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Study on the thermal reactions of [60]fullerene with amino acids and amino acid esters[†]

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Thermal reactions of [60]fullerene with a series of amino acids and amino acid esters under aerobic and dark conditions have been investigated. Fulleropyrrolidines can be obtained from these reactions although an aldehyde is not added purposely. Possible reaction mechanisms involving uncommon C–N bond cleavages have been proposed to generate aldehydes, which then react with amino acids and amino acid esters to provide azomethine ylides, followed by 1,3-dipolar cycloaddition to [60]fullerene affording fulleropyrrolidines. Control experiments support our proposed mechanisms, and elucidate the innate nature of C–N bond cleavages of amino acids and amino acid esters.

Introduction

Many years have elapsed since the discovery of fullerenes, but their chemical modifications continue to be of great appeal due to the potential applications of fullerene derivatives in many fields of life and materials sciences.1 Amino acids and their derivatives are important biological molecules. They have been utilized to functionalize [60]fullerene (C₆₀) via the Prato reaction, *i.e.*, 1,3-dipolar cycloaddition of azomethine ylides to C_{60} .² Azomethine ylides are generated in situ from the decarboxylation of iminium salts derived from the condensation of amino acids with aldehydes/ketones or from the formal 1.2-H shift of imines of amino acid esters. The formed fulleropyrrolidine derivatives have recently been employed as the acceptors of photovoltaic solar cells with a power conversion efficiency (PCE) of up to 3.44% and are superior to methyl [6,6]-phenyl-C61-butylate ([C60]-PCBM) under the same conditions.³ Fulleropyrrolidines can also be further transformed to ionic fullerene derivatives and applied to supramolecular assemblies.^{1d}

Compared to the extensive application of amino acids and their derivatives in the Prato reaction, much fewer reports on the functionalization of C_{60} using amino acids or amino acid esters alone have been described. Gan's group and others have

disclosed a few photochemical reactions between amino acid esters and C₆₀.⁴ Gan and co-workers have also investigated the reactions of amino acid esters with C₆₀ in the presence of iodo reagents under ultrasonification.⁵ We have explored the reaction of C₆₀ with amino acid ester hydrochlorides by using triethylamine (Et₃N) as a base to remove HCl.^{6,7} We found that the reaction in CS2 at room temperature proceeded well and afforded fullerene products containing amino acid ester, thioamide and thiourea units.⁶ In contrast, the reaction in refluxing *o*-dichlorobenzene (ODCB) gave fulleropyrrolidine derivatives containing the CH₃CH moiety originated from an unusual C-N bond cleavage of Et₃N. The fulleropyrrolidines were formed through the reaction of C₆₀ with azomethine ylides from amino acids and acetaldehyde, which in turn were generated by the hydrolysis of the amino acid ester hydrochlorides in the presence of Et₃N and by the fragmentation/oxidization of Et₃N, respectively.⁷ On the other hand, studies on the direct thermal reaction of C₆₀ with amino acids themselves are scarce. Gan and co-workers investigated the reaction of C_{60} with glycine or alanine in the presence of sodium hydroxide and obtained complex mixtures of multiadducts.⁸ Herein, we report the direct thermal reactions of C₆₀ with a series of amino acids and amino acid esters in the absence of an aldehyde and under aerobic and dark conditions affording isolable monoadducts.^{9,10} More importantly, we have gained a mechanistic insight into these reactions deduced from the structural motifs of isolated products and control experiments.

Results and discussion

In our preliminary communication,¹⁰ we reported the thermal reaction of C_{60} with a few amino acids. In order to extend the substrate scope and better understand the reaction mechanism,

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[†] Electronic supplementary information (ESI) available: NMR spectra of products **2a-2d**, **4a-4d**, **5b**, **5c**, **7a-7d**, **8a-8c**, **9** and **10c**. See DOI: 10.1039/c2ob26066b

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we have extended the thermal reaction of C_{60} with more amino acids as well as that with amino acid esters. In contrast to the previous experimental procedure,¹⁰ and for rational comparison amongst the reactions of C_{60} with different amino acids and amino acid esters, the concentration of C_{60} was kept constant; a sealed tube instead of a conventional flask was used for the reaction to prevent any loss of the volatile starting materials or *in situ* generated intermediates; the reaction temperature was 180 °C and 10 equiv. of amino acid/amino acid ester were used in most cases.

We first systematically explored the thermal reaction of C_{60} with representative *N*-unsubstituted amino acids, that is, glycine (1a), alanine (1b), phenylalanine (1c) and phenylglycine (1d). Among the employed *N*-unsubstituted amino acids, 1a-1c are naturally occurring amino acids. To avoid the potential interference of a photochemical reaction,⁴ the thermal reaction of C_{60} was conducted under dark conditions. Intriguingly, the products from the thermal reaction of C_{60} with 1a-1d in refluxing ODCB in the presence of air proved to be *N*-unsubstituted fulleropyrrolidines 2a-2d, instead of aminated fullerene derivatives.⁸ It was found that the presence of oxygen was crucial for the thermal reaction, as strict exclusion of oxygen from the reaction mixture by a freeze–pump–thaw procedure failed to provide 2a-2d.¹¹

Table 1 Reaction conditions and product yields for the reaction of C_{60} with $1a-1d^a$



^{*a*} Unless indicated otherwise, all reactions were performed with C₆₀ (0.05 mmol) and 1 (0.5 mmol) in ODCB (6 mL). ^{*b*} Refers to the temperature of oil bath. ^{*c*} Isolated yield; those in parentheses were based on consumed C₆₀. ^{*d*} 1 mmol of **1a** was used. ^{*e*} *Trans/cis* ratio = 1 : 3.6. ^{*f*} *Trans/cis* ratio = 1.1 : 1. ^{*g*} *Cis* isomer only.

 C_{60} with **1a–1d** are listed in Table 1. It should be pointed out that the reactions with both glycine and alanine were much more sluggish and required higher temperature than those with phenylalanine and phenylglycine, probably due to the lower solubility of the former two amino acids.

Products 2a-2d are known compounds, and their identities were confirmed by comparison of their spectra with those reported in the literature.¹²⁻¹⁴ The structures of adducts 2a-2dare the same as those of products formed by the Prato reaction of C₆₀, **1a-1d** and the aldehydes generated *in situ* from **1a-1d** *via* the apparent elimination of the COOH and NH₂ groups. It should be noted that there are *trans* and *cis* isomers depending on the different orientation of the two R substituents when $R \neq H$. Products **2b** and **2c** existed as a mixture of *trans* and *cis* isomers while product **2d** consisted of only the *cis* isomer. The exact same phenomenon was also observed for the corresponding Prato reactions.¹³

Secondly, typical *N*-substituted amino acids, *i.e.*, *N*-methylglycine (sarcosine) (**3a**), *N*-ethylglycine (**3b**), *N*-benzylglycine (**3c**) and *N*-phenylglycine (**3d**), were chosen to react with C_{60} in refluxing ODCB under aerobic and dark conditions. The reaction of C_{60} with **3b** and **3c** afforded two types (**4b**, **5b** and **4c**, **5c**) of monoadducts, whereas that with **3a** and **3d** provided only one type (**4a** and **4d**) of monoadducts. The reaction conditions and product yields for the reaction of C_{60} with **3a–3d** are listed in Table 2.

