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Synthesis of Imidazo[4,5-c][1,2,6]thiadiazine 2-Oxides from Hydrolytes of Xanthines and Determination of Their Vasodilating Activity

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Novel 1,3,6,7-tetrahydro-7-oxoimidazo[4,5-c][1,2,6]thiadiazine 2-oxide derivatives (2a—q) were synthesized by the reaction of imidazole derivatives (1), obtained by alkaline hydrolysis of 1,3,7-trisubstituted xanthines (3), with SOCl₂ in 42—93% yields. Chlorination of 2 with SO₂Cl₂ gave the 5-chloro derivatives (10a, d, i—q), though in low yields. The reaction of 2 with benzoyl chloride derivatives (14) gave 3,7-dihydro-6H-purin-6-one derivatives (15a—d).

Among 2 and 10, compounds having the phenoxyalkyl group at the 1-position exhibited potent vasodilating activity on the mesenteric artery of spontaneously hypertensive rats. In particular, ED_{50} values of the order of 10^{-7} M were obtained with those having a 1-[6-(4-chlorophenoxy)hexyl]-6-propyl (20), 6-hexyl-1-[6-(4-methoxyphenoxy)hexyl] (2p), 1-[6-(4-chlorophenoxy)hexyl]-6-hexyl (2q), 5-chloro-1-[6-(4-chlorophenoxy)hexyl]-6-methyl (10l), 5-chloro-1-[4-(4-methoxyphenoxy)hexyl]-6-propyl (10m), 5-chloro-1-[6-(4-chlorophenoxy)hexyl]-6-propyl (10n), or 5-chloro-1-[6-(4-chlorophenoxy)hexyl]-6-propyl (10n) substituent.

Keywords—xanthine hydrolyte; imidazo[4,5-c][1,2,6]thiadiazine 2-oxide; thionyl chloride; sulfuryl chloride; 3,7-dihydro-6H-purin-6-one; vascular relaxation

The xanthine skeleton is a basic structure of natural alkaloids such as caffeine and theophylline and of synthetic medicines such as oxyetophylline, 1a proxiphylline, 1b and pentoxifylline. In a series of studies on the synthesis of novel compounds having the xanthine skeleton, we have studied the structural conversion of xanthine derivatives and examined the physiological activities of the newly synthesized compounds. $^{2-4}$ It has been found that 1,2,3,7-tetrahydro-6H-purine-6-one derivatives show vasodilating activity. Thus, in the hope of synthesizing compounds having more potent vasodilating activity, we attempted to introduce a sulfoxide group into the xanthine skeleton in place of the carbonyl group at the 2-position, based on the fact that a sulfur atom participates in the active centers of many enzymes that control biological phenomena as well as materials such as acyl carrier protein, coenzyme A and insulin that are essential for the maintenance of life. The present paper deals with the synthesis of imidazo[4,5-c][1,2,6]thiadiazine 2-oxides (2) from the hydrolytes of 1,3,7-trisubstituted xanthines (3) and SOCl₂, and also with the reactivities of 2. The pharmacological activity of 2 and the 5-chloro derivatives (10) is also discussed.

Barluenga *et al.* synthesized 1,2,6-thiadiazine 1-oxide derivatives having an SO group between two amino groups by cyclization of 3-iminoprop-1-enamine derivatives with SOCl₂.⁵⁾ On the other hand, Deyrup *et al.* carried out the reaction of 2-aminoamide derivatives with SOCl₂ to obtain 5-imino-2-oxo-1,2,3-oxathiazolidine derivatives containing an SO group

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between the nitrogen atom of the amino group and the oxygen atom of the amido group.⁶⁾ However, the cyclization of 3-aminoamide-type compounds such as caffeidine (1a) with SOCl₂ has not been investigated, to our knowledge. Thus, we examined the reaction of 1a with SOCl₂.

The reaction of 1a with SOCl₂ in dry pyridine or in dry benzene in the presence of Et₃N gave colorless needles (2a) (mp 106—107 °C) in 86% or 40% yield, respectively (Chart 1a). The elemental analysis and mass spectrum (M⁺ m/z: 214) of **2a** suggested the molecular formula, C₇H₁₀N₄O₂S, which corresponds to the molecular formula of 1a plus SO minus 2H. The infrared (IR) spectrum of 2a showed very strong absorptions at 1665 and 1110 cm⁻¹ but no absorption band corresponding to v_{NH} of 1a. In the proton nuclear magnetic resonance (${}^{1}H$ -NMR) spectrum of 2a, singlet signals were observed at δ 3.34 (3H), 3.45 (3H), 3.96 (3H) and 7.42 (1H) (Tables I and II). These data suggested that the SO group was taken into 1a. In order to clarify whether the SO group combined with the nitrogen or oxygen atom of the amido group, the IR data of 2a were compared with those of N-[3-(1,1-dimethylethyl)-1,2,3oxathiazolidin-5-ylidene]-2-propanamine S-oxide (16) synthesized by Deyrup et al.6) There was as large a difference as $50 \,\mathrm{cm}^{-1}$ between the absorption band due to $v_{\mathrm{C}=N}$ of 16 at 1715 cm⁻¹ and that of **2a** at 1665 cm⁻¹. This large difference was considered to be attributable to the presence of different functional groups. The absorption at 1665 cm⁻¹, which was the strongest among those of the molecules, seemed due to the amido CO group. Therefore, 2a was suggested to be 1,3,6,7-tetrahydro-1,3,6-trimethyl-7-oxoimidazo[4,5-c][1,2,6]thiadiazine 2-oxide, which contains the SO group between the nitrogen atom of the amido group and that of the amino group.

Compound 2a in H₂O was stable at room temperature for at least 24 h, while in aqueous NaOH, 2a was rapidly hydrolyzed to 1a. This hydrolysis was completed in the presence of 2 eq of NaOH and the pH of the solution was decreased. These findings suggested that the SO group of 2a was eliminated as SO₂ and converted to Na₂SO₃. In aqueous NaHCO₃ (3%), 2a was completely hydrolyzed within ca. 36 h. Furthermore, 2a was also hydrolyzed to 1a in aqueous 0.1 n HCl but this hydrolysis took 3d for completion (Chart 1b). The facile hydrolysis of 2a in acidic and alkaline solutions proved that 2a contains a diaminosulfoxide partial structure.

It was already reported that caffeidine-type imidazole derivatives can easily be obtained by the hydrolysis of 1,3,7-trisubstituted xanthines.^{2,7)} Therefore, we attempted the synthesis of various imidazo[4,5-c][1,2,6]thiadiazine 2-oxides (2) from the hydrolyte (1) of 1,3,7-trisubstituted xanthines (3) and SOCl₂.

