

Bioorganic & Medicinal Chemistry 10 (2002) 2193-2198

Synthesis and Anti-tubercular Evaluation of 4-Quinolylhydrazones

Luisa Savini,* Luisa Chiasserini, Alessandra Gaeta and Cesare Pellerano

Dipartimento Farmaco Chimico Tecnologico, Università degli Studi di Siena, Via A. Moro, 53100 Siena, Italy

Received 16 October 2001; revised 20 December 2001; accepted 6 February 2002

Abstract—A series of 4-quinolylhydrazones were synthesized and tested against *Mycobacterium tuberculosis* H37Rv. Preparation of the title compounds was achieved by reaction of 4-quinolylhydrazine and aryl- or heteroaryl-carboxaldehyde. For the most of derivatives interesting antitubercular properties were showed; two compounds (3_2 and 3_{25}), identified as the most active, were tested also against *Mycobacterium avium*. © 2002 Elsevier Science Ltd. All rights reserved.

Introduction

Tuberculosis (TB), an infection of *Mycobacterium tuberculosis* still remains the leading cause of worldwide death among infectious diseases;^{1,2} one-third of the population is infected with *M. tuberculosis* and the World Health Organization (WHO) estimates that within the next 20 years about 30 million people will catch tuberculosis.^{3–7}

The resurgence of TB is associated with the emergence of HIV/AIDS epidemic⁸ and the fast development of multidrug resistant TB bacterial strains.^{9,10} In the developing world, probably 50% of HIV seropositive individuals are co-infected with TB. Therefore taking into account what is reported above there is a pressing need to develop new and more effective anti-tubercular agents.

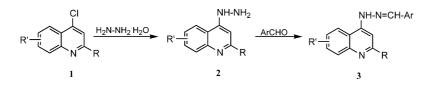
Among the new structures recently reported as potential anti-tubercular of particular interest seems to be some 2,6-bis(alkylthio)4-pyridinecarboxamides,¹¹ *N*-alkyl-1,2-dihydro-2-thioxo-3-pyridinecarbothioamides,¹² 6-chloro-

3 - phenyl - 4 - thioxo - 2H - 1,3 - benzoxazine - 2(3H) - ones, -dithiones¹³ and fluoroquinolones (ciprofloxacin, ofloxacin, levofloxacin, sparfloxacin, gatifloxacin and moxifloxacin).¹⁴

Within a program aimed at developing new quinoline derivatives, we previously synthesized numerous quinolylhydrazones, some of which resulted were endowed with anti-mycoplasmal, anti-bacterial, anti-cestode, anti-viral, anti-tumoral and anti-tubercular properties.^{15–24}

Therefore, as a consequence of previous researches and on the basis of the obtained results, a series of 4-quinolylhydrazones **3**, with various substituents on quinoline nucleus and with aryl- or heteroaryl-hydrazonic moiety, were synthesized as potential anti-tubercular agents.

The new synthesized quinolylhydrazones 3 and the starting quinolylhydrazines 2 were tested against M. *tuberculosis* H37Rv and interesting anti-tubercular properties resulted for most of the title compounds.



R,R', Ar as defined in Table 1

Scheme 1. R,R', Ar as defined in Table 1.

*Corresponding author. Tel.: +39-0577-234304; fax: +39-0577-234333; e-mail: savini@unisi.it

0968-0896/02/\$ - see front matter \odot 2002 Elsevier Science Ltd. All rights reserved. P11: S0968-0896(02)00071-8

Chemistry

The synthesis of 4-quinolylhydrazones **3** was carried out, according to the previously described procedure,¹⁸ by reaction of equimolar amounts of 4-hydrazinoquinoline **2** and the appropriate carboxaldehyde in EtOH, as illustrated in Scheme 1. The 4-quinolylhydrazine 2 necessary for preparation of 4-quinolylhydrazones 3 were obtained from the corresponding 4-chloroquinoline 1 and hydrazine hydrate as we previously described.²⁴

The structures assigned to the synthesized compounds are in good agreement with elemental analyses, IR and