Adducts **4a–4d**, **5b** and **5c** also prove to be fulleropyrrolidines. Among them, products **4a**,¹² **4b**,¹⁵ **4c**¹⁶ and **5c**¹⁷ are known compounds, and their structures are confirmed by comparison of their spectral data with those reported previously. The HRMS spectra of **4d** and **5b** showed correct molecular ions. The ¹H NMR spectrum of **4d** exhibited the signals for a phenyl

Table 2 Reaction conditions and product yields for the reaction of C_{60} with $3a-3d^{a}$

	5 = R'CH ₂
--	---------------------------------

Substrate 3	Reaction time (h)	Yield of 4^{b} (%)	Yield of 5^b (%)
`Ŋ^CO₂H	10	30 (81)	_
3a			
∕_N^CO₂H	2.5	33 (58)	11 (19)
3b			
	28	22 (45)	23 (47)
3c			
Ph_N_CO ₂ H	1.5	36 (71)	_
_c 3d ^c			

^{*a*} Unless indicated otherwise, all reactions were performed with C_{60} (0.05 mmol) and **1** (0.5 mmol) in ODCB (6 mL) at 180 °C. ^{*b*} Isolated yield; those in parentheses were based on consumed C_{60} . ^{*c*} 0.1 mmol of **3d** was used.

group and a singlet at 5.14 ppm for the two methylene groups. The ¹³C NMR spectrum of **4d** displayed 16 lines for the 58 sp²carbons of the C_{60} skeleton along with a peak at 69.23 ppm for the two sp³-carbons of the C_{60} core, consistent with its C_{2v} molecular symmetry. The ¹H NMR spectrum of **5b** gave two doublets at 4.88 and 3.98 ppm with $J_{AB} = 9.2$ Hz for the CH₂ group attached to the fullerene cage, two double quartets at 3.50 (J = 12.0, 7.4 Hz) and 2.73 (J = 12.0, 7.0 Hz) ppm, a triplet at 1.53 ppm (J = 7.2 Hz) for the ethyl group, a quartet at 4.02 ppm (J = 6.3 Hz), and a doublet at 1.95 ppm (J = 6.3 Hz) for the CHCH₃ moiety. The ¹³C NMR spectrum of **5b** revealed 49 peaks in the 136–157 ppm range for the 58 sp²-carbons of the C_{60} cage and two peaks at 75.89 and 69.20 ppm for the two sp³-carbons of the C_{60} skeleton, consistent with its C_1 molecular symmetry. Adducts 4a-4d appeared to be the Prato reaction products of C₆₀, 3 and CH₂O generated in situ from 3 through the apparent extrusion of the COOH and RNH moieties. Meanwhile, **5b** and **5c** might be similarly formed by the reaction of C_{60} with 3 and R'CHO (when $R = R'CH_2$), which was produced via an oxidative cleavage of the R'CH₂-N bond of 3.

Finally, the corresponding *N*-substituted amino acid esters including sarcosine ethyl ester (**6a**), *N*-ethylglycine ethyl ester (**6b**), *N*-benzylglycine ethyl ester (**6c**) and *N*-phenylglycine ethyl ester (**6d**) were allowed to react with C₆₀ under the same conditions as for *N*-substituted amino acids. Although **6c** and **6d** were commercially available, **6a** and **6b** were supplied as hydrochlorides. The *in situ* generation of **6a** and **6b** from their hydrochlorides in the presence of Et₃N during the reaction with C₆₀ may cause hydrolysis of the amino acid ester hydrochlorides.⁷ Therefore, **6a** and **6b** were prepared by the neutralization of sarcosine ethyl ester hydrochloride and *N*-ethylglycine ethyl ester hydrochloride with dilute aqueous solution of K₂CO₃ prior to use. The reaction conditions and product yields for the reaction of C₆₀ with **6a–d** are listed in Table 3.

All adducts 7–10 were identified as fulleropyrrolidines. Among them, products 7a,¹⁷ 8a,^{4a} 9^{4a,17} and 10c¹⁸ are known compounds, and their structures are substantiated by comparison of their spectral data with those reported in the literature. Compounds 7b, 7c, 7d, 8b and 8c exhibited correct molecular weights in their high-resolution mass spectra and the expected chemical shifts as well as the splitting patterns for all protons in their ¹H NMR spectra. In the ¹³C NMR spectrum of 7b–7d, there were no more than 28 peaks in the 135–154 ppm range for the 58 sp²-carbons of the C₆₀ cage and one peak at ~71 ppm for the two sp³-carbons of the C₆₀ skeleton, consistent with their C₂ molecular symmetry. In comparison, 51 lines in the range of 136–157 ppm for the 58 sp²-carbons of the C₆₀ cage along with two lines at *ca*. 71 and 75/76 ppm for the two sp³-carbons of the C₆₀ skeleton were observed in the ¹³C NMR spectrum of 8b and 8c, agreeing with their C₁ molecular symmetry.

The reaction of C_{60} with amino acid esters was much more complex than that with the corresponding amino acids. Adducts 7 and 8 could be envisioned as the products from the reaction of C_{60} with 6 and aldehydes (EtO₂CCHO and R'CHO when R = R' CH₂), which were produced from apparent C–N oxidative cleavages of RNH–CH₂CO₂Et and R'CH₂–NHCH₂CO₂Et (when R = R'CH₂), respectively. Meanwhile, 9 and 10 might be formed from the reaction of C_{60} with the *in situ* generated aldehydes and NH₂CH₂CO₂Et. **Table 3** Reaction conditions and product yields for the reaction of C_{60} with $6a-6d^a$



^{*a*} Unless indicated otherwise, all reactions were performed with C_{60} (0.05 mmol) and **6** (0.5 mmol) in ODCB (6 mL) at 180 °C. ^{*b*} Isolated yield; those in parentheses were based on consumed C_{60} . ^{*c*} Trans isomer only. ^{*d*} 1.0 mmol of **6d** was used. ^{*e*} Trans/cis ratio = 1 : 3.6. ^{*f*} Trans/cis ratio = 1 : 3.5. ^{*g*} Trans/cis ratio = 1 : 3.3.

Primary and secondary amines can react with C_{60} thermally to give aminated products.^{8,19} At first glance, it was surprising that fulleropyrrolidines were obtained from the direct thermal reaction of C_{60} with amino acids or amino acid esters in the absence of a purposely added aldehyde under aerobic and dark conditions. Although the exact pathways for the observed products from the thermal reaction of C_{60} with amino acids **1** and **3** as well as amino acid esters **6** remain to be clarified, structural motifs of the isolated products were sufficient to allow us to deduce the plausible reaction mechanisms (Schemes 1–3). The mechanisms for the formation of all potential intermediates involving C–N bond cleavages of amino acids and amino acid esters are generalized in Schemes 1 and 2, and the subsequent 1,3-dipolar cycloaddition reactions with C_{60} affording fulleropyrrolidines are shown in Scheme 3.

Electron transfer from amino acids/amino acid esters to C_{60} gives cation radical **A** and anion radical C_{60}^{--} (step i),^{7,19} the latter transfers an electron to O_2 to afford O_2^{--} and regenerate C_{60} (step ii).²⁰ Hydrogen abstraction at the α -carbon adjacent to the carbonyl group of **A** by O_2^{--} produces iminium cation **B** and HO_2^{-} (step iii),^{4,7} subsequent proton transfer provides imine **C** and H_2O_2 (step iv). Under heating conditions, H_2O_2 can decompose to H_2O and O_2 . Hydrolysis of **C** generates amine **D** and carbonyl compound **E** (step v).^{7,21} When $R^3 = H$ in **E**, it eliminates CO_2 to give aldehyde **F** upon heating (step vi).²² Meanwhile, hydrogen abstraction at the α -carbon adjacent to R^4 (when $R^1 = R^4CH_2$) of **A** by O_2^{--} attains iminium cation **G** and HO_2^{-} (step vii). Proton transfer from **G** to HO_2^{-} furnishes imine

$$C_{60} + R^{1} \underset{M}{\overset{R^{2}}{\longrightarrow}} C_{02} R^{3} \xrightarrow{ET} C_{60}^{-\cdot} + R^{1} \underset{M}{\overset{H^{2}}{\longrightarrow}} C_{02} R^{3} \qquad (i)$$

$$C_{60}^{-\cdot} + O_{2} \xrightarrow{ET} C_{60} + O_{2}^{-\cdot} \qquad (ii)$$

$$R^{1}_{\underset{\mathbf{A}}{\overset{\mathbf{H}}{\xrightarrow{}}}} \overset{R^{2}}{\underset{\mathbf{A}}{\overset{\mathbf{H}}{\xrightarrow{}}}} + O_{2}^{-} \overset{\mathbf{H}}{\xrightarrow{}} R^{1}_{\underset{\mathbf{A}}{\overset{\mathbf{H}}{\xrightarrow{}}}} \overset{R^{2}}{\underset{\mathbf{A}}{\overset{\mathbf{H}}{\xrightarrow{}}}} + HO_{2}^{-} \quad (iii)$$