Compounds 3b-q were synthesized as follows. Compounds 3b-e, i were synthesized with reference to the literature.⁸⁾ O-Alkylation of $4^{9)}$ with CH_3I in the presence of NaH in dimethylformamide (DMF) at room temperature afforded 3f. Bromohexyltheobromine $(5)^{10)}$ was condensed with ethyl acetoacetate, followed by keto degradation to give 3g. Bromopropyltheobromine $(6)^{9)}$ was treated with morpholine in the presence of K_2CO_3 in acetone to form 3h. Refluxing of 7a and phenoxyalkylbromides (8a-c) in the presence of NaOH in EtOH- H_2O gave 3j-l. According to the method reported previously,⁴⁾ 3c, $r^{8c)}$ were refluxed in $NH_2NH_2 \cdot H_2O$ to afford 1-aminoxanthine derivatives (9b, c), which were

Chart 2

deaminated with NaNO₂ to give 7b, c. ¹¹⁾ Compounds 7b, c were allowed to react with 8a—c under conditions similar to those used for the reaction of 7a and 8 to give 3m—q. The data on newly synthesized xanthine derivatives (3f—h, j—q) are summarized in Tables III and IV. The xanthine derivatives thus obtained (3b—q) gave corresponding imidazole derivatives (1b—q) when refluxed in aqueous $3 \, \text{N}$ NaOH or H₂O-EtOH (3:2) containing 3—4 N NaOH for 6—48 h (Chart 2). The structures of 1b—q were determined by the data given in Tables V and VI.

Since the reaction from 1a to 2a in dry pyridine give a high yield, various caffeidine-type imidazole derivatives (1b-q) were treated with $SOCl_2$ under similar conditions. As a result, the corresponding imidazo[4,5-c][1,2,6]thiadiazine 2-oxide derivatives (2b-q) were obtained in good yields (42-93%): alkyl compounds (2b, c), aralkyl compounds (2d, e), an alkoxyalkyl

compound (2f), an oxoalkyl compound (2g), an aminoalkyl compound (2h) and phenoxyalkyl compounds (2i—q) were thus obtained (Chart 2). The structural assignments of 2b—q were based on the IR spectra showing strong absorption bands at $1665-1680 \,\mathrm{cm}^{-1}$ and $1110-1140 \,\mathrm{cm}^{-1}$, ¹H-NMR spectra, mass spectra and elemental analyses (Tables I and II). The results indicated that imidazo[4,5-c][1,2,6]thiadiazine derivatives could easily be synthesized in high yields by the reaction of caffeidine-type imidazole derivatives with SOCl₂.

Libermann and Rouaix¹²⁾ reported the halogenation at the 8-position of caffeine, which corresponds to the 5-position of **2**. Therefore, we examined the chlorination of **2**. Gentle refluxing of **2a** in $SOCl_2$ for 24 h gave the desired 5-chloro-1,3,6,7-tetrahydro-1,3,6-trimethyl-7-oxoimidazo[4,5-c][1,2,6]thiadiazine 2-oxide (**10a**) in low yield (33%) (Chart 3). An alternative chlorination of **2a** by the use of SO_2Cl_2 in CCl_4 at room temperature for 2 h did not improve the yield of **10a** (31%) (Chart 3). In both reactions, the low yields were due to the formation of many unknown by-products which could mainly be observed near the starting point on thin-layer chromatography (TLC). Compounds **2d**, **i**—**q** were similarly treated with SO_2Cl_2 to give the corresponding **10d**, **i**—**q**, though the yields were low (Chart 3, Tables I and II).

The fact that the SO group is readily eliminated as SO_2 in the hydrolysis of 2a interested us, because it might have implications for the reaction of 2a with electrophilic reagents such as carboxylic acids, aldehydes, and acyl chlorides. Refluxing of 2a in dry acetic acid (AcOH) gave acetylcaffeidine (11) in 59% yield (Chart 3). Heating of 2a and o-nitrobenzaldehyde (12) at $145-150\,^{\circ}$ C gave $13\,(24\%)$ along with SO_2 gas evolution (Chart 3). The facile elimination of the SO group in 2a was considered to have resulted in the formation of 13 as well as 11. The reaction of 2a with an excess of benzoyl chloride (14a) at room temperature did not proceed at all, but at temperatures as high as $150-160\,^{\circ}$ C, an unexpected compound, 3,7-dihydro-3,7-dimethyl-2-phenyl-6H-purine-6-one (15a), was formed in 21% yield (Chart 3, Table VII). The evolution of SO_2 gas was also observed in this reaction. The IR spectrum of 15a showed an absorption band due to a carbonyl group at $1680\,^{\circ}$ cm⁻¹. In the 16a-NMR spectrum, signals due to methyl groups at the 3a- and 3a-positions were observed at 3a-3a- and 3a-3a- and 3a-3a- and 3a- and 3a

155.7, of which two were assigned to the carbon atom of the carbonyl group linked to the imidazole ring and the carbon atom at the 4-position of the imidazole ring, respectively,^{3,13)} and the other was attributed to the newly formed quaternary sp^2 carbon atom. Based on the spectral data mentioned above, the mass spectrum (MS) and elemental analysis data, the structure of 15a was assigned. Heating of 14a and 2c or 2e under similar conditions to those for the formation of 15a gave purine derivatives, 15b and 15c, in yields of 14% and 25%, respectively. The reaction of 14b and 2a also resulted in the formation of 15d (8%) (Chart 3, Table VII), while the reactions of 14a with 2b, d, i afforded black-brown tarry materials.

A plausible reaction mechanism for the formation of 15 is shown in Chart 4 on the basis of the structure of the formed purine derivatives, the reaction conditions (heating of 2 with an excess of 14 at 150—160 °C), and the result of the reaction of 2a with 12. The methyl group at the α -position of the SO group is eliminated by electrophilic attack of the benzoyl group on the oxygen atom of the SO group in 2a to give 2a-1. Next, an additional benzoyl group attacks the nitrogen atom of the amido group to give 2a-2 which is formed by elimination of the benzoyl group substituted at the oxygen atom of the SO group. Finally, SO₂ is eliminated from 2a-2 to form 15a. In this reaction, the initial step from 2a and 14a to 2a-1 is similar to Pummerer's reaction in terms of accompanying elimination of the substituent at the α -position of the SO group. Also, the formation of 15a via 2a-2 can be assumed to correspond to that of 13 from 2a and 12. On the other hand, the reason why 15-type purine derivatives could not be obtained by the reaction of 14a with 2b, d, i having a large group such as benzyl, hexyl or phenoxybutyl group seems to be the difficulty of elimination of such a group.

The synthesized imidazo[4,5-c][1,2,6]thiadiazine derivatives (2, 10) were tested for vasodilating activity using mesenteric arteries of spontaneously hypertensive rats (SHR) according to the method reported previously.³⁾ The ED₅₀ values for the vascular relaxing activity against 30 mm KCl-induced vasocontraction are summarized in Table I. Among the compounds examined, 20, 2p, 2q, 10l, 10m, 10n and 100 were found to have potent vasodilating activity. Though the structure–activity relationship cannot yet be fully discussed due to lack of enough data, the phenoxyalkyl group at the 1-position seemed to have an

5.2

> 20

14.2

> 100

88

> 100

> 100

ED₅₀ (μM)

Compd.

