

# Synthesis and biological evaluation of pyrido[3',2':4,5]furo [3,2-*d*]pyrimidine derivatives as novel PI3 kinase p110 $\alpha$ inhibitors

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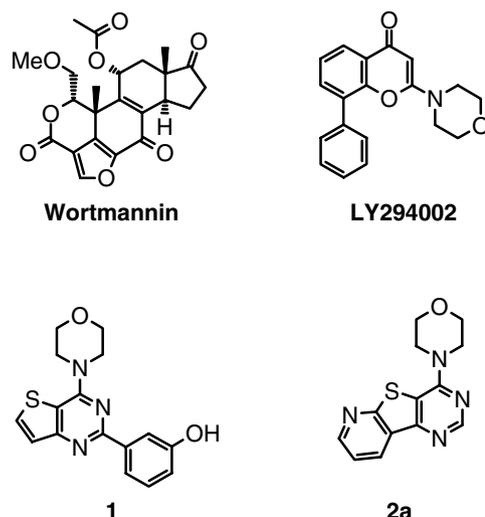
**Abstract**—4-Morpholin-4-ylpyrido[3',2':4,5]thieno[3,2-*d*]pyrimidine **2a** was discovered in our chemical library as a novel p110 $\alpha$  inhibitor with an IC<sub>50</sub> of 1.4  $\mu$ M. By structural modification of **2a**, the 2-aryl-4-morpholinopyrido[3',2':4,5]furo[3,2-*d*]pyrimidine derivative **10e** was discovered as a p110 $\alpha$  inhibitor with approximately 400-fold greater potency than **2a**. Evaluation of isoform selectivity showed that **10e** is a potent inhibitor of p110 $\beta$ . Furthermore, **10e** showed anti-proliferative activity in various cell lines, including multi-drug resistant MCF7/ADR-res cells, and was effective against HeLa human cervical tumor xenografts in nude mice.

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Phosphoinositide 3-kinase (PI3K) is an enzyme that catalyzes phosphorylation of the 3-hydroxyl position of phosphatidylinositides (PIs) and plays a crucial role in mitogenic signal transduction.<sup>1–4</sup> PI3K is negatively regulated by the lipid-phosphatase PTEN, which is one of the most commonly mutated proteins in human cancers,<sup>5–7</sup> and therefore PI3K inhibitors are considered to be potential anti-cancer agents (Fig. 1).

Among the various subtypes of PI3K identified to date,<sup>8–11</sup> class Ia PI3Ks (p110 $\alpha$ , p110 $\beta$ , and p110 $\delta$ ) are known to play critical roles in cell growth and survival.<sup>12</sup> The *PIK3CA* gene that encodes p110 $\alpha$  is amplified and overexpressed in ovarian and other cancers,<sup>13,14</sup> and is mutated in a variety of cancers.<sup>15–18</sup> Therefore, inhibitors of class Ia PI3Ks, and particularly PI3K p110 $\alpha$ , are likely to be useful in cancer treatment.

Several non-selective PI3K inhibitors have been identified, including wortmannin and LY294002,<sup>19–21</sup> but



**Figure 1.** Structures of PI3K inhibitors.

few isoform-specific PI3K inhibitors are available.<sup>22,23</sup> We have reported that the thieno[3,2-*d*]pyrimidine derivative **1** is a highly potent and selective p110 $\alpha$  inhibitor, but it is not effective in vivo because of a poor