$$R^{1}_{\mathbf{N}} \stackrel{R^{2}}{\longrightarrow} CO_{2}R^{3} + HO_{2}^{-} \xrightarrow{\mathbf{R}} R^{1}_{\mathbf{N}} \stackrel{R^{2}}{\longrightarrow} CO_{2}R^{3} + H_{2}O_{2} \quad (iv)$$

$$\begin{array}{cccc} R^{1} & \stackrel{R^{2}}{\longrightarrow} & R^{1}NH_{2} & + & \stackrel{R^{2}}{\longrightarrow} & CO_{2}R^{3} \\ C & D & E \end{array}$$
 (v)

$$\overset{R^2}{\stackrel{\frown}{\longrightarrow}} \underset{E}{\overset{CO_2R^3}{\longrightarrow}} \overset{\text{when } R^3 = H}{\overset{\frown}{\longrightarrow}} R^2 CHO + CO_2$$
 (vi)

$$R^{1} \stackrel{\text{H}^{2}}{\stackrel{\text{H}^{2}}}{\stackrel{\text{H}^{2}}}\stackrel{\text{H}^{2}}{\stackrel{\text{H}^{2}}}\stackrel{\text{H}^{2}}{\stackrel{\text{H}^{2}}}\stackrel{\text{H}^{2}}\stackrel{\text{H}^{2}}\stackrel{\text{H}^{2}}\stackrel{\text{H}^{2}}\stackrel{\text{H}^{2}}\stackrel{\text{H}^{2}}\stackrel{\text{H}^{2}}\stackrel{\text{H}^{2}}\stackrel{\text{H}^{2}}}\stackrel{\text{H}^{2}}\stackrel{\text{$$

Scheme 1 Possible mechanisms for C–N bond cleavages of amino acids and amino acid esters.

$$\mathbb{R}^{1}_{\underset{\mathbf{H}}{\mathsf{N}}_{\mathsf{L}}} \xrightarrow{\mathbb{R}^{2}}_{\underset{\mathbf{H}}{\mathsf{N}}_{\mathsf{L}}} \xrightarrow{\mathbb{R}^{1}_{\underset{\mathbf{H}}{\mathsf{N}}_{\mathsf{M}}}} \mathbb{R}^{2}_{\underset{\mathbf{H}}{\mathsf{M}}_{\mathsf{M}}}} (xii)$$

$$\begin{array}{cccc} R^1 & \stackrel{H_2O}{\longrightarrow} & R^1 N H_2 + R^2 C HO \\ N & D & F \end{array}$$
 (xiv)

Scheme 2 Alternative pathway to generate aldehydes from amino acids.

H and H_2O_2 (step viii), subsequent hydrolysis of **H** produces aldehyde I and amino acid/amino acid ester J (step ix).^{7,21}

An alternative pathway leading to the formation of aldehydes R^2CHO (F) from amino acids is shown in Scheme 2. Electron

(1) For amino acids **1a-1d**,
$$R^1 = R^3 = H$$
, $R^2 = R$

$$C_{60} + RCHO + \underset{H_2N}{R} \xrightarrow{R} CO_2H \xrightarrow{\Delta} R^{R} NH (xv)$$

(2) For amino acids **3a-3d**, $R^1 = R$, $R^2 = R^3 = H$

$$C_{60} + CH_2O + R N_3 CO_2H \rightarrow I N R (xvi)$$

when R = R'CH₂

$$C_{60} + R'CHO + \frac{R}{N} CO_2 H \xrightarrow{R'} N R \quad (xvii)$$

$$I' \qquad 3$$

(3) For amino acid esters **6a-6d**, $R^1 = R$, $R^2 = H$, $R^3 = Et$

$$C_{60} + O CO_{2}Et + R H CO_{2}Et \rightarrow CO_{2}Et$$
when R = R'CH₂

$$C_{60} + R'CHO + R H CO_{2}Et \rightarrow R' CO_{2}Et \rightarrow R' CO_{2}Et$$

$$C_{60} + R'CHO + R H_{2}N CO_{2}Et \rightarrow R' CO_{2}Et$$

$$C_{60} + O CO_{2}Et + H_{2}N CO_{2}Et \rightarrow R' CO_{2}Et$$

$$C_{60} + R'CHO + H_{2}N CO_{2}Et \rightarrow R' CO_{2}Et$$

$$C_{60} + R'CHO + H_{2}N CO_{2}Et \rightarrow R' CO_{2}Et$$

$$C_{60} + R'CHO + H_{2}N CO_{2}Et \rightarrow R' CO_{2}Et$$

$$C_{60} + R'CHO + H_{2}N CO_{2}Et \rightarrow R' CO_{2}Et$$

$$C_{60} + R'CHO + H_{2}N CO_{2}Et \rightarrow R' CO_{2}Et$$

$$C_{60} + R'CHO + H_{2}N CO_{2}Et \rightarrow R' CO_{2}Et$$

Scheme 3 1,3-Dipolar cycloaddition reactions with C_{60} affording fulleropyrrolidines.

transfer from a zwittionic amino acid to C_{60} affords the cationic radical **K** and $C_{60}^{-..}$ (step x),^{4b} followed by the same step ii to form $O_2^{-..}$ and regenerate C_{60} (step ii). Proton transfer from **K** to $O_2^{-..}$ furnishes radical **L** and HO₂. (step xi). Loss of CO₂ from **L** gives radical **M** (step xii),²³ from which a hydrogen abstraction by HO₂. provides imine **N** (step xiii). Hydrolysis of imine **N** produces amine R¹NH₂ (**D**) and R²CHO (**F**) (step xiv).^{7,21}

For amino acids **1a–1d**, $R^1 = R^3 = H$, $R^2 = R$, aldehyde RCHO (**F'**) is generated through steps i–vi and/or steps x–xiv. The 1,3-dipolar reaction of C₆₀ with **1a–1d** and aldehyde **F'** leads to fulleropyrrolidines **2a–2d** (step xv). We previously found that the *cis/trans* isomeric ratio of **2b** and **2c** was heavily dependent on the reaction conditions.¹³ The reaction of C₆₀ with **1b** and acetaldehyde in refluxing toluene (110 °C) for 27 h led to 21% of *trans-***2b** and 5% of *cis-***2b**, and the isolated isomeric mixture of **2b** was then heated in refluxing ODCB (180 °C) for 12 h to afford a mixture dominated by the *cis* isomer.¹³ The

direct reaction of C_{60} with **1b** at 200 °C for 2 d gave *cis*-**2b** as the major product (Table 1, entry 2). When the same reaction was conducted at lower temperature (180 °C for 4 d), *trans*-**2b** and *cis*-**2b** were obtained in 8% and 6% yields, respectively. A similar behavior was also observed for the reaction with **1c**. Thus, a higher reaction temperature tended to give a higher *cis/ trans* isomeric ratio in our current cases. These experimental results support the above proposed mechanisms for the formation of aldehydes from amino acids, followed by 1,3-dipolar cycloaddition with C₆₀ to produce fulleropyrrolidines.¹³

For amino acids **3a–3d**, $R^1 = R$, $R^2 = R^3 = H$, formaldehyde CH₂O is formed via steps i-vi and/or steps x-xiv, while aldehyde R'CHO (CH₂O, CH₃CHO and PhCHO for 3a, 3b and 3c, respectively) can be produced through steps vii–ix when R = R'CH₂. The cycloaddition reaction of C₆₀ with **3a-3d** and CH₂O gives 4a-4d (step xvi), while the reaction of C₆₀ with 3a-3c and R'CHO provides 5a-5c (step xvii). In the case of 3a, product 5a was the same as adduct 4a. Therefore, more pathways leading to 4a may operate simultaneously. Similar steps vii-ix are impossible for 3d, of which the phenyl group is directly attached to the nitrogen atom. As a result, the thermal reaction of C_{60} with 3d afforded 4d only. It should be noted that it was possible for H_2NCH_2COOH to react with $CH_2O/R'CHO$ and C_{60} to give the corresponding N-unsubstituted fulleropyrrolidines. However, no more than a trace amount of, if any, N-unsubstituted fulleropyrrolidines could be isolated from the thermal reaction of C₆₀ with 3a-3d because the in situ generated CH₂O and R'CHO (I') preferred to react with a large excess of amino acids 3a-3d rather than the relatively small amount of the in situ formed H₂NCH₂COOH to furnish an azomethine ylide, which then underwent [2 + 3] cycloaddition reaction with C₆₀ to provide 4a-4d and 5b-5c, respectively.