2a

25

> 100

4

43

TABLE I. Physicochemical Properties, Elemental Analysis and Vascular Relaxing Activity of Imidazo[4,5-c][1,2,6]thiadiazine Derivatives (2,10)

	Analysis (%) Calcd (Found) C H N	39.24 4.70 26.15	284.1306 ^{a)} (284.1285)	50.68 7.09 19.70	53.78 4.86 19.30	(53.50 4.84 19.30) (53.50 4.84 19.30)	314.1441 ^{a)}	52.92 7.11 16.46 (52.64 7.08 16.35)	47.69 6.46 21.39	55.16 5.79 16.08 (54.95 5.73 16.08)	53.95 5.86 14.80	6.45 6.46
	Formula	$C_7H_{10}N_4O_2S$	$C_{12}H_{20}N_4O_2S$	$C_{12}H_{20}N_4O_2S$	$C_{13}H_{14}N_4O_2S$	$C_{13}H_{14}N_4O_2S$	$C_{13}H_{22}N_4O_3S$	$C_{15}H_{24}N_4O_3S$	$C_{13}H_{21}N_5O_3S$	$C_{16}H_{20}N_4O_3S$	$C_{17}H_{22}N_4O_4S$	$C_{19}H_{26}N_4O_4S$
	Yield (%)	98	49	74	93	74	84	47	78	72	74	64
$\begin{array}{c c} R_1 \\ R_1 - N \\ O \\ M_2 \\ M_4 \end{array}$	mp (°C) (Appearance)	106—107	(Colorless needles)	(Cotonics on) 62—63	(Coloriess prisms) 154—155	(Colorless needles) (113—114 (Colorless needles)	(Colorless oil)	87—8 (Colorless needles)	(Colorless prisms)	(Colorless prisms) 80—81 (Colorless prisms)	103—104	(Colorless needles) $71-72$ (Colorless needles)
	R	Ħ	Н	Н	Н	н	н	Н	Н	н	H	Н
	${\bf R}_2$	Me	Me	C_6H_{13}	Me	$\langle \bigcirc \rangle$ CH ₂	Me	Me	Me	Me	Me	Me
	R	Me	C_6H_{12}	Me	(О) сн,	Me	$_{ m MeCHC_4H_8}^{ m OMe}$	MeCOC ₇ H ₁₄	Ó NC3H,	⟨O⟩oc₄H ₈	$MeO\langle\bigcirc\rangle OC_4H_8$	$MeO(\bigcirc)OC_6H_{12}$

7K

Zh

7

77	$CI(\bigcirc)OC_6H_{12}$	Me	H	95—98	79	$C_{18}H_{23}CIN_4O_3S$	52.61 5.64 13.63	1.9
2m	$MeO\langle\bigcirc\rangle OC_4H_8$	Pr	Ħ	(Colorless needles)	71	$\mathrm{C_{19}H_{26}N_{4}O_{4}S}$	(52.64 5.67 13.60) 406.1674^a	6.6
2n	$MeO\langle\bigcirc\rangle OC_6H_{12}$	Pr	Н	(Pale yellow oil)	99	$C_{21}H_{30}N_4O_4S$	(406.1666) 434.1986 ^{a)}	1.4
ě		ſ	;	(Pale yellow oil)	ç	Š	(434.2007)	
9	CI (OC6H12	ቷ	II.	62—63 (Colorless cotton)	7.5	$C_{20}H_{27}CIN_4O_3S$	54.72 6.20 12.76 (54 68 6 15 12 77)	0.47
2p	$MeO\langle\bigcirc\rangle OC_6H_{12}$	C_6H_{13}	Н	56—57	68	$C_{24}H_{36}N_4O_4S$	60.48 7.61 11.75	0.37
) ((Colorless cotton)			(60.56 7.63 11.88)	
2 4	$CI(\bigcirc)OC_6H_{12}$	C_6H_{13}	н	38—39	72	$C_{23}H_{33}CIN_4O_3S$	480.19604)	0.29
109	Me	Me	5	(Coloriess cotton)	33	S O NIO H O	(480.1960) 33 81 3 65 22 53	
100	a (3	(Colorless needles)	C	C7119C11V4C25	(33.89 3.54 22.53)	
10d	$\langle \bigcirc \rangle$ CH ₂	Me	כ כ	117—118	59	$C_{13}H_{13}CIN_4O_2S$		28
				(Colorless needles)			3.93	
10i	(U)0C₄H ₈	Me	ひ	69—71	30	$C_{16}H_{19}CIN_4O_3S$	50.19 5.00 14.63	13
				(Colorless needles)			4.94	
10j	$MeO\langle \bigcirc \rangle OC_4H_8$	Me	じ	59—61	23	$C_{17}H_{21}CIN_4O_4S$	5.13	14.2
				(Colorless needles)			5.03	
10k	$MeO\langle \bigcup OC_6H_{12}$	Me	ひ	81—82	22	$C_{19}H_{25}CIN_4O_4S$		3.7
,				(Colorless needles)				
<u>10</u>	CI(U)OC,H12	Me	ت ت	81—82	70	$C_{18}H_{22}Cl_2N_4O_3S$	48.54 4.98 12.58	0.45
;				(Colorless needles)			(48.69 4.95 12.43)	
10m	$MeO\langle \bigcup \rangle OC_4H_8$	Pr	ひ		20	$C_{19}H_{25}CIN_4O_4S$	440.1283^{a}	0.67
<u>=</u>	MeO(\)OC.H.,	ģ	7	(Pale yellow oil)	00	S O'ND "H"D	(440.1284) 468.1599^{a}	0.35
			; ;	(Pale yellow oil)	ì	27-17-67-17-0	(468.1606))
100	$CI\langle\bigcirc\rangle$ OC_6H_{12}	Pr	C	83—84	23	$C_{20}H_{26}Cl_2N_4O_3S$	50.74 5.53 11.83	0.62
	((Colorless needles)			(50.83 5.55 11.68)	
10p	$MeO\langle \bigcirc \rangle OC_6H_{12}$	C_6H_{13}	Ü	4849	25	$C_{24}H_{35}CIN_4O_4S$		7.0
ç		(7	(Colorless needles)	į	i i	6.79	í
bor	CI COCon 12	C_6H_{13}	ご	8485	2 4	C23H32Cl2N4O3S	97.9	> 20
				(Coloriess needles)			(55.51 6.20 10.86)	

a) Determined by high-resolution mass spectrometry. Upper figure, calcd for M+; lower figure, found.

