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Total Synthesis and Biological Evaluation of Auripyrones A and B

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The total synthesis of auripyrones was achieved by using a novel aldol-type reaction of γ -pyrone as a key step. From this synthetic work, we have established the stereostructure and absolute configuration of auripyrone B. Furthermore, the cytotoxicities of auripyrones against a panel of 39 human cancer cell lines (termed JFCR39) at the Japanese Foundation for Cancer Research were investigated. The patterns of the differential cytotoxicities of auripyrones were COMPARE negative, suggesting that they inhibit cancer cell proliferation through a novel mechanism.

A number of significant biologically active y-pyronecontaining compounds from marine animals have been reported.¹ The structural features of these γ -pyrone-containing compounds are asymmetric centers at the neighboring positions of the γ -pyrone part. In 1996, Yamada and co-workers reported the isolation of auripyrones A (1) and B (2) from the sea hare Dolabella auricularia (Aplysiidae), which exhibit cytotoxicity against HeLa S₃ cells with IC₅₀ values of 0.26 and 0.48 μ g mL⁻¹, respectively (Figure 1).² The relative stereochemistry of the core unit of auripyrones A (1) and B (2) was determined by an extensive analysis of coupling constants and the NOESY correlations. The main structural features of auripyrones are a γ -pyrone ring and a spiroacetal moiety. The unique structures of auripyrones, in conjunction with their biological activities, have made them attractive synthetic targets. Several groups have reported approaches to the synthesis of auripyrones. In 2006, Perkins and Lister were the first to achieve the total synthesis of auripyrone A (1), featuring spiroacetalization.³ This synthesis established the absolute configuration of auripyrone A (1). In 2009, Jung and Salehi-Rad achieved the total synthesis of auripyrone A (1) by using a tandem non-aldol aldol/Paterson aldol process.⁴ The next year, we preliminarily reported the total synthesis of auripyrones A (1) and B (2) by using a novel diastereoselective aldol-type reaction of 2,6diethyl-3,5-dimethyl-4-pyrone (8).⁵ From this synthetic work, we determined the absolute configuration of auripyrone B (2). Very recently, Jung and co-workers reported the total synthesis



Figure 1. Structures of auripyrones A (1) and B (2).

of auripyrone B (2) using their previous strategy.⁶ We describe herein the detailed synthesis of auripyrones A (1) and B (2) and their cytotoxicities against a panel of 39 human cancer cell lines.

Results and Discussion

Our retrosynthetic analyses of auripyrones A (1) and B (2) are shown in Scheme 1. Auripyrones A (1) and B (2) can be obtained from triketone **3** by using spiroacetalization. Triketone **3** can be derived from C1–C13 segment **4** and C14–C20 segment **5** by using the aldol coupling reaction. Asymmetric centers at C11 and C12 of C1–C13 segment **4** can be constructed from aldehyde **7** by asymmetric crotylboration. Aldehyde **7**, which possesses an asymmetric center at the neighboring position of γ -pyrone, can be synthesized from 2,6-diethyl-3,5-dimethyl-4-pyrone (**8**) and the optically active aldehyde **9** by diastereoselective aldol-type reaction as a key step.^{7,8}

Synthesis of C1–C13 Segment. Previously, we reported diastereoselective aldol-type reaction of γ -pyrone (8) with NaHMDS⁷ and the Mukaiyama aldol-type reaction of silyl enol

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Scheme 1. Retrosynthetic analyses of auripyrones A (1) and B (2).

ether of γ -pyrone 8.⁸ These approaches have the benefits of straightforward access even to complex molecules and of the construction of two stereogenic centers at once. Therefore, we planned the synthesis of auripyrones A (1) and B (2) starting from this aldol-type reaction between γ -pyrone (8) and optically active aldehyde 10.9 First, we tried the aldol-type reaction with NaHMDS to afford the desired compound 11 in 47% yield along with other diastereomers (21%) (Scheme 2). Although we attempted a Mukaiyama aldol-type reaction between the silyl enol ether of γ -pyrone 8 and aldehyde 10 by using TiCl₄ to give the desired aldol adduct 11, the yield and diastereoselectivity were low (desired aldol adduct 11: 17% yield, other diastereomers: 21%). The configuration of 11 was determined from ¹HNMR of the corresponding acetonide derivative A including NOESY correlations (Scheme 3). Protection of the secondary hydroxy group in aldol adduct 11 gave a TBS ether, which was transformed into aldehyde 12 by removal of the trityl group and oxidation of the resulting primary hydroxy group. Aldehyde 12 was converted into homoallylic alcohol 14 as a single diastereomer via Brown crotylation using the (E)-crotyl boronate 13.¹⁰ The stereochemistry of 14 was determined as follows. Homoallylic alcohol 14 was converted into 1,3acetonide **B**, the stereochemistry of which was confirmed to be syn by the ¹³C chemical shifts of two acetonide methyls ($\delta_{\rm C}$ 19.2 and 29.8) (Scheme 4).¹¹ Furthermore, 1,3-acetonide B was

converted into 1,3-acetonide **C**, and the configuration at C11 and C12 in **C** was determined by ${}^{1}\text{H}{-}{}^{1}\text{H}$ coupling constants and NOESY correlations. Introduction of the isovaleryl group at C11 in homoallylic alcohol **14** and dihydroxylation of the terminal olefin afforded a diol, which was converted into aldehyde **15** as a C1–C13 segment. This two-step procedure was superior to the direct Lemieux–Johnson conditions¹² in both yield and reproducibility because of the instability of aldehyde **15**.

Synthesis of C14–C20 Segment. Next, we prepared C14–C20 segment **17** (Scheme 5). Oxidation of commercially available (*S*)-2-methy-1-butanol gave aldehyde **16**.¹³ The aldol reaction of aldehyde **16** and 3-pentanone with LDA followed by the protection of the resulting secondary hydroxy group as the TES ether gave C14–C20 segment **17** as a diastereomeric mixture. This segment **17** was used for the next reaction without separation of diastereomers because the configurations of these newly generated stereocenters were lost by oxidation and epimerization in subsequent steps.

Total Synthesis of Auripyrone A. With C1–C13 segment **15** and C14–C20 segment **17** in hand, we next tried the aldol coupling reaction between two segments. First, we attempted the aldol coupling reaction between model compounds **18** and **17** with LDA, LHMDS, and NaHMDS, but the yields were low (9–43%) (Table 1). We then examined the deuteration of C14–



(500 MHz, CDCl₃) (500 MHz, CDCl₃)

Scheme 3. Determination of stereochemistry of 11.

C20 segment 17, as depicted in Table 2. Interestingly, treatment of 17 with LDA or LHMDS, followed by the addition of D₂O, did not give the deuterated compound 20 (Entries 1 and 2). In Entries 3 and 4, the enolate of 17 prepared with KHMDS and NaHMDS, respectively, gave a deuterated compound 20 in 70% yields in both cases. However, as we reported that γ pyrone 8 is easily deprotonated by using metal bis(trimethylsilyl)amides (deuteration yield: >95%),^{7,8} model compound **18** (C2 position) was expected to be more easily deprotonated at the C2 position of γ -pyrone than C14–C20 segment 17 was at the C14 position, reducing the yield of the aldol adduct. Therefore, we attempted an aldol reaction with a weaker base. The Mukaiyama Sn(II)-promoted aldol reaction¹⁴ between C1-C13 segment 15 and C14-C20 segment 17 using Sn(OTf)₂ and Et₃N afforded coupling compound 21 in 99% yield as a diastereomeric mixture (Scheme 6).

Next, we tried to synthesize auripyrone A (1) from coupling compound **21** (Scheme 7). Selective removal of the TES group in **21** gave a diol, which was oxidized into triketone **22** as an equilibrium mixture of the keto and enol forms. Removal of the TBS group in triketone **22** by HF•py was accompanied by spiroacetalization to give auripyrone A (1) as a single isomer. The synthetic auripyrone A (1) is identical in all respects to the natural product.²

Stereocontrol in the spiroacetalization to auripyrone A (1) can be explained as follows (Figure 2). Triketone 22 was converted into hemiacetals 23a and 23b. The stereochemistry of spiro carbon (C13) in 23a and 23b was controlled by the double anomeric effect. The C14 methyl group in hemiacetal 23a was epimerized into the equatorial position (hemiacetal 23b) so as to avoid a 1,3-diaxial interaction between the C12 and C14 methyl groups in 23a via β -diketone form 22.



Scheme 4. Determination of stereochemistry of 14.



Scheme 5. Synthesis of C14–C20 segment 17.









Scheme 6. Aldol coupling reaction between C1–C13 segment 15 and C14–C20 segment 17.

Table 2. Deuteration of Ketone 17							
0	OTES 1) base (1.1 equiv) THF, –78 °C	O OTES				
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$							
		D 1	X7 11 C 1 / / 1				
Entry	Base	ketones	Yield of deuterated compound $20/\%^{a}$				
Entry 1	Base LDA	ketones quant.	$\frac{\text{Yield of deuterated}}{\text{compound } 20/\%^{a)}}{0}$				
Entry 1 2	Base LDA LHMDS	quant.	$\begin{array}{c} \text{Yield of deuterated} \\ \hline \text{compound } 20/\%^{\text{a})} \\ \hline 0 \\ 0 \\ \end{array}$				
Entry 1 2 3	Base LDA LHMDS KHMDS	quant. quant. quant.	Yield of deuterated compound $20/\%^{a}$ 0 0 70				

a) The deuteration yield was calculated by ¹H NMR.

Total Synthesis of (2'S)- and (2'R)-Auripyrones B. We next tried the synthesis of (2'S)- and (2'R)-auripyrones B to determine the absolute structure of auripyrones B (2). Therefore, we planned the conversion of auripyrone A (1) into B (2) by removal of the isovaleryl group at C11 in auripyrone A (1) and subsequent esterification with optically active 2-methylbutanoic acid. Thus, hydrolysis of the isovaleryl group of auripyrone A (1) with LiOH in aqueous methanol was attempted (Scheme 8). However, we could not obtain the desired deacyl derivative 24, but a bis(pyrone) compound 25. Bis(pyrone) compound 25 was thought to be produced by deprotonation at C14 and elimination.

On the other hand, Jung and co-workers reported that the hydrolysis of auripyrone A (1) under similar conditions (KOH(aq)/THF) gave the C14-epimerized elimination compounds 26 and 27 (Scheme 9).⁶ From these results, auripyrone A (1) was shown to be easily deprotonated at the C14 position under basic conditions, causing side reactions.





Scheme 7. Total synthesis of auripyrone A (1).



Figure 2. Spiroacetalization of triketone 22.



Scheme 8. Hydrolysis of auripyrone A (1).



Scheme 9. Hydrolysis of auripyrone A (1) by Jung and co-workers.

Therefore, we attempted the synthesis of auripyrone B (2) by using our synthetic strategy for auripyrone A (1) (Scheme 10). Esterification between homoallylic alcohol 14 and (*S*)-2methylbutanoic (28)¹⁵ by using Yamaguchi conditions¹⁶ gave compound 29. The terminal olefin in 29 was transformed into a diol, which was oxidatively cleaved to afford aldehyde 30. The coupling reaction between aldehyde 30 and C14–C20 segment 17 via the Mukaiyama Sn(II)-promoted aldol reaction¹⁴ gave the coupling product 31 as a diastereomeric mixture. Removal of the TES group in 31 and subsequent oxidation of the diol group afforded triketone 32 as a mixture of the keto and enol forms, a precursor of spiroacetalization. Removal of the TBS group in triketone 32 by HF•py and a spontaneous spiroacetalization gave (2'*S*)-auripyrone B (33).

Also, (2'R)-auripyrone B (36) was synthesized from homoallylic alcohol 14 in the same manner with (*R*)-2-methylbutanoic (34) (Scheme 11).

Determination of the Absolute Configuration of Auri**pyrone B.** With both diastereomers (2'S)-auripyrone B (33)and (2'R)-auripyrone B (36) in hand, the determination of configuration of C2' in auripyrone B was determined by comparing the ¹HNMR spectra of synthetic samples with those reported for the natural auripyrone B (2) (Figure 3). The chemical shifts of protons in the acyl group (H4', H5') in (2'S)auripyrone B (33) were different from those of (2'R)-auripyrone B (36). The ¹H NMR data of (2'S)-auripyrone B (33) were in good agreement with those of the natural one. Furthermore, comparison of the optical rotations of synthetic (2'S)-auripyrone B (33) and natural auripyrone B (2) determined the absolute configuration of auripyrone B (2): The optical rotation of synthetic (2'S)-auripyrone B (33) { $[\alpha]_D^{25} = +43$ (c = 0.29, CHCl₃)} was in accordance with that of the natural one² $\{[\alpha]_{D}^{25} = +39 \ (c = 0.14, \text{ CHCl}_3)\}$ to clarify the absolute configuration of auripyrone B (2) as depicted in Figure 4.

