conveniently summarized in the diagram depicted in Figure 1. Herein the logarithm of the cmc, crc, and cvc for the different surfactants are plotted versus the sum of the hydrophobic fragmental constants $(\Sigma f_i)^{18}$ of the 1-alkyl substituents. Different linear relationships for the three types of aggregates separate concentration ranges for the various aggregate morphologies.

In summary, the present approach demonstrates that the aggregate morphology within a series of 1-methyl-4- $(C_{12}$ -alkyl)pyridinium iodides of almost equal alkyl chain hydrophobicity is primarily determined by the shape of the surfactant molecule. Shape selectivity also governs the aggregation behavior of 1-alkyl-4-*n*-dodecylpyridinium iodides in which the volume of the core is modified through back bending of a sufficiently long 1-alkyl substituent into the hydrophobic interior of the aggregate. It is our contention that systematic studies of alkyl chain packing will become a major activity in surface chemistry. Further studies along these lines are currently underway in our laboratory.

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Flavin-Catalyzed Oxidation of Amines and Sulfur Compounds with Hydrogen Peroxide

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Mammalian liver contains liver microsomal FAD-containing monooxygenase (EC 1.14.13.8, FADMO), which oxygenates various amines.¹ The enzymatic oxygenation seems to involve the following catalytic cycle.^{2,3a} Oxygenation of enzyme-bound reduced flavin (Enz(FIH₂)) with molecular oxygen gives 4ahydroperoxyflavin (Enz(4a-FIHOOH)), which undergoes monooxygenation of substrates to give 4a-hydroxyflavin (Enz(4a-FIHOH)). Dehydration of Enz(4a-FIHOH) gives oxidized flavin (Enz(Fl_{ox})) (rate-determining step),³ which is reduced to Enz-(FIH₂). The mechanism of FADMO has been extensively studied by using 4a-hydroperoxyisoalloxazines (4a-FIOOH), and much understanding has been gained;^{4,5} however, the catalytic recycling step is still ambiguous.

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Table I. Flavin-Catalyzed Oxidation of Amines and Sulfur Compounds^a

substrate	product ^b	isolated yield, %	turnover number
Bu ₂ NH	BuN ⁺ (O ⁻)=CHPr	61	12
$(PhCH_2)_2NH$	PhCH ₂ N ⁺ (O ⁻)=CHPh	40	8
NH	N [±] O ⁻	70	14
(PhCH ₂) ₂ NOH	PhCH ₂ N ⁺ (O ⁻)=CHPh	83 ^{c,e}	8
Bu ₂ S	Bu₂S→O	99∿	99
Ph ₂ S	Ph ₂ S→O	96°	10
$(PhCH_2)_2S$	(PhCH ₂) ₂ S→O	97 ^{c,d,e}	19
(PhCH ₂) ₂ S→O	(PhCH ₂)SO ₂	98 ^{c,d}	10
$(PhCH_2)_2S$	(PhCH ₂)SO ₂	96 ^d	19

^aA mixture of substrate (1 mmol), $FlEt^+ClO_4^-$ (0.1 mmol), and H_2O_2 (2 mmol) in methanol was allowed to react at room temperature under argon. ^bSatisfactory IR, NMR, mass spectral data, and analyses have been obtained. ^c H_2O_2 (1 mmol). ^d CH_2Cl_2 . ^e $FlEt^+ClO_4^-$ (5 mol %). ^f $FlEt^+ClO_4^-$ (1 mol %).

Scheme I



We have found that 4a-hydroxy-5-alkylflavins are readily transformed into 4a-hydroperoxyflavins upon treatment with hydrogen peroxide. This result leads to the finding of the novel catalytic oxidation which may correspond to FADMO.