For amino acid esters **6a–6d**, $R^1 = R$, $R^2 = H$, $R^3 = Et$, EtO₂CCHO (**E'**) and R'CHO (**I'**, when $R = R'CH_2$) were obtained *via* steps i–v and vii–ix, respectively. Substrate **6d** bearing the Ph–N moiety cannot undergo steps vii–ix, and thus is unable to produce the corresponding R'CHO (**I'**) and NH₂CH₂CO₂Et (**J'**). The reaction of C₆₀ with **6a–6d** and **E'** affords **7a–7d** (step xviii), and that with **6a–6c** and **I'** furnishes **8a–8c** (step xix). For **6a–6c**, C₆₀ can react with the *in situ* generated **J'** and EtO₂CCHO or R'CHO to provide **9** or **10** (steps xx and xxi), respectively. As expected, **9** was indeed obtained in all cases of **6a–6c**. However, only **10c** could be isolated because much more benzaldehyde and **J'** from **6c** might be produced than aldehydes (formaldehyde and acetaldehyde) and **J'** from **6a** and **6b** (*vide infra*).

To better understand the product distribution for the thermal reaction of C_{60} with amino acids or amino acid esters, we performed additional experiments. Control experiments showed that the reaction of C_{60} with 1 equiv. of **3a**, 1 equiv. of NH₂CH₂CO₂H and 1 equiv. of CH₂O at 180 °C for 7 min gave **4a** in 18% yield along with only a trace amount of **2a** (Scheme 4). In comparison, the reaction of C_{60} with 1 equiv. of **3c**, 1 equiv. of NH₂CH₂CO₂H and 1 equiv. of CH₂O at 180 °C for 7 min afforded **4c** in 16% yield together with a very small amount of **2a** (Scheme 5); meanwhile the reaction of C_{60} with 1 equiv. of PhCHO at 180 °C for 7 min afforded **5c** in 27% yield along with a trace amount of the corresponding *N*-unsubstituted fulleropyrrolidine



Scheme 4 Thermal reaction of C_{60} with 1a, 3a and CH_2O .



Scheme 5 Thermal reaction of C_{60} with 1a, 3c and CH_2O .



Scheme 6 Thermal reaction of C_{60} with 1a, 3c and PhCHO.

(Scheme 6). These experiments indicated that the reactivity of *N*-substituted glycine was higher than that of glycine. Therefore, it is understandable that we could not identify the *N*-unsubstituted fulleropyrrolidines from the reaction of C_{60} with **3a–3d** after taking into account the very small amount of the *in situ* generated glycine relative to that of **3a–3d**.

It was reported that fulleropyrrolidine 4a was unexpectedly formed from the reaction of C₆₀ with sarcosine and gossypol.²⁴ Product 4a was also obtained from the reaction of C₆₀ with sarcosine and 4,6-dimethoxysalicylaldehyde^{25a} or hydroxyacetaldehyde.^{25b} However, the authors did not provide any explanation of how 4a was produced in their cases. Our current work shows that 4a can be formed directly from the reaction of C_{60} with sarcosine via the aforementioned mechanisms, thus elucidating the origin for the formation of 4a reported in the literature.^{24,25} Control experiments showed that the reaction of C₆₀ with 2 equiv. of 3a under typical Prato reaction conditions (refluxing toluene, 2 h) afforded only a trace amount of 4a. The same reaction in refluxing chlorobenzene for 2 h still gave a very small amount of 4a. However, when the reaction time was prolonged to 48 h, as in the reaction of C_{60} with sarcosine and hydroxyacetaldehyde,^{25b} 4a was obtained in 20% yield along with 78% of unreacted C_{60} . These results suggest that although the formation of fulleropyrrolidines from the direct reaction of C₆₀ with amino acids under typical Prato reaction conditions is negligible, a

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significant amount of fulleropyrrolidines can be generated at high reaction temperature and long reaction time.

It is obvious from the above results and discussion that both N-substituted glycines (R'CH₂NHCH₂CO₂H) and N-substituted glycine esters (R'CH2NHCH2CO2Et) could undergo C-N bond cleavage at either side of the NH group. To further explore the innate nature of the C-N bond cleavage and product distribution, we have conducted additional control experiments. We found that the reaction of C₆₀ with 1 equiv. of CH₃CH₂NHCH₂CO₂H (3b), 1 equiv. of CH₂O and 1 equiv. of CH₃CHO at 180 °C for 6 min produced 4b and 5b in 23% and 8% yields, respectively (Scheme 7). The product ratio of 4b/5b was 3:1 and happened to be the same as that from the direct thermal reaction of C_{60} with **3b** (Table 2, entry 2), suggesting that the same molar amount of CH₂O and CH₃CHO was generated from 3b. In comparison, the reaction of C₆₀ with 1 equiv. of CH₃CH₂-NHCH₂CO₂Et (6b), 1 equiv. of EtO₂CCHO and 1 equiv. of CH₃CHO at 180 °C for 10 min produced 7b and 8b in 38% and 3% yields, respectively (Scheme 8). Combined with the product yields (7b and 8b in 31% and 13% yields, respectively) for the thermal reaction of C_{60} with **6b** (Table 3, entry 2), it could be deduced that EtO₂CCHO and CH₃CHO were generated in situ from **6b** in a molar ratio of 1:5. Thus, the CH₃CH₂–NH bond was more easily to break via steps vii-ix in Scheme 1 than the NH-CH₂CO₂Et bond via steps i-v in Scheme 1. If 3b in its thermal reaction with C₆₀ proceeded only via the same routes as 6b, it was expected that CH₂O and CH₃CHO should be formed also in a molar ratio of 1:5, in sharp contrast to the observed ratio of 1:1. Consequently, other routes, *i.e.*, steps x-xiv, must operate and contribute to the formation of CH2O. The same processes shown in Scheme 2 should also occur for 1a-1d, 3a, 3c and **3d** to provide the corresponding aldehydes.

On the other hand, the reaction of C_{60} with 1 equiv. of PhCH₂NHCH₂CO₂H (**3c**), 1 equiv. of CH₂O and 1 equiv. of PhCHO at 180 °C for 4 min afforded **4c** in 22% yield together with a trace amount of **5c** (Scheme 9), while the reaction of C_{60}



Scheme 7 Thermal reaction of C_{60} with **3b**, CH_2O and CH_3CHO .



Scheme 8 Thermal reaction of C_{60} with 6b, EtO₂CCHO and CH₃CHO.

Scheme 9 Thermal reaction of C_{60} with 3c, CH_2O and PhCHO.



Scheme 10 Thermal reaction of C_{60} with 6c, EtO₂CCHO and PhCHO.

with 1 equiv. of PhCH₂NHCH₂CO₂Et (6c), 1 equiv. of EtO₂CCHO and 1 equiv. of PhCHO at 180 °C for 3 min afforded 7c in 31% yield along with a trace amount of 8c (Scheme 10). The observation of 5c (23%) and 8c (10%) in the direct reaction of C₆₀ with 3c and 6c (entry 3, Tables 2 and 3), respectively, suggested that the PhCH2-NH bond was much more facile to cleave than the NH-CH2CO2H and NH-CH2CO2Et bonds in substrates 3c and 6c, respectively, and thus provided significantly more PhCHO than CH₂O (for 3c)/EtO₂CCHO (for 6c). The reactivity for the hydrogen abstraction in step vii should follow the order of CH₃–NH < CH₃CH₂–NH < PhCH₂–NH. Therefore, more benzaldehyde is expected to form from 3c than acetaldehyde from 3b due to the more reactive benzylic position of the former. It is reasonable to assume that the amount of the in situ generated CH2O via steps i-vi is not strongly affected by the N-alkyl group of 3b and 3c. Consequently, the product ratio of 5c/4c is expected to be higher than that of 5b/4b, fully consistent with our experimental results (Table 2, entries 2 and 3). Similar to amino acids 3, the formation of EtO₂CCHO via steps i-v is little affected by the N-alkyl group of 6, while the production of R'CHO via steps vii-ix should be more favorable for 6c than that for 6b, which in turn is more facile than for 6a, corroborating the observed product distribution of 8c/7c > 8b/7b >8a/7a (see Table 3). In addition, product 10 was observed only in the case of 6c, further supporting that much more PhCHO from 6c was produced than CH₂O from 6a and CH₃CHO from 6b, respectively.