Biological Evaluation of Auripyrones A and B. Table 3 shows the cytotoxic activities of auripyrones A (1) and B (2), 2'-*epi* auripyrone B (**36**), and bis(pyrone) compound **25** against HeLa S₃ cells. Auripyrones A (1) showed an IC₅₀ value of cytotoxic activity equivalent to that of auripyrone B (2). The

IC₅₀ values of synthetic auripyrones A (1) and B (**33**) were about 10-fold larger (less cytotoxic) than the reported IC₅₀ values of natural auripyrones,² maybe because of the condition of the HeLa S₃ cells. 2'-epi Auripyrone B (**36**) showed cytotoxicity, with an IC₅₀ of $3.4 \,\mu g \,m L^{-1}$. From these results, the structure of the O^{11} -acyl group in auripyrones was shown to be unimportant for cytotoxicity. On the other hand, bis(pyrone) compound **25** showed no cytotoxicity even at $10 \,\mu g \,m L^{-1}$. This result suggested that the spiro structure of auripyrones is essential for their cytotoxicity.

The cytotoxicities of synthetic auripyrones A (1) and B (33) were evaluated against a panel of 39 human cancer cell lines (termed JFCR39) at the Japanese Foundation for Cancer Research (Table 4). Auripyrones A (1) and B (33) each showed broad cytotoxicity in the panel, respectively. On the basis of the COMPARE analysis,¹⁷ the patterns of the differential cytotoxicities of auripyrones A (1) and B (33) suggested that they inhibited cancer cell proliferation through a novel mechanism.

Conclusion

In conclusion, we have achieved the total synthesis of auripyrones A (1) (2.6% overall yield in 13 steps) and B (2) (2.8% overall yield in 13 steps), featuring a diastereoselective aldol-type reaction between 2,6-diethyl-3,5-dimethyl-4-pyrone (8) and optically active aldehyde 10 to construct three contiguous stereocenters. From this synthetic work, we have established the stereostructure and absolute configuration of auripyrone B (2). Furthermore, we have investigated the cytotoxicities of auripyrones A (1) and B (2) were COMPARE-negative, suggesting that they inhibit cancer cell proliferation through a novel mechanism. Further studies on the synthesis of O^{11} -modified probe molecules of auripyrones for use in the search for target biomolecules are currently in progress.

Experimental

General. All reagents and dry solvents were used as obtained from commercial supplies unless otherwise noted. Organic solvents for moisture-sensitive reactions were distilled



Scheme 10. Total synthesis of (2'S)-auripyrone B (33).



Scheme 11. Total synthesis of (2'R)-auripyrone B (36).

by standard procedure. Column chromatography was performed using silica gel (75–200 or $45-75\,\mu$ m). All moisturesensitive reactions were performed under an atmosphere of argon or nitrogen, and the starting materials were azeotropically dried with benzene before use. Optical rotations were measured on digital polarimeter at room temperature, using



Figure 3. Partial 1 HNMR spectra (600 MHz, C₆D₆) of synthetic auripyrone B and of the natural one.



Figure 4. Absolute stereochemistry of auripyrone B (2).

Table 3.	Cytotoxicities of A	uripyrones and	Their Analogues
Agains	st HeLa S ₃ Cells		

	$IC_{50} \text{ values}/\mu g m L^{-1}$
Auripyrone A (1)	3.1 (lit. 0.26)
Auripyrone B (33)	3.1 (lit. 0.48)
2'-epi Auripyrone B (36)	3.4
Bis(pyrone) compound 25	>10

the sodium D line. Infrared (IR) spectra were recorded on a FT IR system and only selected peaks are reported. ¹H and ¹³C nuclear magnetic resonance (NMR) spectra were run at various field strengths as indicated. The ¹H and ¹³C chemical shifts (δ) are reported in parts per million (ppm) downfield relative to tetramethylsilane (TMS), CDCl₃ ($\delta_{\rm H}$ 7.26 and $\delta_{\rm c}$ 77.0) or C₆D₆ ($\delta_{\rm H}$ 7.16 and $\delta_{\rm c}$ 128.0). Coupling constants (*J*) are reported in Hz. Multiplicities are reported using the following abbreviations: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet. High-resolution mass spectra (HRMS) were recorded by electrospray ionization (ESI)/time-of-flight (TOF) experiments. Melting points are uncorrected.

		$GI_{50}{}^{a),b)}/\mu M$	
	Cell line	Auripyrone	Auripyrone
		A (1)	B (33)
Breast	HBC-4	4.3	3.5
	BSY-1	2.0	1.9
	HBC-5	5.2	3.6
	MCF-7	4.5	3.7
	MDA-MB-231	4.3	2.5
Central nervous	U251	4.5	3.0
system	SF-268	15>	4.5
	SF-295	3.9	2.9
	SF-539	12	2.4
	SNB-75	3.1	2.0
	SNB-78	8.5	3.4
Colon	HCC2998	2.6	1.8
	KM-12	3.4	2.6
	HT-29	3.3	2.8
	HCT-15	4.6>	2.6
	HCT-116	3.4	2.3
Lung	NCI-H23	3.5	2.9
	NCI-H226	4.6	3.8
	NCI-H522	2.4	1.8
	NCI-H460	4.7	3.2
	A549	5.1	3.2
	DMS273	3.5	2.4
	DMS114	3.5	2.2
Melanoma	LOX-IMVI	5.5	2.2
Ovary	OVCAR-3	2.9	2.1
	OVCAR-4	3.6	2.8
	OVCAR-5	10>	3.7
	OVCAR-8	6.7	3.6
	SK-OV-3	13	3.7
Kidney	RXF-631L	7.3	3.0
	ACHN	5.7	3.5
Stomach	St-4	4.4	3.7
	MKN1	2.8	2.5
	MKN7	2.1	2.3
	MKN28	3.0	2.5
	MKN45	2.8	2.3
	MKN74	2.6	2.6
Prostate	DU-145	20>	4.4
	PC-3	3.5	2.9
MG-MID ^{c)}		-5.35	-5.55
Delta ^a		0.36	0.19
Range ^{e)}		1	0.4

Table 4. GI₅₀ Values of Auripyrones A (1) and B (**33**) against 39 Human Cancer Cell Lines

a) Concentrations for the inhibition of cell growth at 50% relative to control. b) Cell growth was determined according to the sulforhodamine B assay. c) Mean GI_{50} value in all of the cell lines tested. d) Difference in the GI_{50} value between the most-sensitive cells and the MG-MID value. e) Difference in the log GI_{50} value between the most- and least-sensitive cells.

Aldol 11. To a stirred solution of 2,6-diethyl-3,5-dimethyl-4-pyrone (8) (200 mg, 1.11 mmol) in THF (1.5 mL) was added NaHMDS (1.0 M solution in THF, 1.15 mL, 1.15 mmol) at -78 °C. After being stirred at -78 °C for 1.5 h, a solution of aldehyde 10 (275 mg, 0.832 mmol) in THF (1.5 mL) was added via cannula. The mixture was stirred at -78 °C for 2.3 h, diluted with saturated aqueous NH₄Cl (10 mL), and extracted with EtOAc (3 \times 20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (30 g, hexane–EtOAc 7:1 \rightarrow 3:1) to give a diastereomeric mixture of aldols (290 mg, 68%) and recovered 2,6-diethyl-3,5dimethyl-4-pyrone (8) (74.5 mg, 37%). Further purification of the diastereomeric mixture of aldols by column chromatography on silica gel (FL-60D, 26g, benzene-acetone 20:1; FL-60D, 12 g, benzene-acetone 20:1) gave aldol 11 (199 mg, 47%) and an inseparable mixture of undesired diastereomers (90.6 mg, 21%) as colorless powder, respectively: $R_f = 0.33$ (1:1 hexane/EtOAc); mp 136–138 °C; $[\alpha]_{D}^{23}$ –7.2 (c 0.90, CHCl₃); IR (CHCl₃): 3500 (br), 1655, 1595 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 7.45–7.40 (m, 6H), 7.34–7.20 (m, 9H), 4.10 (dd, J = 9.7, 3.2 Hz, 1H), 3.36–3.19 (m, 2H), 3.07 (dg, J = 9.7, 7.2 Hz, 1H), 2.62 (q, J = 7.6 Hz, 2H), 2.10–1.80 (m, 1H), 1.96 (s, 3H), 1.93 (s, 3H), 1.22 (t, J = 7.6 Hz, 3H), 1.12 (d, J = 7.2 Hz, 3H), 1.02 (d, J = 7.1 Hz, 3H), A signal due toone proton (OH) was not observed; ¹³C NMR (67.8 MHz, CDCl₃): § 179.5, 164.7, 163.7, 143.6 (3C), 128.4 (6C), 127.7 (6C), 126.9 (3C), 119.1, 117.6, 86.7, 74.1, 67.2, 38.8, 34.8, 24.7, 14.7, 11.2, 9.6, 9.5, 9.2; HRMS (ESI): m/z 533.2664, calcd for $C_{34}H_{38}O_4Na [M + Na]^+$ 533.2668.

Aldehyde 12. To a stirred solution of aldol 11 (1.61 g, 3.16 mmol) in DMF (3 mL) were added imidazole (2.10 g, 30.1 mmol) and tert-butyldimethylsilvl chloride (2.40 g, 15.9 mmol) at room temperature. The mixture was stirred at 70 °C for 22 h, diluted with H₂O (30 mL) at room temperature, and extracted with Et₂O (3×50 mL). The combined extracts were washed with brine, dried over Na2SO4, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (60 g, hexane-EtOAc $10:1 \rightarrow 5:1 \rightarrow 1:1$) to give tert-butyldimethylsilyl ether 11a (1.95 g, 99%) as a yellow amorphous solid; $R_f = 0.73$ (1:1 hexane/EtOAc); mp 37-39 °C; $[\alpha]_D^{25}$ +4.1 (*c* 0.405, CHCl₃); IR (CHCl₃): 1655, 1595 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 7.45–7.36 (m, 6H), 7.35–7.20 (m, 9H), 4.06 (d, J = 8.9 Hz, 1H), 3.20–3.00 (m, 3H), 2.68 (dq, J = 15.2, 7.6 Hz, 1H), 2.56 (dq, J = 15.2, 7.6 Hz, 1H), 2.05–1.88 (m, 1H), 1.94 (s, 3H), 1.94 (s, 3H), 1.25 (t, J = 7.6 Hz, 3H), 1.13 (d, J = 7.0 Hz, 3H), 0.94 (d, J =6.8 Hz, 3H), 0.63 (s, 9H), -0.35 (s, 3H), -0.48 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 179.6, 164.7, 163.6, 144.1 (3C), 128.5 (6C), 127.5 (6C), 126.7 (3C), 119.3, 117.9, 86.5, 73.6, 66.6, 40.3, 36.8, 25.8 (3C), 24.7, 18.1, 15.1, 11.3, 10.1, 9.8, 9.5, -4.6, -4.7; HRMS (ESI): m/z 647.3532, calcd for $C_{40}H_{52}O_4SiNa \ [M + Na]^+ 647.3533;$ Anal. Found: C, 76.47; H, 8.47%. Calcd for C₄₀H₅₂O₄: C, 76.88; H, 8.39%.