The treatment of 4a-FlEtOH with 30% aqueous hydrogen peroxide (10 equiv) in methanol at room temperature under argon gave 4a-FlEtOOH in 82% isolated yield.⁶ Considering this facile formation of 4a-FlEtOOH, 4a-FlEtOH-catalyzed oxidation of substrates with hydrogen peroxide should occur. Indeed, typically, the reaction of dibutylamine (1) with aqueous H_2O_2 in methanol



4a-FIEtOH

4a-FIEtOOH

in the presence of 10 mol % of 4a-FlEtOH gave N-butylidenebutylamine N-oxide (2) in 48% isolated yield (turnover number 10). The catalyst is not limited to 4a-FlEtOH, and flavins such as 4a-FlEtOOH,⁷ FlEt⁺ClO₄^{-,8} 5-ethyl-1,5-dihydro-3-methyllumiflavin (FlEtH),⁸ FMNHEt,⁹ and FMNHMe⁹ can be used as an active catalyst, although the flavins which have no substituent at the 5(N)-position such as 3-methyllumiflavin,¹⁰ riboflavin, and

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FMN are ineffective. It is noteworthy that FADMO undergoes the oxidation, although Enz(4a-FlHOOH) has no substituent at the 5(N)-position. Charge-transfer complexation between Enz-(4a-FIHOOH) and NAD(P)⁺ retards the elimination of hydrogen peroxide to lead to the oxidation.^{2,3a}

We used $FlEt^+ClO_4^-$ as a catalyst because of its efficiency and stability. The representative results of the $FlEt^+ClO_4^-$ -catalyzed oxidation of amines and sulfur compounds are summarized in Table I. Nitrones¹¹ and sulfoxides, which are versatile synthetic intermediates, can be prepared in a highly efficient manner. Sulfoxides are also oxidized, although the rate of the oxidation is much slower than that of sulfides.

The present catalytic reaction can be rationalized by assuming Scheme I. 4a-FlEtOOH reacts with substrate (S) to give oxidized substrate (SO) and 4a-FlEtOH (the second-order rate constant: k_5').^{4a,5,7} 4a-FlEtOH undergoes ionization (k_1, k_2) to give FlEt⁺ which reacts with hydrogen peroxide to afford 4a-FlEtOOH (k_3 , k_4), where k_1 , k_2 , k_3 , and k_4 are pseudo-first-order rate constants.

In order to gain insight into the mechanism, the FlEt+-ClO₄-catalyzed monooxygenation of methyl phenylsulfide (3) with hydrogen peroxide has been investigated in detail by using a solution of H₂O₂ (15 mM), FlEt⁺ClO₄⁻ (0.25 mM), and 3 (0.1-2.0 M) in methanol at 30 °C. The observed initial rate $(v, k_5' \times [S])$ \times [4a-FlEtOOH]) of the formation of methyl phenylsulfoxide (4) has been determined by GLC analysis of 4. The maximum rate of the reaction $(V_{\rm max})$ and the substrate concentration that produced half-maximal rate (K_m) were obtained to be 83 ± 6 mM/h and 4.0 \pm 0.3 M, respectively, from Woolf double reciprocal plot. Under the same conditions, 4a-FlEtOOH reacts with **3** to give **4** and 4a-FlEtOH. The second-order rate constant k_5' in MeOH (30 °C) was determined to be 0.18 M⁻¹ s⁻¹ by monitoring the disappearance of 4a-FlEtOOH at 370 nm. The first-order rate constant of the decomposition of 4a-FlEtOOH (k_6) in MeOH (30 °C) to give 10a-spirohydantoin 5¹² has been determined to be $1.6 \times 10^{-5} \text{ s}^{-1}$ by monitoring the disappearance of the UV absorption of 4a-FlEtOOH at 370 nm. Since k_6 is negligible, V_{max} , K_{m} , and the concentration of FlEt⁺ cation $([FlEt^+])$ can be represented by the following equations: V_{max} = 0.25 mM × $k_1k_3/(k_1 + k_2 + k_3) = 83 \pm 6$ mM/h, $K_m = (k_1k_3 + k_1k_4 + k_2k_4)/(k_1 + k_2 + k_3)/k_5' = 4.0 \pm 0.3$ M, [FlEt⁺] = $0.25 \text{ mM} \times (k_1 k_4 + k_1 k_5' [\text{MeSPh}]) / (k_1 k_3 + (k_1 + k_2) k_4 + (k_1$ $+ k_2 + k_3 k_5$ [MeSPh]). Concentration of FlEt⁺ was determined by the UV spectra of the reaction mixture (545 nm).¹³ The pseudo-first-order rate constant for the reaction of FIEt⁺ClO₄⁻ with aqueous 30% H₂O₂ in MeOH (15 mM solution) to give 4a-FlEtOOH was determined by stopped-flow spectrophotometer to be 5.7 s⁻¹. These results lead to solve the above equations, giving $k_1 = 0.11 \text{ s}^{-1}, k_2 = 0.46 \text{ s}^{-1}, k_3 = 2.5 \text{ s}^{-1}, \text{ and } k_4 = 3.2 \text{ s}^{-1}$ Therefore, the rate-determining step is the formation of FlEt⁺ from 4a-FlEtOH.

