 °C; $[\alpha]^{25}_{D} = -4.6$ $(c 0.34, CHCl_3)$; ¹H NMR (300 MHz, CDCl₃) δ 4.83–4.66 (m, 1H), 4.71 (bs, 1H), 2.38-2.22 (comp, 2H), 2.10-0.96 (comp, 27H), 0.91 (d, J = 6.3 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.77 (s, 3H), 0.66 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) & 210.3, 166.3, 73.4, 56.6, 56.4, 56.0, 53.7, 46.6, 46.3, 42.9, 40.9, 39.4, 39.5, 37.9, 36.3, 36.0, 35.6, 28.0, 27.9, 27.0, 26.3, 23.9, 23.7, 22.8, 22.5, 21.4, 18.6, 13.0, 11.9; IR (film) 2109 (C=N₂), 1709 (C=O), 1689 (C=O) cm⁻¹. Anal. Calcd for C29H46N2O3: C, 74.00; H, 9.85; N, 5.95. Found: C, 73.80; H, 9.83; N, 5.69.

Diazo Decomposition of 5α **-Cholestan-6-on-3** β **-yl Diazoacetate (16).** To a stirred solution of Rh₂(5*R*-MEPY)₄ (0.9 mg, 0.0011 mmol) in dry CH₂Cl₂ (5 mL) heated at reflux under a nitrogen atmosphere was added diazoacetate 16 (50.0 mg, 0.107 mmol) in dry CH₂Cl₂ (3 mL) over 5 h via syringe pump. The reaction mixture was heated at reflux for an additional 0.5 h to ensure completion of the reaction. After cooling to room temperature, the reaction was filtered through a short silica plug, which was subsequently washed with 10 mL of 35% ethyl acetate in hexanes. The solvent was removed under reduced pressure to afford 43 mg of product (90% yield) consisting of γ -lactone 17 [2-(3 β -hydroxy-5 α -cholestan-6-on-2 α -yl)acetic acid lactone] and β -lactone **18** [3-(3 β -hydroxy-5 α -cholestan-6-on-3 β yl)acetic acid lactone]. The relative ratio of products in the crude reaction mixture was determined by ¹H NMR spectroscopy. Column chromatography on silica gel (2% ethyl acetate in CH₂Cl₂) afforded 20 mg of 17 and 12 mg of 18 (25% yield). The Rh₂(4S-MEOX)₄ catalyzed reaction of diazoacetate 12 afforded 35 mg of 18 (74% yield). 4-(3 β -Hydroxy-5 α -cholestan-6-on-4 α -yl)acetic acid lactone did not form to an appreciable extent in any decomposition.

2-(3 β -Hydroxy-5α-cholestan-6-on-2α-yl)acetic acid lactone (17): mp 173–176 °C; [α]³¹_D = +3.3 (*c* 1.25, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 3.78 (ddd, *J* = 11.8, 10.8, 3.9 Hz, 1H), 2.51 (dd, *J* = 15.4, 5.8 Hz, 1H, lactone α-H_{eq}), 2.39–2.32 (comp, 2H), 2.23 (t, *J* = 15.4 Hz, 1H, lactone α-H_{eq}), 2.39–2.32 (comp, 2H), 2.23 (t, *J* = 15.4 Hz, 1H, lactone α-H_{ax}), 2.24–1.72 (comp, 6H), 1.62–1.04 (comp, 19H), 1.00 (q, *J* = 9.1 Hz, 1H), 0.92 (d, *J* = 6.5 Hz, 3H), 0.87 (d, *J* = 6.7 Hz, 3H), 0.86 (d, *J* = 6.7 Hz, 3H), 0.85 (s, 3H), 0.67 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 209.1, 176.1, 84.8, 57.3, 56.6, 56.1, 53.9, 46.5, 43.0, 42.9, 39.9, 33.6, 39.5, 39.4, 37.6, 36.0, 35.8, 35.7, 29.7, 28.0, 25.4, 23.9, 23.8, 22.8, 22.5, 21.6, 18.6, 14.7, 12.0; IR (film) 1806 (C=O), 1709 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₉H₄₇O₃ 443.3525, found 443.3523 (M⁺¹).

3-(3 β -Hydroxy-5 α -cholestan-6-on-3 α -yl)acetic acid lactone (18): mp 158–160 °C; [α]³⁰_D = -8.3 (c 0.93, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 3.07 (d, J = 15.8 Hz, 1H, lactone α -H), 3.01 (d, J = 15.8 Hz, 1H, lactone α -H), 2.37 (dd, J = 13.4, 4.5 Hz, 1H), 2.15–2.02 (comp, 3H), 1.96 (t, J = 12.8 Hz, 1H), 1.95–1.76 (m, 4H), 1.65–1.47 (comp, 4H), 1.45–1.04 (comp, 15H), 0.99 (q, J = 9.1 Hz, 1H), 0.91 (d, J = 6.5 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.81 (s, 3H), 0.67 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 209.1, 167.5, 78.8, 56.7, 56.2, 56.1, 53.7, 47.2, 46.5, 43.0, 40.8, 39.5, 39.4, 37.8, 36.1, 35.9, 35.7, 31.4, 30.6, 29.7, 28.0, 24.0, 23.8, 22.8, 22.5, 21.5, 18.6, 12.7, 12.0; IR (film) 1842 (C=O), 1710 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₂₉H₄₉O₂ 443.3525, found 443.3523 (M⁺¹).

