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Design of LFA-1 antagonists based on a 2,3-dihydro-1*H*pyrrolizin-5(7aH)-one scaffold

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Abstract—A new class of lymphocyte function-associated antigen-1 (LFA-1) antagonists is described. Elaboration of the 2,3-dihydro-1*H*-pyrrolizin-5(7aH)-one scaffold resulted in the synthesis of potent inhibitors of the LFA-1/ICAM-1 interaction. Along with the in vitro activity, we present the X-ray crystal structure of the complex of compound **9b**, in a novel binding mode to the I-domain of LFA-1.

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The cell adhesion molecule LFA-1 (lymphocyte functionassociated antigen-1), also known as CD11a/CD18, is known to play a crucial role in many immunological functions including antigen presentation, leukocyte trafficking, B- and T-cell activation. LFA-1 belongs to the β_2 integrin family and is found on all leukocytes.¹ The ligands for LFA-1, the ICAMs 1-3 (intercellular adhesion molecules), are expressed on both leukocytes and the endothelium. In particular, the interaction of LFA-1 with ICAM-1 results in leukocyte adhesion to the endothelium and facilitates their extravasation to tissue sites.^{2,3} The LFA-1/ICAM interaction has been proposed to be an attractive therapeutic target in the treatment of several inflammatory disorders such as psoriasis and transplant rejection.^{4,5} The recent approval of efalizumab (Raptiva[®]), a fully humanized anti-LFA-1 antibody for the treatment of moderate to severe psoriasis, provides excellent validation of the target.⁶

In the search for small molecules that would interfere with the LFA-1/ICAM-1 interaction, we and others

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have disclosed several diverse chemical series of compounds.^{7,8} Some representative structures are shown in Figure 1. NMR and X-ray co-crystallographic studies have established that many of these inhibitors bind to the IDAS site (I-domain allosteric site) of the I-domain located on the CD11a unit.^{7–9} The X-ray structure of our clinical candidate BMS-587101^{7a} in complex with the I-domain of LFA-1 shows the following: (i) the



Figure 1. Representative LFA-1 antagonists reported in the literature.

Keywords: LFA-1 antagonists; 2,3-Dihydro-1*H*-pyrrolizin-5-(7aH)-one; LFA-1/ICAM interaction.

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p-cyanophenyl aromatic ring forms a favorable edge-toface π - π interaction with the 3,5-dichlorophenyl ring; (ii) the urea carbonyl is hydrogen bonded to the amino acid residues Lys-NH₂ 305, Lys-NH₂ 287, and Glu-COOH 284 via a water molecule. Unlike the urea carbonyl, the amide carbonyl does not appear to make any discernible contacts with the protein.

As a continuation of our work^{7b} on the modification of the reported bicyclic[5.5]hydantoin core 1,^{8c} we herein describe exploration of the 2,3-dihydro-1*H*-pyrrolizin-5(7aH)-one core **2** as a suitable surrogate in the design of LFA-1 inhibitors. Replacement of the N-6 nitrogen of the urea with a carbon atom and introduction of unsaturation in ring A provide structural rigidity similar to the bicyclic[5.5]hydantoin class of compounds, while retaining the all important carbonyl (C-5) as well as the 3,5-dichlorophenyl and benzyl groups necessary for high affinity. In addition, the pyrrolizinone core allows for elaboration at the C-7 position, thus providing an understanding of the SAR at this position (Fig. 2).

The synthesis of the 2,3-dihydro-1*H*-pyrrolizin-5(7aH)ones is outlined in Schemes 1 and 2. In our initial approach (Scheme 1), homochiral *N*-boc-L-proline methyl ester (3) was elaborated to give 7-hydroxy pyrrolizin-5one 6 in reasonable overall yield. Compound 6 served as the key intermediate for the synthesis of compounds 7, 9, and 10. Unfortunately, C–C coupling of the *O*-triflate 8 was not general and failed with copper mediated alkyl/aryl Grignards or lithium reagents. This necessitated the development of a more general approach utilizing 3 as the starting material (Scheme 2). The new sequence (Scheme 2) was used to prepare compounds 15–18.