**Keywords:** PI3 kinase; p110 $\alpha$ ; Inhibitor; Anti-cancer agent.

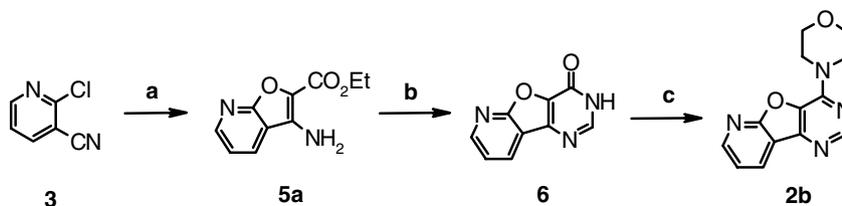
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pharmacokinetic profile due to a short half-life of less than 10 min.<sup>24,25</sup> In screening our chemical library, we identified **2a** as another lead compound with structural similarity to **1**. Through structural modification of **2a**, the novel pyrido[3',2':4,5]furo[3,2-*d*]pyrimidine **10e** was identified as a potent p110 $\alpha$  inhibitor that is also effective in HeLa xenografts in mice. Although biological evaluation of **10e** has been recently reported after our publication of the patent,<sup>26–28</sup> the generation of **10e** and SAR of its derivatives has not yet been described. Herein, we report the synthesis, SAR, and biological data for a series of pyrido[3',2':4,5]furo[3,2-*d*]pyrimidine derivatives as novel PI3K p110 $\alpha$  inhibitors.

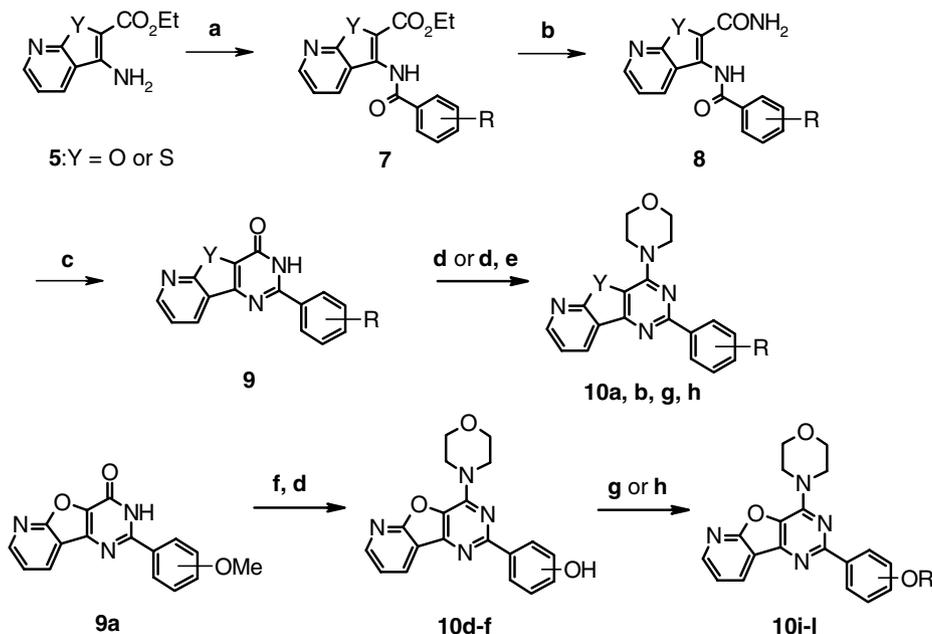
The lead compound **2a** was synthesized according to the literature<sup>29</sup> and its furan analogue, **2b**, was prepared as shown in Scheme 1. Treatment of 2-chloronicotinitrile **3** with **4** in the presence of DBU afforded a bicyclic acid ester **5a**, which was treated with formamide to give a pyrimidone **6**. Chlorination of **6** with phosphorus oxychloride followed by substitution with morpholine gave the desired product **2b**. The synthesis of compounds **10**, each of which has an aryl group on the pyrimidine ring, is illustrated in Scheme 2. Synthetic methods for the derivatives were generally analogous to those previously

described.<sup>24</sup> Acylation of **5** with the appropriate aryl acid chloride, ester hydrolysis, chlorination with SOCl<sub>2</sub>, and subsequent treatment with NH<sub>4</sub>OH afforded the series of amides **8**. Cyclization of **8** under previously reported conditions using MeOH as a solvent<sup>24</sup> provided **9** in low yield due to cleavage of the amide linkage of **8**, while reaction in 2-PrOH instead of MeOH gave cyclized **9** in better yield. Chlorination of **9** with phosphorus oxychloride and substitution with morpholine gave the desired compounds **10a–b** and **10h**. The amino derivative **10g** was synthesized from the corresponding nitro derivative via a reduction with iron. Demethylation of **9a** with HBr, acetylation, chlorination, and treatment with morpholine afforded compounds **10d–f**. Alkylation of the phenol derivative **10e** gave **10i–k**. The acetic acid derivative **10l** was prepared from the corresponding ester derivative by hydrolysis under basic conditions.