Table 1. Physicochemical data and molecular formula of derivatives 2 and 3

Compd	R	R′	Ar	Mp (°C)	Cryst. solvent	Molecular formula
2 ₁ ^a	Н	7-OCH ₃				
$\frac{1}{2}_{2}^{a}$	CH ₃	7-OCH ₃				
2 ₃ ^a	CH ₃	8-OCH ₃				
$2_{4^{a}}$	C_6H_5	6-OCH3				
2 ₄ 2 ₅	Н	7-OC ₂ H ₅		250-253	EtOH	C11H13N3O·HCl
$\mathbf{\hat{2}}_{6_{1}^{a}}^{a}$	CH ₃	$7-OC_2H_5$ 7-OC_2H_5		250 255	LIOII	enning vonter
2 ₇ ^b	Н	6- <i>n</i> .C ₄ H ₉				
27 2 °	CH ₃	6- <i>n</i> .C ₄ H ₉				
28 ^c 29 ^b	Н	6- <i>n</i> .OC ₄ H ₉				
2 9	CH ₃	$6-n.OC_4H_9$				
2_{10}^{c}	H H			287-291	EtOH anhyd.	C II N IICI
2 ₁₁		6-Cyclohexyl		287-291	ElOH annyu.	C ₁₅ H ₁₉ N ₃ ·HCl
2_{12}^{c} 2_{13}^{a}	CH ₃	6-Cyclohexyl				
2_{13}^{a}	Н	7-Cl				
2 ₁₄ ^b	CH ₃	7-Cl				~ ~ ~ ~ ~ ~
2 ₁₅	Н	5,7-Cl		184–187	EtOH	$C_9H_7Cl_2N_3$
2 ₁₆	CH ₃	5,7-Cl		202-205	EtOH	$C_{10}H_9Cl_2N_3$
2_{17}^{a}	CH_3	6-F				
3 ₁	Н	Н	4-OCH ₃ -naphthyl	195 (dec.)	Benzene	$C_{21}H_{17}N_3O$
3 ₂	Н	7-OCH ₃	2-OCH ₃ -naphthyl	118-121	EtOH	$C_{22}H_{19}N_3O_2 \cdot 1/4H_2O$
3 ₃	Н	7-OCH ₃	4-OCH ₃ -naphthyl	235-238	EtOH	$C_{22}H_{19}N_3O_2$
3 ₄	Н	7-OCH ₃	$4 - N(C_2H_5)_2 - C_6H_4$	201 (dec.)	EtOH-H ₂ O	$C_{21}H_{24}N_4O \cdot 1/4H_2O$
3 ₅	CH_3	7-OCH ₃	2-OCH ₃ -naphthyl	202 (dec.)	EtOH	$C_{23}H_{21}N_3O_2 \cdot H_2O$
3 ₆	CH ₃	7-OCH ₃	4-OCH ₃ -naphthyl	286-289	EtOH	C ₂₃ H ₂₁ N ₃ O ₂ ·HCl·H ₂ O
3 ₇	CH ₃	7-OCH ₃	$4 - N(C_2H_5)_2 - C_6H_4$	128-132	EtOH-H ₂ O	$C_{22}H_{26}N_4O\cdot H_2O$
3 ₈	CH ₃	7-OCH ₃	$3,4-(OCH_2O)-C_6H_3$	215 (dec.)	EtOH	C ₁₉ H ₁₇ N ₃ O ₃
3 ₉	CH ₃	8-OCH ₃	4-OCH ₃ -naphthyl	261-262	EtOH-H ₂ O	$C_{23}H_{21}N_{3}O_{2}$
3 ₁₀	Н	$7-OC_2H_5$	2-OCH ₃ -naphthyl	95 (dec.)	EtOH	$C_{23}H_{21}N_{3}O_{2}\cdot 1/2H_{2}O$
3 ₁₁	Н	$7-OC_2H_5$	4-OCH ₃ -naphthyl	200-202	EtOH	$C_{23}H_{21}N_{3}O_{2}$
3 ₁₂	Н	$7-OC_2H_5$	$4-N(C_2H_5)_2-C_6H_4$	119-122	EtOH	$C_{22}H_{26}N_4O \cdot 1/2H_2O$
3 ₁₃	CH ₃	$7-OC_2H_5$	2-OCH ₃ -naphthyl	232-235	EtOH	$C_{24}H_{23}N_3O_2$
3 ₁₄	CH ₃	$7-OC_2H_5$	4-OCH ₃ -naphthyl	231–234	EtOH	$C_{24}H_{23}N_3O_2\cdot 3/4H_2O$