Inhibition of the LFA-1/ICAM binding interaction using the HeLa/HSB assay^{7b} for a limited number of racemic 2,3-dihydro-1H-pyrrolizin-5(7aH)-ones is shown in Table 1. The data clearly suggest that the 2,3-dihydro-1*H*-pyrrolizin-5(7aH)-one core is an excellent surrogate for the bicyclic[5.5]hydantoin system. The enol 6 was essentially inactive, as was the bulky 3-pyrazolyl derivative 17. Compound 7 with a H-bond accepting methoxy group at C-7 displayed sub-micromolar potency in the HeLa/HSB assay. The potency increase was significantly more pronounced for compounds 9 (R = CN) and 18 (R = Et), which displayed IC₅₀s of 40 and 52 nM, respectively, in the HeLa/HSB binding assay. The C-7 SAR for the cyano analog (9) and the ethyl analog (18) was intriguing considering the hydrophilic and hydrophobic nature of these two substituents. These data suggested that either (i) the C-7 position of



Figure 2. Bicyclic[5.5]hydantoin core (1) and 2,3-dihydro-1*H*-pyrrolizin-5(7aH)-one core (2).



Scheme 1. Reagents and conditions: (a) i—LiHMDS, THF, -78 °C, ii—4-cyanobenzyl chloride, THF, 30%; (b) 4 N HCl-dioxane; (c) (3,5-dichlorophenyl)acetic acid, EDCI, DIPEA, DMF, 95%; (d) LiHMDS, THF, 90%; (e) (MeO)₃PO, K₂CO₃, reflux, 10 h, 30%; (f) Tf₂O, DIPEA, DCM, -10 °C, 60%; (g) Pd(PPh₃)₄, Zn(CN)₂, DIPEA, DMF, 195 °C, 10 min microwave, 75%; (h) (CH₃)₂(CN)CuLi, THF, -78 °C, 15%.



Scheme 2. Reagents and conditions: (a) i—LiHMDS, THF, -78 °C, ii—4-bromobenzyl bromide, THF, 85%; (b) LiN(OMe)Me, THF, -10 °C; (c) RMgBr or lithium pyrazol-1-id-3-yllithium, THF rt; (d) 4 N HCl-dioxane; (e) (3,5-dichlorophenyl)acetic acid, EDCI, DIPEA, DMF, 90–95%; (f) 5% KOH in MeOH, reflux (60–80%); (g) Pd(PPh₃)₄, Zn(CN)₂, DMF, 200 °C, 20 min microwave (80%).

the 2,3-dihydro-1*H*-pyrrolizin-5(7aH)-one core tolerates a variety of lipophilic and hydrophobic substituents or (ii) the binding mode for these compounds with the Idomain of LFA-1 may be different. To further understand this discrepancy in the SAR, the cyano analog (9) was co-crystallized with the I-domain of LFA-1.

 Table 1. In vitro activity of racemic 2,3-dihydro-1*H*-pyrrolizin-5(7aH)-ones

Compound	R	Х	HeLa/HSB binding ^a (IC ₅₀ or % inhibition)
BIRT-377			26 nM ^b
6	OH	CN	27% at 1 µM
7	OCH ₃	CN	430 nM
9	CN	CN	40 nM
10	CH ₃	CN	660 nM
15	Et	Br	106 nM
16	Cyclopropyl	Br	200 nM
17	3-Pyrazolyl	Br	30% at 10 μM
18	Et	CN	52 nM

^a Values are averages of at least two determinations.

 ${}^{b}K_{d}$ value in an LFA-1/ICAM-1 binding assay as reported in the literature.⁹

When racemic 9 was co-crystallized with the I-domain of LFA-1, a single enantiomer (S) was found bound to the I-domain. Figure 3 shows the X-ray co-crystal structure of this enantiomer (designated 9b) determined at 1.75 Å resolution.¹⁰ The X-ray co-crystal structure revealed a novel binding mode for compound 9b when compared to the hydantoin class of LFA-1 antagonists such as BMS-587101 and BIRT-377, where the absolute stereochemistry at the equivalent of the C-7a center is $(R)^9$. This 'flipped' binding mode for compound 9b is a direct result of the vinyl cyano group at C-7a forming a hydrogen bonding network through a water molecule to Lys-NH₂ 287 and Lys-NH₂ 305 and Glu-COOH 284, unlike BMS-587101 where the urea carbonyl, which corresponds to the C-5 carbonyl of 9b, forms the same hydrogen bonding network. In addition, the Tyr-OH 166, which is too far from the C-2 carbonyl of BMS-587101 to form a H-bond, has now moved in the direction of the carbonyl of **9b**, and is capable of forming a direct H-bond. Interestingly, the 3,5-dichlorophenyl



Figure 3. X-ray structure of **9b** complexed with the I-domain of LFA-1. Also shown is the initial electron density (magenta: 2Fo-Fc 1) prior to fitting compound **9b**. The water molecule is shown as a red sphere and hydrogen bonds are shown as small cyan spheres. Figure created using PyMol.¹¹



Figure 4. Absolute stereochemistry of 9a and 9b.

group at C-6 is not positioned as deeply in the hydrophobic pocket between α -helices 1 and 7 and β -strands 1, 3, and 4 as was found for BMS-587101.

