

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 17 (2007) 1466-1470

Bis-styrylpyridine and bis-styrylbenzene derivatives as inhibitors for Aβ fibril formation

Seong Rim Byeon,^{a,b} Ji Hoon Lee,^a Ji-Hoon Sohn,^c Dong Chan Kim,^a Kye Jung Shin,^a Kyung Ho Yoo,^a Inhee Mook-Jung,^c Won Koo Lee^b and Dong Jin Kim^{a,*}

^aMedicinal Chemistry Research Center, Korea Institute of Science and Technology, P.O. Box 131, Cheongryang, Seoul 130-650, Republic of Korea ^bDepartment of Chemistry and Interdisciplinary Program of Integrated Biotechnology, Sogang University, Seoul 121-742, Republic of Korea ^cDepartment of Biochemistry and Cancer Research Institute, Seoul National University College of Medicine, Seoul 110-799, Republic of Korea

> Received 18 September 2006; revised 25 October 2006; accepted 28 October 2006 Available online 2 November 2006

Abstract—New bis-styrylpyridine and bis-styrylbenzene derivatives were designed and synthesized. These 34 compounds were evaluated by $A\beta$ fibril formation inhibitory assay using thioflavin T as a dye (named ThT assay). Most of them showed excellent inhibitory activities for $A\beta$ fibril formation at IC₅₀ of 0.1–2.7 μ M which is comparable to curcumin (IC₅₀ of 0.8 μ M). Among them, nine compounds were screened for their cytotoxicities on HT-22 cell by MTT assay at 1, 10, and 50 μ M. In particular, I-7 and II-2 exhibited the best combination of inhibitory activity and compound cytotoxicity. © 2006 Elsevier Ltd. All rights reserved.

Alzheimer disease (AD) is a sort of dementia and Alzheimer patients progressively undergo brain disorder that destroys cognitive functions and daily activities.

Since cholinergic deficit in the central nervous system has been observed in AD patients, inhibitors for acetylcholine esterase (AchE) such as tacrin, donepezil, and rivastigmine are used for the clinical treatment of AD symptoms in current therapeutic approaches. Although these are the only FDA-approved AD therapies on the market, they cannot solve fundamental problems but can be only palliative solutions.¹

Pathomorphologically important hallmarks in AD are senile plaques (SP) composed of amyloid β (A β)²⁻⁴ and neurofibrillary tangles (NFTs). Therefore, reducing deposition of A β plaques in the brain is believed to be pivotal to treat the AD patients.⁵ Based on A β hypothesis,⁶ A β 42 monomers divided by β -, γ -secretases from APP are gradually aggregated to oligomers, protofibrils, fibrils, and plaques. A β 42 aggregates such as oligomers and fibrils deposit in the brain and cause strong neuronal toxicity. On the other hand, it was recently reported that the precursors of fibrils, oligomers might be more toxic than fibrils.⁷

Consequently, we attempted to design and synthesize small molecules that prevent the formation of insoluble A β fibrils by specific binding with A β 42 monomers or relatively smaller oligomers. Based on the previous results,⁸ we synthesized (*E*, *E*)-1,3-bis-3'-methoxy-4'-hydroxystyrylbenzene which was suggested to shift the equilibrium of multimeric A β , inhibiting the pathogenic oligomer or fibril formation. In this research, we designed and synthesized various new bis-styrylpyridine and bis-styrylbenzene derivatives following the below Scheme (Fig. 1).

All the synthesized compounds were evaluated by A β fibril formation inhibitory assay using thioflavin T as a dye (named ThT assay).⁹ IC₅₀ value was estimated after % inhibition screening at 20 μ M, then 2 μ M of the compounds. Among them, nine compounds were screened for their cytotoxicities on HT-22 cell by MTT assay at 1 and 10 μ M (Fig. 4).¹⁰

Keywords: A β fibril formation inhibitor; Alzheimer disease; Bis-styrylpyridine; Bis-styrylbenzene; Amyloid β .