3 ₁₅	CH ₃	$7-OC_2H_5$	$4-N(C_2H_5)_2-C_6H_4$	115-118	EtOH	C ₂₃ H ₂₈ N ₄ O·H ₂ O
3 ₁₆	C_6H_5	6-OCH ₃	4-NHCOCH ₃ -C ₆ H ₄	138 (dec.)	EtOH-H ₂ O	C ₂₅ H ₂₂ N ₄ O ₂
3_{17}^{16}	C_6H_5	6-OCH3	5-NO ₂ -2-furyl	156 (dec.)		$C_{23}H_{21}N_3O_2$
3 ₁₈	H	$6-n.C_4H_9$	4-OCH ₃ -naphthyl	195-196	EtOH-H ₂ O	$C_{25}H_{25}N_{3}O$
3 ₁₉	CH ₃	$6 - n.C_4H_9$	4-OCH ₃ –naphthyl	237-239	EtOH H ₂ O	$C_{26}H_{27}N_{3}O$
3 ₂₀	CH ₃	$6 - n.C_4H_9$	$3,4-(OCH_2O)-C_6H_3$	202-204	EtOH	$C_{22}H_{23}N_3O_2$
3_{20} 3_{21}	Н	6- <i>n</i> .OC ₄ H ₉	$4-OCH_3-naphthyl$	202-204	EtOH	$C_{25}H_{25}N_3O_2$
3_{21}^{3}	CH ₃	6- <i>n</i> .OC ₄ H ₉	4-OCH ₃ –naphthyl	209-210	EtOH-H ₂ O	
3 ₂₂ 3 ₂₃	Н	6-Cyclohexyl	4-OCH ₃ -naphthyl	182–184	EtOH-H ₂ O EtOH	$C_{26}H_{27}N_3O_2 \\ C_{27}H_{17}N_3O$
3_{23}^{23} 3_{24}^{b}	H	6-Cyclohexyl	$1-C_6H_5-2,5-CH_3-3-pyrrolyl$	182-184	LIOII	
324	CH ₃			252 254	E+OULULO	$C_{26}H_{27}N_{3}O$
3 ₂₅		6-Cyclohexyl	C_6H_5	252-254	EtOH-H ₂ O	$C_{23}H_{25}N_3$
3 ₂₆	CH ₃	6-Cyclohexyl	$4-N(C_2H_5)_2-C_6H_4$	163-166	EtOH-H ₂ O	$C_{27}H_{34}N_4$
3 ₂₇	CH ₃	6-Cyclohexyl	4-OCH ₃ -naphthyl	290-292	DMF	C ₂₈ H ₂₉ N ₃ O
3 ₂₈ ^b	CH_3	6-Cyclohexyl	$1-C_6H_5-2,5-CH_3-3-pyrrolyl$	227 220	E-OU	
3 ₂₉	H	7-Cl	2-OCH ₃ -naphthyl	237-239	EtOH	$C_{21}H_{16}CIN_3O$
3 ₃₀	H	7-Cl	4-OCH ₃ -naphthyl	132-136	EtOH	$C_{21}H_{16}ClN_3O\cdot3/4H_2O$
3 ₃₁ ^e	Н	7-Cl	$4-N(C_2H_5)_2-C_6H_4$	245-248	EtOH	$C_{20}H_{21}CIN_4$
3 ₃₂	CH ₃	7-Cl	2-OCH ₃ -naphthyl	259-262	EtOH	$C_{22}H_{18}ClN_3O$
3 ₃₃	CH ₃	7-Cl	4-OCH ₃ -naphthyl	224-226	EtOH	$C_{22}H_{18}ClN_3O$
3 ₃₄	CH_3	7-Cl	$4 - N(C_2H_5)_2 - C_6H_4$	249-252	EtOH	$C_{21}H_{23}ClN_4$
3 ₃₅	Н	5,7-Cl	3,4-(OCH ₂ O)–C ₆ H ₃	223-225	EtOH	$C_{17}H_{11}Cl_2N_3O_2$
3 ₃₆	Н	5,7-Cl	4-OCH ₃ -naphthyl	164–167	EtOH	$C_{21}H_{15}Cl_2N_3O \cdot 1/4H_2Cl_2N_3O \cdot 1/4H_2O \cdot$
3 ₃₇	CH_3	5,7-Cl	3,4-(OCH ₂ O)–C ₆ H ₃	218-220	EtOH	$C_{18}H_{13}Cl_2N_3O_2$
•	CH_3	5,7-Cl	4-OCH ₃ -naphthyl	158-160	EtOH	C ₂₂ H ₁₇ Cl ₂ N ₃ O
3 ₃₈	CH ₃	6-F				