Conclusions

Fulleropyrrolidines are commonly obtained by the Prato reaction, *i.e.*, the reaction of C_{60} with an amino acid or amino acid ester with an aldehyde. In the present work, we have demonstrated that the direct thermal reaction of C_{60} with amino acids or amino acid esters in refluxing ODCB without the purposely added aldehydes under aerobic and dark conditions can also afford

fulleropyrrolidines. Plausible reaction mechanisms involving unusual C–N bond cleavages are proposed to explain the formation of intermediates from the employed amino acids and amino acid esters, and subsequent addition of the azomethine ylides to C_{60} furnishes fulleropyrrolidines. All isolated products from the reaction mixtures can be well explained by the proposed reaction mechanisms. Control experiments shed light on the innate nature of the C–N bond cleavages of amino acids and amino acid esters and support the proposed reaction mechanisms. Our current work also clarifies the origin of the unexpected formation of *N*-methylfulleropyrrolidine from the reaction of C_{60} with sarcosine and gossypol, 4,6-dimethoxysalicylaldehyde or hydroxyacetaldehyde reported in the literature.

Experimental

Thermal reaction of C₆₀ with glycine

A 25 mL tube containing a mixture of C_{60} (36.1 mg, 0.05 mmol) and glycine (76.2 mg, 1.0 mmol) in ODCB (6 mL) was wrapped with aluminum foil, sealed up, and then heated in an oil bath preset at 200 °C for 24 h (monitored by TLC). After removal of the solvent *in vacuo*, flash chromatography of the residue on a silica gel column with carbon disulfide as the eluent to afford unreacted C_{60} (22.7 mg, 63%), then with a mixture of carbon disulfide and dichloromethane to give fulleropyrrolidine $2a^{12}$ (7.4 mg, 19%). ¹H NMR (400 MHz, $CS_2/CDCl_3$): $\delta = 4.78$ (s, 4H).

Thermal reaction of C₆₀ with alanine

By the same procedure as above, the reaction of C₆₀ (36.0 mg, 0.05 mmol) with alanine (46.0 mg, 0.52 mmol) at 200 °C for 48 h gave unreacted C₆₀ (13.4 mg, 37%) and fulleropyrrolidine **2b**¹³ (13.8 mg, 35%) as a mixture of *cis* and *trans* isomers in a ratio of 3.6 : 1. *cis*-**2b**: ¹H NMR (400 MHz, CS₂/CDCl₃): δ = 4.79 (q, *J* = 6.6 Hz, 2H), 2.02 (d, *J* = 6.6 Hz, 6H). *trans*-**2b**: ¹H NMR (400 MHz, CS₂/CDCl₃): δ = 5.06 (q, *J* = 6.8 Hz, 2H), 2.04 (d, *J* = 6.8 Hz, 6H).

Thermal reaction of C₆₀ with phenylalanine

By the same procedure as above, the reaction of C₆₀ (35.9 mg, 0.05 mmol) with phenylalanine (82.2 mg, 0.50 mmol) at 180 °C for 12 h gave unreacted C₆₀ (25.6 mg, 71%) and fulleropyrrolidine $2c^{13}$ (8.7 mg, 19%) as a mixture of *cis* and *trans* isomers in a ratio of 1 : 1.1. *cis*-2c: ¹H NMR (400 MHz, CS₂/CDCl₃): $\delta =$ 7.40 (d, J = 7.2 Hz, 4H), 7.30 (t, J = 7.5 Hz, 4H), 7.21 (t, J = 7.3 Hz, 2H), 4.74 (dd, J = 10.6, 3.0 Hz, 2H), 3.87 (dd, J = 13.4, 3.0 Hz, 2H), 3.35 (dd, J = 13.4, 10.6 Hz, 2H), 2.63 (br s, 1H). *trans*-2c: ¹H NMR (400 MHz, CS₂/CDCl₃): $\delta =$ 7.27–7.24 (m, 10H), 5.11 (dd, J = 11.1, 3.5 Hz, 2H), 3.68 (dd, J = 13.3, 3.5 Hz, 2H), 3.50 (dd, J = 13.3, 11.1 Hz, 2H), 2.63 (br s, 1H).

Thermal reaction of C₆₀ with phenylglycine

By the same procedure as above, the reaction of C_{60} (36.2 mg, 0.05 mmol) with phenylglycine (75.4 mg, 0.50 mmol) at 180 °C

for 10 h gave unreacted C₆₀ (16.1 mg, 44%) and fulleropyrrolidine **2d**¹³ (17.2 mg, 37%); ¹H NMR (400 MHz, CS₂/CDCl₃): δ = 7.95 (d, *J* = 7.2 Hz, 4H), 7.39 (t, *J* = 7.5 Hz, 4H), 7.30 (t, *J* = 7.3 Hz, 2H), 5.95 (s, 2H), 3.11 (br s, 1H).

Thermal reaction of C₆₀ with sarcosine

By the same procedure as above, the reaction of C₆₀ (36.0 mg, 0.05 mmol) with sarcosine (44.3 mg, 0.50 mmol) at 180 °C for 10 h gave unreacted C₆₀ (22.6 mg, 63%) and fulleropyrrolidine **4a**¹² (11.7 mg, 30%). ¹H NMR (300 MHz, CS₂/CDCl₃): δ = 4.37 (s, 4H), 2.97 (s, 3H).

Thermal reaction of C₆₀ with N-ethylglycine

By the same procedure as above, the reaction of C_{60} (36.1 mg, 0.05 mmol) with N-ethylglycine (52.2 mg, 0.51 mmol) at 180 °C for 2.5 h gave unreacted C₆₀ (15.4 mg, 43%) and a mixture of 4b and 5b, which was further separated by recycling HPLC (10 × 250 mm Cosmosil Buckyprep column; flow rate 3 mL min⁻¹; injection volume 5 mL; toluene as eluent) to afford fulleropyrrolidines **4b**¹⁵ (13.1 mg, 33%) and **5b** (4.5 mg, 11%). **4b**: ¹H NMR (400 MHz, CS₂/CDCl₃): δ = 4.40 (s, 4H), 3.15 (q, J = 7.2 Hz, 2H), 1.55 (t, J = 7.2 Hz, 3H). **5b**: ¹H NMR (400 MHz, $CS_2/CDCl_3$): $\delta = 4.88$ (d, J = 9.2 Hz, 1H), 4.02 (q, J = 6.3 Hz, 1H), 3.98 (d, J = 9.2 Hz, 1H), 3.50 (dq, J = 12.0, 7.4 Hz, 1H), 2.73 (dq, J = 12.0, 7.0 Hz, 1H), 1.95 (d, J = 6.3 Hz, 3H), 1.53 (t, J = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CS₂/DMSOd₆ with Cr(acac)₃ as relaxation reagent, all 1C unless indicated): $\delta = 156.69, 154.58, 154.23, 153.42, 147.34$ (2C), 146.98, 146.84, 146.70, 146.43, 146.39, 146.28, 146.26, 146.22, 146.17, 146.06 (2C), 145.92, 145.75, 145.65 (3C), 145.52, 145.43, 145.38 (2C), 145.33, 145.28, 144.90, 144.77, 144.52 (2C), 143.28, 143.16, 142.80, 142.77, 142.74, 142.72, 142.38 (3C), 142.34, 142.29, 142.28, 142.20, 142.18, 142.17, 142.06, 141.85, 141.81, 140.39, 140.32 (2C), 139.88, 137.72, 136.68, 136.13, 135.93, 75.89 (sp³-C of C₆₀), 71.77 (NCHCH₃), 69.20 (sp³-C of C₆₀), 66.49 (CH₂N), 46.64 (NCH₂CH₃), 17.35 (CHCH₃), 13.76 (NCH₂CH₃); FT-IR v/cm⁻¹ (KBr) 2962, 2923, 1506, 1451, 1426, 1377, 1332, 1186, 766, 699, 572, 525; UV-vis (CHCl₃) $\lambda_{\text{max}}/\text{nm}$ (lg ε) 257 (4.92), 323 (4.40), 431 (3.43), 704 (2.55); HRMS (MALDI TF-ICR): *m/z* calcd for C₆₅H₁₁N: 805.0891; found: 805.0895.