^{*} Corresponding author. Tel.: +82 2 958 5142; fax: +82 2 958 5189; e-mail: djk2991@kist.re.kr



I-12: R¹=Br, R²=OMe, R³=NH₂ **I-13:** R¹=H, R²=OMe, R³=OH **I-14:** R¹=Br, R²=OMe, R³=OH **I-15:** R¹=CI, R²=OMe, R³=OH

I-16: R¹=OMe, R²=OMe, R³=OH I-17: R¹=Me, R²=OMe, R³=OH I-18: R¹=NMe₂, R²=OMe, R³=OH



II-11: R¹=Br, R²=OMe, R³=OMe II-16: R¹=I, R²=OMe, R³=OH II-12: R¹=H, R²=H, R³=NMe₂ II-13: R¹=Br, R²=H, R³=NMe₂ II-14: R¹=I, R²=H, R³=NMe₂ II-15: R¹=Br, R²=OMe, R³=OH





Scheme 1. Synthesis of bis-styrylpyridine and bis-stylrylbenzene derivatives. Reagents and conditions: (a) triethylphosphite, 135 °C, 2.5 h; (b) 1 M ^{*t*}KOBu in THF, aldehyde (V-1–7), THF, rt, 0.5 h; (c—i) SnCl₂, EtOH, reflux, 3 h; (c—ii) 1 N HCl, EtOH, reflux, 3 h; (d) NaOMe, (CH₂O)_{*n*}, MeOH, NaBH₄, DMSO, reflux, 2 h; (e) AcOH, (CH₂O)_{*n*}, NaCNBH₃; (f) BBr₃, CH₂Cl₂, rt, 8 h.

Regarding the chemicals, new bis-styrylpyridine and bisstyrylbenzene derivatives were synthesized as shown in Scheme 1. Various synthetic 1,3-bis(bromomethyl)pyridines (or benzenes) (III) were reacted with triethyl phosphite by Arbuzov reaction to give the corresponding 1,3-bis(diethylphosphono-methyl)pyridines (or benzenes) (IV). The Horner–Emmons reaction of phosponates (IV) with synthetic aldehydes (V) (Fig. 2) is the key step to obtain only *trans–trans* (*E*, *E*) isomer in the presence of potassium *tert*-butoxide as a base, respectively.^{11,12} To obtain a series of styryl compounds having hydroxyl



Figure 2. Synthetic aldehydes (V-1–7).



Figure 3. Structures of IMSB and curcumin.

group at C-4 and methoxy at C-3 in the terminal aromatic ring (Fig. 1), PMB-protected intermediates were consecutively deprotected (by c-ii in Scheme 1). ⁸ To obtain C-3 (or C-4) mono or dimethylaminostyrylpyridine (or benzene) derivatives, the intermediates bearing nitro group at C-3 (or C-4) position were reduced (by c-i in Scheme 1), then subsequently were served in reductive amination (by d or e in Scheme 1).¹³ Finally, styryl compounds with both alkylamine and hydroxyl groups were prepared by demethylation using BBr₃ (by f in Scheme 1).^{11,12}

In the design of the target compounds, the features of IMSB (Fig. 3)^{11,12} were considered. Although IMSB has the excellent binding affinity for Aß fibrils, its inhibitory activity for A β fibril formation was relatively low (IC₅₀ of 8μ M) and brain uptake was hampered by the polar carboxylic acid group. Therefore, we replaced the carboxylic acid with a hydroxyl or alkylamine group to increase lipophilicity for penetration of blood-brain barrier (BBB) and enhance in vivo pharmacokinetic stability with high solubility. We then changed *p*-position of the styryl conjugated system to *m*-position as in curcumin.¹⁵ It was reported¹⁶ that low dose of curcumin effectively disaggregates $A\beta$ as well as prevents fibril and oligomer formation in vitro and in vivo (aged Tg 2576 mice). Especially it is widely used as a food additive and has undergone extensive toxicological screening and preclinical investigation in vivo. Therefore, structural similarity of the synthetic compounds with curcumin possesses good opportunity to be a potent AD drug candidate.

Shown in Table 1, 27 compounds have an excellent inhibitory activity with IC_{50} of 0.1–2.7 μ M and the curcumin, a reference compound, was assessed with IC_{50} of 0.8 μ M.