^aRef 24.

^bRef 25. ^cRef 26.

^dRef 15.

^eRef 27.

¹H NMR spectral data. The IR spectra of quinolylhydrazones **3** have a weak band between 3350 and 3170 cm^{-1} due to the stretching vibration of the NH group. In the ¹H NMR spectra a singlet appears between 8.3–9.2 δ ,

Table 2. In vitro evaluation of antimicrobial activity versus *Mycobacterium tuberculosis* H37Rv [minimum inhibitory concentration (MIC) in µg/mL]

Compd		M. tuberculosis			
	MIC (µg/mL)	% Inhibition	MIC (µg/mL)	$\begin{array}{c} IC_{50} \\ (\mu g/mL) \end{array}$	SI
2 ₁ ^b	<12.5	97	12.5		
2 ₂	> 12.5	94	12.5		
2 ₃	> 12.5	54	()5	2 70	0.00
$\frac{2_4}{2}$	< 12.5	99 88	6.25	3.78	0.60
2 ₅ 2 ₆	>12.5 <12.5	80 98	12.5	25.2	2.02
2 ₆ 2 ₇ ^b	<12.5	99	6.25	23.2	2.02
$\frac{2}{2_8}$	<12.5	99	6.25	13.40	2.14
$\frac{1}{2}$	<12.5	99	12.5	7.77	0.62
2 ₁₀	<12.5	99	6.25	5.94	0.95
2 ₁₁	<12.5	99	6.25	10.28	1.64
2 ₁₂	<12.5	100	6.25	5.31	0.85
2 ₁₃	<12.5	98	12.5	53.54	4.28
2 ₁₄	<12.5	97	12.5	76.97	6.16
2 ₁₅	> 12.5	82			
2 ₁₆	>12.5	87			
2 ₁₇	>12.5	69			
3 ₁	< 12.5	100	3.13	3.50	1.10
3 ₂ ^a	< 12.5	100	6.25	73	11.68
3 ₃	< 12.5	100	0.78	7.20 7.20	9.23
3 ₄	<12.5 <12.5	100 100	12.5 3.13	23	0.58 7.00
3 ₅ 3 ₆ ^b	<12.5	99	1.56	25	7.00
3 ₇ ^b	<12.5	100	12.5		
3_{8}	<12.5	100	12.5	5.40	0.43
3 ₉	<12.5	100	1.56	2.70	1.73
3 ₁₀	<12.5	100	3.13	2.70	1.75
3 ₁₁	<12.5	100	6.25	< 4.20	< 0.70
3 ₁₂	<12.5	100	6.25	6.40	1.02
3 ₁₃	<12.5	100	6.25	6.90	1.10
3 ₁₄	<12.5	100	1.56	3.60	2.31
3 ₁₅	<12.5	100	12.5	6.20	0.50
3 ₁₆ ^b	<12.5	95	12.5		
3 ₁₇ ^b	>12.5	72	12.5		
3 ₁₈	<12.5	100	6.25	5.70	0.91
3 ₁₉	<12.5	100	6.25	9.80	1.57
3 ₂₀	< 12.5	100	6.25	6.80	1.09
3 ₂₁	< 12.5	100	6.25	3.40	0.54
3 ₂₂	< 12.5	100	0.78	5.20	6.67
3 ₂₃	< 12.5	100	3.13	13.20	4.22
3_{24} 3_{25}^{a}	<12.5 <12.5	100 100	6.25 6.25	12.60 64	2.02 10.24
3_{25} 3_{26}	<12.5	100	1.56	12.1	7.76
3 ₂₇	<12.5	100	1.56	13.1	8.4
3_{28}	<12.5	100	1.56	6.60	4.23
3_{29}^{28}	<12.5	99	6.25	5.00	
3 ₃₀	<12.5	99	3.13	3.40	1.09
3 ₃₁	<12.5	100	3.13		
3 ₃₂	<12.5	100	6.25	3.30	0.56
3 ₃₃	<12.5	100	0.78	3.40	4.36
3 ₃₄	<12.5	100	6.25	11.90	1.9
3 ₃₅ ^b	>12.5	4			
3 ₃₆ ^b 3 ₃₇ ^b	>12.5	10			
3 ₃₇ ^b	>12.5	4			
3 ₃₈ ^b	>12.5	53			
3 ₃₉ ^b	<12.5	100	3.13	1.90	0.61
Rifampin (RMP)	0.031	90			

^aMIC against *M. tuberculosis* Erdman was 3.13.

^bInsoluble in DMSO, unable to test.

which is typical of the aldehyde proton. Physicochemical properties are summarized in Table 1.

Pharmacology

The in vitro evaluation of antituberculosis activity was carried out at the GWL Hansen's Disease Center within the Tuberculosis Antimicrobial Acquisition Coordinating Facility (TAACF) screening program for the discovery of novel drugs for treatment of tuberculosis. Under the direction of the US National Institute of Allergy and Infectious Diseases (NIAID), Southern Research Institute coordinates the overall program. The purpose of the screening program is to provide a resource whereby new experimental compounds can be tested for their capacity to inhibit the growth of virulent *M. tuberculosis*.