TABLE II. Spectral Data for Imidazo[4,5-c][1,2,6]thiadiazine Derivatives (2, 10)

		TABLE II. Specifal Data	i for fillidazo[4,5-c][1,2,0]thiadiazhic Defivatives (2, 10)
Compd.	MS (m/z) (M ⁺)	IR (cm ⁻¹)	1 H-NMR (δ)
2a	214	1665 (CO), 1110 (SO)	3.34 (3H, s, N_1 -CH ₃), 3.45 (3H, s, N_3 -CH ₃), 3.96 (3H, s, N_6 -CH ₃), 7.42 (1H, s, C_5 -H)
2b	284	1670 (CO), 1130 (SO) ^{a)}	0.89 (3H, t, $J=7$ Hz, CH ₃), 1.1—1.9 (8H, m, CH ₂ ×4), 3.44 (3H, s, N ₃ -CH ₃), 3.63 (1H, q, $J=7$ Hz, N ₁ -CH), 3.94 (3H, s, N ₆ -CH ₃), 4.00 (1H, q, $J=7$ Hz, N ₁ -CH), 7.44 (1H, s, C ₅ -H)
2c	284	1675 (CO), 1125 (SO)	0.88 (3H, t, $J = 7$ Hz, CH ₃), 1.1—1.5 (6H, m, CH ₂ × 3), 1.7—2.0 (2H, m, CH ₂), 3.34 (3H, s, N ₁ -CH ₃), 3.45 (3H, s, N ₃ -CH ₃), 4.25 (1H, t,
2d	290	1665 (CO), 1135 (SO)	J=7 Hz, N ₆ -CH), 4.27 (1H, t, $J=7$ Hz, N ₆ -CH), 7.45 (1H, s, C ₅ -H) 3.44 (3H, s, N ₃ -CH ₃), 3.94 (3H, s, N ₆ -CH ₃), 4.65 (1H, d, $J=16$ Hz, N ₁ -CH), 5.37 (1H, d, $J=16$ Hz, N ₁ -CH), 7.32 (5H, s, Ph), 7.41 (1H, s, C ₅ -H)
2 e	290	1655 (CO), 1135 (SO)	3.34 (3H, s, N ₁ -CH ₃), 3.46 (3H, s, N ₃ -CH ₃), 5.50 (2H, s, N ₆ -CH ₂), 7.33 (5H, s, Ph), 7.43 (1H, s, C ₅ -H)
2f	314	1670 (CO), 1140 (SO) ^{a)}	1.12 (3H, d, $J=6$ Hz, CHCH ₃), 1.3—2.0 (6H, m, CH ₂ × 3), 3.30 (3H, s, OCH ₃ ; 1H, m, CHCH ₃), 3.45 (3H, s, N ₃ -CH ₃), 3.62 (1H, q, $J=7$ Hz, N ₁ -CH), 4.00 (1H, q, $J=7$ Hz, N ₁ -CH), 7.41 (1H, s, C ₅ -H)
2g	340	1710, 1655 (CO),	1.2—1.8 (10H, m, $CH_2 \times 5$), 2.12 (3H, s, $COCH_3$), 2.41 (2H, t, $J=7$ Hz,
		1120 (SO)	$COC\underline{H}_2$), 3.45 (3H, s, N ₃ -CH ₃), 3.61 (1H, q, $J = 7$ Hz, N ₁ -CH), 3.96
			$(3H, s, N_6-CH_3), 3.97 (1H, q, J=7 Hz, N_1-CH), 7.41 (1H, s, C_5-H)$
2h	327	1660 (CO), 1130 (SO)	1.92 (2H, m, CH ₂), 2.30—2.56 (6H, m, NCH ₂ × 3), 3.44 (3H, s, N ₃ -CH ₃), 3.64—3.84 (4H, m, OCH ₂ × 2), 3.88—4.24 (2H, m, N ₁ -CH ₂), 3.96 (3H, s, N ₆ -CH ₃), 7.42 (1H, s, C ₅ -H)
2i	348	1680 (CO), 1250 (COC), 1120 (SO)	1.7—2.1 (4H, m, $CH_2 \times 2$), 3.44 (3H, s, N_3 - CH_3), 3.50—3.80 (1H, m, N_1 - CH), 3.95 (3H, s, N_6 - CH_3), 3.98 (2H, t, $J = 5$ Hz, OCH_2), 4.00—4.30 (1H, m, N_1 - CH), 6.80—7.03 (3H, m, Ph - H_3 , H_4 , H_5), 7.15—7.38 (2H, m, Ph - H_2 , H_6), 7.40 (1H, s, C_5 - H)
2 j	378	1660 (CO), 1225 (COC), 1120 (SO)	1.7—2.1 (4H, m, CH ₂ × 2), 3.44 (3H, s, N ₃ -CH ₃), 3.63 (1H, q, J =7 Hz, N ₁ -CH), 3.75 (3H, s, OCH ₃), 3.95 (3H, s, N ₆ -CH ₃), 3.98 (2H, t, J =6 Hz, OCH ₂), 4.00 (1H, q, J =7 Hz, N ₁ -CH), 6.81 (4H, s, Ph), 7.41 (1H, s, C ₅ -H)
2k	406	1665 (CO), 1225 (COC), 1135 (SO)	1.5—2.0 (8H, m, $CH_2 \times 4$), 3.44 (3H, s, N_3 - CH_3), 3.63 (1H, q, J =7 Hz, N_1 - CH), 3.76 (3H, s, OCH_3), 3.83 (2H, t, J =6 Hz, OCH_2), 3.95 (3H, s, N_6 - CH_3), 4.10 (1H, q, J =7 Hz, N_1 - CH), 6.81 (4H, s, Ph), 7.40 (1H, s, C_5 - H)
21	410	1660 (CO), 1245 (COC), 1125 (SO)	1.3—1.9 (8H, m, CH ₂ × 4), 3.44 (3H, s, N ₃ -CH ₃), 3.64 (1H, q, J =7 Hz, N ₁ -CH), 3.91 (2H, t, J =6 Hz, OCH ₂), 4.00 (1H, q, J =7 Hz, N ₁ -CH), 6.79 (2H, td, J =9, 3 Hz, Ph-H ₂ ,H ₆), 7.21 (2H, td, J =9, 3 Hz, Ph-H ₃ ,H ₅), 7.41 (1H, s, C ₅ -H)
2m	406	1640 (CO), 1220 (COC), 1140 (SO) ^{a)}	0.95 (3H, t, $J=7$ Hz, CH ₃), 1.7—2.1 (6H, m, CH ₂ ×3), 3.45 (3H, s, N ₃ -CH ₃), 3.64 (1H, q, $J=7$ Hz, N ₁ -CH), 3.75 (3H, s, OCH ₃), 3.93 (2H, t, $J=6$ Hz, OCH ₂), 4.04 (1H, q, $J=7$ Hz, N ₁ -CH), 4.21 (1H, t, $J=7$ Hz, N ₆ -CH), 4.25 (1H, t, $J=7$ Hz, N ₆ -CH), 6.81 (4H, s, Ph), 7.45 (1H, s, C ₅ -H)
2n	434	1665 (CO), 1220 (COC), 1135 (SO) ^{a)}	
20	438	1670 (CO), 1240 (COC), 1130 (COC)	0.95 (3H, t, $J=7$ Hz, CH ₃), 1.3—2.1 (10H, m, CH ₂ × 5), 3.45 (3H, s, N ₃ -CH ₃), 3.63 (1H, q, $J=7$ Hz, N ₁ -CH), 3.91 (2H, t, $J=6$ Hz, OCH ₂), 4.01 (1H, q, $J=7$ Hz, N ₁ -CH), 4.21 (1H, t, $J=7$ Hz, N ₆ -CH), 4.25 (1H, q, $J=7$ Hz, N ₆ -CH), 6.79 (2H, td, $J=9$, 3 Hz, Ph-H ₂ ,H ₆), 7.21 (2H, td, $J=9$, 3 Hz, Ph-H ₃ ,H ₅), 7.45 (1H, s, C ₅ -H)