The kinetics of the catalytic oxidation of amines is complex, because an equilibrium between FIEt⁺ and the adduct of substrates, 4a-FlEtS (Scheme I, k_7 , k_8), has to be considered. It is known that secondary amines add to FlEt⁺ to give the 4a-amino adducts.4ª The pseudo-first-order rate constant of the formation of 4a-FlEt-NBu₂ from FlEt⁺ and 1 has been determined to be >10³ s⁻¹. The reaction of 4a-FlEtOH with 1 in MeOH gave 4a-FlEt-NBu₂ quantitatively. The pseudo-first-order rate constants of the formation of 4a-FlEt-NBu₂ (355 nm, ϵ 7100) from 4a-FlEtOH and 1 in MeOH were determined to be constant $(1.5 \times$ 10^{-4} s⁻¹) upon changing the concentration of 1 (0.007-0.10 M) by monitoring the UV absorption of 4a-FlEtOH (355 nm, ϵ 8600).

Apparently, 4a-FlEtOH undergoes ionization to give FlEt⁺, which reacts with 1 to afford 4a-FlEt-NBu₂. Under the same conditions, FlEt⁺ also reacts with H_2O_2 to give 4a-FlEtOOH because of higher nucleophilicity of OOH⁻ in comparison with secondary amines.¹⁴ The v value of the oxidation of 1 (0.2 M, 0.20 mM/h) is smaller than that of 3 (0.2 M, 3.9 mM/h); however, the k_5' value of the oxidation of 1 (0.36 M^{-1} s⁻¹) is larger than that of 3 (0.18 M^{-1} s⁻¹). The k_1 value of 1 (ca. 10⁻⁴ s⁻¹) is smaller than that of 3 (0.11 s⁻¹), and the k_3 value of **1** is larger than that of **3**.¹⁵ The v value of the oxidation of 3 (0.3M) in the presence of 1 (0.04 M) by using 4a-FlEtOH as the catalyst (0.015 M) was determined to be 1.4 mM/h, which is smaller than the v value (2.5 mM/h) obtained under the same conditions by using 4a-FlEt-NBu₂ as the catalyst, indicating that the k_1 value is smaller than the k_8 value. Therefore, the rate-determining step of the oxidation of secondary amines seems to be the formation of $FlEt^+$ ion (k_1) .

The present catalytic oxidation is highly useful, because potential flavin hydroperoxide, which has ca. 10⁴ times oxidizing potential in comparison with hydrogen peroxide,^{4a} can be generated catalytically.16

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Supplementary Material Available: A listing of observed initial rates of formation of 4 (Table S1) (1 page). Ordering information is given on any current masthead page.

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Formation of a Cyclopropyl Eicosanoid via an Allene Oxide in the Coral Plexaura homomalla: Implications for the Biosynthesis of 5,6-trans-Prostaglandin A2

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The Caribbean soft coral Plexaura homomalla produces large quantities of prostaglandin (PG) $A_2(1)$ as the methyl ester acetate (ca. 2-3% of its dry weight) by a pathway which is distinct from the mammalian cyclooxygenase/endoperoxide route.¹ This alternative pathway may involve an 8-lipoxygenase and formation of an allene oxide intermediate, although there is as yet no firm evidence linking either to biosynthesis of the prostaglandins.² Allene oxide 2 was recently isolated from incubation of 8(R)hydroperoxyeicosatetraenoic acid (3) with an acetone powder preparation of P. homomalla.³ It is the facility with which allene oxides can form cyclopentenones⁴ which makes this pathway seem attractive for the biosynthesis of PGA₂.

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