Inhibition of p110 $\alpha$  was determined by a scintillation proximity assay (SPA), as previously reported,<sup>24,25</sup> which gave similar results to those obtained by a conventional TLC method.<sup>21</sup> The lead compound **2a** had an IC<sub>50</sub> of 1.4  $\mu$ M for p110 $\alpha$  in the SPA assay. In this assay, LY294002 inhibited p110 $\alpha$  with an IC<sub>50</sub> of 0.63  $\mu$ M, showing approximately 2-fold greater potency



**Scheme 1.** Reagents and conditions: (a) HOCH<sub>2</sub>CO<sub>2</sub>Et **4**, DBU, EtOH, reflux; (b) formamide, reflux; (c) i) POCl<sub>3</sub>, reflux; ii) morpholine, toluene, reflux.



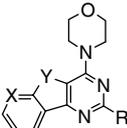
**Scheme 2.** Reagents and conditions: (a) ArCOCl, Et<sub>3</sub>N or pyridine, THF or CHCl<sub>3</sub>; (b) i) 1 N NaOH, EtOH; ii) SOCl<sub>2</sub>, reflux; iii) NH<sub>4</sub>OH, THF; (c) 2 N NaOH, MeOH or 2-PrOH, reflux; (d) i) POCl<sub>3</sub>,  $\Delta$ ; ii) morpholine, toluene or neat, reflux; (e) Fe, NH<sub>4</sub>Cl, aqueous EtOH, reflux; (f) i) HBr, AcOH, reflux; ii) Ac<sub>2</sub>O, AcONa, reflux; (g) R'-Cl, K<sub>2</sub>CO<sub>3</sub>, DMF; (h) i) BrCH<sub>2</sub>CO<sub>2</sub>Et, K<sub>2</sub>CO<sub>3</sub>, DMF; ii) 1 N NaOH, MeOH, THF.

than **2a**. Since the morpholino group is important for p110 $\alpha$  inhibitory activity in derivatives of **1**,<sup>24</sup> the morpholino group in **2a** was retained and other groups in **2a** were modified with the goal of obtaining p110 $\alpha$  inhibitors with greater potency.

First, modifications on the tricyclic ring of **2a** were investigated (Table 1). Replacement of sulfur with oxygen at Y in **2a** resulted in a 3-fold increase in p110 $\alpha$  inhibitory activity (**2b**: IC<sub>50</sub> = 0.56  $\mu$ M), whereas the pyrazole derivative **2c** (Y = NH) was inactive against p110 $\alpha$ . Introduction of a phenyl group at R of the thiophene derivative **2a** retained similar p110 $\alpha$  inhibitory activity (**10a**: IC<sub>50</sub> = 1.7  $\mu$ M), whereas the furan derivative **10b** with a phenyl ring at C2 was about 10-fold more potent than **2a** (**10b**: IC<sub>50</sub> = 0.16  $\mu$ M). The benzothienopyrimidine derivative **2d** (X = C) showed comparable potency to **2a**; however, introduction of a benzene ring at C2 (**10c**) eliminated the p110 $\alpha$  inhibitory activity.