Thermal reaction of C₆₀ with N-benzylglycine

By the same procedure as above, the reaction of C_{60} (36.1 mg, 0.05 mmol) with *N*-benzylglycine (82.5 mg, 0.50 mmol) at 180 °C for 28 h gave unreacted C_{60} (18.3 mg, 51%), fulleropyrrolidines **4c**¹⁶ (9.4 mg, 22%) and **5c**¹⁷ (10.7 mg, 23%). **4c**: ¹H NMR (300 MHz, CS₂/CDCl₃): δ = 7.69 (d, *J* = 7.1 Hz, 2H), 7.46 (t, *J* = 7.3 Hz, 2H), 7.36 (t, *J* = 7.3 Hz, 1H), 4.43 (s, 4H), 4.30 (s, 2H); ¹³C NMR (75 MHz, CS₂/CDCl₃ with Cr(acac)₃ as relaxation reagent): δ = 154.76 (4C), 147.13 (2C), 146.09 (4C), 145.90 (8C), 145.52 (2C), 145.30 (4C), 145.13 (4C), 144.41 (4C), 142.95 (2C), 142.49 (4C), 142.10 (4C), 141.92 (4C), 141.74 (4C), 140.04 (4C), 137.77 (1C, aryl C), 136.15 (4C), 128.68 (2C, aryl C), 128.65 (2C, aryl C), 127.55 (1C, aryl C),

70.54 (2C, sp³-C of C₆₀), 67.47 (2C, CH₂), 58.82 (1C, NCH₂). **5c**: ¹H NMR (400 MHz, CS₂/CDCl₃): δ = 7.93 (br s, 2H), 7.68 (d, J = 7.4 Hz, 2H), 7.50–7.43 (m, 4H), 7.40–7.33 (m, 2H), 5.21 (s, 1H), 4.86 (d, J = 9.4 Hz, 1H), 4.59 (d, J = 13.4 Hz, 1H), 4.16 (d, J = 9.4 Hz, 1H), 3.67 (d, J = 13.4 Hz, 1H); ¹³C NMR (75 MHz, CS₂/CDCl₃ with Cr(acac)₃ as relaxation reagent, all 1C unless indicated): $\delta = 156.32$, 154.04, 153.39, 153.33, 147.29 (2C), 146.79, 146.40, 146.29, 146.26, 146.20, 146.17, 146.14, 146.12, 146.09, 145.93 (2C), 145.74, 145.53 (2C), 145.51, 145.49, 145.35, 145.30, 145.25 (2C), 145.22, 145.13, 144.70, 144.62, 144.39, 144.36, 143.15, 142.98, 142.67, 142.58, 142.55 (2C), 142.34, 142.24, 142.14, 142.12, 142.10, 142.07, 142.01, 141.99, 141.92, 141.82, 141.64, 141.54, 140.18, 140.14, 139.89, 139.45, 137.85 (aryl C), 136.99 (aryl C), 136.84, 136.51, 135.92, 135.71, 129.46 (2C, aryl C), 128.86 (2C, aryl C), 128.82 (2C, aryl C), 128.74 (2C, aryl C), 128.66 (aryl C), 127.56 (aryl C), 81.54 (CHPh), 77.37 (sp³-C of C₆₀), 68.68 (sp³-*C* of C₆₀), 66.69 (*C*H₂), 56.84 (N*C*H₂).

Thermal reaction of C₆₀ with N-phenylglycine

By the same procedure as above, the reaction of C_{60} (36.0 mg, 0.05 mmol) with N-phenylglycine (16.2 mg, 0.11 mmol) at 180 °C for 1.5 h gave unreacted C₆₀ (17.6 mg, 49%) and fulleropyrrolidine 4d (15.1 mg, 36%). ¹H NMR (300 MHz, $CS_2/$ CDCl₃): δ = 7.41 (t, J = 7.4 Hz, 2H), 7.25 (d, J = 7.7 Hz, 2H), 7.00 (t, J = 7.3 Hz, 1H), 5.14 (s, 4H); ¹³C NMR (75 MHz, CS₂/ CDCl₃ with Cr(acac)₃ as relaxation reagent): $\delta = 153.77$ (4C), 146.82 (2C), 145.74 (4C), 145.54 (4C), 145.30 (4C), 145.09 (2C), 145.02 (4C), 144.75 (4C), 144.01 (4C), 142.57 (2C), 142.12 (4C), 141.67 (4C), 141.54 (4C), 141.37 (4C), 139.72 (4C), 135.63 (4C), 129.14 (2C, aryl C), 120.18 (1C, aryl C), 116.10 (2C, aryl C), 69.23 (2C, sp³-C of C₆₀), 62.63 (CH₂N); FT-IR v/cm⁻¹ (KBr) 2921, 1597, 1501, 1466, 1427, 1353, 1216, 1187, 751, 687, 558, 526; UV-vis (CHCl₃) λ_{max} /nm (lg ε) 256 (5.05), 308 (4.55), 430 (3.53), 703 (2.09); HRMS (MALDI TF-ICR): *m/z* calcd for C₆₈H₉N: 839.0735; found: 839.0732.

Thermal reaction of C₆₀ with sarcosine ethyl ester

By the same procedure as above, the reaction of C_{60} (36.0 mg, 0.05 mmol) with sarcosine ethyl ester (57.3 mg, 0.49 mmol) at 180 °C for 4 h gave unreacted C₆₀ (16.2 mg, 45%), $7a^{17}$ (11.0 mg, 24%), 8a^{4a} (3.9 mg, 9%) and 9 as a mixture of cisisomer^{4a,17} (1.1 mg, 2.4%) and *trans*-isomer^{4a} (0.3 mg, 0.7%). **7a**: ¹H NMR (400 MHz, CDCl₃): δ = 5.82 (s, 2H), 4.42 (dq, J = 10.8, 7.1 Hz, 2H), 4.33 (dq, J = 10.8, 7.1 Hz, 2H), 3.10 (s, 3H), 1.28 (t, J = 7.1 Hz, 6H). 8a: ¹H NMR (300 MHz, CS₂/CDCl₃): δ = 4.96 (d, J = 9.5 Hz, 1H), 4.79 (s, 1H), 4.41 (dq, J = 10.8, 7.2 Hz, 1H), 4.31 (dq, J = 10.8, 7.1 Hz, 1H), 4.25 (d, J = 9.5 Hz, 1H), 3.02 (s, 3H), 1.26 (t, J = 7.1 Hz, 3H). *cis*-9: ¹H NMR (400 MHz, CDCl₃): δ = 5.51 (d, J = 13.6 Hz, 2H), 4.48 (t, J = 13.6 Hz, 1H, NH), 4.43 (dq, J = 10.8, 7.1 Hz, 2H), 4.32 (dq, J = 10.8, 7.1 Hz, 2H), 1.26 (t, J = 7.1 Hz, 6H). trans-9: ¹H NMR (400 MHz, CDCl₃): δ = 6.01 (d, J = 9.0 Hz, 2H), 4.42 (dq, J = 10.8, 7.1 Hz, 2H), 4.33 (dq, J = 10.8, 7.1 Hz, 2H), 4.21 (t, J = 9.0 Hz, 1H, NH), 1.28 (t, J = 7.1 Hz, 6H); ¹³C NMR (75 MHz, $CS_2/CDCl_3$ with $Cr(acac)_3$ as relaxation reagent, all 2C unless

indicated): $\delta = 169.17$ (C=O), 153.30, 149.98, 146.47, 145.69, 145.65, 145.35, 145.33, 145.18, 144.87, 144.83 (4C), 144.75, 144.66, 144.61, 143.79, 143.62, 142.46, 142.09, 142.05, 141.55 (6C), 141.49, 141.26, 141.18, 139.41, 138.94, 135.86, 134.92, 75.70 (sp³-C of C₆₀), 73.09 (CHCO₂), 61.24 (OCH₂CH₃), 13.91 (OCH₂CH₃); FT-IR ν /cm⁻¹ (KBr) 2923, 1739, 1566, 1520, 1429, 1373, 1279, 1199, 1027, 942, 803, 751, 592, 527; UV-vis (CHCl₃) λ_{max} nm (lg ε) 251 (4.97), 325 (4.50), 428 (3.47), 693 (2.62); HRMS (MALDI TF-ICR): *m*/*z* calcd for C₆₈H₁₃NO₄: 907.0845; found: 907.0842.

