

Tetrahedron Letters 42 (2001) 4029-4031

TETRAHEDRON LETTERS

Chemoenzymatic total synthesis of (+)-conagenin, a low-molecular-weight immunomodulator

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Abstract—Total synthesis of (+)-conagenin, a low-molecular-weight immunomodulator, was achieved based on enantioselective enzymatic reactions followed by chemoselective reductions. © 2001 Elsevier Science Ltd. All rights reserved.

(+)-Conagenin (CNG, 1), a natural product originally isolated from fermentation broths of Streptomyces roseosporus, has been found to stimulate activated T cells by producing lymphokines as a low-molecularweight immunomodulator.^{1–3} In addition, the improvement of efficacy of antitumor agents, such as mitomycin C and adriamycin, by CNG (1) has also been reported.⁴⁻⁹ These biological activities of CNG (1), possessing an α -substituted serine moiety, has made this compound an intriguing target for total synthesis. An asymmetric total synthesis¹⁰ and an asymmetric formal synthesis¹¹ of CNG (1) have been reported, as have investigations of further syntheses^{12,13} of diastereoisomers of CNG (1). We report herein a successful convergent method for the total synthesis of CNG (1) based on enantioselective enzymatic reactions followed by chemoselective reductions.

The retrosynthetic analysis of CNG (1) is outlined in Scheme 1. (S)-Z- α -Methylserine (5) was synthesized in 94% ee { $[\alpha]_{D}^{26} + 10.1^{\circ}$ (c 0.88, EtOH)} as a colorless

amorphous powder from σ -symmetric prochiral diethyl α -aminomalonate (4) by utilizing our chemoenzymatic methodology based on enantioselective enzymatic hydrolysis followed by chemoselective enantiodivergent reduction.¹⁴ Methylation of 5 with (trimethylsilyl)-diazomethane gave the (*S*)-Z- α -methylserine methylester (6) in 66% yield. Hydrogenolytic deprotection of the *N*-Z group of 6, followed transesterification of the resultant (*S*)- α -methylserine methylester in BnOH and Et₃N (10:1) at 60°C, afforded (*S*)- α -methylserine benzyl ester (7) {colorless oil, $[\alpha]_{D}^{26}$ –9.0° (*c* 0.66, CHCl₃)} in 44% yield from 6, as shown in Scheme 2.

Pentanoic acid **13** was also prepared by the chemoenzymatic sequence illustrated in Scheme 3. Photosensitized oxygenation of 5-methylcyclopentadiene (**8**),^{15,16} followed by in situ reduction with thiourea, furnished σ -symmetric prochiral 5-methyl-2-cyclopentene-1,4-diol (**2**) in 35% yield.¹⁷ Enzymatic acetylation of **2** with lipase AK (Amano Pharmaceutical Co. Ltd) in the



Scheme 1. Retrosynthetic analysis of (+)-conagenin (CNG, 1).

Keywords: amino acids and derivatives; asymmetric synthesis; enzymes and enzyme reactions; natural products; reduction. * Corresponding author. Fax: +81-88-633-9503; e-mail: ynagao@ph2.tokushima-u.ac.jp

$$4 \xrightarrow{a, b} 5 \xrightarrow{c} \xrightarrow{Z-H} \xrightarrow{(S)} \xrightarrow{CO_2Me} \xrightarrow{d, e} \xrightarrow{H_2N} \xrightarrow{(S)} \xrightarrow{CO_2Bn} \xrightarrow{Me} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{OH} \xrightarrow{G} \xrightarrow{OH} \xrightarrow{O$$

Scheme 2. (a) Porcine liver esterase (PLE)/1/15 M phosphate buffer (pH 7.0)–MeCN (10:1)/rt/12 h; (b) LiBH₄/Et₂O–THF (3:1)/reflux/1.5 h; (c) TMSCHN₂/MeOH–benzene (2:7)/rt/4 h; (d) $H_2/10\%$ Pd–C/MeOH/rt/12 h; (e) BnOH–Et₃N (10:1)/ 60°C/2 days.

presence of vinyl acetate as an acyl donor afforded a chiral mono ester 9 {colorless oil, $[\alpha]_{D}^{25}$ -76.3° (c 0.98, $CHCl_3$) in 94% yield with high enantiomeric excess (99% ee).¹⁸ The absolute configuration of **9** was determined by ¹H NMR analysis (400 MHz, CDCl₃) of the [(R)-Mosher ester and (S)-methoxy(trifluoromethyl)phenylacetate (MTPA)] derivatives of 9.19 After benzylation of 9 with BnBr in the presence of Ag₂O in 87% yield, oxidative cleavage of the double bond of the resulting benzyl ether 10 {colorless oil, $[\alpha]_D^{26}$ -4.1° (c 1.01, CHCl₃), followed by alkaline hydrolysis of the acetoxy group, lactonization with cyanuric chloride, and chemoselective reduction of the activated carboxyl group,²⁰ provided the desired primary alcohol 11 {colorless column from Et₂O, mp 61–63°C, $[\alpha]_D^{26}$ +182.4° (c 1.02, CHCl₃)} without isolation of each intermediate (23% yield, four steps). Radical-induced deoxygenation of 11 was performed using Barton's method²¹ to give the corresponding chiral lactone 3 {colorless powder from Et₂O, mp 91–94°C, $[\alpha]_D^{25}$ $+146.7^{\circ}$ (c 1.00, CHCl₃) (46% yield, two steps). The desired stereochemistry of 3 was revealed by NOE analysis (300 MHz, CDCl₃) as follows. When C3-H of 3 was irradiated, an NOE enhancement of the C4- and C5-Me protons was recognized, as shown in Fig. 1. A similar NOE enhancement between the C4- and C5-Me protons of 3 was also observed. Treatment of the lactone 3 with aqueous NaOH in MeOH followed by methylation with MeI gave methyl ester **12** {colorless oil, $[\alpha]_D^{26}$ +89.5° (*c* 1.00, CHCl₃)} in 85% yield. After benzylation of **12** with benzyl 2,2,2-trichloroacetimidate in the presence of BF₃·Et₂O,²² the resulting ester was subjected to alkaline hydrolysis to give pentanoic acid **13** {colorless oil, $[\alpha]_D^{26}$ +3.1° (*c* 1.04, CHCl₃)} in 63% yield from **12**.