Preparation of 5-Cholesten-3β-yl Phenyldiazoacetate (21). To a stirred solution of cholesterol (4.0 mL, 10 mmol) and triethylamine (2.2 mL, 11 mmol) in freshly distilled CH₂- Cl_2 (50 mL) at 0 °C under an argon atmosphere was added phenylacetyl chloride (1.5 mL, 16 mmol) via syringe pump over 10 min. After being stirred at 0 °C for 1 h, the reaction mixture was warmed to room temperature and stirred overnight. The reaction mixture was diluted with CH₂Cl₂ (200 mL) and extracted with 10% aqueous HCl (2 \times 50 mL), water (25 mL), and brine (25 mL). The organic layer was dried over anhydrous MgSO₄, filtered and concentrated under reduced pressure. Purification by column chromatography on silica gel (0-7.5%)ethyl acetate in hexanes) afforded 3.4 g (6.7 mmol, 64% yield) of 5-cholesten-3β-yl phenylacetate: ¹H NMR (600 MHz, CDCl₃) & 7.39-7.22 (comp, 5H), 5.39-32 (m, 1H), 4.65-4.56 (m, 1H), 3.59 (s, 2H), 2.33-2.26 (comp, 2H), 2.05-1.76 (comp, 6H), 1.62–0.87 (comp, 20H), 1.01 (s, 3H), 0.91 (d, J = 6.5 Hz, 3H), 0.91 (d, J = 6.6 Hz, 3H), 0.91 (d, J = 6.6 Hz, 3H), 0.67 (s, 3H).

To a stirred solution of 5-cholesten- 3β -yl phenylacetate (3.4 g, 6.7 mmol) and 4-acetamidobenzenesulfonyl azide (1.8 g, 7.7 mmol) in HPLC grade acetonitrile (50 mL) at 0 °C under an argon atmosphere was added DBU (2.0 mL, 13.4 mmol) portion wise. The reaction mixture was warmed to room temperature and stirred overnight. The reaction mixture was concentrated under reduced pressure, dissolved with ethyl acetate (100 mL), washed with saturated aqueous NH₄Cl (3 × 15 mL), dried (MgSO₄), filtered, and concentrated under reduced pressure. Purification by column chromatography (0–10% ethyl acetate

in hexanes) afforded 2.7 g (5.3 mmol, 80% yield) of phenyldiazoacetate **21** as a yellow solid: mp 129–131 °C; $[\alpha]^{25}_{\rm D} =$ -11.0 (c = 0.95 in CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.48 (d, J = 7.6 Hz, 2H), 7.37 (t, J = 7.6 Hz, 2H), 7.16 (t, J = 7.6 Hz, 1H), 5.42–5.36 (m, 1H), 4.43–4.23 (m, 1H), 2.47–2.32 (comp, 2H), 2.05–1.77 (comp. 6H), 1.71–0.80 (comp. 20H), 1.03 (s, 3H), 0.92 (d, J = 6.5 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.68 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 164.4, 139.4, 128.8, 125.7, 125.5, 123.8, 122.8, 74.7, 63.3, 56.6, 56.1, 50.0, 42.2, 39.7, 39.5, 38.3, 36.9, 36.5, 36.3, 35.8, 31.9, 31.8, 28.2, 28.0, 27.9, 24.2, 23.8, 22.8, 22.5, 21.0, 19.3, 18.7, 11.8; IR (film, cm⁻¹) 2087 (C=N₂) and 1705 (C=O). Anal. Calcd for C₃₅H₅₀N₂O₂: C, 79.2; H, 9.49. Found: C, 79.31; H, 9.56.