To a stirred solution of *tert*-butyldimethylsilyl ether **11a** (165 mg, 0.265 mmol) in Et₂O (3 mL) was added formic acid (6 mL) at room temperature. The mixture was stirred at room temperature for 10 min, poured into saturated aqueous NaHCO₃ (150 mL) at 0 °C, and extracted with Et₂O (3 × 20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated to afford a crude formate. The crude formate was dissolved in methanol (9 mL), and 25% aqueous NH₃ (3 mL) was added to the mixture at room temperature. The

mixture was stirred at room temperature for 20 min, concentrated, diluted with half-saturated brine (10 mL), and extracted with EtOAc ($3 \times 10 \text{ mL}$). The combined extracts were dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10 g, hexane-EtOAc $5:1 \rightarrow 3:1 \rightarrow 1:1$) to give alcohol 11b (92.7 mg, 92%) as a colorless oil: $R_f = 0.37$ (1:1 hexane/EtOAc); $[\alpha]_D^{24}$ –2.2 (c 0.815, CHCl₃); IR (CHCl₃): 3416, 1653, 1592, 1463, 1378 cm⁻¹; ¹HNMR (270 MHz, CDCl₃): δ 4.12 (dd, J = 9.0, 1.6 Hz, 1H), 3.62–3.45 (m, 2H), 3.20 (dq, J = 9.0,7.0 Hz, 1H), 2.68 (dq, J = 15.2, 7.6 Hz, 1H), 2.55 (dq, J =15.2, 7.6 Hz, 1H), 2.05-1.85 (m, 1H), 1.97 (s, 3H), 1.95 (s, 3H), 1.23 (t, J = 7.4 Hz, 3H), 1.15 (d, J = 6.8 Hz, 3H), 0.93 (d, J = 7.0 Hz, 3H), 0.80 (s, 9H), 0.02 (s, 3H), -0.40 (s, 3H),A signal due to one proton (OH) was not observed: ¹³C NMR (67.8 MHz, CDCl₃): δ 179.8, 165.0, 163.9, 119.3, 117.9, 73.1, 65.0, 39.9, 38.6, 25.9 (3C), 24.7, 18.1, 15.2, 11.4, 9.8, 9.5, 9.4, -4.8 (2C); HRMS (ESI): m/z 405.2461, calcd for C₂₁H₃₈O₄-SiNa $[M + Na]^+$ 405.2437.

To a stirred solution of oxalyl chloride (70.0 µL, 0.800 mmol) in CH₂Cl₂ (0.4 mL) was added DMSO (4.0 M solution in CH₂Cl₂, 0.500 mL, 2.00 mmol) at -78 °C. The mixture was stirred at -78 °C for 30 min, and a solution of alcohol 11b (151 mg, 0.394 mmol) in CH₂Cl₂ (1.0 mL) was added via cannula. The mixture was stirred at -78 °C for 15 min, and diisopropylethylamine (0.280 mL, 1.61 mmol) was added. The resulting mixture was stirred at -78 °C for 15 min, warmed to 0 °C, and stirred for an additional 15 min. The mixture was diluted with pH 6.8 phosphate buffer (5 mL) and extracted with EtOAc ($3 \times 10 \text{ mL}$). The combined extracts were washed with brine, dried over Na2SO4, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (6.5 g, hexane–EtOAc $3:1 \rightarrow 1:1$) to give aldehyde 12 (149 mg, 99%) as a colorless oil: $R_f = 0.53$ (1:1 hexane/ EtOAc); $[\alpha]_{D}^{25}$ +29.0 (c 0.813, CHCl₃); IR (CHCl₃): 2827, 2786, 1728, 1655, 1595, 1463, 1426, 1377 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 9.71 (s, 1H), 4.54 (dd, J = 8.9, 1.9 Hz, 1H), 3.20 (dq, J = 8.9, 7.1 Hz, 1H), 2.70 (dq, J = 15.2, 7.6 Hz, 1H), 2.57 (dq, J = 15.2, 7.6 Hz, 1H), 2.52 (dq, J = 1.9, 6.8 Hz, 1H), 1.97 (s, 3H), 1.95 (s, 3H), 1.26 (t, J = 7.4 Hz, 3H), 1.22 (d, J = 6.8 Hz, 3H), 1.17 (d, J = 7.1 Hz, 3H), 0.74 (s, 9H), -0.09 (s, 3H), -0.34 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 203.9, 179.6, 163.9, 163.5, 119.8, 118.2, 72.3, 49.8, 40.1, 25.7 (3C), 24.8, 18.0, 15.1, 11.5, 9.9, 9.6, 6.4, -4.4, -4.9; HRMS (ESI): m/z 403.2275, calcd for C₂₁H₃₆O₄SiNa [M + Na]⁺ 403.2281.

Homoallylic Alcohol 14. To a stirred solution of potassium *tert*-butoxide (455 mg, 4.05 mmol) in THF (1.3 mL) were added *trans*-2-butene (1.20 mL, 12.9 mmol) and *n*-BuLi (1.57 M solution in hexane, 2.55 mL, 4.00 mmol) at -78 °C. After the mixture was stirred at -40 °C for 1 h, a solution of (–)-*B*-methoxydiisopinocampheylborane (2.10 g, 6.64 mmol) in THF (2.1 mL) was added via cannula at -78 °C. After 40 min, boron trifluoride diethyl etherate (0.754 mL, 6.00 mmol) and a solution of aldehyde **12** (780 mg, 2.05 mmol) in THF (2.3 mL) were added successively. The mixture was stirred at -40 °C for 8.3 h and diluted with a 1:1 mixture of Et₂O and EtOH (10 mL). The mixture was warmed up to room temperature and treated with a 1:1 mixture of 20% H₂O₂ and 20% NaOH (10 mL). After being

stirred for 13 h, the mixture was extracted with EtOAc (3 \times 15 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (FL-60D, 25 g, hexane-EtOAc 7:1; FL-60D, 25 g, hexane-EtOAc 7:1; FL-60D, 25 g, hexane–EtOAc 7:1 \rightarrow 5:1 \rightarrow 3:1) to give homoallylic alcohol 14 (636 mg, 71%) as colorless powder: $R_f =$ 0.57 (1:1 hexane/EtOAc); mp 24–26 °C; $[\alpha]_D^{26}$ +9.7 (c 0.600, CHCl₃); IR (CHCl₃): 3429, 1653, 1593, 1463, 1428, 1378 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 5.76 (ddd, J = 17.0, 10.5, 8.6 Hz, 1H), 5.15–5.05 (m, 2H), 4.03 (dd, J = 8.1, 2.4 Hz, 1H), 3.40 (t, J = 6.0 Hz, 1H), 3.28 (dq, J = 8.1, 7.0 Hz, 1H), 2.75-2.50 (m, 2H), 2.45-2.30 (m, 1H), 2.05-1.90 (m, 1H), 1.97 (s, 3H), 1.95 (s, 3H), 1.24 (t, J = 7.6 Hz, 3H), 1.17 (d, J = 7.0 Hz, 3H), 1.02 (d, J = 7.0 Hz, 3H), 1.01 (d, J = 7.0 Hz, 3H), 0.81 (s, 9H), 0.02 (s, 3H), -0.28 (s, 3H), A signal due to one proton (OH) was not observed; ¹³C NMR (67.8 MHz, CDCl₃): δ 179.6, 164.7, 163.6, 139.7, 119.2, 117.9, 116.2, 75.3, 75.0, 41.6, 40.9, 39.0, 26.0 (3C), 24.8, 18.1, 17.5, 14.9, 11.4, 9.9, 9.6, 9.0, -4.1, -4.6; HRMS (ESI): m/z 459.2922, calcd for $C_{25}H_{44}O_4SiNa [M + Na]^+ 459.2907$.

C1-C13 Segment 15. To a stirred solution of homoallylic alcohol 14 (120 mg, 0.275 mmol) in pyridine (1 mL) were added dimethylaminopyridine (3.70 mg, 30.3 µmol) and isovaleryl chloride (0.200 mL, 1.64 mmol) at room temperature. The mixture was stirred at room temperature for 18 h, poured into saturated aqueous NaHCO₃ (50 mL) at 0 °C, and extracted with EtOAc ($3 \times 10 \text{ mL}$). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10 g, hexane-EtOAc $10:1 \rightarrow 5:1 \rightarrow 3:1; 10$ g, hexane-EtOAc $10:1 \rightarrow 5:1$) to give ester **14a** (143 mg, quant) as a colorless oil: $R_f = 0.77$ (1:1 hexane/EtOAc); $[\alpha]_{\rm D}^{27}$ +19.7 (c 1.445, CHCl₃); IR (CHCl₃): 1727, 1654, 1594, 1463, 1377 cm⁻¹; ¹HNMR (270 MHz, CDCl₃): δ 5.73 (ddd, J = 17.5, 10.1, 6.8 Hz, 1H), 5.10–4.95 (m, 3H), 4.04 (d, J = 8.4 Hz, 1H), 3.16 (dq, J = 8.4, 7.1 Hz, 1H), 2.70–2.50 (m, 3H), 2.30–2.05 (m, 3H), 2.10–1.90 (m, 1H), 1.96 (s, 3H), 1.93 (s, 3H), 1.26 (t, J = 7.6 Hz, 3H), 1.09 (d, J = 7.1 Hz, 3H), 0.98 (d, J = 6.2 Hz, 9H), 0.89 (d, J = 7.0 Hz, 3H), 0.83 (s, 9H), 0.07 (s, 3H), -0.34(s, 3H); 13 C NMR (67.8 MHz, CDCl₃): δ 179.6, 172.5, 164.5, 163.6, 138.7, 119.4, 118.0, 115.9, 76.4, 72.6, 43.5, 40.6, 40.2, 37.9, 26.0 (3C), 25.4, 24.8, 22.6, 22.5, 18.2, 18.1, 15.4, 11.3, 9.9, 9.5, 9.3, -3.9, -4.5; HRMS (ESI): m/z 543.3486, calcd for $C_{30}H_{52}O_5SiNa [M + Na]^+ 543.3482$.

To a stirred solution of ester **14a** (143 mg, 0.186 mmol) in acetone (1 mL) and H₂O (1 mL) were added *N*-methylmorpholine oxide (103 mg, 0.879 mmol) and osmium tetroxide (0.1 M solution in *tert*-butanol, 0.500 mL, 50.0 µmol) at room temperature. The mixture was stirred at room temperature for 17 h, diluted with saturated aqueous NaHSO₃ (5 mL) at 0 °C, and extracted with EtOAc (3 × 10 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10 g, hexane–EtOAc 1:1 → 0:1) to give a diastereomeric mixture of diol **14b** (ca. 6:1) (138 mg, 90%) as a colorless oil: $R_f = 0.20$ (major), 0.13 (minor) (1:1 hexane/EtOAc); IR (CHCl₃): 3421, 1726, 1651, 1592, 1464, 1378 cm⁻¹; ¹HNMR (270 MHz, CDCl₃) for major isomer: δ 5.05

(dd, J = 8.4, 6.8 Hz, 1H), 4.07 (dd, J = 7.5, 2.4 Hz, 1H), 3.75– 3.64 (m, 2H), 3.47–3.40 (m, 1H), 3.23 (dq, J = 7.5, 7.3 Hz, 1H), 2.70–2.55 (m, 3H), 2.30–2.05 (m, 6H), 1.99 (s, 3H), 1.95 (s, 3H), 1.24 (t, J = 7.6 Hz, 3H), 1.22 (d, J = 7.3 Hz, 3H), 0.97 (d, J = 6.2 Hz, 6H), 0.92 (d, J = 7.0 Hz, 3H), 0.86 (s, 9H), 0.83 (d, J = 6.8 Hz, 3H), 0.06 (s, 3H), -0.23 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 180.0, 172.6, 164.8, 164.4, 119.5, 118.0, 77.0, 73.4, 72.0, 64.3, 43.6, 40.9, 38.0, 37.6, 26.0 (3C), 25.4, 24.8, 22.5 (2C), 18.2, 14.9, 12.9, 11.4, 10.2, 10.0, 9.6, -3.9, -4.3; HRMS (ESI): m/z 577.3533, calcd for C₃₀H₅₄O₇SiNa [M + Na]⁺ 577.3537.