The 3-hydroxy group on the benzene ring of **1** was previously found to be important for p110 $\alpha$  inhibitor activity.<sup>24</sup> Considering the structural similarity between **1** and **10b**, introduction of a hydroxy group on the benzene ring of **10b** was investigated (Table 2). The 2-hydroxy derivative **10d** showed about a 3-fold decrease in potency compared with **10b**. As expected, the 3-hydroxy derivative **10e** and the 4-hydroxy derivative **10f** were approximately 45- and 6-fold more potent p110 $\alpha$  inhibitors compared with the unsubstituted **10b** (**10e**, **10f**: IC<sub>50</sub> = 3.6, 26 nM, respectively). Since the hydroxy group at the 3-position on the benzene ring of **10e** was especially effective in producing p110 $\alpha$  inhibitory activity, other hydrophilic substituents at C3 on the benzene ring were introduced, based on the possibility of a hydrophilic binding pocket in the binding site. The 3-amino derivative **10g** showed almost the same p110 $\alpha$  activity compared with **10b**. Introduction of a 3-methoxy group at C3 on the benzene ring of **10b** retained the same p110 $\alpha$  inhibitory activity, but most compounds with ether substituents at this position showed increased

Table 1. Inhibition of p110 $\alpha$  by tricyclic derivatives



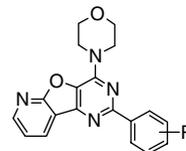
Compound	X	Y	R	IC <sub>50</sub> <sup>c</sup> ( $\mu$ M)
				p110 $\alpha$
LY294002 <sup>a</sup>				0.63
<b>2a</b> <sup>a</sup>	N	S	H	1.4
<b>2b</b> <sup>a</sup>	N	O	H	0.56
<b>2c</b> <sup>a</sup>	N	NH	H	>30
<b>10a</b> <sup>a</sup>	N	S	Ph	1.7
<b>10b</b> <sup>a</sup>	N	O	Ph	0.16
<b>2d</b> <sup>b</sup>	C	S	H	2.4
<b>10c</b> <sup>a</sup>	C	S	Ph	>30

<sup>a</sup> Free base.

<sup>b</sup> HCl salt.

<sup>c</sup> IC<sub>50</sub> values represent means of at least two separate determinations with typical variations of less than  $\pm 20\%$ .

Table 2. Inhibition of p110 $\alpha$  by pyridofuroprymidine derivatives



Compound	R	IC <sub>50</sub> <sup>d</sup> ( $\mu$ M)	
		p110 $\alpha$	A375
<b>10b</b> <sup>a</sup>	H	0.16	NT
<b>10d</b> <sup>a</sup>	2-OH	0.40	>30
<b>10e</b> <sup>b</sup>	3-OH	0.0036	0.33
<b>10f</b> <sup>b</sup>	4-OH	0.026	>30
<b>10g</b> <sup>c</sup>	3-NH <sub>2</sub>	0.14	3.87
<b>10h</b> <sup>b</sup>	3-OMe	0.10	15.78
<b>10i</b> <sup>c</sup>	3-(Piperidinoethoxy)	0.17	9.67
<b>10j</b> <sup>c</sup>	3-(Morpholinoethoxy)	0.016	12.32
<b>10k</b> <sup>c</sup>	3-(Thiomorpholinoethoxy)	0.079	11.76
<b>10l</b> <sup>b</sup>	3-OCH <sub>2</sub> CO <sub>2</sub> H	0.024	>30

NT, not tested.

<sup>a</sup> Free base.

<sup>b</sup> HCl salt.

<sup>c</sup> 2HCl salt.

<sup>d</sup> IC<sub>50</sub> values represent means of at least two separate determinations with typical variations of less than  $\pm 20\%$  for the p110 $\alpha$  enzyme and A375 cell proliferation assays.

activity. Among these compounds, the morpholinoethoxy derivative **10j** and the carboxylic acid derivative **10l** were potent p110 $\alpha$  inhibitors, with IC<sub>50</sub> values of 16 and 24 nM, respectively. Preliminary studies indicated that the inhibitors are ATP competitive but further work is required to investigate the inhibition kinetics in more detail.

The inhibitory activities of these derivatives were evaluated against serum-induced proliferation of A375 human melanoma cells. The 3-hydroxy derivative **10e** showed a potent anti-proliferative effect in vitro with an IC<sub>50</sub> of 0.33  $\mu$ M, whereas compounds **10f**, **10j**, **10k**, and **10l**, which had relatively potent p110 $\alpha$  inhibitory activities in the enzyme assay, did not show cellular activity; it can be speculated that this may be due to poor permeability through the cell membrane.

