

Synthesis and Protein Tyrosine Phosphatase Inhibitory Activity of Dephostatin Analogs

TAKUMI WATANABE, TOMIO TAKEUCHI, MASAMI OTSUKA[†],
SHIN-ICHIRO TANAKA^{††} and KAZUO UMEZAWA^{*,††}

Institute of Microbial Chemistry,
3-14-23 Kamiosaki, Shinagawa-ku, Tokyo 141, Japan

[†]Institute for Chemical Research, Kyoto University,
Uji, Kyoto 611, Japan

^{††}Department of Applied Chemistry, Faculty of Science and Technology,
3-14-1 Hiyoshi, Kohoku-ku, Yokohama 223, Japan

(Received for publication June 23, 1995)

We have synthesized derivatives of dephostatin, a protein tyrosine phosphatase (PTPase) inhibitor, to study the structure-activity relationships of this inhibitor. Inactive analogs revealed some insight into structural requirements for PTPase inhibitory activity of dephostatin. Both a nitroso group and phenolic hydroxyl groups were found to be essential for the inhibitory activity. Among the dephostatin derivatives synthesized, one of the regioisomers of dephostatin showed PTPase inhibitory activity equivalent to that of dephostatin, and also had increased stability.

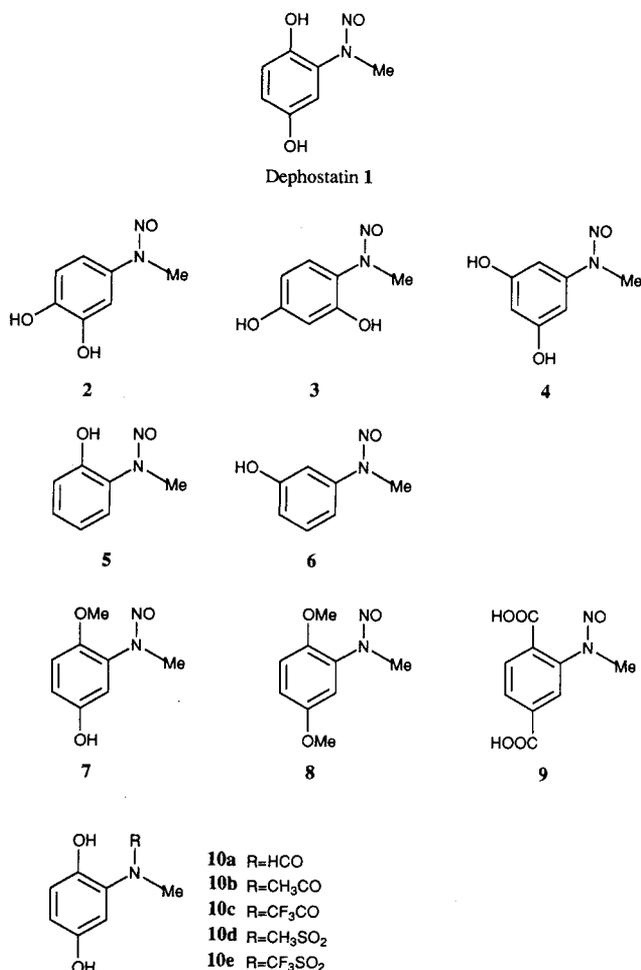
Protein tyrosine phosphatase (PTPase) has been implicated in the regulation of the cell cycle and activation of lymphocytes^{1,2}). A novel protein tyrosine phosphatase inhibitor, dephostatin **1**, was isolated from the culture filtrate of *Streptomyces* sp. MJ742-NF5 in our laboratory^{3,4}). In spite of its potential significance as a biochemical and therapeutic agent, sufficient amounts of dephostatin were difficult to obtain from the natural source. Therefore, we established a practical synthetic route for dephostatin⁵). However, dephostatin is not stable enough, and partial decomposition is frequently observed under storage and under conditions of biological experiments. Therefore, we synthesized several structurally related dephostatin analogs. As lability of dephostatin should arise from its hydroquinone or *N*-nitroso moiety, these structural elements were modified.

Results

Synthesis of Dephostatin Analogs

We synthesized 13 dephostatin analogs as shown in Fig. 1. Among them were some regioisomers of dephostatin (**2**, **3**, and **4**). Then, to clarify the role of the two hydroxyl groups in PTPase inhibitory activity, we synthesized two monohydroxyl analogs (**5** and **6**) and two methyl ether analogs (**7** and **8**) of the inhibitor. A carboxyl derivative (**9**), in which both of the hydroxyl groups were replaced by carboxyl groups, was also examined. Additionally, amide and sulfonamide derivatives

Fig. 1. Dephostatin and its analogs.