TABLE II. (continued)

Compd.	MS (m/z)	IR (cm ⁻¹)	¹ H-NMR (δ)
No.	(M/2)	in j	
2р	476	1680 (CO), 1235 (COC), 1120 (SO)	0.88 (3H, t, $J=7$ Hz, CH ₃), 1.1—2.0 (16H, m, CH ₂ ×8), 3.45 (3H, s, N ₃ -CH ₃), 3.62 (1H, q, $J=7$ Hz, N ₁ -CH), 3.76 (3H, s, OCH ₃), 3.89 (2H, t, $J=6$ Hz, OCH ₂), 4.02 (1H, q, $J=7$ Hz, N ₁ -CH), 4.24 (1H, t, $J=7$ Hz, N ₆ -CH), 4.28 (1H, t, $J=7$ Hz, N ₆ -CH), 6.81 (4H, s, Ph), 7.44 (1H, s, C ₅ -H)
2q	480	1660 (CO), 1240 (COC), 1140 (SO) ^{a)}	0.88 (3H, t, $J=7$ Hz, CH ₃), 1.1—2.0 (16H, m, CH ₂ ×8), 3.45 (3H, s, N ₃ -CH ₃), 3.64 (1H, q, $J=7$ Hz, N ₁ -CH), 3.91 (2H, t, $J=6$ Hz, OCH ₂), 4.00 (1H, q, $J=7$ Hz, N ₁ -CH), 4.24 (1H, t, $J=7$ Hz, N ₆ -CH) 4.28 (1H, t, $J=7$ Hz, N ₆ -CH), 6.79 (2H, td, $J=9$, 3 Hz, Ph-H ₃ ,H ₅), 7.21 (2H, td, $J=9$, 3 Hz, Ph-H ₂ ,H ₆), 7.44 (1H, s, C ₅ -H)
10a	248	1665 (CO), 1135 (SO)	3.33 (3H, s, N ₁ -CH ₃), 3.45 (3H, s, N ₃ -CH ₃), 3.93 (3H, s, N ₆ -CH ₃)
10d	324	1665 (CO), 1140 (SO)	3.41 (3H, s, N ₃ -CH ₃), 3.92 (3H, s, N ₆ -CH ₃), 4.64 (1H, d, $J = 16$ Hz, N ₁ -CH), 5.36 (1H, d, $J = 16$ Hz, N ₁ -CH), 7.32 (5H, s, Ph)
10i	382	1660 (CO), 1245 (COC), 1140 (SO)	1.7—2.0 (4H, m, CH ₂ × 2), 3.41 (3H, s, N ₃ -CH ₃), 3.5—3.8 (1H, m, N ₁ -CH), 3.92 (3H, s, N ₆ -CH ₃), 3.98 (2H, t, J =6 Hz, OCH ₂), 4.0—4.3 (1H, m, N ₁ -CH), 6.80—7.04 (3H, m, Ph-H ₃ ,H ₄ ,H ₅), 7.16—7.38 (2H, m, Ph-H ₂ ,H ₆)
10j	412	1660 (CO), 1240 (COC), 1135 (SO)	1.7—2.0 (4H, m, $CH_2 \times 2$), 3.42 (3H, s, N_3 - CH_3), 3.61 (1H, q, J =7Hz, N_1 - CH), 3.76 (3H, s, OCH_3), 3.92 (3H, s, N_6 - CH_3), 3.99 (2H, t, J =6Hz, OCH_2), 4.16 (1H, q, J =7Hz, N_1 - CH), 6.81 (4H, s, Ph)
10k	440	1670 (CO), 1225 (COC), 1120 (SO)	1.3—1.9 (8H, m, $CH_2 \times 4$), 3.41 (3H, s, N_3 - CH_3), 3.62 (1H, q, J =7Hz, N_1 - CH), 3.76 (3H, s, OCH_3), 3.89 (2H, t, J =6Hz, OCH_2), 3.92 (3H, s, N_6 - CH_3), 4.16 (1H, q, J =7Hz, N_1 - CH), 6.81 (4H, s, Ph)
101	444	1680 (CO), 1245 (COC), 1140 (SO)	1.3—1.9 (8H, m, $CH_2 \times 4$), 3.41 (3H, s, N_3 - CH_3), 3.63 (1H, q, $J = 7$ Hz, N_1 - CH), 3.90 (2H, t, $J = 6$ Hz, OCH_2), 3.92 (3H, s, N_6 - CH_3), 3.95 (1H, q, $J = 7$ Hz, N_1 - CH), 6.79 (2H, td, $J = 9$, 3 Hz, Ph - H_2 , H_6), 7.21 (2H, td, $J = 9$, 3 Hz, Ph - H_3 , H_5)
10m	440	1660 (CO), 1230 (COC), 1140 (SO) ^{a)}	0.97 (3H, t, $J=7$ Hz, CH ₃), 1.8—2.1 (6H, m, CH ₂ × 3), 3.42 (3H, s, N ₃ -CH ₃), 3.60 (1H, q, $J=7$ Hz, N ₁ -CH), 3.75 (3H, s, OCH ₃), 3.93 (2H, t, $J=6$ Hz, OCH ₂), 4.01 (1H, q, $J=7$ Hz, N ₁ -CH), 4.26 (1H, t, $J=7$ Hz, N ₆ -CH), 4.31 (1H, t, $J=7$ Hz, N ₆ -CH), 6.81 (4H, s, Ph)
10n	468	1665 (CO), 1230 (COC), 1140 (SO) ^{a)}	0.97 (3H, t, $J=7$ Hz, CH ₃), 1.3—2.1 (10H, m, CH ₂ × 5), 3.42 (3H, s, N ₃ -CH ₃), 3.61 (1H, q, $J=7$ Hz, N ₁ -CH), 3.76 (3H, s, OCH ₃), 3.89 (2H, t, $J=6$ Hz, OCH ₂), 4.00 (1H, q, $J=7$ Hz, N ₁ -CH), 4.26 (1H, t, $J=7$ Hz, N ₆ -CH), 4.31 (1H, t, $J=7$ Hz, N ₆ -CH), 6.81 (4H, s, Ph)
100	472	1665 (CO), 1240 (COC), 1125 (SO)	0.97 (3H, t, $J=7$ Hz, CH ₃), 1.2—2.1 (10H, m, CH ₂ ×5), 3.42 (3H, s, N ₃ -CH ₃), 3.62 (1H, q, $J=7$ Hz, N ₁ -CH), 3.91 (2H, t, $J=6$ Hz, OCH ₂), 3.99 (1H, q, $J=7$ Hz, N ₁ -CH), 4.26 (1H, t, $J=7$ Hz, N ₆ -CH), 4.30 (1H, t, $J=7$ Hz, N ₆ -CH), 6.79 (2H, td, $J=9$, 3 Hz, Ph-H ₂ ,H ₆), 7.21 (2H, td, $J=9$, 3 Hz, Ph-H ₃ ,H ₅)
10p	510	1660 (CO), 1225 (COC), 1140 (SO)	0.89 (3H, t, $J=7$ Hz, CH ₃), 1.1—2.0 (16H, m, CH ₂ × 8), 3.42 (3H, s, N ₃ -CH ₃), 3.61 (1H, q, $J=7$ Hz, N ₁ -CH), 3.76 (3H, s, OCH ₃), 3.89 (2H, t, $J=6$ Hz, OCH ₂), 3.99 (1H, q, $J=7$ Hz, N ₁ -CH), 4.28 (1H, t, $J=7$ Hz, N ₆ -CH), 4.33 (1H, t, $J=7$ Hz, N ₆ -CH), 6.81 (4H, s, Ph)
10q	514	1660 (CO), 1240 (COC), 1140 (SO)	0.88 (3H, t, $J=7$ Hz, CH ₃), 1.2—2.0 (16H, m, CH ₂ ×8), 3.42 (3H, s, N ₃ -CH ₃), 3.62 (1H, q, $J=7$ Hz, N ₁ -CH), 3.91 (2H, t, $J=6$ Hz, OCH ₂), 3.99 (1H, q, $J=7$ Hz, N ₁ -CH), 4.28 (1H, t, $J=7$ Hz, N ₆ -CH), 4.32 (1H,
	-		t, $J=7$ Hz, N ₆ -CH), 6.79 (2H, td, $J=9$, 3 Hz, Ph-H ₂ ,H ₆), 7.21 (2H, td, $J=9$, 3 Hz, Ph-H ₃ ,H ₅)