Further evaluation of compound **10e** was performed, since this was the most potent p110 $\alpha$  inhibitor in the series and was effective as an anti-proliferative agent in vitro. To examine selectivity for p110 $\alpha$ , the inhibitory activity of **10e** was evaluated against other PI3K isoforms. As shown in Table 3, **10e** was also a potent inhibitor of p110 $\beta$  (class Ia), which is also a potential target for cancer treatment.<sup>12</sup> Although **10e** showed excellent selectivity for p110 $\alpha$  over p110 $\gamma$  (class Ib), the selectivity for p110 $\alpha$  over PI3K C2 $\beta$  (class II) was not very high. It is of interest that the thienopyrimidine derivative **1**, which is structurally similar to **10e**, is a more isoform-selective p110 $\alpha$  inhibitor than **10e**. Concerning selectivity against protein kinases, **10e** was inactive at 100  $\mu$ M against protein kinases such as KDR, PKA, PKC $\alpha$ , and cyclin E/CDK<sub>2</sub>. Recent publications reported that **10e** also inhibited other kinases such as DNA-PK or mTOR.<sup>27,28</sup> Although we have not yet determined the

**Table 3.** Selectivity of **10e** against PI3Ks and protein kinases

Compound	IC <sub>50</sub> <sup>c</sup> (μM)							
	p110α	p110β	p110γ	PI3K C2β	KDR	PKA	PKCα	Cyclin E/CDK <sub>2</sub>
LY294002 <sup>a</sup>	0.63	0.34	1.6	2.1	NT	NT	NT	NT
<b>10e</b> <sup>b</sup>	0.0036	0.0030	0.25	0.010	>100	>100	>100	>100
<b>1</b> <sup>b</sup>	0.0025	0.016	0.66	0.22	91	3.4	466	28

NT, not tested.

<sup>a</sup> Free base.

<sup>b</sup> HCl salt.

<sup>c</sup> IC<sub>50</sub> values represent means of at least two separate determinations with typical variations of less than ±20%.

**Table 4.** Inhibition of human tumor cell proliferation in vitro by **10e**

Compound	IC <sub>50</sub> <sup>b</sup> (μM)				
	A375 (Melanoma)	HeLa (Cervix)	A549 (Lung)	MCF7 (Breast)	MCF7 ADR-res (Breast)
<b>10e</b> <sup>a</sup>	0.33	0.54	0.54	0.18	0.13

<sup>a</sup> HCl salt.

<sup>b</sup> IC<sub>50</sub> values represent means of at least two separate determinations with typical variations of less than ±20%.

inhibitory activity of **10e** and its derivatives against mTOR and DNA-PK, the inhibitory potencies of the compounds in A375 melanoma cell proliferation assay were well correlated with those in a PKB phosphorylation assay (data not shown). This observation suggested that the tumor suppressive actions of **10e** and its derivatives may depend mainly on PI3K inhibition. The involvement of mTOR and DNA-PK inhibition should be further investigated.

Evaluation of the effects of **10e** in several human tumor cell lines showed that **10e** exerted anti-proliferative activity at submicromolar concentrations (Table 4). Notably, **10e** was active in MCF7 ADR-res human breast cancer cells, which display a high level of resistance to doxorubicin and several other anti-cancer agents due to overexpression of P-glycoprotein.<sup>30,31</sup> Finally, the activity of **10e** was evaluated in vivo using a HeLa human cervical tumor xenograft model in mice. The half-life of **10e**, as measured by HPLC/MS/MS, was 4.1 h, when administered intraperitoneally. **10e** significantly suppressed tumor growth by 45% in this mod-

el when dosed intraperitoneally at 100 mg/kg daily for 2 weeks, and did not cause any associated weight loss (Fig. 2).