To a stirred solution of diol 14b (138 mg, 0.249 mmol) in acetone (1 mL) and H₂O (1 mL) was added NaIO₄ (103 mg, 0.390 mmol). The mixture was stirred at room temperature for 15 min, diluted with half-saturated brine (5 mL), and extracted with EtOAc ($3 \times 10 \text{ mL}$). The combined extracts were dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (8 g, hexane-EtOAc $3:1 \rightarrow 2:1 \rightarrow 1:1$) to give C1–C13 segment 15 (94.3 mg, 72%) as a colorless oil and recovered diol 14b (18.1 mg, 13%); 15: $R_f = 0.66$ (1:1 hexane/EtOAc); $[\alpha]_D^{26} + 17.7$ (c 0.951, CHCl₃); IR (CHCl₃): 1725, 1653, 1594, 1463, 1427, 1377 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 9.67 (d, J = 3.2 Hz, 1H), 5.26 (dd, J = 8.9, 3.2 Hz, 1H), 4.06 (dd, J = 8.4, 2.1 Hz, 1H), 3.19 (dg, J = 8.4, 6.8 Hz, 1H), 2.80–2.55 (m, 3H), 2.26– 2.04 (m, 4H), 1.97 (s, 3H), 1.95 (s, 3H), 1.26 (t, J = 7.1 Hz, 3H), 1.15 (d, J = 6.5 Hz, 3H), 1.13 (d, J = 6.8 Hz, 3H), 0.98 (d, J = 6.2 Hz, 6H), 0.96 (d, J = 6.8 Hz, 3H), 0.82 (s, 9H), 0.03 (s, 3H), -0.32 (s. 3H); ¹³C NMR (67.8 MHz, CDCl₃); δ 202.8, 179.5, 172.3, 164.0, 163.8, 119.5, 118.1, 75.1, 73.4, 47.6, 43.3, 40.5, 38.8, 26.0 (3C), 25.5, 24.8, 22.5 (2C), 18.1, 15.3, 11.8, 11.4, 10.1, 9.9, 9.6, -3.9, -4.5; HRMS (ESI): m/z 545.3268, calcd for $C_{29}H_{50}O_6SiNa [M + Na]^+$ 545.3274.

C14-C20 Segment 17. To a stirred solution of LDA (5.21 mmol) prepared from diisopropylamine (0.730 mL, 5.21 mmol) and n-BuLi (1.63 M solution of in hexane, 3.20 mL, 5.22 mmol) in THF (6.0 mL) was added 3-pentanone (0.5 mL, 4.73 mmol) at -78 °C. After the mixture was stirred at -78 °C for 40 min, a solution of (S)-2-methylbutyraldehyde 16 (493 mg, 5.72 mmol) in THF (3.0 mL) was added via cannula. The mixture was stirred at -78 °C for 1 h, diluted with saturated aqueous NH₄Cl (10 mL), and extracted with Et_2O (3 × 10 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (FL-60D, 50 g, hexane–Et₂O 10:1 \rightarrow 7:1) to give a diastereomeric mixture of aldol 16a (728 mg, 89%) as a colorless oil: $R_f = 0.63$, 0.38 (1:1 hexane/EtOAc); IR (CHCl₃): 3494, 1699, 1603, 1460, 1409, 1380 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 3.69 (ddd, J = 8.1, 5.9, 3.0 Hz, 0.51H), 3.61 (ddd, J = 6.7, 2.8, 2.8 Hz, 0.26H), 3.26 (ddd, J = 7.6, 5.9, 5.9 Hz, 0.23H), 2.88 (d, J = 3.0 Hz, 0.3H), 2.87–2.63 (m, 1.2H), 2.63–2.36 (m, 2H), 2.27 (d, J = 5.9 Hz, 0.5H), 1.88–1.65 (m, 0.6H), 1.65–1.38 (m, 1.7H), 1.38-1.20 (m, 0.7H), 1.20-1.10 (m, 6H), 0.96-0.75 (m, 6H); ¹³C NMR (67.8 MHz, CDCl₃): δ 217.1, 216.6, 216.5, 78.3, 75.7, 74.4, 48.6, 47.4, 47.0, 37.7, 36.8, 36.6, 36.2, 36.0, 34.8, 26.8, 24.9, 23.3, 16.1, 14.9, 14.1, 12.3, 11.8, 11.6, 10.9, 9.2, 7.7, 7.5, 7.5; HRMS (ESI): m/z 195.1359, calcd for $C_{10}H_{20}O_2Na [M + Na]^+$ 195.1361.

To a stirred solution of aldol 16a (322 mg, 1.87 mmol) and imidazole (389 mg, 5.71 mmol) in DMF (4.6 mL) was added triethylsilyl chloride (0.470 mL, 2.81 mmol) at room temperature. The mixture was stirred at room temperature for 75 min, diluted with H₂O (15 mL), and extracted with Et₂O (2 \times 10 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (15 g, hexane-Et₂O $200:1 \rightarrow 100:1 \rightarrow 20:1$) to give a diastereomeric mixture of C14–C20 segment 17 (507 mg, 95%) as a yellow oil: $R_f = 0.78$ (3:1 hexane/EtOAc); IR (CHCl₃): 1712, 1459, 1412, 1380 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 3.89 (dd, J = 8.5, 1.8 Hz, 0.21H), 3.88 (dd, J = 6.8, 1.9 Hz, 0.66H), 3.84 (dd, J = 8.9, 5.4 Hz, 0.13H), 2.86-2.65 (m, 1H), 2.65-2.35 (m, 2H), 1.55-1.30 (m, 2H), 1.30–1.10 (m, 1H), 1.10–0.80 (m, 21H), 0.65– 0.45 (m, 6H); 13 C NMR (67.8 MHz, CDCl₃): δ 214.4, 214.2, 214.0, 78.6, 77.5, 77.0, 49.9, 49.7, 49.3, 40.3, 38.3, 37.9, 37.1, 36.8, 35.4, 26.9, 24.7, 23.7, 15.8, 14.2, 14.0, 13.4, 12.7, 12.5, 12.4, 12.3, 7.8, 7.5, 7.4, 7.1, 7.1, 5.5, 5.4, 5.3; HRMS (ESI): m/z 309.2206, calcd for $C_{16}H_{34}O_2SiNa [M + Na]^+$ 309.2226.

Aldol 21. To a stirred solution of $Sn(OTf)_2$ (75.0 mg, 0.180 mmol) in CH₂Cl₂ (0.6 mL) were added triethylamine (27.0 µL, 0.200 mmol) and a solution of C14-C20 segment 17 (40.0 mg, 0.140 mmol) in CH₂Cl₂ (0.4 mL) at $-78 \degree$ C, and the mixture was stirred at -78 °C for 1.3 h. After a solution of C1-C13 segment 15 (21.9 mg, 0.042 mmol) in CH₂Cl₂ (0.4 mL) was added via cannula, the reaction mixture was stirred at -78 °C for 3 h and diluted with pH 6.8 phosphate buffer (5 mL). The mixture was filtered through a pad of Celite, and the Celite was rinsed with EtOAc ($4 \times 5 \text{ mL}$). The filtrate and rinses were combined, and the layers were separated. The aqueous layer was extracted with EtOAc $(3 \times 10 \text{ mL})$. The organic layer and extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (4 g, hexane-EtOAc $10:1 \rightarrow$ $3:1 \rightarrow 2:1$) to give a diastereometric mixture of aldol 21 (33.6 mg, 99%) as a colorless oil: $R_f = 0.73, 0.70, 0.64$ (1:1 hexane/ EtOAc); IR (CHCl₃): 3503, 1725, 1653, 1592, 1462, 1426, 1378 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 5.12 (d, J = 9.5 Hz, 1H), 4.38 (d, J = 8.4 Hz, 1H), 4.04 (d, J = 9.5 Hz, 1H), 3.93– 3.75 (m, 2H), 3.74–3.70 (m, 1H), 3.33–2.55 (m, 7H), 2.25–2.05 (m, 4H), 1.96 (s, 3H), 1.94 (s, 3H), 1.30-1.19 (m, 6H), 1.18-1.06 (m, 6H), 1.05-0.92 (m, 21H), 0.90-0.80 (m, 15H), 0.59 (q, J = 8.1 Hz, 6H), 0.02 (s, 3H), -0.38 (s, 3H), A signal dueto one proton (OH) was not observed; ¹³C NMR (67.8 MHz, CDCl₃): § 220.0, 219.1, 216.5, 179.7 (3C), 179.6, 172.7 (2C), 172.6, 165.3 (2C), 165.2, 164.4, 164.1, 163.4, 119.6, 119.2 (2C), 117.9 (2C), 79.2, 78.5, 78.1, 77.2, 76.5, 75.7, 74.3, 73.0 (2C), 71.8, 71.1, 70.2, 69.6, 51.2, 49.0, 48.9, 48.3, 48.0, 46.8, 43.6, 43.5 (2C), 41.3, 41.2, 40.0, 39.9, 38.6, 38.1, 38.0 (2C), 37.9, 35.6, 35.3 (2C), 35.0, 26.8, 26.2, 26.0, 25.5 (2C), 25.4, 24.9, 24.7, 24.4, 22.6, 18.3, 18.2 (2C), 16.4, 15.9, 15.6 (2C), 15.3, 15.1, 15.0, 14.9, 14.8, 14.2, 14.1, 13.8, 13.5 (2C), 12.6, 12.5, 12.3, 11.7, 11.1 (2C), 10.0, 9.9 (2C), 9.8, 9.6, 9.0, 8.7, 7.8, 7.1, 7.0, 5.5, 5.4 (3C), 5.2, -3.7, -3.9, -4.1, -4.3; HRMS (ESI): m/z 831.5604, calcd for C₄₅H₈₄O₈Si₂Na [M + Na]⁺ 831.5602.

Triketone 22. Aldol **21** (122 mg, 0.151 mmol) was treated with a 4:4:1 mixture of acetic acid, H₂O, and THF (4.5 mL) at

room temperature for 6.5 h. The mixture was poured into saturated aqueous NaHCO₃ (70 mL) at 0 °C and extracted with EtOAc $(3 \times 20 \text{ mL})$. The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (6 g, hexane-EtOAc $3:1 \rightarrow 2:1 \rightarrow 1:1$) to give a diastereomeric mixture of diol 21a (76.4 mg, 73%) as a colorless oil: $R_f = 0.74, 0.65, 0.58, 0.48$ (1:1 hexane/EtOAc); IR (CHCl₃): 3503, 1724, 1653, 1593, 1463, 1427, 1378 cm⁻¹; ¹H NMR $(270 \text{ MHz}, \text{ CDCl}_3)$: δ 5.12 (d, J = 9.1 Hz, 1H), 4.28 (d, J =8.4 Hz, 1H), 3.94 (d, J = 9.5 Hz, 1H), 3.70–3.55 (m, 1H), 3.20– 3.10 (m, 1H), 3.15–2.80 (m, 3H), 2.70–2.50 (m, 4H), 2.30–2.05 (m, 5H), 1.96 (s, 3H), 1.95 (s, 3H), 1.55-1.40 (m, 2H), 1.30-1.20 (m, 6H), 1.20-1.10 (m, 9H), 1.05-0.95 (m, 9H), 0.95-0.80 (m. 15H), 0.04 (s. 3H), -0.31 (s. 3H); 13 C NMR (67.8 MHz, CDCl₃): δ 220.7, 219.5, 179.7, 179.5, 173.1, 172.6, 165.0, 164.0, 163.9, 163.6, 119.5, 119.3, 119.2, 118.0, 117.9 (2C), 77.2, 76.4, 76.3 (2C), 74.7, 74.1, 73.1, 72.6, 70.4, 51.1, 49.1, 47.0, 45.7, 43.7, 43.6, 43.4, 41.2, 40.5, 37.9, 37.8, 37.3, 36.3, 36.1, 34.7, 26.8, 26.7, 26.0 (2C), 25.5 (2C), 25.4, 24.9, 24.7, 22.6, 22.5 (2C), 18.3, 18.2, 18.1 (2C), 15.4, 15.3, 15.2, 14.9, 13.6, 12.0 (2C), 11.9, 11.5, 11.2, 11.1, 10.0 (2C), 9.9, 9.8, 9.5, 9.3, 9.1, 8.7, 7.1, -3.9, -4.0, -4.2, -4.3, -4.5; HRMS (ESI): m/z 717.4735, calcd for C₃₉H₇₀O₈SiNa [M + Na]⁺ 717.4738.