Condensation of α -methylserine derivative 7 with pentanoic acid **13** accompanied by 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDC·HCl) in the presence of *N*-methylmorpholine (NMM) and 1-hydroxybenzotriazole (HOBt) in DMF furnished the protected CNG **14** {colorless oil, $[\alpha]_D^{26} + 22.7^\circ$ (*c* 0.52, CHCl₃)} as an enantiomerically pure form in 57% yield (Scheme 4). Finally, compound **14** was submitted to catalytic hydrogenolysis on Pd–C to give CNG **(1)** {colorless powder, $[\alpha]_D^{25} + 48.4^\circ$ (*c* 0.40, MeOH), lit.¹ $[\alpha]_D^{27} + 55.4^\circ$, lit.¹⁰ $[\alpha]_D^{26} + 48.7^\circ$ (*c* 0.43, MeOH)} in 95% yield. The accomplishment of this total asymmetric synthesis was confirmed by the identity of all physicochemical data of the synthesized and the natural CNG **(1)**.²³

In summary, we have described a convergent and chemoenzymatic total synthesis of (+)-conagenin (CNG, 1). This methodology will also be applicable for the synthesis of a variety of stereoisomers of CNG (1).



Figure 1. Selected ${}^{1}H-{}^{1}H$ NOE enhancements (300 MHz ${}^{1}H$ NMR, CDCl₃) for 3.



Scheme 3. (a) $O_2/hv/rose$ bengal/AcONa/MeOH/-78°C/80 min; (b) $(NH_2)_2CS/MeOH/rt/13$ h; (c) lipase AK/CH₂=CHOAc/THF/rt/7 h; (d) Ag₂O/BnBr/KI/toluene/rt/15.5 h; (e) NaIO₄/KMnO₄/Na₂CO₃/acetone-H₂O (5:6)/rt/1 h; (f) 1N NaOH/MeOH/rt/1 h; (g) H⁺; (h) cyanuric chloride/NMM/DME/rt/3 h; (i) NaBH₄/0°C/10 min; (j) PhOCSCl/DMAP/CH₂Cl₂/rt/4 h; (k) AIBN/n-Bu₃SnH/toluene/75°C/1 h; (l) 1N NaOH/MeOH/rt/1 h; (m) H⁺ (pH 6–7); (n) K₂CO₃/MeI/acetone/rt/3.5 h; (o) BnOCNHCCl₃/BF₃·Et₂O/cyclopentane/rt/2 h; (p) 1N K₂CO₃/MeOH/reflux/2 h.

Scheme 4. (a) EDC·HCl/NMM/HOBt/DMF/rt/6 h; (b) $H_2/10\%$ Pd–C/MeOH/rt/5 days.

Acknowledgements

This work was partially supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science, Sports and Culture, Japan. The authors are grateful to Professor S. Hakakeyama (Nagasaki University) for useful discussions and for kindly providing (+)-conagenin and its spectral data. They would also like to thank Amano Pharmaceutical Co. Ltd for the supply of lipase AK.

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- Iversen, T.; Bundle, D. R. J. Chem. Soc., Chem. Commun. 1981, 1240–1241.
- 23. Synthesized (+)-conagenin (CNG, 1): ¹H NMR (400 MHz, CD₃OD) δ 0.93 (3 H, d, J=7.1 Hz), 1.22 (3 H, d, J = 6.3 Hz), 1.50 (3 H, s), 1.89 (1 H, dq, J = 2.4, 6.3 Hz), 3.84 (1 H, d, J=11.0 Hz), 3.85 (1 H, q, J=6.1 Hz), 4.04 $(1 \text{ H}, d, J=11.0 \text{ Hz}), 4.15 (1 \text{ H}, d, J=2.4 \text{ Hz}); {}^{13}\text{C NMR}$ (75 MHz, CD₃OD) δ 8.4, 20.0, 21.2, 43.7, 62.8, 66.1, 71.2, 75.1, 175.8, 176.9. The synthesized methyl ester of (+)-conagenin (CNG, 1) also had spectroscopic properties identical to the authentic sample:¹⁰ ¹H NMR (400 MHz, CDCl₃) δ 0.92 (3 H, d, J=7.1 Hz), 1.26 (3 H, d, J=6.3 Hz), 1.55 (3 H, s), 1.8 (2 H, brs), 2.09 (1 H, tq, J=2.1, 7.1 Hz), 2.40 (1 H, brs), 3.81 (3 H, s), 3.84 (1 H, d, J=11.2 Hz), 3.9 (1 H, brs), 4.06 (1 H, d, J=11.2 Hz), 4.35 (1 H, d, J=2.2 Hz), 7.51 (1 H, brs); ¹³C NMR (75 MHz, CDCl₃) & 4.9, 20.5, 21.9, 40.3, 53.0, 61.9, 66.8, 72.2, 77.2, 173.0, 173.3