## Formal Total Synthesis of (-)-5,6-Dihydrocineromycine B

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**Abstract:** An efficient and highly convergent formal total synthesis of the 14-membered macrolide (–)-5,6-dihydrocineromycine B is achieved. Key reaction sequences include a Sharpless asymmetric epoxidation followed by esterification for the formation of a fully functionalized acyclic precursor, Corey–Bakshi–Shibata reduction, and ring-closing metathesis, respectively.

Key words: antibiotic macrolide, (-)-5,6-dihydrocineromycine B, Sharpless asymmetric epoxidation, CBS reduction, ring-closing metathesis

Macrolide antibiotics belong to a class of polyketide natural products that exhibit a wide range of biological activities.<sup>1</sup> These antibiotics are usually produced by certain species of *Streptomyces* and possess a macrolactone ring and double bonds. The cineromycins and 5,6-dihydrocineromycine B are perhaps the best examples of these interesting macrolides that have been isolated from *Streptomyces* sp. (strain Gö 40/10).<sup>2</sup> The cineromycins and albocyclins-18 are structurally related antibiotic natural products which display potent antibiotic activity against *Staphylococci*. As shown in Figure 1, structural features of (–)-5,6-dihydrocineromycine B (1) encompasses four stereogenic centers, two hydroxyls and two carbon–carbon double bonds which are embedded in a 14-membered lactone ring.



Figure 1 Structures of cineromycin B and its derivatives

*SYNLETT* 2012, 23, 2677–2681 Advanced online publication: 18.10.2012 DOI: 10.1055/s-0032-1317346; Art ID: ST-2012-D0589-L © Georg Thieme Verlag Stuttgart · New York The unique structures of these macrolides, in conjunction with their potent biological activities, have made them attractive synthetic targets.<sup>3</sup> To date two total syntheses<sup>4</sup> of 5,6-dihydrocineromycine B (1) have appeared in the literature.

As part of our continuous endeavors in the synthesis of bioactive macrolides,<sup>5</sup> we were attracted to the structural features and antibiotic activity of (–)-5,6-dihydrocineromycine B (1). Herein, we report the formal total synthesis of 1, utilizing Sharpless asymmetric epoxidation, Yamaguchi esterification, Corey–Bakshi–Shibata (CBS) reduction, and ring-closing metathesis (RCM) as key steps.

Our retrosynthetic analysis of 1 is shown in Scheme 1. As indicated, the cyclic framework could be constructed by RCM of 5, which in turn could be obtained by the Yamaguchi esterification of the building blocks – alkenes 6 and 7. Fragment 6 could be derived from ester 18 via oxidation followed by Grignard reaction, which is conceived to be obtained through a four-step sequence starting from the inexpensive propane-1,3-diol (10). Similarly, we envisioned that fragment 7 can be obtained from but-2-yne-1,4-diol (12) through Sharpless asymmetric epoxidation and other functional-group transformations.

As outlined retrosynthetically in Scheme 1, the synthesis of fragment 6 commenced from commercially available propane-1,3-diol (10) which was converted into the  $\alpha$ , $\beta$ unsaturated ester 13 by following a known procedure (Scheme 2).<sup>6</sup> Treatment of  $\alpha,\beta$ -unsaturated ester 13 with DIBAL-H at 0 °C yielded allyl alcohol 14 in 95% yield. Sharpless asymmetric epoxidation<sup>7</sup> of 14 using (+)-diisopropyl L-tartrate, titanium isopropoxide, and TBHP at -20 °C furnished 15 in 95% yield with an excellent enantioselectivity of 98% ee. Regioselective opening of epoxide 15 using LAH provided the corresponding<sup>8</sup> 1,2-diol 9 in 92% yield. The primary hydroxyl group in 9 was oxidized using 2-iodoxybenzoic acid (IBX)<sup>9</sup> to afford an aldehyde, which was immediately subjected to Horner-Wadsworth-Emmons olefination<sup>10</sup> to give **16** exclusively as the E-isomer (J = 15.6 Hz) in 90% yield.