Biological tests have been performed according to the method described in refs 28–30 and the results are reported in Tables 2–4.

Results and Discussion

From the primary screening against *M. tuberculosis* H37Rv the maiority of the tested compounds showed an inhibitory activity between 95 and 100%, therefore these compounds were submitted to a second level assay to evaluate the actual minimum inhibitory concentration (MIC) and to assess cytotoxicity towards the VERO cell line and selectivity index (SI), defined as the ratio of the measured IC₅₀ in VERO cells to the MIC.

On the basis of the obtained data we can remark the interesting anti-tubercular properties showed by some quinolylhydrazones **3**. The activity is significantly affected by substituents both on quinoline nucleus and hydrazonic moiety. On quinoline nucleus the most effective substituents resulted 6-cyclohexyl, 7-methoxy, 7-ethoxy and 7-chloro, while, as for the hydrazonic moiety, greater effectiveness resulted for *para*- and

Table 3. Evaluation of anti-Mycobacterium avium activity

Compd	MIC $(\mu g/mL)^a$	SI	
3 ₈	3.13	23.32	
3 ₉	25	0.29	
$3_8 3_9 3_{25}$	3.13	20.45	

^aMICs are determined in the MABA against a strain of *M. avium* (A TCC 25291).

 Table 4.
 Macrofage assay

Compd	$MIC \; (\mu g/mL)$	SI	EC ₉₀	EC ₉₉	EC ₉₀ /MIC
$ 3_2 3_9 3_{25} $	6.25	11.68	0.65	2.00	0.10
	0.78	9.23	>1	> 1	> 1.28
	6.25	10.24	0.80	1.61	0.13

The columns labeled EC_{90} and EC_{99} list the concentrations effecting 90 and 99% reduction in residual mycobacterial growth after 7 days compared to untreated controls. Compounds with $EC_{90} > 16 \times MIC$ (as reported in the column labeled EC_{90}/MIC) are considered inactive.

ortho-methoxynaphtyl substituents. MIC values for those compounds ranged from 0.78 to $3.13 \,\mu\text{g/mL}$ (Table 2). The chlorodisubstitution led to inactive derivatives ($\mathbf{3}_{35-39}$). As far as the substitution in position 2 of quinoline nucleus is concerned the methyl appears to be most favourable group. Quinolylhydrazines **2**, even if endowed with appreciable inhibiting action, showed a SI generally below 4, with the only exception of $\mathbf{2}_{13}$ and $\mathbf{2}_{14}$ with a SI = 4.28 and 6.16 respectively, while for some quinolylhydrazones ($\mathbf{3}_{2,3,5,22,25,26,27}$) a significantly higher SI was found (Table 2).

Three compounds (3_2 , 3_3 and 3_{25}), with a high SI value (11.68, 9.23, and 10.24, respectively), were then tested for efficacy in vitro in a TB-infected macrophage model, showing good values of EC₉₀ and EC₉₉. Furthemore, these compounds were evaluated for their inhibitory activity against a single strain of *M. avium*, an opportunistic pathogen which has been associated with tuberculosis in patients infected by HIV (Table 3). As for compounds 3_2 and 3_{25} , which showed a SI of 23.32 and 20.45 respectively, further evaluation in additional *M. avium* assays is in progress.

It is important to point out the low toxicity particularly shown by quinolylhydrazones $\mathbf{3}_2$ and $\mathbf{3}_{25}$ active against *M. tuberculosis* H37Rv and *M. avium*.

In conclusion, for the development of our research, it seems advisable to synthesize new derivatives with suitably chosen substituents both on the quinoline nucleus and hydrazonic moiety. Additional tests are required in order to obtain more definitive and clear answers from structure–activity relationships.

Experimental

Chemistry

Melting points were determined on a Köfler blok or on a Büchi 510 apparatus and are uncorrected. Elemental analyses were performed on a Perkin-Elmer 240 C Elemental Analyzer by Laboratories of Dipartimento Farmaco Chimico Tecnologico, Università di Siena (Italy) and the data for C, H, and N are within $\pm 0.4\%$ of the theoretical values. IR spectra were determined in Nujol mull on a Perkin-Elmer FT-IR 1600 spectrophotometer. ¹H NMR spectra were recorded on a Varian XL-200 or AC200 Bruker instruments. The chemical shifts (δ) are relative to Me₄Si used as internal standard; the following abbreviations were used: s = singlet, t = triplet, dd = double doublet, q = quartet, qt = quintet,st = sextet, m = multiplet, u = unresolved, b = broad. The coupling constants J were in Hertz. TLC on silica-gel plates (Merck, 60, F_{254}) was used for purity check and reaction monitoring. Column chromatography on silica gel (Merck, 70-230 mesh and 230-400 mesh ASTH for flash chromatography) was applied when necessary, to isolate and purify the different reactions products.