Thermal reaction of C₆₀ with *N*-ethylglycine ethyl ester

By the same procedure as above, the reaction of C_{60} (36.1 mg, 0.05 mmol) with N-ethylglycine ethyl ester (67.8 mg, 0.52 mmol) at 180 °C for 2 h gave unreacted C₆₀ (16.2 mg, 45%), 7b (14.5 mg, 31%), 8b (5.6 mg, 13%) and 9 as a mixture of cis-isomer^{4a,17} (1.4 mg, 3.0%) and trans-isomer^{4a} (0.4 mg, 0.9%). **7b**: ¹H NMR (400 MHz, CDCl₃): δ = 5.92 (s, 2H), 4.41 (dq, J = 10.8, 7.1 Hz, 2H), 4.31 (dq, J = 10.8, 7.1 Hz, 2H), 3.52 (dq, J = 11.8, 7.5 Hz, 1H), 3.16 (dq, J = 11.8, 7.0 Hz, 1H), 1.57 (t, J = 7.3 Hz, 3H), 1.28 (t, J = 7.1 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃ with Cr(acac)₃ as relaxation reagent, all 2C unless indicated): $\delta = 170.56$ (C=O), 153.85, 151.14, 147.54, 146.51, 146.45, 146.19 (4C), 146.14, 145.95, 145.74 (4C), 145.61, 145.44, 145.39, 144.66, 144.62, 143.20, 142.84, 142.76, 142.33, 142.25, 142.11, 142.02, 141.95, 141.87, 140.12, 139.68, 137.28, 136.61, 74.21 (CHCO₂), 71.64 (sp³-C of C₆₀), 61.63 (OCH₂CH₃), 43.40 (1C, NCH₂CH₃), 14.45 (OCH₂CH₃), 13.71 (1C, NCH₂CH₃). FT-IR v/cm⁻¹ (KBr) 2970, 1727, 1509, 1427, 1366, 1337, 1263, 1166, 1093, 1012, 802, 573, 523; UV-vis (CHCl₃) λ_{max} /nm (lg ε) 256 (5.00), 311 (4.52), 429 (3.53), 695 (2.84); HRMS (MALDI TF-ICR): m/z calcd for C₇₀H₁₇NO₄: 935.1158; found: 935.1165. 8b: ¹H NMR (400 MHz, CS₂/ CDCl₃): $\delta = 5.56$ (s, 1H), 5.40 (q, J = 6.4 Hz, 1H), 4.35 (dq, J =10.8, 7.1 Hz, 1H), 4.27 (dq, J = 10.8, 7.1 Hz, 1H), 3.43 (dq, J = 12.0, 7.5 Hz, 1H), 3.10 (dq, J = 12.0, 7.0 Hz, 1H), 1.90 (d, J = 6.4 Hz, 3H), 1.52 (t, J = 7.3 Hz, 3H), 1.29 (t, J = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CS₂/DMSO-d₆ with Cr(acac)₃ as relaxation reagent, all 1C unless indicated): $\delta = 169.90$ (C=O), 156.25, 154.46, 153.92, 151.48, 147.48, 147.35, 146.83, 146.81, 146.77, 146.45, 146.44, 146.41, 146.31, 146.20, 146.17, 146.09, 146.06, 145.82, 145.65, 145.63 (2C), 145.61, 145.49, 145.45, 145.42, 145.37 (2C), 145.24, 144.80 (2C), 144.63, 144.47, 143.26, 143.14, 142.84, 142.78 (2C), 142.70, 142.40, 142.33, 142.32 (2C), 142.23, 142.18 (2C), 142.07, 141.97, 141.91, 141.84 (2C), 140.42, 140.30, 139.89, 139.66, 137.67, 137.08, 136.51, 136.31, 75.05 (sp³-C of C₆₀), 73.42 (CHCO₂), 71.36 (sp³-C of C₆₀), 67.20 (CHCH₃), 61.02 (OCH₂CH₃), 42.25 (NCH₂CH₃), 18.15 (CHCH₃), 14.99 (OCH₂CH₃), 14.73 (NCH₂CH₃). FT-IR v/cm⁻¹ (KBr) 2964, 2921, 1729, 1509, 1450, 1429, 1377, 1336, 1169, 574, 524; UV-vis (CHCl₃) λ_{max} /nm (lg ε) 256 (4.93), 320 (4.40), 430 (3.43), 701 (2.46); HRMS (MALDI TF-ICR): m/z calcd for C₆₈H₁₅NO₂: 877.1103; found: 877.1112.

Thermal reaction of C₆₀ with *N*-benzylglycine ethyl ester

By the same procedure as above, the reaction of C_{60} (35.9 mg, 0.05 mmol) with *N*-benzylglycine ethyl ester (96.7 mg,

0.50 mmol) at 180 °C for 40 min gave unreacted C₆₀ (16.9 mg, 47%), 7c (8.2 mg, 16%), 8c (5.0 mg, 10%) and 9 as a mixture of *cis*-isomer^{4a,17} (4.5 mg, 10%) and *trans*-isomer^{4a} (1.5 mg, 3%), and 10c¹⁸ (0.3 mg, 0.7%). 7c: ¹H NMR (300 MHz, CDCl₃): δ = 7.76 (d, J = 7.2 Hz, 2H), 7.47 (t, J = 7.5 Hz, 2H), 7.39 (t, J = 7.2 Hz, 1H), 5.80 (s, 2H), 4.74 (d, J = 13.1 Hz, 1H), 4.37–4.30 (m, 4H), 4.14 (d, J = 13.1 Hz, 1H), 1.25 (t, J = 6.9Hz, 6H); ¹³C NMR (75 MHz, CS₂/CDCl₃ with Cr(acac)₃ as relaxation reagent, all 2C unless indicated): $\delta = 169.82$ (C=O), 153.49, 150.73, 147.17, 146.17, 146.11, 145.84 (4C), 145.78, 145.66, 145.41, 145.39, 145.27, 145.09, 145.03, 144.32, 144.28, 142.85, 142.51, 142.43, 141.98, 141.92, 141.82, 141.67, 141.62, 141.50, 139.79, 139.30, 137.35 (1C, aryl C), 136.82, 136.10, 128.78 (aryl C), 128.56 (aryl C), 127.68 (1C, aryl C), 73.05 (CHCO₂), 71.26 (sp³-C of C₆₀), 61.18 (OCH₂CH₃), 52.27 (1C, NCH₂), 14.27 (OCH₂CH₃); FT-IR v/cm⁻¹ (KBr) 2923, 1731, 1567, 1520, 1375, 1279, 1146, 1095, 1027, 941, 803, 751, 679, 592, 526; UV-vis (CHCl₃) λ_{max} nm (lg ε) 256 (5.01), 310 (4.52), 429 (3.36), 693 (1.81); HRMS (MALDI TF-ICR): m/z calcd for C₇₅H₁₉NO₄: 997.1314; found: 997.1308. **8c**: ¹H NMR (300 MHz, CS₂/CDCl₃): δ = 7.89 (br s, 2H), 7.65 (d, J = 7.3 Hz, 2H), 7.46–7.28 (m, 6H), 6.64 (s, 1H), 5.43 (s, 1H), 4.50 (d, J = 13.8 Hz, 1H), 4.39 (dq, J = 10.7, 7.1 Hz, 1H), 4.28 (dq, J =10.7, 7.1 Hz, 1H), 4.05 (d, J = 13.8 Hz, 1H), 1.26 (t, J = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CS_2 /acetone-d₆ with Cr(acac)₃ as relaxation reagent, all 1C unless indicated): $\delta = 170.34$ (COO), 156.23, 154.25, 153.96, 151.31, 147.67, 147.54, 147.06, 146.87, 146.62 (2C), 146.51, 146.45, 146.37, 146.33, 146.29, 146.20 (2C), 145.92, 145.83 (3C), 145.78, 145.58, 145.53, 145.49, 145.45 (2C), 145.36, 144.96, 144.86, 144.74, 144.63, 143.40, 143.25, 142.98, 142.87 (2C), 142.82, 142.45 (3C), 142.41, 142.36, 142.29, 142.28 (2C), 142.03, 141.99, 141.90, 141.85, 140.41, 140.34, 139.80, 139.73, 138.25, 137.59, 137.34, 136.58, 136.42, 136.38, 129.35 (4C, aryl C), 129.22 (2C, aryl C), 128.87 (3C, aryl C), 128.22 (1C, aryl C), 77.32 (CHPh), 76.02 (sp³-C of C₆₀), 73.04 (CHCO₂), 70.89 (sp³-C of C₆₀), 61.30 (OCH_2CH_3) , 51.90 (NCH_2) , 14.93 (OCH_2CH_3) ; FT-IR ν/cm^{-1} (KBr) 1731, 1493, 1454, 1428, 1188, 1166, 1138, 1027, 906, 732, 700, 575, 527; UV-vis (CHCl₃) λ_{max} nm (lg ε) 257 (4.98), 311 (4.53), 431 (3.44), 703 (1.90); HRMS (MALDI TF-ICR): *m*/*z* calcd for C₇₈H₁₉NO₂: 1001.1416; found: 1001.1422. **10c**: ¹H NMR (400 MHz, CS₂/CDCl₃): δ = 7.76 (d, J = 7.2 Hz, 2H), 7.43 (t, J = 7.5 Hz, 2H), 7.35 (t, J = 7.4 Hz, 1H), 5.85 (s, 1H), 5.61 (s, 1H), 4.43 (dq, J = 10.8, 7.1 Hz, 1H), 4.34 (dq, J = 10.8, 7.1 Hz, 1H), 1.29 (t, J = 7.1 Hz, 3H).