Compound 9 was separated into its individual enantiomers 9a and 9b employing a Chiralpak AD column and the compounds tested for LFA-1/ICAM binding inhibition in the HeLa/HSB assay. As expected, there was a marked difference in the inhibition potential of these enantiomers. The faster eluting enantiomer, 9a, was much less active and displayed an IC₅₀ of 1.4 μ M, while the slower eluting enantiomer, 9b, had an IC₅₀ of 15 nM (Fig. 4). The absolute stereochemistry for 9b was established as *S* at the 7a position employing vibrational circular dichroism (VCD) spectroscopic analysis.¹² This result is consistent with the absolute stereochemistry of the compound in the X-ray co-crystal structure (vide supra).

The accommodation of **9b** in a binding mode opposite to that of the hydantoin class of LFA-1 antagonists is evidence of the flexibility of the I-domain of the LFA-1 protein. Since only **9** was subjected to X-ray crystallographic analysis, we cannot be certain that this opposite 'flipped' binding mode is an inherent property of the 2,3dihydro-1*H*-pyrrolizin-5(7aH)-one core or just unique to the C-7 cyano substituted compounds in this series. Racemic **18**, containing an ethyl group at C-7, was equipotent to **9** (IC₅₀ = 52 nM) in the HeLa/HSB binding assay. The lack of hydrogen-bonding ability of the ethyl group does not preclude the possibility that this compound may be binding in the 'traditional' mode similar to the hydantoin class of LFA-1 antagonists.

In conclusion, we have designed a novel class of LFA-1/ ICAM antagonists based on the 2,3-dihydro-1*H*-pyrrolizin-5(7aH)-one scaffold. X-ray crystal structural analysis of our most active compound **9b** revealed a 'flipped' binding mode to the I-domain of the LFA-1 protein. The stereochemistry of **9b** was found to be 7a(S), which is opposite to that found in the hydantoin class of LFA-1 antagonists.

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References and notes

- Harris, E. S.; McIntyre, T. M.; Prescott, S. M.; Zimmerman, G. A. J. Biol. Chem 2000, 275, 23409.
- Hogg, N.; Henderson, R.; Leitinger, B.; McDowall, A.; Porter, J.; Stanley, P. Immunol. Rev. 2002, 186, 164.
- Shimaoka, M.; Xiao, T.; Liu, J.-H.; Yang, Y.; Dong, Y.; Jun, C.-D.; McCormack, A.; Zhang, R.; Joachimiak, A.; Takagi, J.; Wanh, J.-H.; Springer, T. A. *Cell* 2003, *112*, 99.
- 4. Liu, G. Drugs Future 2001, 26, 767.
- 5. Dedrick, R. L.; Walicke, P.; Garovoy, M. Transpl. Immunol. 2002, 9, 181.
- Cather, J. C.; Cather, J. C.; Menter, A. Expert Opin. Biol. Ther. 2003, 3, 361.
- 7. (a) Potin, D.; Launay, M.; Monatlik, F.; Malabre, P.; Fabreguettes, M.; Fouquet, A.; Maillet, M.; Nicolai, E.; Dorgeret, L.; Chevallier, F.; Besse, D.; Dufort, M.; Caussade, F.; Ahmad, S. Z.; Stetsko, D. S.; Skala, S.; Davis, P. M.; Balimane, P.; Patel, K.; Yang, Z.; Marathe, P.; Postelneck, J.; Townsend, R. M.; Goldfarb, V.; Sheriff, S.; Einspahr, H.; Kish, K.; Malley, M. F.; DiMarco, J. D.; Gougoutas, J. Z.; Kadiyala, P.; Cheney, D. L.; Tejwani, R. W.; Murphy, D. K.; Mcintyre, K. W.; Yang, X.; Chao, S.; Leith, L.; Xiao, Z.; Mathur, A.; Chen, B. C.; Wu, D.-R.; Traeger, S. C.; McKinnon, M.; Barrish, J. C.; Robl, J. A.; Iwanowicz, E. J.; Suchard, S. J.; Dhar, T. G. M. J. Med. Chem. 2006, 49, 6946; (b) Potin, D.; Launay, M.; Nicolai, E.; Fabreguette, M.; Malabre, P.; Caussade, F.; Besse, B.; Skala, S.; Stetsko, D. K.; Todderud, G.; Beno, B. R.; Cheney, D. L.; Chang, C. J.; Sheriff, S.; Hollenbaugh, D. L.; Barrish, J. C.; Iwanowicz, E. J.; Suchard, S. J.; Dhar, T. G. M. Bioorg. Med. Chem. Lett. 2005, 15, 1161.
- (a) Kelly, T. A.; Jeanfavre, D. D.; McNeil, D. W.; Woska, R. J.; Reilly, L. P.; Mainolfi, E. A.; Kishimoto, K. M.; Nabozny, G. H.; Zinter, R.; Bormann, B.; Rothlein, R. J. Immunol. 1999, 163, 5173–5177; (b) Weitz-Schmidt, G.; Welzenbach, K.; Brinkmann, V.; Kamata, T.; Kallen, J.;