All the reagents were of the best available commercial quality and were used without further purification. The

preparation of the unknown 4-quinolylhydrazines ($\mathbf{2}_5$, $\mathbf{2}_{11}$, $\mathbf{2}_{15}$, $\mathbf{2}_{16}$) are also described in this section; analytical and physicochemical data are reported in Table 1. Spectroscopic data (IR and ¹H NMR) are fully consistent with the proposed structures.

7-Ethoxy-4-hydrazinoquinoline hydrochloride (2₅**).** Equimolar amounts of 7-ethoxy-4-chloroquinoline and hydrazine hydrate (85%) in anhydrous ethanol were heated under reflux for 7–8 h. The corresponding hydrazinoquinoline hydrochloride precipitated already in the course of refluxing and completely after cooling. The obtained precipitate was purified by recrystallization (65% yield).

6-Cyclohexyl-4-hydrazinoquinoline hydrochloride (211). The compound was obtained, according to the method described for 2_5 , by refluxing equimolar amounts of 6-cyclohexyl-4-chloroquinoline and hydrazine hydrate (85%) for 8 h (61% yield).

4-Hydrazino-5,7-dichloroquinoline (2_{15}). 4,5,7,-Trichloroquinoline (21.5 mmol, 5 g) and hydrazine hydrate (85%) (25 mL) in anhydrous ethanol were refluxed for 9–10 h. After cooling and dilution with water the 4-hydrazino-5,7-dichloroquinoline (2_{15}) precipitated, it was collected, and recrystallized (60% yield).

2-Methyl-4-hydrazino-5,7-dichloroquinoline (2₁₆). The compound was obtained by refluxing, for 3–4 h, the 2-methyl-4,5,7-trichloroquinoline (20.29 mmol, 5 g) and hydrazine hydrate (85%) (25 mL) under the same conditions described above for compound 2_{15} (80% yield).

General procedure for the preparation of 4-quinolylhydrazones (3)

The title compounds were prepared from 4-quinolylhydrazine or its HCl salt and the appropriate carboxaldehyde in EtOH for 1–2 h. After cooling and diluting with H₂O the quinolylhydrazone **3** precipitated from the reaction mixture; it was collected and purified by crystallization from the suitable solvent (Table 1) (70– 90% yields). In those instances where 4-hydrazinoquinoline hydrochloride was employed, an equimolar amount of NaOAc was added to the mixture to liberate the free base for reaction. The ¹H NMR spectral data of some representative 4-quinolylhydrazones are here described.

37. (DMSO- d_6) δ 1.09 (t, 6H, 2 CH_3CH_2N); 2.44 (s, 3H, CH₃); 3.35 (q, 4H, 2 CH₃ CH_2N); 3.84 (s, 3H, OCH₃); 6.68 (d, 2H, PhH, $J_{orto} = 8.6$); 6.97–7.08 (m, 3H, ArH and H-3 quinoline); 7.53 (d, 2H, PhH, $J_{orto} = 8.6$); 8.12 (d, 1H, H-5, $J_{5-6} = 9.2$); 8.19 (s, 1H, CH=N); 10.55 (bs, 1H, NH, D₂O exchangeable).

313. (DMSO- d_6) δ 1.38 (s, 3H, CH_3CH_2O); 2.48 (s, 3H, CH₃); 4.00 (s, 3H, OCH₃); 4.14 (q, 2H, CH₃ CH_2O); 7.01–7.11 (m, 3H, H-3 quinoline and 2 ArH); 7.38–7.52 (m, 2H, ArH); 7.63 (t, 1H, ArH); 7.88–8.00 (m, 2H, ArH); 8.24 (d, 1H, ArH, J_{orto} = 9.06); 9.07 (s, 1H, CH=N); 9.30 (d, 1H, ArH, J_{orto} = 8.6); 10.98 (bs, 1H, NH, D₂O exchangeable).