Thermal reaction of C₆₀ with ethyl N-phenylglycinate

By the same procedure as above, the reaction of C₆₀ (36.3 mg, 0.05 mmol) with ethyl *N*-phenylglycinate (182.4 mg, 1.02 mmol) at 180 °C for 75 h gave unreacted C₆₀ (18.1 mg, 50%) and fulleropyrrolidine **7d** (5.3 mg, 11%). ¹H NMR (300 MHz, CDCl₃): δ = 7.45 (t, *J* = 7.7 Hz, 2H), 7.33 (d, *J* = 7.8 Hz, 2H), 7.12 (t, *J* = 7.2 Hz, 1H), 6.54 (s, 2H), 4.26–4.16 (m, 4H), 1.15 (t, *J* = 7.1 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃ with Cr(acac)₃ as relaxation reagent, all 2C unless indicated): δ = 169.81 (*C*=O), 153.04, 150.17, 147.20, 146.13, 146.09, 145.82 (4C), 145.46, 145.40 (4C), 145.32 (4C), 145.06 (3C), 144.99,

144.26, 144.19, 142.82, 142.46, 142.38, 141.93, 141.86, 141.62 (4C), 141.50, 141.46, 139.89, 139.33, 136.57, 135.88, 129.21 (aryl C), 122.13 (1C, aryl C), 118.68 (aryl C), 74.12 (*C*HCO₂), 70.75 (sp³-*C* of C₆₀), 61.57 (*O*CH₂CH₃), 13.89 (*O*CH₂CH₃). FT-IR ν /cm⁻¹ (KBr) 2923, 1734, 1598, 1498, 1461, 1368, 1340, 1292, 1262, 1180, 1015, 757, 692, 576, 526; UV-vis (CHCl₃) λ_{max} /nm (lg ε) 257 (5.03), 310 (4.52), 429 (3.41), 691 (2.14); HRMS (MALDI TF-ICR): *m*/*z* calcd for C₇₄H₁₇NO₄: 983.1158; found: 983.1163.

Reaction of C₆₀ with sarcosine, glycine and formaldehyde

A 25-mL tube containing a mixture of C_{60} (72.1 mg, 0.10 mmol), sarcosine (8.9 mg, 0.10 mmol), glycine (7.5 mg, 0.10 mmol) and formaldehyde solution (7.2 µL, 0.10 mmol) in ODCB (10 mL) was wrapped with aluminum foil, sealed up, and ultrasonicated until C_{60} was dissolved, and then heated in an oil bath preset at 180 °C for 7 min. The usual workup gave unreacted C_{60} (48.1 mg, 67%), **4a**¹² (14.3 mg, 18%) and a trace amount of **2a**.¹²

Reaction of C₆₀ with N-benzylglycine, glycine and formaldehyde

By the same procedure as above, the reaction of C_{60} (72.1 mg, 0.10 mmol) with *N*-benzylglycine (16.5 mg, 0.10 mmol), glycine (7.5 mg, 0.10 mmol) and formaldehyde solution (7.2 μ L, 0.10 mmol) at 180 °C for 7 min gave unreacted C_{60} (41.4 mg, 57%), **4c**¹⁶ (13.8 mg, 16%), and a trace amount of **2a**.¹²

Reaction of C₆₀ with *N*-benzylglycine, glycine and benzaldehyde

By the same procedure as above, the reaction of C_{60} (72.0 mg, 0.10 mmol), *N*-benzylglycine (16.5 mg, 0.10 mmol), glycine (7.5 mg, 0.10 mmol) and benzaldehyde (10 µL, 0.10 mmol) at 180 °C for 7 min gave unreacted C_{60} (45.3 mg, 63%), **5**c¹⁷ (25.4 mg, 27%) and a trace amount of the corresponding *N*-unsubstituted fulleropyrrolidine.

Reaction of C_{60} with *N*-ethylglycine, formaldehyde and acetaldehyde

By the same procedure as above, the reaction of C_{60} (71.9 mg, 0.10 mmol), *N*-ethylglycine (10.3 mg, 0.10 mmol), formaldehyde solution (7.2 µL, 0.10 mmol) and acetaldehyde solution (12 µL, 0.10 mmol) at 180 °C for 6 min gave unreacted C_{60} (28.7 mg, 40%), **4b**¹⁵ (18.5 mg, 23%) and **5b** (6.4 mg, 8%).

Reaction of C_{60} with *N*-ethylglycine ethyl ester, ethyl glyoxalate and acetaldehyde

By the same procedure as above, the reaction of C_{60} (72.0 mg, 0.10 mmol), *N*-ethylglycine ethyl ester (13.1 mg, 0.10 mmol), ethyl glyoxalate solution (20.5 µL, 0.10 mmol) and acetaldehyde solution (12 µL, 0.10 mmol) at 180 °C for 10 min gave unreacted C_{60} (34.5 mg, 48%), **7b** (35.5 mg, 38%) and **8b** (2.6 mg, 3%).

Reaction of C₆₀ with *N*-benzylglycine, formaldehyde and benzaldehyde

By the same procedure as above, the reaction of C₆₀ (36.1 mg, 0.05 mmol), *N*-benzylglycine (8.4 mg, 0.05 mmol), formaldehyde (3.6 μ L, 0.05 mmol) and benzaldehyde (5.2 μ L, 0.05 mmol) in ODCB (6 mL) at 180 °C for 4 min gave unreacted C₆₀ (27.1 mg, 75%), **4c**¹⁶ (9.4 mg, 22%) and a trace amount of **5c**.¹⁷

Reaction of C_{60} with *N*-benzylglycine ethyl ester, ethyl glyoxalate and benzaldehyde

By the same procedure as above, the reaction of C_{60} (35.9 mg, 0.05 mmol), *N*-benzylglycine ethyl ester (9.7 mg, 0.05 mmol), ethyl glyoxalate (10 µL, 0.05 mmol) and benzaldehyde (5.2 µL, 0.05 mmol) in ODCB (6 mL) at 180 °C for 3 min gave unreacted C_{60} (21.0 mg, 58%), **7c** (15.3 mg, 31%) and a trace amount of **8c**.

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