To a stirred solution of diol 21a (76.4 mg, 0.110 mmol) in CH₂Cl₂ (3.0 mL) was added Dess-Martin periodinane (184 mg, 0.434 mmol) at room temperature. The mixture was stirred at room temperature for 20 min: diluted with a 1:1:2 mixture of saturated aqueous NaHCO₃, saturated aqueous Na₂S₂O₃, and H₂O (4 mL); and extracted with Et₂O (2 \times 10 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (6g, hexane-EtOAc $5:1 \rightarrow 3:1 \rightarrow 1:1$) to give a keto-enol equilibrium mixture of triketone 22 (63.7 mg, 83%) as a colorless amorphous solid: $R_f = 0.61$ (1:1 hexane/EtOAc); mp 26–28 °C; $[\alpha]_D^{25}$ +16.9 (c 0.586, CHCl₃); IR (CHCl₃): 1724, 1653, 1593, 1463, 1378 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 16.93 (s, 0.06H), 16.80 (s, 0.04H), 16.66 (s, 0.10H), 5.50-5.39 (m, 0.3H), 5.32-5.22 (m, 0.4H), 5.20–5.08 (m, 0.3H), 4.15–3.66 (m, 2H), 3.50– 3.30 (m, 0.5H), 3.25-3.00 (m, 1.5H), 2.80-2.50 (m, 2.8H), 2.25-1.80 (m, 13H), 1.40-0.70 (m, 39H), 0.10-0.05 (m, 3H). $\{(-0.10)-(-0.32)\}$ (m, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 210.1, 207.9, 206.2, 203.9, 202.8, 198.9, 198.4, 197.6, 195.0, 193.6, 193.5, 191.7, 191.4, 190.2, 189.5, 179.6, 179.5 (2C), 174.1, 172.3 (2C), 172.1, 172.0 (2C), 171.8, 171.7, 166.2, 165.7, 164.7, 164.6, 164.5 (2C), 164.4, 164.2 (2C), 164.0 (2C), 163.9, 128.1, 119.6 (2C), 119.5 (2C), 119.4 (2C), 117.8 (3C), 117.7, 109.5, 108.4, 105.8, 105.4, 105.0, 104.4, 104.2, 103.8, 103.5, 75.5, 75.4, 74.8, 74.6, 74.5, 74.3, 74.1, 73.9, 73.8, 73.6, 73.5, 58.2, 58.1, 57.7, 57.1, 53.2, 52.6, 51.8, 50.5, 47.3, 47.2, 46.9, 46.6, 46.5, 46.2, 46.0, 45.9 (2C), 45.8, 44.7, 43.4, 43.3 (2C), 43.2, 41.9, 40.3, 40.2 (2C), 40.1, 39.9, 39.8, 39.7, 39.5, 39.4, 39.3, 38.7, 38.6, 38.4, 38.3, 26.8, 26.7, 26.1, 25.9, 25.7 (2C), 25.4, 25.3 (2C), 25.0, 24.9, 22.5, 22.4, 20.9, 18.2, 18.1, 18.0, 16.9, 16.3, 16.0, 15.9, 15.8, 15.6, 15.4, 15.2, 14.4, 14.2, 14.1, 14.0, 13.9, 13.8, 13.5, 13.3, 13.2 (2C), 13.0, 12.5, 12.2 (2C), 12.1, 11.7, 11.6, 10.7, 10.4, 10.3,

10.0, 9.5, 9.3, 9.0, 7.6, -3.7, -3.9, -4.0, -4.2, -4.7; HRMS (ESI): m/z 713.4419, calcd for $C_{39}H_{66}O_8SiNa$ [M + Na]⁺ 713.4425.

Auripyrone A (1). A solution of triketone 22 (32.5 mg, 47.0 µmol) in a 5:3:7 mixture of HF pyridine, pyridine, and THF (5.0 mL) was stirred at 60 °C for 14.3 h. The mixture was poured into saturated aqueous NaHCO₃ (150 mL) at 0 °C and extracted with EtOAc $(3 \times 20 \text{ mL})$. The combined extracts were washed with brine, dried over Na2SO4, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (5 g, hexane-EtOAc $5:1 \rightarrow 2:1 \rightarrow 1:1$) and by preparative HPLC [Develosil ODS-HG-5 (250×20 mm), flow rate 5 mL min⁻¹; detection, UV 215 nm, solvent 80% aqueous MeOH] to give auripyrone A (1) (5.9 mg, 22%) as a colorless solid: $R_f = 0.37$ (1:1 CH₂Cl₂/Et₂O); $[\alpha]_D^{25} + 47.3$ (c 0.390, CHCl₃); IR (CHCl₃): 1725, 1655, 1623, 1598, 1462, 1387 cm⁻¹; ¹H NMR (600 MHz, C₆D₆): δ 4.92 (dd, J = 3.3, 3.3 Hz, 1H), 3.96 (dd, J = 10.3, 2.1 Hz, 1H), 2.75 (dg, J = 10.3, 2.1 Hz, 1H)J = 10.3, 7.0 Hz, 1H), 2.32 (q, J = 7.0 Hz, 1H), 2.31 (dq, J =15.0, 7.5 Hz, 1H), 2.25 (ddq, J = 7.5, 7.5, 3.8 Hz, 1H), 2.21– 2.16 (m, 1H), 2.16–2.11 (m, 2H), 2.09 (dg, J = 15.0, 7.5 Hz, 1H), 2.03 (s, 3H), 1.98 (s, 3H), 1.85–1.80 (m, 2H), 1.64 (s, 3H), 1.56–1.48 (m, 1H), 1.40–1.33 (m, 1H), 1.12 (d, J = 7.0 Hz, 3H), 0.99 (d, J = 6.8 Hz, 3H), 0.91 (d, J = 6.2 Hz, 3H), 0.90 (d, J = 6.5 Hz, 3H), 0.89 (t, J = 7.6 Hz, 3H), 0.83 (t, J = 7.5 Hz, 3H), 0.77 (d, J = 7.0 Hz, 3H), 0.73 (d, J = 7.0 Hz, 3H), 0.63 (d, J = 7.1 Hz, 3H); ¹³C NMR (150 MHz, C₆D₆): δ 191.8, 178.5, 172.0, 166.2, 162.1, 161.5, 121.3, 118.1, 107.8, 105.0, 75.2, 70.5, 44.5, 44.0, 37.2, 36.6, 34.1, 31.7, 26.4, 26.3, 24.7, 22.5, 22.3, 16.0, 12.1, 12.0, 11.9, 11.0, 10.8, 9.7, 9.5, 9.0, 8.1; HRMS (ESI): m/z 581.3446, calcd for C₃₃H₅₀O₇Na [M + Na]⁺ 581.3454.

Bis(pyrone) Compound 25. To a stirred solution of auripyrone A (1) (2.5 mg, 4.5 µmol) in MeOH (2 mL) was added 1 M aqueous LiOH (2 mL) at room temperature. The mixture was stirred at room temperature for 3.5 h, diluted with saturated aqueous NH₄Cl (5 mL) at 0 °C, and extracted with EtOAc (3 \times 20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (0.6 g, hexane-EtOAc $1:1 \rightarrow 0:1$) to give bis(pyrone) compound 25 (1.7 mg, 81%) as colorless crystals: $R_f = 0.38$ (1:1 hexane/acetone); $[\alpha]_{D}^{25}$ +21.7 (c 0.138, CHCl₃); IR (CHCl₃): 3422, 1653, 1593, 1460, 1426, 1381 cm⁻¹; ¹H NMR (600 MHz, CDCl₃): δ 4.01 (dd, J = 13.8, 9.7 Hz, 2H), 3.23– 3.16 (m, 2H), 3.02 (br s, 1H), 2.96–2.85 (m, 2H), 2.68 (dq, J = 15.2, 7.6 Hz, 1H), 2.62 (dq, J = 15.2, 7.6 Hz, 1H), 2.00– 1.90 (m, 1H), 2.00 (s, 3H), 1.99 (s, 3H), 1.96 (s, 3H), 1.94 (s, 3H), 1.70–1.55 (m, 2H), 1.25 (t, J = 7.6 Hz, 3H), 1.23 (d, J = 7.6 Hz, 3H), 1.19 (d, J = 7.0 Hz, 3H), 1.18 (d, J = 6.8 Hz, 3H), 1.05 (d, J = 6.8 Hz, 3H), 0.90 (t, J = 7.3 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 179.7, 179.6, 165.9, 164.2, 163.5, 163.4, 120.1, 119.8, 118.3, 118.1, 79.0, 78.8, 39.3 (2C), 37.0, 34.5, 27.7, 24.8, 17.9, 14.6, 14.4, 11.9, 11.4, 9.7 (2C), 9.5 (2C), 3.8; HRMS (ESI): m/z 497.2882, calcd for C₂₈H₄₂O₆Na $[M + Na]^+$ 497.2879.

Ester 29. To a stirred solution of homoallylic alcohol 14 (94.2 mg, 0.216 mmol), dimethylaminopyridine (1.00 g, 8.19 mmol), (*S*)-2-methylbutanoic acid (28) (0.234 mL, 2.15 mmol),

and triethylamine (0.602 mL, 4.53 mmol) in toluene (18 mL) was added 2,4,6-trichlorobenzoyl chloride (0.742 mL, 4.75 mmol) at -78 °C. The mixture was stirred at 0 °C for 1.3 h, diluted with saturated aqueous NaHCO₃ (15 mL) at 0 °C, and extracted with EtOAc $(3 \times 10 \text{ mL})$. The combined extracts were washed with brine, dried over Na2SO4, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10 g, hexane-EtOAc $5:1 \rightarrow 3:1$) to give ester **29** (104 mg, 93%) as a colorless oil: $R_f = 0.77$ (1:1 hexane/EtOAc); $[\alpha]_{D}^{25}$ +21.5 (c 1.316, CHCl₃); IR (CHCl₃): 1724, 1654, 1594, 1462, 1426, 1378 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 5.78 (ddd, J = 17.3, 10.0, 7.8 Hz, 1H), 5.12–4.99 (m, 3H), 4.08 (d, J = 8.6 Hz, 1H), 3.17 (dq, J = 8.6, 7.3 Hz, 1H), 2.72–2.52 (m, 3H), 2.45–2.30 (m, 1H), 2.00–1.92 (m, 1H), 1.97 (s, 3H), 1.96 (s, 3H), 1.73 (ddg, J = 14.0, 7.0, 7.0 Hz, 1H), 1.48 (ddg, J = 14.0, 7.0, 7.0 Hz, 1H), 1.26 (t, J = 7.6 Hz, 3H), 1.20 (d, J = 7.3 Hz, 3H), 1.09 (d, J = 7.3 Hz, 3H), 0.98 (d, J = 6.8 Hz, 3H), 0.94 (t, J = 7.6 Hz, 3H), 0.89 (d, J = 6.8 Hz, 3H), 0.83 (s, 9H), 0.07 (s, 3H), -0.33 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 179.6, 176.0, 164.4, 163.5, 138.7, 119.4, 118.0, 116.0, 76.2, 72.6, 41.6, 40.6, 40.3, 37.9, 26.6, 26.0 (3C), 24.8, 18.2, 18.1, 17.2, 15.5, 11.9, 11.4, 9.9, 9.5, 9.3, -3.8, -4.4; HRMS (ESI): m/z 543.3485, calcd for C₃₀H₅₂O₅SiNa $[M + Na]^+$ 543.3482.