a) IR spectrum was measured in Nujol.

important role. Favorable activity was observed in compounds having a chlorine atom on the benzene ring (compare 21 and 2k, 2n and 2o, and 2p and 2q), those having the $(CH_2)_6$ alkyl chain (compare 2j and 2k, and 2m and 2n), and those having the 6-hexyl group as an alkyl group (compare 2a and 2e, 2k, 2n and 2p, and 2l, 2o and 2q). However, compounds having the chlorine atom at the 5-position showed both potentiating and agonizing actions on the induced relaxation and the effect of the substituent was unclear.

Experimental

All melting points are uncorrected. IR spectrophotometry was performed by the KBr disc method with a Hitachi 285 spectrophotometer unless otherwise stated. Mass spectra were measured with a JEOL JMS-DX 300 spectrometer. 1 H- and 13 C-NMR spectra (1 H-NMR at 89.55 MHz and 13 C-NMR at 22.50 MHz) were recorded with a JEOL JMN-FX90Q spectrometer using tetramethylsilane as an internal standard. Unless otherwise stated, CDCl₃ was used as the solvent. Chemical shifts are given in δ values (ppm) and the abbreviations of signal patterns are as follows: s, singlet; d, doublet; dd, doublet of doublets; t, triplet; td, doublet of triplets; q, quartet; qd, doublet of quartets; m, multiplet; md, doublet of multiplets; br, broad. Column chromatography was done on silica gel (Wakogel C-200). TLC was performed on precoated silica gel plates (Kieselgel 60, F_{254} , Merck). All organic extracts were dried over anhydrous Na_2SO_4 .

1-Amino-3-methyl-7-propylxanthine (9b) — A mixture of 3r (105 g, 0.47 mol) and hydrazine hydrate (1.25 l, large excess) was refluxed for 8 h and cooled. The reaction mixture was extracted with CHCl₃ and the CHCl₃ layer was evaporated *in vacuo* to give a residue, which was purified by column chromatography using CHCl₃-EtOH (20:1, v/v) as the eluent to afford 43.9 g (42%) of 9b as colorless needles (from Et₂O-EtOH), mp 122—123 °C. *Anal.* Calcd for C₉H₁₃N₅O₂: C, 48.42; H, 5.87; N, 31.37. Found: C, 48.28; H, 5.82; N, 31.37. IR v_{max}^{KBr} cm⁻¹: 3310, 3240 (NH₂),

TABLE III. Physicochemical Properties and Elemental Analysis of 1,3,7-Trisubstituted Xanthine Derivatives (3)

Compd.	mp (°C)	Yield	Formula	Analysis (%) Calcd (Found)			
No.	(Appearance)	(%)		C H N			
3f		73	$C_{14}H_{22}N_4O_3$	294.1691 ^{a)}			
	(Colorless oil)			(294.1684)			
3g	140—141	59	$C_{16}H_{24}N_4O_3$	59.98 7.55 17.49			
•	(Colorless prisms)			(59.75 7.54 17.40			
3h·HCl	256—259	77	$C_{14}H_{22}ClN_5O_3$	48.90 6.45 20.37			
	(Colorless prisms)			(48.71 6.40 20.58			
3j	88—89	58	$C_{18}H_{22}N_4O_4$	60.32 6.19 15.63			
·	(Colorless needles)			(60.22 6.14 15.65			
3k	113—114	40	$C_{20}H_{26}N_4O_4$	62.16 6.78 14.50			
	(Colorless needles)			(62.39 6.79 14.27			
31	97—98	61	$C_{19}H_{23}ClN_4O_3$	58.38 5.93 14.33			
	(Colorless prisms)			(58.28 5.95 14.27			
3m	61—62	65	$C_{20}H_{26}N_4O_4$	62.16 6.78 14.50			
	(Colorless needles)			(62.12 6.75 14.35			
3n	56—57	47	$C_{22}H_{30}N_4O_4$	63.75 7.30 13.52			
	(Colorless needles)			(63.90 7.44 13.52			
30	50—51	51	$C_{21}H_{27}ClN_4O_3$	60.21 6.50 13.37			
	(Colorless prisms)			(60.26 6.64 13.65			
3p	4546	71	$C_{25}H_{36}N_4O_4$	456.2734 ^{a)}			
	(Colorless prisms)			(456.2763)			
3q	51—52	84	$C_{24}H_{33}CIN_4O_3$	460.2239a)			
	(Colorless prisms)			(460.2232)			

a) Determined by high-resolution mass spectrometry. Upper figure, calcd for M⁺ and lower figure, found.