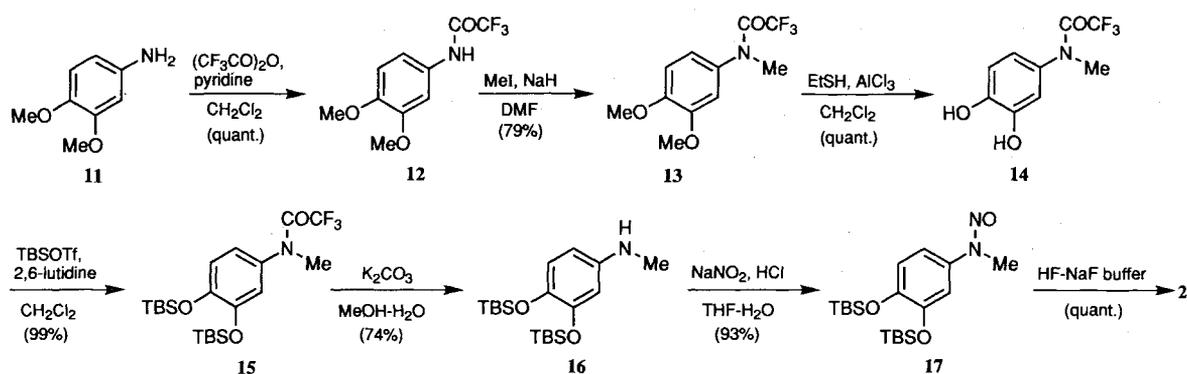
(10a to 10e), in place of nitrosamine, were tested to investigate whether the labile and potentially toxic nitroso moiety could be removed from the overall structure.

Attempts to synthesize 2, 3 and 4 by applying our synthetic route to dephostatin⁵⁾ starting from corresponding dimethoxyanilines were not successful, therefore, modifications of reaction conditions and/or protecting groups were needed.

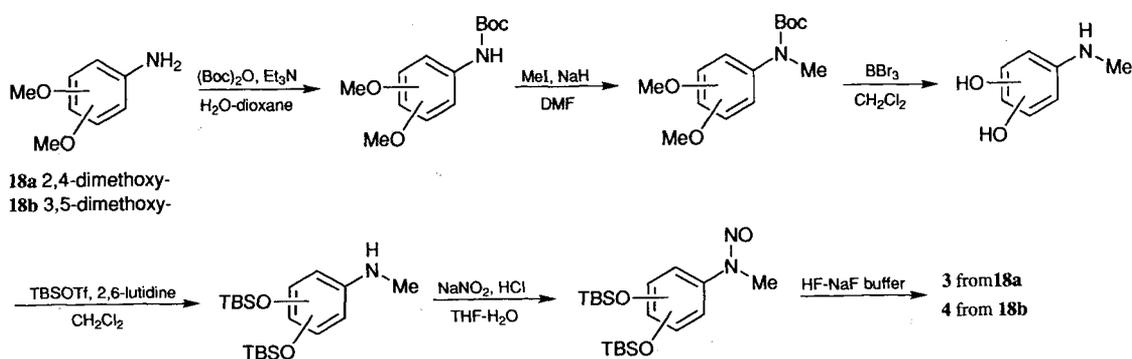
For the synthesis of 2 (Scheme 1), we used 3,4-dimethoxyaniline 11 as a starting material. The trifluoroacetyl group was chosen as a protecting group

for the amino group because its stability under acid promoted ether cleavage (conversion from 13 to 14). Acylation of 11 was conducted with trifluoroacetic anhydride and pyridine to give 12 in quantitative yield. Treatment of 12 with NaH in DMF followed by addition of MeI gave *N*-methylated product 13 in 79% yield. In the next step, two methyl ether groups were cleaved. This deprotection could be accomplished with BBr₃ without damaging the amide moiety, giving 14, although the yield was not satisfactory (37%). Instead, reaction with ethanethiol (EtSH) in the presence of AlCl₃⁶⁾ resulted in deprotection in quantitative yield. Silylation of 14

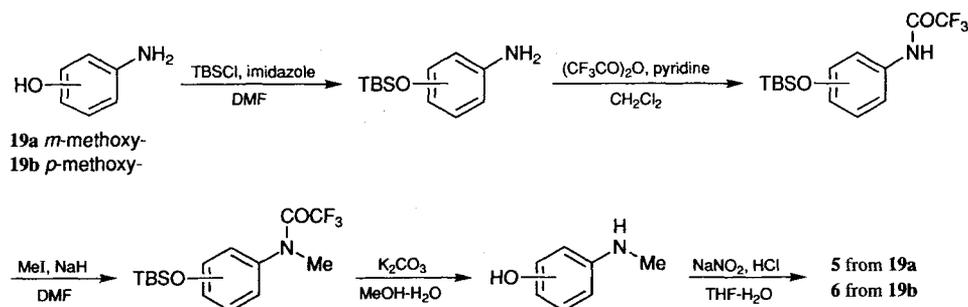
Scheme 1. Synthesis of 2.



Scheme 2. Synthesis of 3 and 4.