C1-C13 Segment 30. To a stirred solution of ester 29 (65.8 mg, 0.126 mmol) in acetone (1 mL) and H₂O (1 mL) were added N-methylmorpholine oxide (55.0 mg, 0.469 mmol) and osmium tetroxide (0.1 M solution in tert-butanol, 0.300 mL, 30.0 umol) at room temperature. The mixture was stirred at room temperature for 14 h, diluted with saturated aqueous NaHSO₃ (10 mL) at 0 °C, and extracted with EtOAc (3 \times 10 mL). The combined extracts were washed with saturated aqueous NaHSO3 and brine, dried over Na2SO4, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (7 g, hexane–EtOAc $1:1 \rightarrow 0:1$) to give a diastereomeric mixture of diol 29a (ca. 6:1) (65.5 mg, 94%) as a colorless oil: $R_f = 0.18$ (major), 0.10 (minor) (1:1 hexane/ EtOAc); IR (CHCl₃): 3424, 1724, 1651, 1591, 1462, 1380 cm⁻¹; ¹HNMR (270 MHz, CDCl₃) for major isomer: δ 5.07 (dd, J = 8.6, 3.5 Hz, 1H), 4.10 (dd, J = 8.1, 1.9 Hz, 1H), 3.80-3.62 (m, 2H), 3.50–3.38 (m, 1H), 3.24 (dq, J = 8.1, 7.3 Hz, 1H), 2.64 (dq, J = 7.6, 3.0 Hz, 2H), 2.63–2.55 (m, 1H), 2.43– 2.05 (m, 4H), 1.99 (s, 3H), 1.95 (s, 3H), 1.72 (ddq, J = 14.0, 7.0, 7.0 Hz, 1H), 1.47 (ddq, J = 14.0, 7.0, 7.0 Hz, 1H), 1.24 (t, J = 7.3 Hz, 3H), 1.19 (d, J = 7.3 Hz, 3H), 1.18 (d, J = 7.0 Hz, 3H), 0.93 (t, J = 7.0 Hz, 3H), 0.91 (d, J = 6.8 Hz, 3H), 0.85 (s, 9H), 0.82 (d, J = 7.3 Hz, 3H), -0.18 (s, 3H), -0.24 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 179.9, 176.1, 164.8, 164.4, 119.5, 118.0, 76.9, 73.3, 71.9, 64.4, 41.6, 40.9, 38.0, 37.6, 26.6, 26.0 (3C), 24.8, 18.2, 17.1, 15.1, 12.9, 11.9, 11.4, 10.2, 10.0, 9.6, -3.9, -4.3; HRMS (ESI): m/z 577.3558, calcd for $C_{30}H_{54}O_7SiNa [M + Na]^+ 577.3537.$

To a stirred solution of diol **29a** (88.2 mg, 0.159 mmol) in acetone (1 mL) and H₂O (1 mL) was added NaIO₄ (73.2 mg, 0.277 mmol). The mixture was stirred at room temperature for 15 min, diluted with half saturated brine (5 mL), and extracted with EtOAc (3×10 mL). The combined extracts were washed with saturated aqueous NaHSO₃ and brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified

by column chromatography on silica gel (8 g, hexane-EtOAc $3:1 \rightarrow 2:1 \rightarrow 1:1$) to give C1–C13 segment **30** (68.0 mg, 82%) as a colorless oil: $R_f = 0.73$ (1:1 hexane/EtOAc); $[\alpha]_D^{25} + 16.2$ (c 1.000, CHCl₃); IR (CHCl₃): 1725, 1654, 1594, 1462, 1426, 1378 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 9.69 (d, J = 3.2 Hz, 1H), 5.26 (dd, J = 9.5, 3.5 Hz, 1H), 4.07 (dd, J = 8.4, 2.2 Hz, 1H), 3.20 (dq, J = 8.4, 7.0 Hz, 1H), 2.80–2.30 (m, 4H), 2.17– 2.06 (m, 1H), 1.98 (s, 3H), 1.95 (s, 3H), 1.71 (ddg, J = 14.4, 7.2, 7.2 Hz, 1H), 1.47 (ddq, J = 14.4, 7.2, 7.2 Hz, 1H), 1.26 (t, J = 7.6 Hz, 3H), 1.19 (d, J = 7.3 Hz, 3H), 1.15 (d, J = 8.1 Hz, 3H), 1.12 (d, J = 6.5 Hz, 3H), 0.96 (d, J = 7.0 Hz, 3H), 0.94 (t, J = 7.3 Hz, 3H), 0.82 (s, 9H), 0.04 (s, 3H), -0.32 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 202.7, 179.5, 175.9, 164.0, 163.8, 119.6, 118.1, 75.0, 73.6, 47.7, 41.4, 40.4, 38.8, 26.5, 26.0 (3C), 24.8, 18.2, 17.1, 15.5, 11.8, 11.7, 11.4, 10.0, 9.9, 9.6, -3.9, -4.5; HRMS (ESI): m/z 523.3432, calcd for $C_{29}H_{50}O_6Si [M + H]^+ 523.3455.$

Aldol 31. To a stirred solution of Sn(OTf)₂ (220 mg, 0.528 mmol) in CH₂Cl₂ (1.4 mL) were added triethylamine (0.0700 mL, 0.527 mmol) and a solution of C14-C20 segment 17 (110 mg, 0.384 mmol) in CH_2Cl_2 (0.7 mL) at -78 °C. After the mixture was stirred at -78 °C for 1.3 h, a solution of aldehyde 30 (68.0 mg, 0.130 mmol) in CH₂Cl₂ (0.7 mL) was added via cannula. The reaction mixture was stirred at -78 °C for 2.5 h, diluted with pH 6.8 phosphate buffer (10 mL), and filtrated through a pad of Celite. This Celite was rinsed with EtOAc ($4 \times 5 \text{ mL}$). The filtrate and rinses were combined, and the layers were separated. The aqueous layer was extracted with EtOAc ($3 \times 10 \text{ mL}$). The organic layer and extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10 g, hexane-EtOAc $10:1 \rightarrow 3:1 \rightarrow 2:1$) to give a diastereomeric mixture of aldol 31 (104 mg, 99%) as a colorless oil: $R_f = 0.83$, 0.77 (1:1 hexane/EtOAc); IR (CHCl₃): 3509, 1723, 1653, 1592, 1461, 1426, 1379 cm⁻¹; ¹HNMR (270 MHz, CDCl₃): δ 5.11 (d, J = 10.3 Hz, 1H), 4.39 (d, J =8.6 Hz, 1H), 4.06 (d, J = 9.5 Hz, 1H), 4.02–3.80 (m, 1H), 3.76– 3.65 (m, 1H), 3.20-3.05 (m, 1H), 2.95-2.25 (m, 6H), 2.13-2.06 (m, 1H), 1.95 (s, 3H), 1.93 (s, 3H), 1.80-1.60 (m, 1H), 1.52-1.33 (m, 2H), 1.30-1.00 (m, 15H), 1.00-0.85 (m, 18H), 0.85-0.70 (m, 15H), 0.60 (q, J = 7.8 Hz, 6 H), 0.55-0.48 (m, 2H),0.01 (s, 3H), -0.39 (s, 3H); 13 C NMR (67.8 MHz, CDCl₃): δ 220.0, 179.7, 179.5, 176.1, 175.8, 165.2, 163.9, 163.8, 163.4, 119.5, 119.1, 118.0, 117.9, 79.0, 78.1, 77.2, 75.7, 75.6, 72.9, 70.1, 60.3, 48.9, 46.9, 41.6, 41.4, 41.3, 40.3, 40.0, 38.7, 37.8, 35.4, 26.8, 26.6, 26.5, 26.0, 25.9, 24.8, 24.7, 24.4, 21.0, 18.3, 18.1, 17.2, 16.4, 15.6, 15.5, 15.0, 14.2, 12.2, 11.9, 11.7, 11.4, 11.1, 10.0, 9.9, 9.8, 9.5, 8.6, 8.2, 7.1, 7.0, 5.5, 5.4, 5.3, 5.2, -3.9, -4.3, -4.5; HRMS (ESI): m/z 831.5596, calcd for $C_{45}H_{84}O_8Si_2Na [M + Na]^+ 831.5602.$

Triketone 32. Aldol **31** (59.6 mg, 73.6 µmol) was treated with a 4:4:1 mixture of acetic acid, H₂O, and THF (4.5 mL) at room temperature for 13.3 h. The mixture was poured into saturated aqueous NaHCO₃ (70 mL) at 0 °C and extracted with EtOAc (3 × 20 mL). The combined extracts were washed with saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10 g, hexane–EtOAc $3:1 \rightarrow 2:1 \rightarrow 1:1$) to give a diastereomeric mixture of diol **31a** (45.9 mg, 90%) as a colorless oil: $R_f = 0.67, 0.60, 0.50$ (1:1 hexane/EtOAc); IR (CHCl₃): 3523, 1723, 1652, 1592, 1462, 1427, 1379 cm^{-1} ; ¹H NMR (270 MHz, CDCl₃): δ 5.10 (d, J = 9.5 Hz, 1H), 4.32–4.24 (m, 1H), 3.95 (d, J = 8.9 Hz, 1H), 3.70-3.55 (m, 1H), 3.25-3.10 (m, 1H), 3.08-2.75 (m, 2H), 2.75-2.50 (m, 4H), 2.50-2.28 (m, 1H), 1.95 (s, 3H), 1.93 (s, 3H), 1.80-1.62 (m, 2H), 1.58-1.35 (m, 2H), 1.30-1.05 (m, 15H), 1.05–0.75 (m, 27H), 0.03 (s, 3H), -0.33 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 227.9, 220.8, 179.7, 176.1 (2C), 165.1, 165.0, 164.9, 163.6 (3C), 119.6, 119.2, 117.9, 117.7, 78.1, 77.2, 74.7, 74.3, 73.0, 70.2, 52.6, 50.2, 47.1, 46.9, 45.7, 45.5, 45.3, 44.7, 43.8, 41.6, 41.2, 41.1, 37.9, 37.3, 37.0, 36.4 (2C), 34.6, 33.7, 26.6, 26.1, 25.9, 25.0, 24.8, 18.3 (2C), 18.2, 17.2, 15.5, 15.4, 15.2, 15.0, 14.8, 12.5, 11.9, 11.2, 11.1 (2C), 10.0, 9.9 (2C), 9.6, 9.1, 8.7, -3.9, -4.2, -5.3; HRMS (ESI): m/z 717.4725, calcd for C₃₉H₇₀O₈SiNa [M + Na]⁺ 717.4738.

To a stirred solution of diol 31a (34.0 mg, 48.9 µmol) in CH₂Cl₂ (1.5 mL) was added Dess-Martin periodinane (86.6 mg, 204 µmol) at room temperature. The mixture was stirred at room temperature for 25 min; diluted with a 1:1:2 mixture of saturated aqueous NaHCO₃, saturated aqueous Na₂S₂O₃, and H_2O (4 mL); and extracted with Et₂O (2 × 10 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10g, hexane-EtOAc $5:1 \rightarrow 3:1 \rightarrow 1:1$) to give a keto-enol equilibrium mixture of triketone 32 (32.1 mg, 95%) as a colorless amorphous solid: $R_f = 0.63$ (1:1 hexane/EtOAc); $[\alpha]_D^{25} + 15.9$ (c 1.325, CHCl₃); IR (CHCl₃): 1724, 1652, 1593, 1462, 1427, 1379 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 16.93 (s, 0.06H), 16.81 (s, 0.04H), 16.66 (s, 0.10H), 5.52-5.36 (m, 0.3H), 5.35-5.21 (m, 0.3H), 5.20-5.06 (m, 0.4H), 4.15-3.60 (m, 1.8H), 3.58-3.00 (m, 2H), 2.80-2.45 (m, 3H), 2.40-2.18 (m, 1H), 2.20-1.55 (m, 12H), 1.55–1.14 (m, 11H), 1.14–0.75 (m, 28H), {(0.10)– (-0.06)} (m, 3H), {(-0.10)-(-0.35)} (m, 3H); ¹³C NMR (67.8) MHz, CDCl₃): δ 210.1, 206.1, 204.0, 203.9, 198.6, 197.6, 195.1, 194.0, 193.7, 191.4 (2C), 179.7, 179.6, 179.5, 177.8 (2C), 176.0, 175.9, 175.7 (2C), 175.6, 164.9, 164.8, 164.7 (2C), 164.6 (2C), 164.5 (2C), 164.4, 164.3 (2C), 164.2, 164.1, 164.0, 119.7, 119.6, 119.5 (2C), 119.4, 117.8 (3C), 109.4, 108.2, 106.0, 105.6, 105.1, 104.3, 104.0, 103.8, 103.5, 75.5, 75.4, 74.7, 74.4, 73.9, 73.7, 73.1, 57.6, 52.7, 50.4, 47.2, 46.8, 46.6, 46.2, 46.1, 46.0, 45.9, 44.8, 44.7, 42.0, 41.5, 41.4, 41.3 (2C), 41.2, 41.1, 40.3, 40.0, 39.8, 39.6 (2C), 39.5, 39.4 (2C), 39.3, 39.1, 38.7, 38.6, 38.5 (2C), 38.2, 26.9, 26.7 (2C), 26.5 (2C), 26.1, 26.0, 25.6, 24.9, 21.0, 18.4, 18.3, 18.2, 18.1, 16.9, 16.8, 16.4, 16.3, 16.1, 16.0, 14.6, 14.2, 14.0, 13.9, 13.8, 13.3, 13.2, 13.1, 13.0, 12.6, 12.2, 12.0, 11.9, 11.8 (2C), 11.6 (2C), 10.7, 10.5 (2C), 10.1 (2C), 10.0, 9.6, 9.3, 7.7, -3.5, -3.7, -3.8, -4.0, -4.4; HRMS (ESI): m/z 713.4432, calcd for $C_{39}H_{66}O_8SiNa [M + Na]^+ 713.4425.$