The tertiary hydroxyl group in **16** was protected as a TES ether by using TESCl and triethylamine in dichloromethane to give compound **8** in 94% yield and subsequent oxidative removal<sup>11</sup> of the PMB group by using DDQ in dichloromethane, pH buffer (9:1 ratio) afforded **17** in 90% yield. Oxidation of primary alcohol **17** by Dess–Martin periodinane<sup>12</sup> in dichloromethane gave an aldehyde in quantitative yield which was treated with isopropenylmagnesium bromide in THF to afford the secondary alcohol in a 1:1 diastereomeric mixture and further Dess–



Scheme 1 Retrosynthetic analysis of (-)-5,6-dihydrocineromycine B (1) leading to the key building blocks 5-7

Martin oxidation of the secondary alcohol yielded ketone **18** in 97% yield.

Finally, compound **18** was subjected to ester hydrolysis using LiOH in THF–MeOH–H<sub>2</sub>O (4:4:2) and THF–H<sub>2</sub>O (1:1) at 0 °C to room temperature to give the corresponding acid **6** with low yields. The reaction was sluggish and resulted in multiple spots by TLC. At this stage, we opted for the Me<sub>3</sub>SnOH-mediated nonacidolytic and nonnucleophilic hydrolysis<sup>13</sup> conditions in dichloroethane at 80 °C. As anticipated, the hydrolysis proceeded smoothly to afford the expected acid **3** in 98% yield. We believe this could be a general mild method for the benign cleavage of esters within labile substrates.

The synthesis of building block 7 started from but-2-yne-1,4-diol (12) which was converted to allylic alcohol 19 by a known procedure.<sup>14</sup> Sharpless asymmetric epoxidation<sup>7</sup> of allylic alcohol 19 using (-)-DIPT resulted in pure epoxide 20 in 95% yield with 98% ee. Ring opening of the epoxide was carried out by following Tius<sup>15</sup> protocol using MeLi and CuI in Et<sub>2</sub>O, which yielded the mixture of 1,3diol and undesired 1,2-diol (6:1). Subsequently, the minor isomer was readily removed by treating the mixture with sodium periodate to yield 1,3-diol 21 in 71% yield. After debenzylation of 21 by H<sub>2</sub>, 10% Pd/C and MeOH at 25 °C for twelve hours afforded triol 22<sup>16</sup> in 95% yield, the 1,2protected hvdroxvl groups were using 22 dimethoxypropane<sup>17</sup> and PTSA in  $CH_2Cl_2$  at 25 °C to give 23 in 90% yield. Tosylation of the primary alcohol in 23 followed by the Grignard reaction with allylmagnesium bromide in the presence of stoichiometric<sup>18</sup> CuI led to the formation of allylated product 24 in 84% yield. Acid hy-



**Scheme 2** *Reagents and conditions*: (a) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, 30 min, 95%; (b) (+)-DIPT, Ti(Oi-Pr)<sub>4</sub>, *t*-BuOOH, CH<sub>2</sub>Cl<sub>2</sub>, -20 °C, 95%; (c) LAH, THF, 0 °C, 30 min, 92%; (d) (i) IBX, DMSO, THF, r.t., 1 h; (ii) (EtO)<sub>2</sub>POCH<sub>2</sub>CO<sub>2</sub>Et, *n*-BuLi, THF, 0 °C, 30 min, 90%; (e) TESCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to r.t., 6 h, 94%; (f) DDQ, CH<sub>2</sub>Cl<sub>2</sub>-H<sub>2</sub>O (9:1), 30 min, 90%; (g) (i) DMP, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to r.t., 1 h; (ii) C<sub>3</sub>H<sub>3</sub>MgBr, THF, -40 °C, 30 min, 70%; (iii) DMP, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to r.t., 1 h, 97%; (h) Me<sub>3</sub>SnOH, DCE, 80 °C, 10 h, 98%.

Synlett 2012, 23, 2677-2681

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**Scheme 3** *Reagents and conditions*: (a) (-)-DIPT, Ti(O*i*-Pr)<sub>4</sub>, *t*-BuOOH, CH<sub>2</sub>Cl<sub>2</sub>, -20 °C, 95%; (b) Me<sub>2</sub>CuLi, Et<sub>2</sub>O, -50 °C to 25 °C, 1,3-diol-1,2-diol (6:1); NaIO<sub>4</sub>, H<sub>2</sub>O, 71%; (c) H<sub>2</sub>, Pd/C, MeOH, 12 h, 95%; (d) 2,2-dimethoxypropane, PTSA, CH<sub>2</sub>Cl<sub>2</sub>, 6 h, 90%; (e) TsCl, pyridine, CH<sub>2</sub>Cl<sub>2</sub>, 12 h, 93%; (f) AllylMgBr, CuI, Et<sub>2</sub>O, 0 °C to r.t., 12 h, 82%; (g) AcOH–THF–H<sub>2</sub>O (1:1:1), 8 h, 60 °C, 92%; (h) TsCl, Bu<sub>2</sub>SnO, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 10 h, 90%; (i) LAH, THF, 0 °C, 2 h, 85%.

drolysis followed by the tosylation of the primary hydroxyl group in **25** was accomplished **26** in 90% using the Martinelli<sup>19</sup> procedure (Bu<sub>2</sub>SnO, TsCl). Finally, reductive cleavage of sulfonate **26** by LAH in THF afforded key fragment **7** in 85% yield.