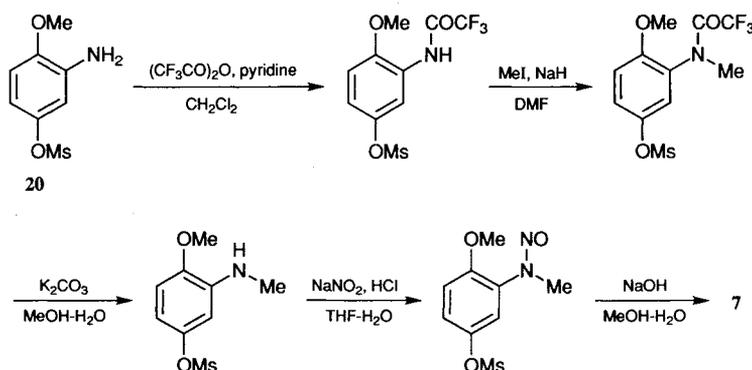
Scheme 3. Synthesis of 5 and 6.



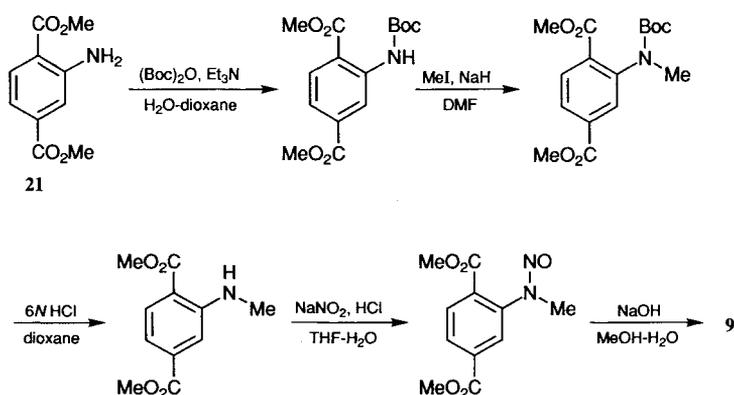
with *tert*-butyldimethylsilyl trifluoromethanesulfonate (TBSOTf) and 2,6-lutidine proceeded efficiently (99% yield). Subsequent treatment of **15** with aqueous K_2CO_3 yielded the deamide product **16**. Hydrolysis of silyl ether was a competing reaction in this step, and an appropriate choice of solvent was necessary to minimize this side reaction. The reaction system composed of 7% K_2CO_3

and MeOH gave a satisfactory result (74% yield). Nitrosation of **16** was accomplished under the standard conditions with $NaNO_2$ -HCl to give **17** in 93% yield. Final desilylation with HF-NaF buffer⁷⁾ gave the desired dephostatin analog **2** in quantitative yield. Detailed description of this synthesis is given in the experimental part.

Scheme 4. Synthesis of 7.



Scheme 5. Synthesis of 9.



Scheme 6. Synthesis of 10a~10e.

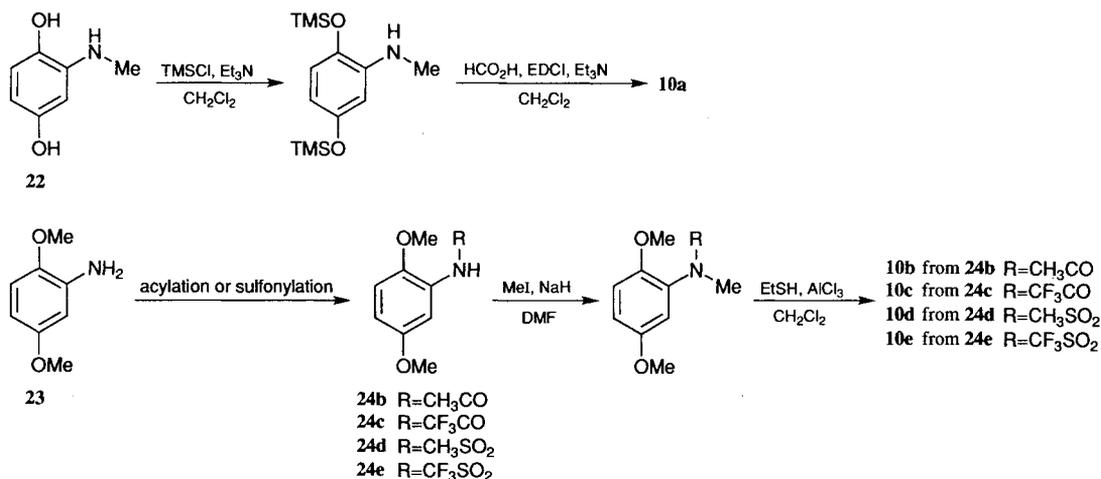


Table 1. PTPase inhibitory activities of dephostatin and its analogs.

Compound	IC ₅₀ (μg/ml)
1	1.8
2	3.0
3	>30
4	>30
5	>30
6	>30
7	>30
8	>30
9	>30
10a	>30
10b	>30
10c	>30
10d	>30
10e	>30

Analogs **3** and **4** were synthesized in similar manners as the case of dephostatin (Scheme 2). Other derivatives, **5** and **6**, **7**, **9** and **10a**~**10e**, were synthesized as shown in Schemes 3, 4, 5 and 6, respectively. Compound **8** was synthesized as already reported⁵). In scheme 3, the TBS group was removed without HF in the condition to eliminate the trifluoroacetyl group.