(2'S)-Auripyrone B (Natural-Type Auripyrone B) (33). A solution of triketone 32 (26.5 mg, 38.3μ mol) in a 5:3:7 mixture of HF•pyridine, pyridine, and THF (10.0 mL) was stirred at 60 °C for 17 h. The mixture was poured into saturated aqueous NaHCO₃ (150 mL) at 0 °C and extracted with EtOAc (3 × 20 mL). The combined extracts were washed with saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄, filtered,

and concentrated. The residual oil was purified by column chromatography on silica gel (6 g, hexane-EtOAc $5:1 \rightarrow$ $2:1 \rightarrow 1:1$) and by preparative HPLC [Develosil ODS-HG-5 $(250 \times 20 \text{ mm})$, flow rate 5 mL min⁻¹; detection, UV 215 nm, solvent 80% aqueous MeOH] to give (2'S)-auripyrone B (natural-type auripyrone B) (33) (3.6 mg, 17%) as colorless crystals: $R_f = 0.33$ (1:1 CH₂Cl₂/Et₂O); $[\alpha]_D^{25}$ +43.4 (c 0.293, CHCl₃); IR (CHCl₃): 1726, 1655, 1623, 1597, 1462, 1387 cm⁻¹; ¹H NMR (600 MHz, C₆D₆): δ 4.91 (dd, J = 3.4, 3.4 Hz, 1H), 3.97 (dd, J = 10.3, 2.1 Hz, 1H), 2.74 (dq, J = 10.3, 7.0 Hz, 1H), 2.43 (ddg, J = 7.0, 7.0, 7.0 Hz, 1H), 2.32 (g, J = 7.0 Hz, 1H), 2.32–2.24 (m, 2H), 2.09 (dq, J = 15.0, 7.5 Hz, 1H), 2.03 (s, 3H), 1.98 (s, 3H), 1.88-1.70 (m, 3H), 1.66 (s, 3H), 1.58–1.45 (m, 1H), 1.43–1.38 (m, 1H), 1.37–1.30 (m, 1H), 1.16 (d, J = 7.0 Hz, 3H), 1.13 (d, J = 7.0 Hz, 3H), 0.99 (d, J = 6.8 Hz, 3H), 0.89 (t, J = 7.5 Hz, 3H), 0.87 (t, J = 7.6 Hz, 3H), 0.83 (t, J = 7.5 Hz, 3H), 0.77 (d, J = 7.0 Hz, 3H), 0.72 (d, J = 7.0 Hz, 3H), 0.63 (d, J = 7.1 Hz, 3H); ¹³C NMR (150 MHz, C₆D₆): δ 191.8, 178.5, 175.9, 166.2, 162.0, 161.5, 121.3, 118.1, 107.8, 105.1, 75.2, 70.6, 44.5, 41.6, 37.2, 36.6, 34.1, 31.7, 27.4, 26.4, 24.7, 16.9, 15.9, 12.1, 12.0, 11.8, 11.8, 10.8, 10.7, 9.7, 9.4, 9.0, 8.2; HRMS (ESI): m/z 581.3472, calcd for $C_{33}H_{50}O_7Na [M + Na]^+ 581.3454.$

Ester 35. To a stirred solution of homoallylic alcohol 14 (149.5 mg, 0.342 mmol), dimethylaminopyridine (660 mg, 5.40 mmol), (R)-2-methylbutanoic acid (34) (0.123 mL, 1.13 mmol), and triethylamine (0.317 mL, 2.39 mmol) in toluene (9 mL) was added 2,4,6-trichlorobenzoyl chloride (0.374 mL, 2.39 mmol) at -78 °C. The mixture was stirred at 0 °C for 2.5 h, diluted with saturated aqueous NaHCO₃ (15 mL) at 0 °C, and extracted with EtOAc ($3 \times 10 \text{ mL}$). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10 g, hexane–EtOAc 5:1 \rightarrow 3:1) to give ester 35 (182 mg, quant) as a colorless oil: $R_f = 0.77$ (1:1 hexane/EtOAc); $[\alpha]_D^{25}$ +14.7° (c 1.228, CHCl₃); IR (CHCl₃): 1724, 1654, 1594, 1462, 1426, 1378 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 5.73 (ddd, J = 17.0, 10.0, 7.0 Hz, 1H, 5.10–4.97 (m, 3H), 4.08 (d, J = 8.4 Hz, 1H), 3.16 (dq, J = 8.1, 7.6 Hz, 1H), 2.72–2.52 (m, 3H), 2.40 (q, J = 6.8 Hz, 1H), 2.00–1.90 (m, 1H), 1.96 (s, 3H), 1.94 (s, 3H), 1.75 (ddg, J = 14.0, 7.0, 7.0 Hz, 1H), 1.46 (ddg, J = 14.0, 7.0, 7.0 Hz, 1H), 1.25 (t, J = 7.6 Hz, 3H), 1.21 (d, J = 7.6 Hz, 3H), 1.08 (d, J = 7.0 Hz, 3H), 0.97 (d, J = 6.5 Hz, 3H), 0.94 (t, J = 7.3 Hz, 3H), 0.88 (d, J = 6.8 Hz, 3H), 0.82 (s, 9H), 0.06 (s, 3H), -0.35 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 179.6, 176.0, 164.4, 163.5, 138.7, 119.5, 118.1, 116.0, 76.3, 72.6, 41.6, 40.7, 40.2, 38.0, 26.8, 26.0 (3C), 24.8, 18.2, 18.1, 16.9, 15.5, 11.9, 11.4, 9.9, 9.5, 9.2, -3.8, -4.4; HRMS (ESI): m/z 543.3471, calcd for C₃₀H₅₂O₅SiNa $[M + Na]^+$ 543.3482.

C1–C13 Segment 35b. To a stirred solution of ester 35 (93.5 mg, 0.180 mmol) in acetone (1 mL) and H₂O (1 mL) were added *N*-methylmorpholine oxide (70.0 mg, 0.598 mmol) and osmium tetroxide (0.1 M solution in *tert*-butanol, 0.400 mL, 40.0 μ mol) at room temperature. The mixture was stirred at room temperature for 7 h, diluted with saturated aqueous NaHSO₃ (15 mL) at 0 °C, and extracted with EtOAc (3 × 20 mL). The combined extracts were washed with saturated aqueous NaHSO₃ and brine, dried over Na₂SO₄, filtered, and

concentrated. The residual oil was purified by column chromatography on silica gel (7 g, hexane–EtOAc $1:1 \rightarrow 0:1$) to give a diastereomeric mixture of diol 35a (ca. 9:1) (84.5 mg, 85%) as a colorless oil: $R_f = 0.18$ (major), 0.10 (minor) (1:1 hexane/ EtOAc): IR (CHCl₂): 3422, 1724, 1651, 1591, 1462, 1380 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 5.05 (dd, J = 8.9, 3.5 Hz, 1H), 4.09 (dd, J = 9.7, 4.0 Hz, 1H), 3.75–3.64 (m, 2H), 3.47– 3.38 (m, 1H), 3.23 (dq, J = 7.6, 7.6 Hz, 1H), 2.64 (dq, J = 7.8, 3.5 Hz, 2H), 2.50-2.25 (m, 2H), 2.20-2.05 (m, 1H), 1.99 (s, 3H), 1.95 (s, 3H), 1.75 (ddg, J = 14.0, 7.0, 7.0 Hz, 1H), 1.45 (ddg, J = 14.0, 7.0, 7.0 Hz, 1H), 1.20 (d, J = 7.0 Hz, 3H), 1.17(d, J = 8.1 Hz, 3H), 1.14 (d, J = 7.0 Hz, 3H), 0.89 (d, J =7.3 Hz, 3H), 0.87 (d, J = 6.8 Hz, 3H), 0.81 (s, 9H), 0.80 (d, J = 8.4 Hz, 3H), 0.04 (s, 3H), -0.28 (s, 3H), signals due to two protons (OH) were not observed: ¹³C NMR (67.8 MHz. CDCl₃): δ 180.0, 176.1, 164.9, 164.4, 119.5, 118.0, 76.9, 73.3, 71.9, 64.4, 41.5, 40.9, 38.0, 37.5, 26.7, 26.0 (3C), 24.8, 18.2, 16.9, 15.0, 13.0, 11.8, 11.3, 10.1, 9.9, 9.6, -3.9, -4.3; HRMS (ESI): m/z 555.3722, calcd for C₃₀H₅₅O₇Si [M + H]⁺ 555.3717.

To a stirred solution of diol 35a (144 mg, 0.264 mmol) in acetone (1.5 mL) and H₂O (1.5 mL) was added NaIO₄ (120 mg, 0.445 mmol). The mixture was stirred at room temperature for 10 min, diluted with half saturated brine (5 mL), and extracted with EtOAc $(3 \times 10 \text{ mL})$. The combined extracts were dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (8 g, hexane-EtOAc $3:1 \rightarrow 2:1 \rightarrow 1:1$) to give aldehyde **35b** (96.8 mg, 70%) as a colorless oil: $R_f = 0.73$ (1:1 hexane/EtOAc); $[\alpha]_D^{25} + 11.0$ (c 1.000, CHCl₃); IR (CHCl₃): 1726, 1654, 1594, 1462, 1426, 1378 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 9.68 (d, J = 3.5 Hz, 1H), 5.26 (dd, J = 9.1, 5.8 Hz, 1H), 4.07 (dd, J = 8.4, 1.0 Hz, 1H), 3.19 (dq, J = 8.4, 7.0 Hz, 1H), 2.80–2.35 (m, 4H), 2.18– 2.05 (m, 1H), 1.97 (s, 3H), 1.95 (s, 3H), 1.72 (ddq, J = 14.4, 7.2, 7.2 Hz, 1H), 1.49 (ddg, J = 14.4, 7.2, 7.2 Hz, 1H), 1.26 (t, J = 7.6 Hz, 3H), 1.18 (d, J = 7.3 Hz, 3H), 1.14 (d, J = 7.0 Hz, 3H), 1.12 (d, J = 6.5 Hz, 3H), 0.95 (d, J = 7.6 Hz, 3H), 0.94 (t, J = 7.3 Hz, 3H), 0.82 (s, 9H), 0.04 (s, 3H), -0.32 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 202.7, 179.5, 175.8, 163.9, 163.8, 119.5, 118.1, 75.0, 73.3, 47.6, 41.3, 40.4, 38.9, 26.7, 26.0 (3C), 24.8 (2C), 18.2, 16.7, 15.4, 11.8, 11.4, 10.0, 9.9, 9.5, -3.9, -4.5; HRMS (ESI): m/z 523.3436, calcd for C₂₉H₅₁O₆Si $[M + H]^+$ 523.3455.

To a stirred solution of Sn(OTf)₂ (320 mg, Aldol 35c. 0.768 mmol) in CH₂Cl₂ (2 mL) were added triethylamine (0.100 mL, 0.753 mmol) and a solution of C14-C20 segment 17 (162 mg, 0.565 mmol) in CH₂Cl₂ (1 mL) at -78 °C. After the mixture was stirred at -78 °C for 1 h, a solution of aldehyde 35b (96.8 mg, 0.185 mmol) in CH₂Cl₂ (1 mL) was added via cannula. The reaction mixture was stirred at -78 °C for 3.5 h, diluted with pH 6.8 phosphate buffer (15 mL), and filtered through a pad of Celite. This Celite was rinsed with EtOAc $(4 \times 5 \text{ mL})$. The filtrate and rinses were combined, and the layers were separated. The aqueous layer was extracted with EtOAc $(3 \times 10 \text{ mL})$. The organic layer and extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10 g, hexane-EtOAc $10:1 \rightarrow 3:1 \rightarrow 2:1$) to give a diastereomeric mixture of aldol 35c (139 mg, 93%) as a

colorless oil: $R_f = 0.87$, 0.81 (1:1 hexane/EtOAc); IR (CHCl₃): $3507, 1723, 1694, 1653, 1592, 1462, 1426, 1379 \,\mathrm{cm}^{-1};$ ¹H NMR (270 MHz, CDCl₃): δ 5.12 (d, J = 10.0 Hz, 1H), 4.41 (d, J = 8.4 Hz, 1H), 4.06 (d, J = 9.5 Hz, 1H), 4.02–3.82 (m, 1H), 3.75–3.70 (m, 1H), 3.13 (t, J = 7.6 Hz, 1H), 3.00–2.25 (m, 5H), 2.12-2.05 (m, 1H), 1.96 (s, 3H), 1.94 (s, 3H), 1.82-1.65 (m, 1H), 1.52-1.30 (m, 2H), 1.30-1.00 (m, 15H), 1.00- $0.70 \text{ (m, 36H)}, 0.58 \text{ (q, } J = 8.0 \text{ Hz, 6H)}, 0.02 \text{ (s, 3H)}, -0.38 \text{ (s, 3H$ 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 220.0, 217.1, 179.7 (2C), 176.1 (2C), 165.3, 165.1, 163.5 (2C), 119.2, 119.1, 117.9 (2C), 78.3, 78.1, 77.2, 75.7, 73.0, 72.9 (2C), 70.1, 49.7, 48.9, 48.6, 47.4, 46.9, 41.6, 41.3, 41.2, 40.0, 38.0, 37.8, 36.6, 36.2, 36.0, 35.3, 26.8, 26.2, 26.1, 26.0, 25.9, 24.7, 24.4, 23.3, 18.3, 16.9, 16.2, 15.6, 15.0, 14.9, 14.1 (2C), 12.4 (2C), 12.3, 12.2 (2C), 11.9. 11.1. 11.0. 9.8. 9.6. 7.6. 7.5. 7.4. 7.0. 6.9. 6.6. 5.9. 5.5. 5.4, 5.3, 5.3, -3.8, -4.3; HRMS (ESI): m/z 831.5591, calcd for $C_{45}H_{84}O_8Si_2Na [M + Na]^+ 831.5602$.