Table 1.	Biological a	ctivity of	bis-styryipyridines	(I-I-I8) and	bis-styryibenzenes	(II-I–I0) IC	or inhibition of Ap	42 fibril formation

Compound	Inhibition of fibril formation $IC_{50} (\mu M)^a$	Compound	Inhibition of fibril formation $IC_{50} (\mu M)^a$
I-1	0.6	II-1	0.6
I-2	1.1	II-2 ¹⁷	0.6
I-3	0.5	II-3	1.0
I-4	0.9	II-4	0.7
I-5	2.3	II-5	0.3
I-6	0.7	II-6	1.3
I-7 ¹⁷	0.5	II-7	1.1
I-8	0.9	II-8	2.7
I-9	0.8	П-9	>20
I-10	0.9	II-10	1.8
I-11 ^b	-	II-11 ^b	$\simeq 20$
I-12	1.1	II-12	>20
I-13	0.8	П-13	>20
I-14	1.1	II-14	>20
I-15	1.1	II-15	0.1
I-16	1.7	II-16 ^b	
I-17	1.4	Curcumin	0.8
I-18	1.9	Acridine orange ¹⁴	0.6
		IMSB	8

IC₅₀ was calculated from nonlinear regression by Graphpad Prism software.

^a ThT assay.

^b These were not suitable to ThT assay because of their own high-fluorescence interference effect.



Figure 4. Compound (finally $10 \,\mu$ M) cytotoxicities by MTT assay on HT-22 cell. The cells were treated with the compounds in media with 10% fetal bovine serum and 1% penicillin/streptomycin (totally 0.5% DMSO). In control, the cells were only treated with the same media (0.5% DMSO).

In the structure–activity relationship, the central pyridine or benzene did not affect the IC_{50} and R^1 substituents in the central ring also did not show a meaningful effect (**I-13–18** in Table 1). The value of IC_{50} was increased slightly in order of halogen < Me < OMe < NMe₂. The compounds (**II-12, 13**, and **14**) with only one substituent (dimethylamine) at C-4 position in the terminal aromatic ring showed lower activity than compound (**II-3**) with two substituents (dimethylamine and methoxy) both at C-3 and C-4.

Changing –OMe at C-4 (II-11) to –OH (II-15) or –NH₂ (II-10) significantly enhanced the inhibitory activity. These results suggest that the presence of two electron donating groups both at C-3 and C-4 positions probably plays a critical role to display higher activity, and at least one of them at C-3 or C-4 is necessary to be – NH₂, –NHMe, –NMe₂ or OH.¹² When R³ substituent was replaced with NH₂, NMe₂ or N(CH₂ CH₂)₂, inhibitory activity was decreased from 0.6, 1.0 to >20 μ M (in II-1, 3, and 9).

Selected nine compounds were initially screened at low concentration of 1 μ M for the compound cytotoxicity and then at high concentration of 10 μ M. In Figure 4, **II-2** (150%) showed a predominant relative cell viability compared with control (100%) and curcumin (116%) at 10 μ M. And HT-22 neuronal cells are mostly safe to the other compounds at high dose.

In conclusion, most of the synthesized compounds in this study inhibit $A\beta$ 42 fibril formation near sub- μ M range of IC₅₀ and most of the compounds do not show their own cytotoxicities on HT-22 neuronal cell even at high concentration. In particular, I-7 and II-2 exhibited the best combination of inhibitory activity and compound cytotoxicity. Based on these results, they are more potent $A\beta$ fibril formation inhibitors than curcumin.

Additional assays including in vitro BBB penetration, protection effect of toxicity induced by $A\beta$ 42 fibrils, in vivo pharmacokinetics, and behavior tests on APP/ PSI mice are currently in progress.

Acknowledgments

We are grateful to the Ministry of Science and Technology (MOST) and Ministry of Commerce, Industry and Energy (MCIE) of Korea for financial support.

References and notes

- 1. Cutler, N. R.; Sramek, J. J. Progr. Neuro-Psychopharmacol. Biol. Psychiatr. 2001, 25, 27.
- 2. Meyer-luemen, M. Nat. Neurosci. 2003, 6, 1.
- 3. Nussbaum, R. L.; Ellis, C. E. N. Engl. J. Med. 2003, 348, 1356.
- 4. Hardy, J.; Selkoe, D. J. Science 2002, 297, 353.
- (a) Thorsett, E. D.; Latimer, L. H. Curr. Opin. Chem. Biol. 2000, 4, 377; (b) Vickers, J. C.; Dickson, T. C.; Adlard, P. A.; Sounders, H. L.; King, C. E.; McCormack, G. Prog. Neurobiol. 2000, 60, 139.
- (a) Hammarstrom, P.; Wiseman, R. L.; Powers, E. T.; Kelly, J. W. *Science* 2003, *299*, 713; (b) Dominquez, D. I.; De Strooper, B. *Trends Pharmacol. Sci.* 2002, *23*, 324.
- (a) Lashuel, H. A.; Petre, B. M.; Wall, J.; Simon, M.; Nowak, R. J.; Walz, T.; Lansbury, P. T. *J. Mol. Biol.* 2002, 322, 1089; (b) Volles, M. J.; Lansbury, P. T. *Biochemistry* 2003, 42, 7871.
- Lee, K. H.; Shin, B. H.; Shin, K. J.; Kim, D. J.; Yu, J. Biochem. Biophys. Res. Commun. 2005, 328, 816.
- 9. Revine, H., III Arch. Biochem. Biophys. **2002**, 404, 106, In vitro ThT assay: the final concentration of $A\beta$ 42 (Bachem) was 25 μ M. The excitation and emission wavelengths were 450 and 482 nm with 10 nm slit (LS-55 luminescence spectrometer: Perkin-Elmer).
- 10. MTT assay: MTT(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) was added into the HT-22 cell.