PTPase Inhibitory Activity of Dephostatin Analogs

PTPase inhibitory activities of the above analogs were examined by the method described previously³). The IC₅₀ value of each analog is listed in Table 1. Among them, only **2** has showed inhibitory activity equivalent to that of dephostatin.

Discussion

Since all the amide and sulfonamide analogs had no effect on the PTPase, the *N*-nitroso moiety of dephostatin must play a crucial role in PTPase inhibition. An equally important structural factor is the presence of two unmasked hydroxyl groups on the benzene ring. So far only 2,5- and 3,4-substitutions (dephostatin and **2**, respectively) gave the inhibitory activity. Monohydroxyl and methoxy analogs had no inhibitory activity. Replacement of hydroxyl groups by carboxyl groups, which are similarly anionic, could not maintain the PTPase inhibitory activity.

Compound **2** was found to be more stable than dephostatin in the medium commonly used for cell culture. Dephostatin largely decomposed within 2 hours in DULBECCO's modified EAGLE's medium, whereas **2** was stable for at least 24 hours (manuscript in preparation). Compound **2** can also be stored for at least several months in a refrigerator without apparent decomposition. Since it was shown to inhibit PTPase *in situ* (unpublished result), **2** may be easier to handle than dephostatin for

biological experiments. As **7** is only monomethylated at the 2-hydroxyl group and has no effect on PTPase, it should be a suitable inactive analogue for use in mechanistic studies.

Experimental

General

Melting points were determined with a Yanagimoto micro melting point apparatus and were uncorrected. IR spectra were measured with a Hitachi I-5020 FT-IR spectrometer or Hitachi Model 260-10 spectrophotometer. HRFAB-MS and FAB-MS were taken by JEOL JMS-SX 102. NMR spectra were recorded with a JEOL JNM-EX 400 spectrometer.

Synthesis of Dephostatin Analog **2**

3,4-Dimethoxy-*N*-trifluoroacetylaniline (**12**)

To a solution of 3,4-dimethoxyaniline **11** (10.04 g, 65.5 mmol) in dichloromethane (CH₂Cl₂) were added pyridine (5.82 ml, 72.1 mmol) and trifluoroacetic anhydride (10.0 ml, 72.1 mmol) successively under argon at 0°C. The solution was stirred at room temperature for 2 hours. After addition of CH₂Cl₂ to the reaction mixture, the organic layer was washed with 1 N HCl and brine successively, then dried over Na₂SO₄ and concentrated *in vacuo*. The residue was purified by elution from a silica gel column with EtOAc-*n*-hexane (1:1) to give 16.30 g of **12** (65.4 mmol, quantitative yield) as a pale yellow powder: mp 101~103°C. IR (KBr) ν_{\max} 3287, 1705, 1605, 1518, 1217, 1181, 841, 802 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.81 (1H, br), 7.30 (1H, d, *J*=2.4 Hz), 6.99 (1H, dd, *J*=8.8, 2.4 Hz), 6.86 (1H, d, *J*=8.8 Hz), 3.90 (3H, s), 3.89 (3H, s). ¹³C NMR (100 MHz, CDCl₃) δ 154.7 (q, *J*=36.7 Hz), 149.3, 147.3, 128.4, 115.8 (q, *J*=288.7 Hz), 112.9, 111.3, 105.0, 56.1, 56.0. FAB-MS (positive): *m/z* 249 (M⁺). Anal. Calcd for C₁₀H₁₀F₃NO₃: C, 48.20, H, 4.06, N, 5.62. Found: C, 48.12, H, 4.20, N, 5.60.

3,4-Dimethoxy-*N*-methyl-*N*-trifluoroacetylaniline (**13**)

To a suspension containing 1.45 g of 60% NaH and 10 ml of DMF was added a solution of **12** (6.00 g, 24.1 mmol) in 20 ml of DMF dropwise under argon at -23°C and the suspension was stirred at 0°C for 1 hour. Then, to the solution was added MeI (3.00 ml, 48.2 mmol) at -23°C, and the solution was stirred at room temperature for 1 hour. The solution was poured onto cold 1 N HCl and was extracted with EtOAc twice. The organic layer was washed with brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography with EtOAc-*n*-hexane (1:3 to 1:1) to give 5.00 g of **13** (19.0 mmol, 79% yield) as a white powder: mp 84~86°C. IR (KBr) ν_{\max} 3445, 2843, 1700, 1518, 1213 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 6.86 (1H, d, *J*=8.8 Hz), 6.82 (1H, dd, *J*=8.8, 2.4 Hz), 6.73 (1H, d, *J*=2.4 Hz), 3.91 (3H, s), 3.88 (3H,

s), 3.34 (3H, s). ^{13}C NMR (100 MHz, CDCl_3) δ 157.1 (q, $J=36.8$ Hz), 149.4, 133.4, 119.7, 116.4 (q, $J=288.6$ Hz), 111.0, 110.6, 56.1, 56.0, 39.9. FAB-MS (positive): m/z 263 (M^+). Anal. Calcd for $\text{C}_{11}\text{H}_{12}\text{F}_3\text{NO}_3$: C, 50.20, H, 4.60, N, 5.32. Found: C, 50.11, H, 4.73, N, 5.24.