The key fragment 7 can also be synthesized from 4-penten-1-ol (27) shown in Scheme 4. Swern oxidation and subsequent Wittig homologation of 27 yielded  $\alpha,\beta$ -unsaturated ester 28 in 90% yield exclusively as the *E*-isomer (*J* = 15.6 Hz). The ester was then subjected to reduction by DIBAL-H and afforded allylic alcohol 29, which was further subjected to Sharpless epoxidation using (–)-DIPT and resulted in pure epoxide 30 in 75% yield with 95% ee. Regioselective ring opening of epoxide 30 with trimethylaluminium<sup>20</sup> in dichloromethane gave 1,2-diol 25, which was converted into key fragment 7 as shown in Scheme 3.

With the two key fragments **6** and **7** in hand, the stage was now set for their assembly and elaboration into (–)-5,6-dihydrocineromycine B (**1**) as shown in Scheme 5. Our fragment assembly began with coupling of fragments **6** and **7**. Esterification of **6** and **7** under Yamaguchi conditions<sup>21</sup> yielded **31** in 97% yield.

The  $\alpha$ , $\beta$ -unsaturated ketone **31** was then treated with (*R*)-(+)-2-methyl-CBS oxazaborolidine<sup>22</sup> and BH<sub>3</sub>·DMS at -40 °C to furnish allyl alcohol **5** with an *S*-configuration in 90% yield with 98% de.<sup>23</sup> With a wealth of literature<sup>24</sup>

available for sterically crowded or functionalized dienes to undergo RCM,<sup>24</sup> we next submitted the diene **5** to 25 mol% of Grubbs' second-generation catalyst in refluxing dichloromethane. As expected, the RCM proceeded smoothly to afford compound **32** in 40% yield. This latestage intermediate has been already converted into the target compound in one step.<sup>4b</sup> All the intermediate compounds including macrolide **32** were fully characterized by IR, <sup>1</sup>H NMR, <sup>13</sup>C NMR, and mass spectral data.<sup>25</sup>

In summary, we have achieved an efficient and elegant formal synthesis of (–)-5,6-dihydrocineromycine B. The synthesis features key fragments which were constructed employing Sharpless asymmetric epoxidation and CBS reduction followed by ring-closing metathesis. A Yamaguchi esterification protocol was utilized to construct the backbone of the natural product, in which the 14-membered lactone moiety was constructed by the ring-closing metathesis. We believe that the presented synthetic method could be of value in the development of novel 14-membered lactone ring analogues for 5,6-dihydrocineromycine B research.

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**Scheme 4** *Reagents and conditions*: (a) (i) Swern oxidation; (ii) Ph<sub>3</sub>PCHCOOEt, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 6 h, 90% (2 steps); (b) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C; (c) (-)-DIPT, Ti(Oi-Pr)<sub>4</sub>, *t*-BuOOH, CH<sub>2</sub>Cl<sub>2</sub>, -20 °C, 95%; (d) Me<sub>3</sub>Al, toluene, 0 °C to r.t., 6 h, 86%.

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Scheme 5 *Reagents and conditions*: (a) benzene, 2,4,6-trichlorobenzoyl chloride, Et<sub>3</sub>N then 7, DMAP, 30 min, 97%; (b) (*R*)-2-methyl-CBS oxazaborolidine, THF, 0 °C, BH<sub>3</sub>·DMS, 3 h, 90%, 98% de; (c) Grubbs II (25 mol%), CH<sub>2</sub>Cl<sub>2</sub>, reflux, 48 h, 40%.