Triketone 35e. Aldol 35c (82.5 mg, 102 µmol) was treated with a 4:4:1 mixture of acetic acid, H₂O, and THF (4.5 mL) at room temperature for 13 h. The mixture was poured into saturated aqueous NaHCO₃ (70 mL) at 0 °C and extracted with EtOAc $(3 \times 20 \text{ mL})$. The combined extracts were washed with saturated aqueous NaHCO3 and brine, dried over Na2SO4, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (10g, hexane-EtOAc $3:1 \rightarrow 2:1 \rightarrow 1:1$) to give a diastereometric mixture of diol **35d** (60.2 mg, 84%) as a colorless oil: $R_f = 0.68, 0.61, 0.45$ (1:1 hexane/EtOAc); IR (CHCl₃): 3493, 1723, 1652, 1592, 1462, 1427, 1379 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 5.12 (d, J = 9.5 Hz, 1H), 4.32 (d, J = 8.1 Hz, 1H), 3.96 (d, J =8.9 Hz, 1H), 3.70-3.55 (m, 1H), 3.25-3.10 (m, 1H), 3.05-2.75 (m, 2H), 2.70–2.30 (m, 4H), 2.20–2.00 (m, 1H), 1.97 (s, 3H), 1.94 (s, 3H), 1.80-1.65 (m, 2H), 1.60-1.35 (m, 2H), 1.30-1.05 (m, 15H), 1.05–0.75 (m, 27H), 0.04 (s, 3H), -0.31 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃): δ 220.8, 179.7, 176.2, 165.0, 163.6, 119.3, 118.0, 77.2, 74.8, 73.1, 70.3, 47.1, 47.0, 45.7, 41.6, 41.2, 39.9, 39.1, 38.1, 37.4, 36.3, 36.1, 34.4, 33.3, 30.6, 26.8 (2C), 26.1, 26.0, 25.9, 25.0, 24.8, 18.3 (3C), 17.1, 16.9, 16.8, 15.4, 15.2, 14.9, 14.8 (2C), 11.9 (2C), 11.2, 9.8, 9.6, -3.9, -4.2; HRMS (ESI): m/z 717.4747, calcd for C₃₉H₇₀O₈-SiNa $[M + Na]^+$ 717.4738.

To a stirred solution of diol 35d (60.2 mg, 86.6 µmol) in CH₂Cl₂ (2 mL) was added Dess-Martin periodinane (260 mg, 471 µmol) at room temperature. The mixture was stirred at room temperature for 35 min; diluted with a 1:1:2 mixture of saturated aqueous NaHCO₃, saturated aqueous Na₂S₂O₃ and H₂O (4 mL); and extracted with Et₂O (2×10 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (6 g, hexane-EtOAc $5:1 \rightarrow 3:1 \rightarrow 1:1$) to give a keto-enol equilibrium mixture of triketone **35e** (51.8 mg, 87%) as a colorless oil: $R_f = 0.66$ (1:1 hexane/EtOAc); $[\alpha]_{D}^{25}$ +23.4 (c 1.036, CHCl₃); IR (CHCl₃): 1724, 1653, 1593, 1462, 1427, 1379 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 16.94 (s, 0.07H), 16.81 (s, 0.06H), 16.67 (s, 0.18H), 5.55-5.39 (m, 0.3H), 5.32-5.25 (m, 0.4H), 5.20-5.08 (m, 0.3H), 4.15-3.60 (m, 1.69H), 3.55-3.30 (m, 0.7H), 3.30-3.00 (m, 1.3H), 2.80-2.45 (m, 3H), 2.45-2.20 (m, 1H), 2.20-1.55 (m, 12H), 1.50–1.05 (m, 18H), 1.05–0.70 (m, 21H), {(0.10)–

(2'R)-Auripyrone B (2'-epi-Auripyrone B) (36). A solution of triketone 35e (51.8 mg, 75.1 µmol) in a 5:3:7 mixture of HF. pvridine, pvridine, and THF (10.0 mL) was stirred at 60 °C for 19.5 h. The mixture was poured into saturated aqueous NaHCO₃ (150 mL) at 0 °C and extracted with EtOAc (3 \times 20 mL). The combined extracts were washed with saturated aqueous NaHCO3 and brine, dried over Na2SO4, filtered, and concentrated. The residual oil was purified by column chromatography on silica gel (6 g, hexane-EtOAc $5:1 \rightarrow 2:1 \rightarrow 1:1$) and by preparative HPLC [Develosil ODS-HG-5 (250×20 mm), flow rate 5 mL min⁻¹; detection, UV 215 nm, solvent 80% aqueous MeOH] to give (2'R)-auripyrone B (2'-epiauripyrone B) (36) (10.3 mg, 25%) as colorless crystals: $R_f = 0.37$ (1:1 CH₂Cl₂/Et₂O); $[\alpha]_D^{25}$ +35.7 (*c* 0.837, CHCl₃); IR (CHCl₃): 1726, 1655, 1623, 1461, 1426, 1386 cm⁻¹; ¹H NMR (600 MHz, C_6D_6): δ 4.90 (dd, J = 3.4, 3.4 Hz, 1H), 3.95 (dd, J = 10.3, 2.2 Hz, 1H), 2.74 (dq, J = 10.3, 7.0 Hz, 1H), 2.43 (ddq, J = 7.0, 7.0, 7.0 Hz, 1H), 2.35–2.11 (m, 3H), 2.09 (dq, J = 15.0, 7.4 Hz, 1H), 2.02 (s, 3H), 1.97 (s, 3H), 1.88-1.65 (m, 3H), 1.64 (s, 3H), 1.58-1.48 (m, 1H), 1.43-1.32 (m, 2H), 1.13 (d, J = 7.0 Hz, 3H), 1.11 (d, J = 7.0 Hz, 3H), 0.99 (d, J = 6.8 Hz, 3H), 0.89 (t, J = 7.4 Hz, 3H), 0.88 (t, J = 7.6 Hz, 3H), 0.83 (t, J = 7.5 Hz, 3H), 0.77 (d, J = 7.1 Hz, 3H), 0.71 (d, J = 7.0 Hz, 3H), 0.63 (d, J = 7.1 Hz, 3H); ¹³C NMR (150 MHz, C_6D_6): δ 191.9, 178.6, 175.8, 166.3, 162.2, 161.6, 121.4, 118.2, 107.9, 105.1, 75.1, 70.7, 44.6, 42.0, 37.3, 36.6, 34.1, 31.8, 27.0, 26.3, 24.8, 17.0, 16.0, 12.2, 12.0 (2C), 11.9, 11.0, 10.8, 9.8, 9.5, 9.0, 8.2; HRMS (ESI): m/z 581.3443, calcd for $C_{33}H_{50}O_7Na [M + Na]^+$ 581.3454.

Cytotoxic Activity Assay. Stock cultures of HeLa S₃ cells were maintained in Eagle's Minimum Essential Medium containing Earle's Balanced Salts and 10% fetal bovine serum (DS Pharma Biomedical Co., Ltd.) and 1% antibiotic–antimycotic mixed stock solution {penicillin (10000 units mL⁻¹), streptomycin (10 mg mL⁻¹), and amphotericin B ($25 \mu g mL^{-1}$)} at 37 °C under 5% CO₂.

For the purpose of the experiment, 2×10^3 cells suspended in 100 µL of medium per well were plated in a 96-well plate. After 12 h incubation at 37 °C under 5% CO₂ to allow cell attachment, compounds in 100 µL of medium were added to the wells at different concentrations and incubated for 96 h under the same conditions. After 3 h of the MTT (1.44 mg mL⁻¹, 50 µL) addition to each well, the medium/MTT mixtures were removed, and the formazan crystals formed were dissolved in 150 µL of DMSO per well. After 30 min, optical absorbance at 540 nm were measured with a microplate reader. The cytotoxic effects of each compound were obtained as IC₅₀ values.

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Supporting Information

¹H and ¹³C NMR spectra of all new compounds. This material is available free of charge on the Web at http://www.csj.jp/journals/bcsj/.

References

1 Review: a) S. Yamamura, S. Nishiyama, *Bull. Chem. Soc. Jpn.* **1997**, *70*, 2025. b) P. Sharma, K. J. Powell, J. Burnley, A. S. Awaad, J. E. Moses, *Synthesis* **2011**, 2865.

2 K. Suenaga, H. Kigoshi, K. Yamada, *Tetrahedron Lett.* **1996**, *37*, 5151.

3 T. Lister, M. V. Perkins, Angew. Chem., Int. Ed. 2006, 45,

2560.

4 M. E. Jung, R. Salehi-Rad, Angew. Chem., Int. Ed. 2009, 48, 8766.

5 I. Hayakawa, T. Takemura, E. Fukasawa, Y. Ebihara, N. Sato, T. Nakamura, K. Suenaga, H. Kigoshi, *Angew. Chem., Int. Ed.* **2010**, *49*, 2401.

6 M. E. Jung, M. Chaumontet, R. Salehi-Rad, Org. Lett. 2010, 12, 2872.

7 T. Sengoku, T. Takemura, E. Fukasawa, I. Hayakawa, H. Kigoshi, *Tetrahedron Lett.* **2009**, *50*, 325.

8 T. Takemura, I. Hayakawa, E. Fukasawa, T. Sengoku, H. Kigoshi, *Tetrahedron* **2012**, *68*, 6477.

9 M. J. Gaunt, A. S. Jessiman, P. Orsini, H. R. Tanner, D. F. Hook, S. V. Ley, *Org. Lett.* **2003**, *5*, 4819.

10 H. C. Brown, K. S. Bhat, J. Am. Chem. Soc. 1986, 108, 293.

11 S. D. Rychnovsky, B. Rogers, G. Yang, J. Org. Chem. 1993, 58, 3511.

12 R. Pappo, D. S. Allen, Jr., R. U. Lemieux, W. S. Johnson, J. Org. Chem. 1956, 21, 478.

13 J. D. White, G. L. Bolton, A. P. Dantanarayana, C. M. J. Fox, R. N. Hiner, R. W. Jackson, K. Sakuma, U. S. Warrier, *J. Am. Chem. Soc.* **1995**, *117*, 1908.

14 T. Mukaiyama, R. W. Stevens, N. Iwasawa, *Chem. Lett.* **1982**, 353.

15 N. M. Kelly, R. G. Reid, C. L. Wellis, P. L. Winton, *Tetrahedron Lett.* **1996**, *37*, 1517.

16 J. Inanaga, K. Hirata, H. Saeki, T. Katsuki, M. Yamaguchi, Bull. Chem. Soc. Jpn. **1979**, *52*, 1989.

17 T. Yamori, A. Matsunaga, S. Sato, K. Yamazaki, A. Komi, K. Ishizu, I. Mita, H. Edatsugi, Y. Matsuba, K. Takezawa, O. Nakanishi, H. Kohno, Y. Nakajima, H. Komatsu, T. Andoh, T. Tsuruo, *Cancer Res.* **1999**, *59*, 4042.