3,4-Dihydroxy-*N*-methyl-*N*-trifluoroacetylaniline (**14**)

To a solution of AlCl_3 (2.53 g, 19.0 mmol) and ethanethiol (19.0 ml) in 30 ml of CH_2Cl_2 solution was added **13** (1.00 g, 3.80 mmol) under argon at 0°C , and the solution was stirred for 5 hours⁶. Then, the solution was concentrated to dryness. The residue was chromatographed on silica gel eluted with EtOAc-*n*-hexane (1 : 5 to 1 : 1), and 893 mg of **14** (3.80 mmol, quantitative yield) was obtained as a white powder: mp $115\sim 116^\circ\text{C}$. IR (KBr) ν_{max} 3447, 3368, 1680, 1520, 1225, 1206, 1167 cm^{-1} . ^1H NMR (400 MHz, acetone- d_6) δ 8.29 (2H, br), 6.88 (1H, d, $J=8.3$ Hz), 6.87 (1H, d, $J=2.9$ Hz), 6.74 (1H, dd, $J=8.3, 2.9$ Hz), 3.27 (3H, s). ^{13}C NMR (100 MHz, acetone- d_6) δ 157.4 (q, $J=35.0$ Hz), 147.0, 146.8, 134.2, 120.4, 118.1 ($J=288.6$ Hz), 116.5, 116.0, 40.4. HRFAB-MS (positive): calcd for $\text{C}_9\text{H}_8\text{F}_3\text{NO}_3$ 235.0456, found 235.0447 (M^+).

3,4-Bis[(*tert*-butyldimethylsilyloxy)]-*N*-methyl-*N*-trifluoroacetylaniline (**15**)

To a solution of **14** (507 mg, 2.15 mmol) in 15 ml of CH_2Cl_2 were successively added 2,6-lutidine (1.90 ml, 16.3 mmol) and *tert*-butyldimethylsilyl trifluoromethanesulfonate (1.78 ml, 7.74 mmol) under argon at 0°C , and the solution was stirred at room temperature for 12 hours. The solution was poured into water and extracted with EtOAc twice. The organic layer was washed with saturated NaHCO_3 and brine successively, then, dried over Na_2SO_4 and concentrated to dryness. The resulting dark brown oil was purified by silica gel column chromatography with EtOAc-*n*-hexane (1 : 20 to 1 : 10) as eluent to give 887 mg of colorless needles **15** (2.13 mmol, 99%): mp $88\sim 89^\circ\text{C}$. IR (KBr) ν_{max} 3438, 2861, 1713, 1510, 1327, 1254, 909, 843 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 6.82 (1H, d, $J=9.3$ Hz), 6.67~6.71 (2H, m), 3.31 (3H, s), 0.99 (9H, s), 0.98 (9H, s), 0.22 (6H, s), 0.22 (6H, s). ^{13}C NMR (100 MHz, CDCl_3) δ 147.1 (q, $J=36.8$ Hz), 147.7, 147.3, 133.7, 121.0, 120.4, 120.3, 116.4 (q, $J=286.7$ Hz), 39.8, 25.9, 25.8, 18.5, 18.4, -4.1, -4.3. HRFAB-MS (positive): calcd for $\text{C}_{21}\text{H}_{37}\text{F}_3\text{NO}_3\text{Si}_2$ 464.2264, found 464.2235 ($\text{M}+\text{H}^+$).

3,4-Bis[(*tert*-butyldimethylsilyloxy)]-*N*-methylaniline (**16**)

Compound **15** was dissolved in 10 ml of aqueous methanol solution of K_2CO_3 (7% K_2CO_3 -MeOH = 2 : 5). The solution was stirred at room temperature for 3 hours. Then, the reaction mixture was extracted with CH_2Cl_2 twice. The organic layer was dried over Na_2SO_4 and concentrated *in vacuo*. The residue was purified by elution from a silica gel column with EtOAc-*n*-hexane (1 : 15 to 1 : 10) to give 142 mg of **16** (0.387 mmol, 74%

yield) as colorless needles: mp $34\sim 35^\circ\text{C}$. IR (KBr) ν_{max} 3428, 2857, 1617, 1514, 1275, 1251, 840 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 6.67 (1H, d, $J=8.8$ Hz), 6.17 (1H, d, $J=2.9$ Hz), 6.12 (1H, dd, $J=8.8, 2.9$ Hz), 2.76 (3H, s), 0.98 (9H, s), 0.97 (9H, s), 0.19 (6H, s), 0.15 (6H, s). ^{13}C NMR (100 MHz, CDCl_3) δ 147.3, 144.2, 138.6, 121.5, 106.0, 105.5, 31.4, 26.0, 26.0, 18.5, 18.4, -4.1, -4.2. HRFAB-MS (positive): calcd for $\text{C}_{19}\text{H}_{37}\text{NO}_3\text{Si}_2$ 367.2363, found 367.2365 (M^+).