318. (DMSO-*d*₆) δ 0.96 (t, 3H, *CH*₃CH₂CH₂CH₂CH₂-); 1.39 (st, 2H, CH₃*CH*₂CH₂CH₂-); 1.72 (qt, 2H, CH₃CH₂*CH*₂CH₂-); 2.81 (t, 2H, CH₃CH₂CH₂*CH*₂-); 4.06 (s, 3H, OCH₃); 7.13 (d, 1H, H-3 quinoline, *J*₃₋₂ = 8.2); 7.31 (bs, 1H, ArH); 7.52–7.79 (m, 5H, ArH); 7.93 (d, 1H, ArH, *J*_{orto} = 8.1); 8.16 (s, 1H, ArH); 8.29 (d, 1H, H-2 quinoline, *J*₂₋₃ = 8.2); 8.98 (ud, 2H, 1ArH and CH=N); 10.95 (bs, 1H, NH, D₂O exchangeable).

323. (DMSO- d_6) δ 1.39–1.63 and 1.92–2.27 (2m, 10H, cyclohexyl); 2.51–2.72 (m, 1H cyclohexyl); 4.06 (s, 3H, OCH₃); 7.13 (d, 1H, H-3 quinoline J_{3-2} =8.4); 7.62–7.80 (m, 6H, ArH); 7.94 (d, 1H, ArH); 8.17 (ud, 1H, H-5 quinoline); 8.30 (d, 1H, H-2 quinoline, J_{2-3} =8.4); 9.01 (bs, 2H, ArH and CH=N); 11.70 (bs, 1H, NH, D₂O exchangeable).

3₃₇. (DMSO- d_6) δ 2.56 (s, 3H, CH₃); 6.10 (s, 2H, OCH₂O); 7.36 (dd, 2H, H-3 quinoline and 2 PhH); 7.46–7.79 (m, 4H ArH); 8.32 (s, 1H, CH=N); 10.51 (bs, 1H, NH, D₂O exchangeable).

Biological evaluation

In vitro evaluation of anti-tuberculosis activity. Primary screening was conducted at $12.5 \,\mu\text{g/mL}$ against *M. tuberculosis* H37Rv in BACTEC 12B medium using a broth microdilution assay, the Microplate Alamar Blue Assay.²⁸ Compounds exhibiting fluorescence were tested in the BACTEC 460 radiometric system too.²⁹ Compounds showing $\geq 90\%$ inhibition in the primary screening were considered active and then retested at lower concentration against *M. tuberculosis* H37Rv to determine the actual minimum inhibitory concentration (MIC), using MABA.

The MIC is defined as the lowest concentration effecting a reduction in fluorescence of 90% relative to the controls. Compounds were tested also for cytotoxicity (IC₅₀) in a VERO cell line at concentration equal to and greater than the MIC for *M. tuberculosis* H37Rv. After 72 h exposure, viability was assessed on the basis of cellular conversion of MTT into a formazan product using the Promega Cell Titer 96 Non-radioactive Cell Proliferation assay. The selectivity index (SI) was also determined, it is considered significant when >4. Rifampin (RMP) was used as reference compound.

Macrophage assay. Compounds with a MIC $\leq 6.25 \,\mu\text{g/mL}$ and a SI > 10 were then tested to evaluate efficacy in vitro in a TB-infected macrophage model.³⁰ The EC₉₀ and EC₉₉ are defined as the concentrations effecting 90 and 99% reduction in residual mycobacterial growth after 7 days, compared to untreated controls. Compounds with EC₉₀ > 16×MIC are considered inactive.

Acknowledgements

Anti-tubercular data presented here were provided by the Tuberculosis Antimicrobial Acquisition and Coordinating Facility (TAACF), Southern Research Institute, Frederick, MD, USA, through a research and development contract of our University with the US National Institute of Allergy and Infectious Diseases. Thanks are addressed to Dr. Joseph A. Maddry (TAACF) for his collaboration. This work was supported by financial assistance from Italian Murst (ex 60% and ex 40%).

References and Notes

1. Murray, C. J. L.; Styblo, K.; Rouillon, A. In *Disease Control Priorities in Developing Countries*; Jamison, D. T., Mosely, W. H., Eds.; Oxford University Press: New York, 1993; p 233.