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- (25) Spectral Data for Representative New Compounds **Compound 9**: colorless oil;  $[\alpha]_D^{25}$  –5.5 (*c* 1, CHCl<sub>3</sub>). IR (KBr): 3399, 2933, 1612, 1513, 1248, 1035, 772 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.17 (d, 2 H, J = 8.30 Hz), 6.80 (d, 2 H, J = 9.00 Hz), 4.38 (s, 2 H), 3.74 (s, 3 H), 3.40 (t, 2 H, J = 5.20 Hz), 3.35-3.21 (m, 2 H), 1.70-1.39 (m, 4 H),1.06 (s, 3 H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 159.1, 129.2, 113.6, 72.5, 72.4, 70.4, 69.5, 54.8, 35.2, 24.0, 23.1. ESI-MS:  $m/z = 277 [M + Na]^+$ . ESI-HRMS: m/z calcd for C14H22NaO4: 277.1416; found: 277.1410. **Compound 16**: colorless oil;  $[\alpha]_D^{25}$  +13.7 (*c* 1, CHCl<sub>3</sub>). IR (KBr): 3446, 2926, 1722, 1655, 1513, 1219, 1035, 772 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.19 (d, 2 H, J = 8.49 Hz), 6.92-6.77 (m, 3 H), 5.97 (d, 1 H, J = 15.67 Hz), 4.40 (s, 2 H), 4.16 (q, 2 H, J = 7.17, 14.16 Hz), 3.77 (s, 3 H), 3.47-3.32 (m, 3 H), 1.75–1.56 (m, 4 H), 1.33–1.22 (m, 6 H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ = 166.3, 159.1, 154.4, 129.7, 129.1, 118.9, 113.6, 72.6, 72.1, 70.0, 59.9, 54.8, 39.5, 28.1, 24.3, 14.3. ESI-MS: *m/z* = 345 [M + Na]<sup>+</sup>. ESI-HRMS: *m/z* calcd for C<sub>18</sub>H<sub>26</sub>NaO<sub>5</sub>: 345.1678; found: 345.1667. **Compound 17**: colorless oil;  $[\alpha]_D^{25} + 10.6$  (*c* 1, CHCl<sub>3</sub>). IR (KBr): 3424, 2955, 1720, 1656, 1459, 1299, 1061, 772 cm<sup>-1</sup>.

Synlett 2012, 23, 2677-2681

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(–)-5,6-Dihydrocineromycine B **2681** 

<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ = 6.83 (d, 1 H, *J* = 15.1 Hz), 5.88 (d, 1 H, *J* = 15.1 Hz), 4.18 (q, 2 H, *J* = 6.79, 14.35 Hz), 3.59 (t, 2 H, *J* = 6.04 Hz), 1.67–1.52 (m, 4 H), 1.39 (s, 3 H), 1.31 (t, 3 H, *J* = 6.79 Hz) 0.97 (t, 9 H, *J* = 8.30, 15.86 Hz), 0.61 (q, 6 H, *J* = 7.55, 15.86 Hz). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ = 166.4, 154.4, 119.0, 75.1, 62.9, 60.2, 39.8, 27.9, 27.4, 14.4, 7.2, 6.8. ESI-MS: *m/z* = 339 [M + Na]<sup>+</sup>. ESI-HRMS: *m/z* calcd for C<sub>16</sub>H<sub>32</sub>NaO<sub>4</sub>Si: 339.1968; found: 339.1961.

**Compound 6**: colorless oil;  $[\alpha]_D^{25} - 13.5$  (*c* 1, CHCl<sub>3</sub>). IR (KBr): 3414, 2924, 1706, 1454, 1219, 1018, 770 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 6.92$  (d, 1 H, *J* = 15.86 Hz), 5.95 (d, 1 H, *J* = 15.10 Hz), 5.91 (s, 1 H), 5.73 (s, 1 H), 2.83– 2.69 (m, 1 H), 2.66–2.52 (m, 1 H), 1.95–1.81 (m, 5 H), 1.43 (s, 3 H), 0.97 (t, 9 H, *J* = 8.30 Hz), 0.61 (q, 6 H, *J* = 7.55, 15.86 Hz). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 200.6$ , 172.0, 156.7, 144.6, 124.1, 118.8, 74.8, 37.0, 31.9, 28.2, 17.8, 7.2, 6.7. ESI-MS: *m/z* = 349 [M + Na]<sup>+</sup>. ESI-HRMS: *m/z* calcd for C<sub>17</sub>H<sub>30</sub>NaO<sub>4</sub>Si: 349.1811; found: 349.1820. **Compound 11**: colorless oil,  $[\alpha]_D^{25}$ –2.5 (*c* 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 7.79$  (d, 2 H, *J* = 7.90 Hz),