3,4-Bis[(*tert*-butyldimethylsilyloxy)]-*N*-methyl-*N*-nitrosoaniline (**17**)

To a solution of **16** (130 mg, 0.353 mmol) in 15 ml of THF were added 3 ml of 1 N HCl and NaNO_2 (26.7 mg, 0.387 mmol) successively at 0°C . The solution was stirred at 0°C for 3 hours. Then, the reaction mixture was extracted with CH_2Cl_2 twice. The organic layer was dried over Na_2SO_4 and concentrated *in vacuo*. The residue was purified by silica gel column chromatography eluting with EtOAc-*n*-hexane (1 : 15 to 1 : 10) to give 131 mg of **17** (0.329 mmol, 93% yield) as a bright yellow powder: mp $37\sim 38^\circ\text{C}$. IR (KBr) ν_{max} 3461, 2861, 1603, 1518, 1445, 1345, 905, 841 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 7.08 (1H, d, $J=1.5$ Hz), 6.91 (2H, d, $J=1.5$ Hz), 3.41 (3H, s), 1.01 (9H, s), 1.00 (9H, s), 0.23 (12H, s). ^{13}C NMR (100 MHz, CDCl_3) δ 147.5, 146.6, 136.1, 121.1, 113.0, 112.3, 32.0, 25.9, 18.5, -4.1. HRFAB-MS (positive): calcd for $\text{C}_{19}\text{H}_{37}\text{N}_2\text{O}_3\text{Si}_2$ 397.2343, found 397.2338 ($\text{M}+\text{H}^+$).

3,4-Dihydroxy-*N*-methyl-*N*-nitrosoaniline (**2**)

To a solution of **17** (58.3 mg, 0.147 mmol) in 6 ml of THF was added 1.5 ml of NaF-HF buffer (pH 4.98, prepared as reported⁷) under argon at 0°C . The solution was stirred at room temperature for 24 hours. The reaction mixture was extracted with CHCl_3 three times. The organic layer was dried over Na_2SO_4 and concentrated *in vacuo*. The residue was purified on preparative TLC developed with MeOH- CHCl_3 (1 : 5) to give 24.5 mg of **2** (0.147 mmol, quantitative yield) as a brown powder: mp $120\sim 122^\circ\text{C}$. IR (KBr) ν_{max} 3405, 1607, 1537, 1466, 1368, 1256, 1192, 870, 801 cm^{-1} . ^1H NMR (400 MHz, acetone- d_6) δ 8.32 (2H, br), 7.12 (1H, d, $J=2.4$ Hz), 6.88~6.95 (2H, m), 3.35 (3H, s). ^{13}C NMR (100 MHz, acetone- d_6) δ 147.0, 146.0, 136.8, 116.7, 112.7, 109.1, 32.6. HRFAB-MS (positive): calcd for $\text{C}_7\text{H}_9\text{N}_2\text{O}_3$ 169.0613, found 169.0608 ($\text{M}+\text{H}^+$).

Physico-chemical Data of Other Dephostatin Analogs

2,4-Dihydroxy-*N*-methyl-*N*-nitrosoaniline (**3**)

Brown powder: mp $116\sim 118^\circ\text{C}$. IR (KBr) ν_{max} 3347, 1609, 1518, 1503, 1424, 1372, 1244, 1169, 835, 791 cm^{-1} . ^1H NMR (400 MHz, acetone- d_6) δ 8.91 (1H, s), 8.65 (1H, s), 7.14 (1H, d, $J=8.3$ Hz), 6.59 (1H, d, $J=2.4$ Hz), 6.47 (1H, dd, $J=8.3, 2.4$ Hz), 3.33 (3H, s). ^{13}C NMR (100 MHz, acetone- d_6) δ 160.3, 153.6, 128.9, 124.1, 108.2, 104.8, 35.4. HRFAB-MS (positive): calcd for $\text{C}_7\text{H}_9\text{N}_2\text{O}_3$

169.0613, found 169.0603 (M+H)⁺.

3,5-Dihydroxy-*N*-methyl-*N*-nitrosoaniline (4)

Brown powder: mp 164~166°C. IR (KBr) ν_{\max} 3094, 1611, 1526, 1422, 1364, 1275, 1175, 851, 789 cm⁻¹. ¹H NMR (400 MHz, acetone-*d*₆) δ 8.64 (2H, s), 6.61 (2H, d, *J*=2.0 Hz), 6.37 (1H, t, *J*=2.0 Hz), 3.37 (3H, s). ¹³C NMR (100 MHz, acetone-*d*₆) δ 160.4, 145.6, 102.5, 99.3, 31.7. HRFAB-MS (positive): calcd for C₇H₉N₂O₃ 169.0613, found 169.0604 (M+H)⁺.