- 2. Styblo, K. Bull: Int. Union Tuberc. 1990, 65, 24.
- 3. Baker, R. W.; Mitscher, L. A.; Arain, T. M.; Shawar, R.;
- Stover, C. K. Ann. Rep. Med. Chem. 1996, 31, 161.
- 4. Dunkan, K. Exp. Opin. Ther. Pat. 1997, 7, 129.
- 5. Niccolai, D.; Tarsi, L.; Thomas, R. J. Chem. Commun. 1997, 2333.
- 6. Horsburgh, C. R., Jr. New Engl. J. Med. 1991, 324, 1332.
- 7. Heifets, L. Antimicrob. Agents Chemotr. 1996, 40, 1759.
- 8. Harries, A. D.; Mahler, D. *TB/HIV, A Clinical Manual*; World Healt Organization: Geneve, 1996.
- 9. Hamilton, C. D. Curr. Infect. Dis. Rep. 1999, 1, 80.
- 10. Migliori, G. B.; Ambrosetti, M.; Fattorini, L.; Penati, V.;
- Vaccarino, P.; Besozzi, G.; Ortona, L.; Saltini, C.; Orefici, G.; Moro, M. L.; Lona, E.; Cassone, A. Int. J. Tuberc. Lung Dis.
- **2000**, *4*, 940.
- 11. Miletin, M.; Hartl, J.; Odlerova, Z.; Machacek, M. Pharmazie 1997, 52, 558.

12. Pagani, G.; Pregnolato, M.; Ubiali, D.; Terreni, M.; Piersimoni, C.; Scaglione, F.; Fraschini, F.; Rodriguez Gascon, A.; Pedraz Munoz, J. L. *J. Med, Chem.* **2000**, *43*, 199.

13. Waisser, K.; Gregor, J.; Kubicova, L.; Klimesova, V.; Kunes, J.; Machacek, M.; Kaustova, J. *Eur. J. Med. Chem.* **2000**, *35*, 733.

14. (a) Gootz, T.D.; Brigthy, K.E. In *The Quinolones*; Andriole, V. T. Ed.; Academic: New York, 1998; p 29. (b) Fass, R. J. *Antimicrob. Agents Chemother*. **1997**, *41*, 1818.

- 15. Berkoff, C. E.; Craig, P. M.; Giordan, B. P.; Pellerano, C. Arzneim.-Forsch. 1973, 23, 830.
- 16. Actor, P.P.; Pellerano, C.E.G. US Patent 3,646,019, Feb. 29, 1972.
- 17. Pellerano, C.; Savini, L.; Berkoff, C. E.; Thomas, J.; Actor, P. Il Farmaco- Ed. Sc 1975, 30, 965.
- 18. Thomas, J.; Berkoff, C. E.; Flagg, W. B.; Gallo, J. J.; Haff, R. F.; Pinto, C. A.; Pellerano, C.; Savini, L. *J. Med. Chem.* **1975**, *18*, 245.
- 19. Bartolucci, C.; Cellai, L.; Di Filippo, P.; Brizzi, V.; Pellerano, C.; Savini, L.; Benedetto, A.; Elia, G. *Il Farmaco* **1992**, *47*, 945.
- 20. Pellerano, C.; Savini, L.; Massarelli, P. *Il Farmaco* 1985, 40, 645.
- 21. Savini, L.; Massarelli, P.; Chiasserini, L.; Sega, A.; Pellerano, C.; Barzi, A.; Nocentini, G. *Eur. J. Med. Chem.* **1995**, *30*, 547.
- 22. Savini, L.; Massarelli, P.; Chiasserini, L.; Nencini, C.; Pellerano, C. *Il Farmaco* **1997**, *52*, 609.
- 23. Pellerano, C.; Savini, L.; Selvolini, L. Boll. Chim. Farm 1978, 117, 721.
- 24. Pellerano, C.; Brizzi, V.; Savini, L. Atti Acc. Fisiocr. Siena 1971, 3, 253.
- 25. Pellerano, C.; Savini, L.; Brizzi, V. Il Farmaco-Ed. Sc 1985, 40, 486.

- 26. Savini, L.; Massarelli, P.; Pellerano, C. Il Farmaco 1993, 48, 515.
- Singh, T.; Stein, R. G.; Biel, J. H. J. Med. Chem. 1969, 12, 801.
 Collins, L.; Franzblau, S. G. Antimicrob. Agents Chemother. 1997, 41, 1004.
- 29. Franzblau, S. G.; Witzig, R. S.; McLaughlin, J. C.;

Torres, P.; Madico, G.; Hernandez, A.; Degnan, M. T.; Cook, M. B.; Quenzer, V. K.; Freguson, R. M.; Gilman, R. H. J.

Clin. Microb. 1998, 36, 362. 30. Skinner, P. S.; Furney, S. K.; Jacobs, M. R.; Klopman, G.; Ellner, J. J.; Orme, I. M. Antimicrob. Agents Chemother. **1994**, *38*, 2557.