TABLE IV. Spectral Data for 1,3,7-Trisubstituted Xanthine Derivatives (3)

Compd. No.	$MS(M^+)$ m/z	IR (cm ⁻¹)	1 H-NMR (δ)
3f	294	1700, 1660 (CO) ^{a)}	1.12 (3H, d, $J = 6$ Hz, CHC $\underline{\text{H}}_3$), 1.3—1.8 (6H, m, CH $_2 \times 3$), 3.16—3.42 (1H, m, C $\underline{\text{H}}_{0}$ CH $_3$), 3.30 (3H, s, OCH $_3$), 3.57 (3H, s, N $_3$ -CH $_3$),
3g	320	1700, 1660 (CO)	3.99 (3H, s, N_7 -CH ₃ , 2H, t, J =7Hz, N_1 -CH ₂), 7.51 (1H, s, C_8 -H) 1.2—1.9 (10H, m, CH ₂ × 5), 2.13 (3H, s, COCH ₃), 2.42 (2H, t, J =7Hz, COCH ₂), 3.57 (3H, s, N_3 -CH ₃), 3.99 (3H, s, N_7 -CH ₃ ; 2H, t, J =7Hz, N_1 -CH ₂), 7.51 (1H, s, C_8 -H)
3h·HCl	307	2700—2300 (NH ⁺),	1.97—2.32 (2H, m, CH ₂), 3.2—3.4 (4H, m, OCH ₂ × 2), 3.51 (3H, s,
		1710, 1660 (CO)	N_3 -CH ₃), 3.6—4.3 (8H, m, NCH ₂ ×4), 3.96 (3H, s, N ₇ -CH ₃), 7.51 (1H, s, C ₈ -H) ^{b)}
3j	358	1705, 1665 (CO),	1.7 - 1.9 (4H, m, CH ₂ × 2), 3.57 (3H, s, N ₃ -CH ₃), 3.75 (3H, s,
•		1240 (COC)	OCH ₃), 3.87—4.16 (4H, m, OCH ₂ , N ₁ -CH ₂), 6.80 (4H, s, Ph), 7.49 (1H, s, C_8 -H)
3k	386	1700, 1665 (CO), 1230 (COC)	1.3—1.9 (8H, m, $CH_2 \times 4$), 3.57 (3H, s, N_3 - CH_3), 3.76 (3H, s, OCH_3), 3.89 (2H, t, $J=6$ Hz, OCH_2), 3.97 (3H, s, N_7 - CH_3), 4.01
		1250 (COC)	(2H + I - 7Hz) N CH) 6.91 (4H a Db) 7.49 (1H a C H)
31	390	1720, 1675 (CO),	$(2H, t, J=7Hz, N_1-CH_2)$, 6.81 (4H, s, Ph), 7.48 (1H, s, C_8 -H)
	370	1255 (COC)	1.3—1.9 (8H, m, $CH_2 \times 4$), 3.57 (3H, s, N_3 - CH_3), 3.91 (2H, t, $J = 6H_7$, OCH_3), 3.92 (3H, s, N_3 - CH_3), 4.91 (2H, t, J -
		1233 (COC)	6 Hz, OCH ₂), 3.98 (3H, s, N ₇ -CH ₃), 4.01 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 6.79 (2H, td, $J=9$, 3 Hz, Ph-H ₂ ,H ₆), 7.20 (2H, td, $J=9$, 3 Hz, Ph-H ₃ ,H ₅), 7.50 (1H, s, C ₈ -H)
3m	386	1700, 1660 (CO), 1240 (COC)	0.95 (3H, t, $J=7$ Hz, CH ₃), 1.7—2.1 (6H, m, CH ₂ × 3), 3.58 (3H, s, N ₃ -CH ₃), 3.75 (3H, s, OCH ₃), 3.95 (2H, t, $J=6$ Hz, OCH ₂), 4.09 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 4.25 (2H, t, $J=7$ Hz, N ₇ -CH ₂), 6.81 (4H, s, Ph), 7.52 (1H, s, C ₈ -H)
3n	414	1705, 1655 (CO), 1240 (COC)	0.95 (3H, t, $J=7$ Hz, CH ₃), 1.3—2.1 (10H, m, CH ₂ × 5), 3.58 (3H, s, N ₃ -CH ₃), 3.76 (3H, s, OCH ₃), 3.89 (2H, t, $J=6$ Hz, OCH ₂), 4.02 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 4.24 (2H, t, $J=7$ Hz, N ₇ -CH ₂), 6.81 (4H, s, Ph), 7.52 (1H, s, C ₈ -H)
30	418	1700, 1645 (CO), 1240 (COC)	0.95 (3H, t, J =7 Hz, CH ₃), 1.3—2.1 (10H, m, CH ₂ × 5), 3.58 (3H, s, N ₃ -CH ₃), 3.91 (2H, t, J =6 Hz, OCH ₂), 4.01 (2H, t, J =7 Hz, N ₁ -CH ₂), 4.24 (2H, t, J =7 Hz, N ₇ -CH ₂), 6.79 (2H, td, J =9, 3 Hz, Ph-
3р	456	1710, 1660 (CO), ^{a)} 1230 (COC)	H_2, H_6), 7.21 (2H, td, $J=9$, 3 Hz, Ph- H_3, H_5), 7.52 (1H, s, C_8 -H) 0.88 (3H, t, $J=7$ Hz, CH ₃), 1.1—2.0 (16H, m, CH ₂ × 8), 3.58 (3H, s, N ₃ -CH ₃), 3.76 (3H, s, OCH ₃), 3.89 (2H, t, $J=6$ Hz, OCH ₂), 4.02 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 4.27 (2H, t, $J=7$ Hz, N ₇ -CH ₂), 6.81 (4H, $J=7$ Hz, $J=7$
3q	460	1705, 1660 (CO), ^{a)} 1240 (COC)	s, Ph), 7.51 (1H, s, C ₈ -H) 0.88 (3H, t, J =7 Hz, CH ₃), 1.1—2.0 (16H, m, CH ₂ ×8), 3.58 (3H, s, N ₃ -CH ₃), 3.91 (2H, t, J =6 Hz, OCH ₂), 4.02 (2H, t, J =7 Hz, N ₁ -CH ₂), 4.27 (2H, t, J =7 Hz, N ₇ -CH ₂), 6.79 (2H, td, J =9, 3 Hz, Ph-H ₂ ,H ₆), 7.20 (2H, td, J =9, 3 Hz, Ph-H ₃ ,H ₅), 7.52 (1H, s, C ₈ -H)

a) IR spectrum was measured in Nujol. b) ¹H-NMR spectrum was measured in D₂O.