2-Hydroxy-*N*-methyl-*N*-nitrosoaniline (5)

Brown powder: mp 106~107°C. IR (KBr) ν_{\max} 3115, 1603, 1516, 1472, 1418, 1375, 1233, 752 cm⁻¹. ¹H NMR (400 MHz, acetone-*d*₆) δ 7.99 (1H, s), 6.97~7.36 (4H, m), 3.56 (3H, s). ¹³C NMR (100 MHz, acetone-*d*₆) δ 152.4, 131.8, 131.1, 127.9, 121.3, 118.2, 35.0. HRFAB-MS (positive): calcd for C₇H₉N₂O₂ 153.0664, found 153.0676 (M+H)⁺.

3-Hydroxy-*N*-methyl-*N*-nitrosoaniline (6)

Brown powder: mp 125~127°C. IR (KBr) ν_{\max} 3206, 1611, 1599, 1472, 1431, 1372, 1246, 868 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.32~7.37 (2H, m), 7.00 (1H, br), 6.90~6.98 (2H, m), 3.50 (3H, s). ¹³C NMR (100 MHz, CDCl₃) δ 160.0, 145.1, 131.6, 115.4, 111.5, 107.6, 31.8. HRFAB-MS (positive): calcd for C₇H₉N₂O₂ 153.0664, found 153.0666 (M+H)⁺.

5-Hydroxy-2-methoxy-*N*-methyl-*N*-nitrosoaniline (7)

Brown powder: mp 114~116°C. IR (KBr) ν_{\max} 3187, 2840, 1520, 1470, 1424, 1364, 1225, 1088, 862, 815 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 6.94 (3H, br), 5.67 (1H, br), 3.93 (3H, s), 3.40 (3H, s). ¹³C NMR (100 MHz, CDCl₃) δ 150.4, 146.2, 131.1, 117.1, 113.9, 113.5, 56.3, 35.7. HRFAB-MS (positive): calcd for C₈H₁₁N₂O₃ 183.0770, found 183.0777 (M+H)⁺.

2,5-Dimethoxy-*N*-methyl-*N*-nitrosoaniline (8)

Colorless needles, mp 83~84°C. IR (KBr) ν_{\max} 2840, 1515, 1485, 1445, 1390, 1280, 1080, 1045, 1015, 865, 815, 735 cm⁻¹. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.19 (1H, d, *J*=9.3 Hz), 7.07 (1H, dd, *J*=9.3, 3.3 Hz), 3.79 (3H, s), 3.76 (3H, s), 3.27 (3H, s). ¹³C NMR (100 MHz, CDCl₃) δ 153.6, 146.9, 131.7, 115.2, 113.2, 112.4, 56.3, 55.9, 35.0. FAB-MS (positive): *m/z* 197 (M+H)⁺. Anal. Calcd for C₉H₁₂N₂O₃: C, 55.09, H, 6.16, N, 14.28. Found: C, 55.31, H, 6.16, N, 14.31.

2-(*N*-Methyl-*N*-nitrosoamino)-1,4-benzenedicarboxylic acid (9)

Yellow powder: mp 210~212°C. IR (KBr) ν_{\max} 3065, 2813, 2618, 1728, 1701, 1503, 1439, 1422, 1258 cm⁻¹. ¹H NMR (400 MHz, acetone-*d*₆) δ 8.16 (1H, dd, *J*=8.3, 1.5 Hz), 8.05~8.08 (2H, m), 3.42 (3H, s). ¹³C NMR (100 MHz, acetone-*d*₆) δ 167.7, 167.1, 142.8, 136.1, 133.1, 132.5, 130.7, 128.0, 35.8. HRFAB-MS (negative): calcd for C₉H₇N₂O₅ 223.0355, found 223.0346 (M-2H)⁻.

2,5-Dihydroxy-*N*-methyl-*N*-formylaniline (10a)

Brown amorphous. IR (KBr) ν_{\max} 3300, 1659, 1609, 1514, 1460, 1366, 1196, 1090, 1007, 816, 779 cm⁻¹. ¹H NMR (400 MHz, acetone-*d*₆) δ 8.13 (3H, br), 6.85 (1H, d, *J*=8.8 Hz), 6.69 (1H, dd, *J*=8.8, 2.9 Hz), 6.65 (1H, d, *J*=2.9 Hz), 3.12 (3H, s). ¹³C NMR (100 MHz, acetone-*d*₆) δ 163.8, 151.6, 146.3, 130.5, 118.2, 116.1, 115.1, 32.4. HRFAB-MS (positive): calcd for C₈H₁₀NO₃ 168.0661, found 168.0669 (M+H)⁺.

2,5-Dihydroxy-*N*-methyl-*N*-acetylaniline (10b)

White powder: mp 201~204°C. IR (KBr) ν_{\max} 3355, 3264, 1644, 1524, 1456, 1252, 816, 787 cm⁻¹. ¹H NMR (400 MHz, acetone-*d*₆) δ 8.15 (2H, br), 6.86 (1H, d, *J*=8.8 Hz), 6.73 (1H, dd, *J*=8.8, 2.9 Hz), 6.69 (1H, d, *J*=2.9 Hz), 3.09 (3H, s), 1.77 (3H, s). ¹³C NMR (100 MHz, acetone-*d*₆) δ 170.5, 151.7, 146.7, 132.7, 118.1, 116.7, 115.9, 35.6, 21.8. HRFAB-MS (positive): calcd for C₉H₁₂NO₃ 182.0817, found 182.0828 (M+H)⁺.