7.34 (d, J = 7.90 Hz), 4.19–4.13 (m, 1 H), 4.03–3.96 (m, 2 H), 3.84 (q, 1 H, J = 6.30, 14.30 Hz), 3.58 (t, 1 H, J = 7.10 Hz), 2.45 (s, 3 H), 1.96–1.87 (m, 1 H), 1.31 (s, 3 H), 1.27 (s, 3 H), 0.90 (d, 3 H, J = 7.10 Hz). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 144.6$ , 132.9, 129.7, 127.8, 109.0, 76.3, 72.2, 67.6, 37.0, 26.5, 25.3, 21.5, 12.8. ESI-MS: m/z = 337 [M + Na]<sup>+</sup>. ESI-HRMS: m/z calcd for C<sub>15</sub>H<sub>22</sub>NaO<sub>5</sub>S: 337.1086; found: 337.1091.

**Compound 7**: colorless oil;  $[\alpha]_D^{25}$  –12.8 (*c* 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 5.85–5.65 (m, 1 H), 5.05–4.85

(m, 2 H), 3.62 (pent, 1 H, J = 6.04, 12.08, 18.12 Hz), 2.27– 2.06 (m, 1 H), 2.05–1.86 (m, 2 H), 1.66–1.42 (m, 1 H), 1.23– 1.13 (m, 1 H), 1.10 (d, 3 H, J = 6.04 Hz), 0.87 (d, 3 H, J = 6.79 Hz). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 138.73$ , 114.53, 71.26, 39.41, 31.73, 31.50, 19.39, 14.50. **Compound 31**: colorless oil;  $[\alpha]_D^{25}$  –17.2 (*c* 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 6.78$  (d, 1 H, J = 15.70 Hz), 5.93-5.84 (m, 2 H), 5.77-5.69 (m, 2 H), 5.02-4.38 (m, 3 H), 2.80-2.69 (m, 1 H), 2.67-2.58 (m, 1 H), 2.21-2.09 (m, 1 H), 2.06-1.97 (m, 1 H), 1.91-1.83 (m, 5 H), 1.79-1.69 (m, 1 H), 1.56-1.45 (m, 1 H), 1.41 (s, 3 H), 1.27-1.23 (m, 1 H), 1.20 (d, 3 H, J = 6.87 Hz), 0.96 (t, 9 H, J = 7.85 Hz), 0.92 (d, 3 H, J)J = 6.87 Hz), 0.61 (q, 6 H, J = 7.85, 15.70 Hz). <sup>13</sup>C NMR (75) MHz, CDCl<sub>3</sub>): δ = 201.47, 166.14, 153.78, 144.47, 138.58, 124.34, 119.72, 114.63, 74.67, 74.17, 37.26, 36.80, 32.09, 31.78, 31.29, 28.09, 17.73, 16.07, 14.60, 7.12, 6.71. ESI-MS:  $m/z = 459 [M + Na]^+$ . ESI-HRMS: m/z calcd for C25H44NaO4Si: 459.2907; found: 459.2910. **Compound 5**: colorless oil;  $[\alpha]_D^{25}$  –21.7 (*c* 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 6.86$  (d, 1 H, J = 15.48 Hz), 5.93–5.84 (m, 2 H), 5.91 (d, 1 H, J = 15.48 Hz), 5.87–5.69 (m, 1 H), 5.07-4.81 (m, 5 H), 4.02 (t, 1 H, J = 5.28 Hz),2.30-2.30 (m, 1 H), 2.23-1.93(m, 2 H), 1.69 (s, 3 H), 1.64-1.44 (m, 4 H), 1.38 (s, 3 H), 1.30–1.23 (m, 2 H), 1.19 (d, 3 H, J = 6.7 Hz), 0.96 (t, 9 H, J = 7.55 Hz), 0.92 (d, 3 H, J = 6.79 Hz), 0.61 (q, 6 H, J = 7.55, 15.86 Hz). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ = 166.17, 154.27, 147.27, 128.58, 127.58, 126.56, 114.75, 111.27, 75.81, 74.96, 74.10, 39.02 36.83, 31.81, 31.34, 29.79, 29.11, 27.71, 17.64, 16.12, 14.68, 7.16, 6.79. ESI-MS: *m/z* 461 [M + Na]<sup>+</sup>. ESI-HRMS: *m/z* calcd for C25H46NaO4Si: 461.3063; found: 461.3044.

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