Cell viability was estimated by measuring the absorbance of MTT formazan at 570 nm with a microplate reader. All compounds were tested as the HCl salt form. Compound **II-3** was tested in the state of a free-form and a HCl-salt form as an example.

- Lee, C. W.; Zhuang, Z. P.; Kung, M. P.; Plössl, K.; Skovronsky, D.; Gur, T.; Hou, C.; Trojanowski, J. Q.; Lee, W. M. Y.; Kung, H. F. J. Med. Chem. 2001, 44, 2270.
- 12. Mathis, C. A.; Wang, Y.; Klunk, W. E. Curr. Pharm. Design 2004, 10, 1469.
- 13. Ono, M.; Kung, M. P.; Hou, C.; Kung, H. F. Nucl. Med. Biol. 2002, 29, 633.
- Suemoto, T.; Okamura, N.; Shiomitsu, T.; Suzuki, M.; Shimadzu, H.; Akatsu, H.; Yamamota, T.; Kudo, Y.; Sawada, T. *Neurosci. Res.* 2004, 48, 65.
- Yang, F.; Lim, G. P.; Begum, A. N.; Ubeda, O. J.; Simmons, M. R.; Ambegaokar, S. S.; Chen, P.; Kayed, R.; Glabe, C. G.; Frautschy, S. A.; Cole, G. M. J. Biol. Chem. 2005, 280, 5892.
- 16. Chainani-Wu, N. J. Altern. Complement Med. 2003, 9, 161.
- 17. Selected data. Compound I-7: ¹H NMR (CD₃OD, 300 MHz) δ 2.86 (s, 6H), 3.93 (s, 6H), 6.59 (d, J = 7.83, 2H), 7.03 (d, J = 16.15, 2H), 7.12 (d, J = 8.35, 2H), 7.07 (s, 2H), 7.37 (d, J = 7.81, 2H), 7.52 (d, J = 16.14, 2H), 7.67 (t, J = 7.51, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ 30.16, 55.46, 106.78, 108.65, 118.45, 122.29, 124.05, 125.33, 133.35, 136.58, 139.98, 146.87, 156.18; HRMS m/z Calcd for $C_{25}H_{27}N_3O_2$ (M)⁺ 401.2103. Found: 401.2182. Anal. Calcd for $C_{25}H_{27}N_3O_2$: C, 74.79; H, 6.78; N, 10.47. Found: C, 74.51; H, 6.77; N, 10.12. II-2: ¹H NMR (CDCl₃, 300 MHz) & 2.91 (s, 6H), 3.93 (s, 6H), 6.58 (d, J = 8.09, 2H), 6.94 (d, J = 16.23, 2H), 7.01 (d, J = 1.64, 2H), 7.07 (dd, J = 8.20, 1.71, 2H), 7.11 (d, J = 16.25, 2H), 7.33 (t, J = 4.63, 1H), 7.34 (d, J = 1.18, 2H), 7.61 (s, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ 30.44, 55.49, 106.66, 109.30, 121.11, 123.10, 124.40, 124.52, 127.36, 128.84, 129.39, 138.63, 139.11, 147.12; HRMS m/z Calcd for C₂₆H₂₈N₂O₂ (M)⁺ 400.2151, Found 400.2151; Anal. Calcd for C₂₆H₂₈N₂O₂: C, 77.97; H, 7.05; N, 6.99. Found: C, 78.41; H, 7.18; N, 6.57.