2,5-Dihydroxy-*N*-methyl-*N*-trifluoroacetylaniline (10c)

Brown powder: mp 158~160°C. IR (KBr) ν_{\max} 3401, 1688, 1518, 1458, 1341, 1244, 1215, 1198, 1161, 823, 783 cm⁻¹. ¹H NMR (400 MHz, acetone-*d*₆) δ 8.23 (2H, br), 6.76~6.87 (3H, m), 3.09 (3H, s). ¹³C NMR (100 MHz, acetone-*d*₆) δ 158.1 (q, *J*=34.9 Hz), 151.7, 147.6, 129.2, 118.4, 118.3, 117.9 (q, *J*=288.7 Hz), 116.9, 38.5. HRFAB-MS (positive): calcd for C₉H₉F₃NO₃ 236.0535, found 236.0531 (M+H)⁺.

2,5-Dihydroxy-*N*-methyl-*N*-methanesulfonylaniline (10d)

White powder: mp 116~118°C. IR (KBr) ν_{\max} 3476, 1510, 1316, 1201, 1145, 820, 787 cm⁻¹. ¹H NMR (400 MHz, acetone-*d*₆) δ 8.01 (2H, br), 6.81 (1H, d, *J*=8.3 Hz), 6.81 (1H, d, *J*=2.9 Hz), 6.71 (1H, dd, *J*=8.3, 2.9 Hz), 3.23, (3H, s), 2.96 (3H, s). ¹³C NMR (100 MHz, acetone-*d*₆) δ 151.4, 148.0, 129.3, 118.0, 117.8, 117.1, 38.2, 37.3. FABHR-MS (positive): calcd for C₈H₁₂NO₄S 218.0487, found 218.0497 (M+H)⁺.

2,5-Dihydroxy-*N*-methyl-*N*-trifluoromethanesulfonylaniline (10e)

White powder: mp 131~133°C. IR (KBr) ν_{\max} 3476, 3320, 1518, 1379, 1325, 1227, 1194, 970, 820 cm⁻¹. ¹H NMR (400 MHz, acetone-*d*₆) δ 8.10~8.70 (2H, br), 6.76~6.88 (3H, m), 3.41, (3H, s). ¹³C NMR (100 MHz, acetone-*d*₆) δ 151.6, 148.5, 127.3, 121.8 (q, *J*=324.5 Hz), 118.8, 118.5, 118.0, 40.2. HRFAB-MS (positive): calcd for C₈H₈F₃NO₄S 271.0126, found 271.0123 (M+H)⁺.

Acknowledgments

We thank Dr. H. NAGANAWA and Dr. R. SAWA, Institute of Microbial Chemistry, for valuable suggestions on structure determination.

References

- 1) GOULD, K. L.; S. MORENO, N. K. TONKS & P. NURSE: Complementation of the mitotic activator p80^{cdc25}, by a human protein-tyrosine phosphatase. *Science* 250: 1573~1576, 1990
- 2) KORETZKY, G. A.; J. PICUS, T. SCHULTZ & A. WEISS: Tyrosine phosphatase CD45 is required for T-cell antigen receptor and CD2-mediated activation of a protein tyrosine kinase and interleukin 2 production. *Proc. Natl. Acad. Sci. U.S.A.* 88: 2037~2044, 1991
- 3) IMOTO, M.; H. KAKEYA, T. SAWA, C. HAYASHI, M. HAMADA, T. TAKEUCHI & K. UMEZAWA: Dephostatin, a novel protein tyrosine phosphatase inhibitor produced by *Streptomyces*. I. Taxonomy, isolation, and characterization. *J. Antibiotics* 46: 1342~1346, 1993
- 4) KAKEYA, H.; M. IMOTO, Y. TAKAHASHI, H. NAGANAWA, T. TAKEUCHI & K. UMEZAWA: Dephostatin, a novel protein tyrosine phosphatase inhibitor produced by *Streptomyces*. II. Structure determination. *J. Antibiotics* 46: 1716~1719, 1993
- 5) WATANABE, T.; T. TAKEUCHI, M. OTSUKA & K. UMEZAWA: Total synthesis of dephostatin, a novel protein tyrosine phosphatase inhibitor. *J. Chem. Soc., Chem. Commun.* 437~438, 1994
- 6) NODE, M.; K. NISHIDE, K. FUJI & E. FUJITA: Hard acid and soft nucleophile system. 2. Demethylation of methyl ether of alcohol and phenol with an aluminum halide-thiol system. *J. Org. Chem.* 45: 4275~4277, 1980
- 7) KENDALL, P. M.; J. V. JOHNSON & C. E. COOK: Synthetic route to an aromatic analogue of strigol. *J. Org. Chem.* 44: 1421~1424, 1979