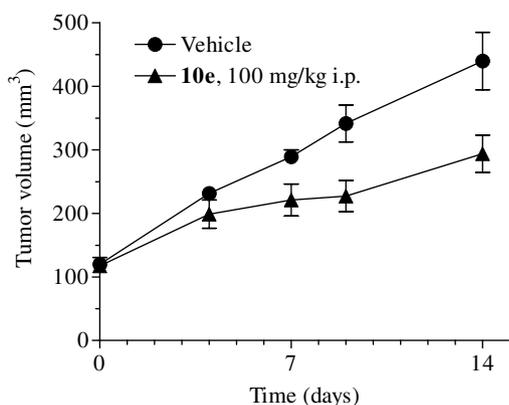
In summary, structural modification of 4-morpholino-pyrido[3',2':4,5]thieno[3,2-*d*]pyrimidine **2a** led to the discovery of **10e** as a p110α inhibitor of approximately 400-fold greater potency than **2a**. Compound **10e** is also a potent inhibitor of p110β and showed selectivity over several tested protein kinases. Furthermore, **10e** exhibited anti-tumor activity against the MCF7 ADR-res tumor cell line, which shows multi-drug resistance to anti-cancer drugs, and was effective in a HeLa human cervical cancer xenograft model without causing weight loss.

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### References and notes

- Leever, S. J.; Vanhaesebroeck, B.; Waterfield, M. D. *Curr. Opin. Cell Biol.* **1999**, *11*, 219.
- Lawlor, M. A.; Alessi, D. R. *J. Cell Sci.* **2001**, *114*, 2903.
- Vanhaesebroeck, B.; Leever, S. J.; Ahmadi, K.; Timms, J.; Katso, R.; Driscoll, P. C.; Woscholski, R.; Parker, P. J.; Waterfield, M. D. *Annu. Rev. Biochem.* **2001**, *70*, 535.
- Cantley, L. C. *Science* **2002**, *296*, 1655.
- Hopkins, K. *Science* **1998**, *282*, 1027.
- Maehama, T.; Dixon, J. E. *Trends Cell Biol.* **1999**, *9*, 125.
- Simpson, L.; Parsons, R. *Exp. Cell Res.* **2001**, *264*, 29.
- Domin, J.; Waterfield, M. D. *FEBS Lett.* **1997**, *410*, 91.
- Vanhaesebroeck, B.; Waterfield, M. D. *Exp. Cell Res.* **1999**, *253*, 239.



**Figure 2.** Effect of compound **10e** on the growth of HeLa human cervical tumor xenografts. Compound **10e** (100 mg/kg) suspended in 20% hydroxypropyl-β-cyclodextrin/saline was administered intraperitoneally daily for 2 weeks to nude mice carrying a subcutaneous HeLa xenograft. Error bars show ±SE.

