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LAMPTEROMYCES BIOLUMINESCENCE --- 5

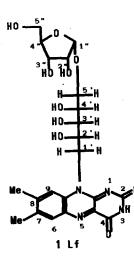
CHEMICAL SYNTHESIS OF LAMPTEROFLAVIN AS MUSHROOM LIGHT EMITTER

Minoru Isobe,* Hiroyuki Takahashi and Toshio Goto Laboratory of Organic Chemistry, School of Agriculture, Nagoya University Chikusa, Nagoya 464, Japan

Abstract: Lampteroflavin was synthesized through coupling between 5-trityl-2,3-(p-methoxy)benzylidene-1- β -D-ribofuranosyl trichloroacetimidate and 2',4'-(p-methoxy)benzylidene derivative of riboflavin to produce the crucial alpha riboside. Chloroethyl group was removed under neutral condition.

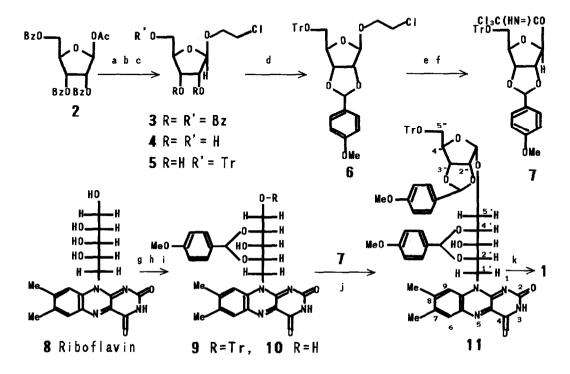
Lampteroflavin (Lf) was deduced as in 1 as the light emitter in bioluminescence of moon night mushroom, Lampteromyces japonicus.¹ We became interested in synthesizing 1 for confirmation of the stereochemistry and then for studies on the bioluminescent mechanism. The synthesis includes crucial alpha glycosidation onto ribose² with riboflavin at the 5'-position on its ribityl side chain. The significant protecting groups were selected to be trityl and p-methoxybenzylidene since they had to be deprotected either under non-reductive or non-basic condition due to the instability of the isoaloxazine ring. These protective groups in 7 and 10 survived under the coupling condition and they were removed after the coupling with no glycosidic cleavage.

The starting material, tribenzoate of 1-acetoxy-D-ribose 2 was converted into the chloroethyl- β -glycoside 3 aiming at a later deglycosidation under neutral condition. The



benzoyl groups were hydrolyzed into the triol 4, and the 5~ hydroxy group was selectively tritylated to give the diol 5 [PMR δ 5.00 ppm (H-1; s)]. The corresponding p-methoxybenzylidene δ [m/z 574, 576] was a single isomer judging from benzylidene proton at δ 5.72 ppm as a singlet. The chloroethyl glycoside was treated with sodium sulfinate and then with trichloroacetonitrile to afford the trichloroacetimidate 7 [PMR δ 6.37 ppm (H-1; s)].³ On the other hand, riboflavin 8 was protected 5'-trityl-2',4'-benzylidene derivative 9 [PMR & 5.60 ppm 88 (benzylidene-H; s)].4 After removing the trityl group in 9, the coupling was facilitated between 7 and 10 with BF3-OEt2 in dry dichloromethane at ambient temperature to produce the α glycoside 11 [5 5.17(H1"; d, J= 3.4Hz); FAB/MS m/z 987(M+1), 988, 989].14 The protective groups were cleaved in dichloromethane with trifluoroacetic acid without glycosidic bond

cleavage^{1b} to yield *1* as precipitates [m/z 509 (M+1), 510, 511].¹ The synthetic lampteroflavin as well as its hexa-acetate were identical in PMR (500 MHz), MS, HPLC etc. with those of the natural lampteroflavin, *1*.



Scheme 1

a) HOCH₂CH₂Cl/CSA rt 12hr; b) 1% KOH/MeOH rt 1hr; c) TrCl/Py 80°C 1.5hr (67% overall yield); d) p-methoxybenzaldehyde dimethylacetal/PPTS/DMF rt 4hr (96%); e) PhSO₂Na KI/DMF 90°C 5hr; f) Cl₃CCN DBU/CH₂Cl₂ 0°C 3hr (63% overall yield); g) TrCl/Py, 110°C 1hr (64%); h) p-methoxybenzaldehyde dimethylacetal/PPTS/DMF rt 3hr (96%); i) TFA/CH₂Cl₂ rt 0.5hr (60%); j) BF₃-OEt₂ CH₂Cl₂ rt 2hr; k) TFA/CH₂Cl₂ rt 3hr (48% overall yield).

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- 4. The signal of H-3' found at δ 3.5 ppm shifted to δ 5.21 ppm (t, J= 9.2) after the acetylation.

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