Diazo Decomposition of 5-Cholesten-3*β*-yl Phenyldiazoacetate. The procedure for the diazo decomposition of 21 with Rh₂(OAc)₄ is representative. To a stirred solution of Rh₂-(OAc)₄ (2.1 mg, 0.0047 mmol) in dry CH₂Cl₂ (5 mL) heated at reflux under an argon atmosphere was added phenyldiazoacetate 2 (250 mg, 0.47 mmol) in dry CH₂Cl₂ (3 mL) over 2 h via syringe pump. The reaction mixture was heated at reflux for an additional 30 min upon completion of the addition. After cooling to room temperature, the reaction was filtered through a short silica plug, which was subsequently washed with 10 mL of 25% ethyl acetate in hexanes. The solvent was removed under reduced pressure to afford 239 mg (quantitative) of the crude product as an off-white crystalline solid. ¹H NMR analysis of the reaction mixture revealed 3-(3 β -hydroxy-5cholesten- 3α -yl)-2-phenylacetic acid lactones (22 and 23) was 68% of the reaction mixture as a mixture of diastereomers. Column chromatography on silica gel (eluent: 5% ethyl acetate in hexanes) afforded 149 mg (63%) of 3-(3 β -hydroxy-5-cholesten-3ayl)-2-phenylacetic acid lactones (22 and 23) as a crystalline white solid, the diastereomer ratio of which was determined by ¹H NMR (600 MHz) to be 22:78. Isolation of analytical samples of β -lactones **22** and **23** was achieved by (silica gel loading ratio 100:1, eluent: 1-2% ethyl acetate in hexanes) to afford each as a white solid film.

3-(3β-Hydroxy-5-cholesten-3α-yl)-2S-phenylacetic acid lactone (22): ¹H NMR (600 MHz, CDCl₃) δ 7.37–7.30 (comp, 3H), 7.16 (dd, J = 8.0, 1.7 Hz, 2H), 5.63-5.59 (m, 1H), 4.46 (s, 1H, α -H β -lactone), 3.05 (dddd, J = 13.3, 2.6, 2.6, 2.4 Hz, 1H, C4-H_{ax}), 2.15 (dd, J = 13.3, 2.8 Hz, 1H, C4-H_{eq}), 2.12-2.04 (m, 1H), 1.98 (ddd, J = 18.5, 13.9, 4.7 Hz, 1H, C2-H_{ax}), 1.96-1.91 (m, 1H), 1.88-1.80 (m, 1H), 1.80-1.74 (m, 1H, C2-H_{eq}), 1.70–0.95 (comp, 19H), 1.01 (s, 3H), 0.89 (d, J = 6.6 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.73 (ddd, J = 16.0, 12.2, 4.7 Hz, 1H), 0.66 (s, 3H), 0.40 (ddd, J = 18.3, 14.0, 4.3 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 169.7, 138.7, $130.9,\ 129.5,\ 128.6,\ 128.2,\ 124.2,\ 83.8,\ 63.6,\ 56.6,\ 56.2,\ 50.4,$ 44.6, 42.3, 39.5, 36.2, 35.7, 35.2, 32.1, 31.9, 29.7, 28.2, 28.0, 27.9, 24.3, 23.8, 22.8, 22.6, 21.0, 19.2, 18.7, 11.8; IR (CDCl₃) 1816 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₃₅H₅₁O₂ 503.3889, found 503.3889 (M⁺¹).

3-(3β-Hydroxy-5-cholesten-3α-yl)-2*R*-phenylacetic acid **lactone (23):** ¹H NMR (600 MHz, CDCl₃) δ 7.33–7.25 (comp, 3H), 7.15 (dd, J = 8.3, 1.5 Hz, 2H), 4.48 (s, 1H, α -H β -lactone), 4.13-4.09 (m, 1H), 2.66 (dddd, J = 13.8, 2.9, 2.8, 2.5 Hz, 1H, C4- H_{ax}), 2.30 (dddd, J = 17.1, 14.2, 3.9, 1.0 Hz, 1H, C2- H_{ax}), 2.15 (dd, J = 13.8, 2.8 Hz, 1H, C4–H_{eq}), 2.07–1.99 (comp, 2H), 1.96 (dddd, J = 12.9, 3.3, 3.2, 3.2 Hz, 1H, C2-H_{eq}), 1.85-1.76 (m, 1H), 1.70-1.63 (m, 1H), 1.56-1.05 (comp, 18H), 1.04-0.85 (comp, 3H), 0.99 (s, 3H), 0.92 (d, J = 6.6 Hz, 3H), 0.87 (d, J =6.7 Hz, 3H), 0.86 (d, J = 6.6 Hz, 3H), 0.81 (ddd, J = 16.6, 11.9, 5.2 Hz, 1H), 0.64 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 169.6, 135.7, 130.8, 129.6, 128.1, 128.0, 124.0, 82.8, 63.8, 56.6, 56.2, 50.2, 42.3, 39.7, 39.5, 38.1, 36.4, 36.2, 36.1, 35.7, 33.8, 31.9, 31.7, 28.2, 28.0, 24.2, 23.8, 22.8, 22.5, 21.1, 19.2, 18.7, 11.8; IR (CDCl₃) 1817 (C=O) cm⁻¹; HRMS (FAB⁺) calcd for C₃₅H₅₁O₂ 503.3889, found 503.3889 (M⁺¹).

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Supporting Information Available: The methodology used for stereochemical assignments of C–H insertion prod-

ucts and ${}^{1}H/{}^{13}C$ spectra for 9, 10, 13a, 14a, 15a, 13b, 14b, 15b, 17, 18, 22, and 23. This material is available free of charge via the Internet at http://pubs.acs.org.

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