Bruns, C.; Cottens, S.; Takada, Y.; Hommel, U. Nat. Med. 2001, 7, 687; (c) Sircar, I.; Furth, P.; Teegarden, B. R.; Morningstar, M.; Smith, N.; Griffith, R. WO 0130781 (to Tanabe Seiyaku), 2001; (d) Liu, G.; Huth, J. R.; Olejniczak, E. T.; Mendoza, R.; DeVries, P.; Leitza, S.; Reilly, E. B.; Okasinski, G. F.; Fesik, S. W.; von Geldern, T. W. J. Med. Chem. 2001, 44, 1202; (e) Gadek, T. R.; Burdick, D. J.; McDowell, R. S.; Stanley, M. S.; Marsters, J. C., Jr.; Paris, K. J.; Oare, D. A.; Reynolds, M. E.; Ladner, C.; Zioncheck, K. A.; Lee, W. P.; Gribling, P.; Dennis, M. S.; Skelton, N. J.; Tumas, D. B.; Clark, K. R.; Keating, S. M.; Beresini, M. H.; Tilley, J. W.; Presta, L. G.; Bodary, S. C. Science 2002, 295, 1086; (f) Wu, J.-P.; Emeigh, J.; Gao, A. A.; Goldberg, D. R.; Kuzmich, D.; Miao, C.; Potocki, I.; Qian, K. C.; Sorcek, R. J.; Jeanfavre, D. D.; Kishimoto, K.; Mainolfi, E. A.; Nabozny, G.; Peng, C.; Reilly, P.; Rothlein, R.; Sellati, R. H.; Woska, J. R.; Chen, S.; Gunn, J. A.; O'Brien, D.; Norris, S. H.; Kelly, T. A. J. Med. Chem. 2004, 47, 5356; (g) Wattanasin, S.; Kallen, J.; Myers, S.; Guo, Q.; Sabio, M.; Ehrhardt, C.; Albert, R.; Hommel, U.; Weckbecker, G.; Welzenbach, K.; Weitz-Schmidt, G. Bioorg. Med. Chem. Lett. 2005, 15, 1217.

- Last-Barney, K.; Davidson, W.; Cardozo, M.; Frye, L. L.; Grygon, C. A.; Hopkins, J. L.; Jeanfavre, D. D.; Pav, S.; Qian, C.; Stevenson, J. M.; Tong, L.; Zindell, R.; Kelly, T. A. J. Am. Chem. Soc. 2001, 123, 5643.
- 10. PDB ID code 207N.
- DeLano, W.L. The PyMol Molecular graphics System. 2002. DeLano Scientific, San Carlos, CA, US. http://www.pymol.org>.
- (a) Stephens, P. J.; Aamouche, A.; Devlin, F. J.; Superchi, S.; Donnoli, M. I.; Rosini, C. J. Org. Chem. 2001, 66, 2671; (b) Freedman, T. B.; Cao, X.; Phillips, L. M.; Cheng, P. T. W.; Dalterio, R.; Shu, Y.-Z.; Zhang, H.; Zhao, N.; Shukla, R. B.; Tymiak, A.; Gozo, S. K.; Nafie, L. A.; Gougoutas, J. Z. Chirality 2006, 18, 746.