10. Vanhaesebroeck, B.; Leever, S. J.; Panayotou, G.; Waterfield, M. D. *Trends Biochem. Sci.* **1997**, *22*, 267.
11. Fruman, D. A.; Meyers, R. E.; Cantley, L. C. *Annu. Rev. Biochem.* **1998**, *67*, 481.
12. Vivanco, I.; Sawyers, C. L. *Nat. Rev. Cancer* **2002**, *2*, 489.
13. Shayesteh, L.; Lu, Y.; Kuo, W.-L.; Baldocchi, R.; Godfrey, T.; Collins, C.; Pinkel, D.; Powell, B.; Mills, G. B.; Gray, J. W. *Nat. Genet.* **1999**, *21*, 99.
14. Ma, Y.-Y.; Wei, S.-J.; Lin, Y.-C.; Lung, J.-C.; Chang, T.-C.; Whang-Peng, J.; Liu, J. M.; Yang, D.-M.; Yang, W. K.; Schen, C.-Y. *Oncogene* **2000**, *19*, 2739.
15. Samuels, Y.; Wang, Z.; Bardelli, A.; Silliman, N.; Ptak, J.; Szabo, S.; Yan, H.; Gazdar, A.; Powell, S. M.; Riggins, G. J.; Willson, J. K.; Markowitz, S.; Kinzler, K. W.; Vogelstein, B.; Velculescu, V. E. *Science* **2004**, *304*, 554.
16. Campbell, I. G.; Russell, S. E.; Choong, D. Y.; Montgomery, K. G.; Ciavarella, M. L.; Hooi, C. S.; Cristiano, B. E.; Pearson, R. B.; Phillips, W. A. *Cancer Res.* **2004**, *64*, 7678.
17. Kang, S.; Bader, A. G.; Vogt, P. K. *Proc. Natl. Acad. Sci. U.S.A.* **2005**, *102*, 802.
18. Parsons, D. W.; Wang, T.-L.; Samuels, Y.; Bardelli, A.; Cummins, J. M.; DeLong, L.; Silliman, N.; Ptak, J.; Szabo, S.; Willson, J. K.; Markowitz, S.; Kinzler, K. W.; Vogelstein, B.; Lengauer, C.; Velculescu, V. E. *Nature* **2005**, *436*, 792.
19. Schultz, R. M.; Merriman, R. L.; Andis, S. L.; Bonjouklian, R.; Grindey, G. B.; Rutherford, P. G.; Gallegos, A.; Massey, K.; Powis, G. *Anticancer Res.* **1995**, *15*, 1135.
20. Wymann, M. P.; Bulgarelli-Leva, G.; Zvelebil, M. J.; Piroola, L.; Vanhaesebroeck, B.; Waterfield, M. D.; Panayotou, G. *Mol. Cell Biol.* **1996**, *16*, 1722.
21. Vlahos, C. J.; Matter, W. F.; Hui, K. Y.; Brown, R. F. *J. Biol. Chem.* **1994**, *269*, 5241.
22. Ward, S.; Sotsios, Y.; Dowden, J.; Bruce, I.; Finan, P. *Chem. Biol.* **2003**, *10*, 207.
23. Ward, S. G.; Finan, P. *Curr. Opin. Pharmacol.* **2003**, *3*, 426.
24. Hayakawa, M.; Kaizawa, H.; Moritomo, H.; Koizumi, T.; Ohishi, T.; Okada, M.; Ohta, M.; Tsukamoto, S.; Parker, P.; Workman, P.; Waterfield, M. *Bioorg. Med. Chem.* **2006**, *14*, 6847.
25. Hayakawa, M.; Kaizawa, H.; Kawaguchi, K.; Ishikawa, N.; Koizumi, T.; Ohishi, T.; Yamano, M.; Okada, M.; Ohta, M.; Tsukamoto, S.; Raynaud, F. I.; Waterfield, M. D.; Parker, P.; Workman, P. *Bioorg. Med. Chem.* **2007**, *15*, 403.
26. Hayakawa, M.; Kaizawa, H.; Moritomo, H.; Kawaguchi, K.; Koizumi, T.; Yamano, M.; Matsuda, K.; Okada, M.; Ohta, M. U.S. Patent 6,608,053 B2, **2003**.
27. Knight, Z. A.; Gonzalez, B.; Feldman, M. E.; Zunder, E. R.; Goldenberg, D. D.; Williams, O.; Loewith, R.; Stokoe, D.; Balla, A.; Toth, B.; Balla, T.; Weiss, W. A.; Williams, R. L.; Shokat, K. M. *Cell* **2006**, *125*, 733.
28. Fan, Q.-W.; Knight, Z. A.; Goldenberg, D. D.; Yu, W.; Mostov, K. E.; Stokoe, D.; Shokat, K. M.; Weiss, W. A. *Cancer Cell* **2006**, *9*, 341.
29. Schneller, S. W.; Clough, F. W. *J. Heterocycl. Chem.* **1974**, *11*, 975.
30. Fairchild, C. R.; Ivy, S. P.; Kao-Shan, C.-S.; Whang-Peng, J.; Rosen, N.; Israel, M. A.; Melera, P. W.; Cowan, K. H.; Goldsmith, M. E. *Cancer Res.* **1987**, *47*, 5141.
31. Scudiero, D. A.; Monks, A.; Sausville, E. A. *J. Natl. Cancer Inst.* **1998**, *90*, 862.