1710, 1650 (CO). ¹H-NMR (CDCl₃) δ : 0.94 (3H, t, J = 7 Hz, CH₂C $\underline{\text{H}}_3$), 1.73—2.12 (2H, m, CH₂C $\underline{\text{H}}_2$ CH₃), 3.62 (3H, s, N₃-CH₃), 4.28 (2H, t, J = 7 Hz, N₇-CH₂), 4.99 (2H, br s, NH₂), 7.59 (1H, s, C₈-H). MS m/z: 223 (M⁺).

1-Amino-7-hexyl-3-methylxanthine (9c) — Treatment of 3c (60 g, 0.23 mol) with hydrazine hydrate (1.2 l, large excess) as described for 9b gave 9.2 g (15%) of 9c as colorless needles (from Et₂O–EtOH), mp 115—116 °C. Anal. Calcd for C₁₂H₁₉N₅O₂: C, 54.32; H, 7.22; N, 26.40. Found: C, 54.35; H, 7.22; N, 26.38. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3310, 3250 (NH₂), 1710, 1660 (CO). ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J = 7 Hz, CH₃(CH₂)₅), 1.1—1.5 (6H, m, CH₂ × 3), 1.7—2.1 (2H, m, N₇-CH₂–CH₂), 3.63 (3H, s, N₃-CH₃), 4.30 (2H, t, J = 7 Hz, N₇-CH₂), 4.51 (2H, br s, NH₂), 7.56 (1H, s, C₈-H). MS m/z: 265 (M⁺).

3-Methyl-7-propylxanthine (7b) —A solution of NaNO₂ (28.8 g, excess) in H_2O (150 ml) was added dropwise to a solution of 9b (40 g, 0.18 mol) in AcOH- H_2O (9:5 v/v, 1.12 l) with stirring at room temperature, and stirring was continued for 6 h. The reaction mixture was evaporated *in vacuo* to give a residue, which was washed with H_2O to afford 36.1 g (96%) of 7b. Anal. Calcd for $C_9H_{12}N_4O_2$: C_7 , 51.92; C_7 , 51.92; C_7 , 51.91; C_7 , 705.

TABLE V. Physicochemical Properties and Elemental Analysis of Imidazole Derivatives (1)

Compd. No.	mp (°C) (Appearance)	Yield (%)	Formula	Analysis (%) Calcd (Found)			
140.	(Appearance)	(/₀)		C	H	N	
1b pierie acid	164—165 (Yellow needles)	38	$C_{12}H_{22}N_4O \cdot C_6H_3N_3O_7$	46.25 (46.04		20.98	
1e · picric acid	114—116 (Yellow needles)	63	$C_{18}H_{22}N_4O\cdot C_6H_3N_3O_7$	•	5.39	20.98	
1d	110—111 (Colorless needles)	38	$C_{12}H_{16}N_4O$	63.92 (63.63	6.60	22.93	
1e	113—114 (Colorless needles)	52	$C_{13}H_{16}N_4O$	63.92 (63.81	6.60	22.93	
1f · picric acid	144—145 (Yellow needles)	29	$C_{13}H_{24}N_4O_2\cdot C_6H_3N_3O_7$	45.87 (45.65	5.47	19.71	
1g · picric acid	118—119 (Yellow needles)	31	$C_{15}H_{26}N_4O_2\cdot C_6H_3N_3O_7$	48.18 (47.99	5.58	18.73	
1h·2HCl	209—212 (Colorless needles)	26	$C_{13}H_{23}N_5O_2\cdot 2HCl$	44.07 (43.85	7.11	19.77	
1i · picric acid	137—138 (Yellow needles)	34	$C_{16}H_{22}N_4O_2\cdot C_6H_3N_3O_7$	49.72 (49.42	4.71	18.45	
1j	48—49 (Colorless needles)	47	$C_{17}H_{24}N_4O_3$	61.42	7.28	16.86	
1k · picric acid	119—120 (Yellow needles)	42	$C_{19}H_{28}N_4O_3\cdot C_6H_3N_3O_7$	50.93	5.30	16.63	
11 · picric acid	138—139 (Yellow needles)	22	$C_{18}H_{25}CIN_4O_2 \cdot C_6H_3N_3O_7$	48.53 (48.48	4.75	16.51	
1m·picric acid	82—83 (Yellow needles)	26	$C_{19}H_{28}N_4O_3\cdot C_6H_3N_3O_7$	50.93 (51.23	5.30	16.63	
1n · picric acid	99—100 (Yellow needles)	48	$C_{21}H_{32}N_4O_3\cdot C_6H_3N_3O_7$	52.51 (52.40	5.71	15.88	
10	66—67 (Colorless needles)	38	$\mathrm{C}_{20}\mathrm{H}_{29}\mathrm{ClN}_4\mathrm{O}_2$	61.14 (61.16	7.44	14.26	
1p	35—36 (Colorless needles)	24	$C_{24}H_{38}N_4O_3$	66.95 (66.76	8.90	13.01	
1q	55—56 (Colorless cotton)	29	$\mathrm{C_{23}H_{35}ClN_4O_2}$	63.51 (63.43	8.11	12.88	

7-Hexyl-3-methylxanthine (7c)—Treatment of **9c** (23 g, 0.087 mol) with NaNO₂ (13.9 g, excess) as described for **7b** gave 20.4 g (94%) of **7c**. Anal. Calcd for $C_{12}H_{18}N_4O_2$: C, 57.58; H, 7.25; N, 22.38. Found: C, 57.46; H, 7.26; N, 22.43.

1-(5-Methoxyhexyl)-3,7-dimethylxanthine (3f)——A suspension of 4 (5.6 g, 2 mmol) and NaH (60% in oil, 0.9 g, 2.3 mmol) in dry DMF (40 ml) was stirred for 1 h at room temperature, and then CH₃I (20 g, large excess) was added. Stirring was continued for 16 h. The reaction mixture was evaporated *in vacuo* to give a residue, which was purified by column chromatography using CHCl₃-EtOH (20:1, v/v) as the eluent to afford 4.3 g of 3f. Data are summarized in Tables III and IV.

3,7-Dimethyl-1-(8-oxononyl)xanthine (3g)—A solution of 5 (15 g, 0.046 mol), ethyl acetoacetate (6.2 g, 0.048 mol) and NaOEt (prepared from Na (1.0 g, 0.043 mol)) in absolute EtOH (100 ml) was refluxed for 6 h and evaporated in vacuo to give a residue, to which 1 N NaOH (60 ml) was added. The mixture was stirred for 2.5 h at room temperature and then acidified with diluted H_2SO_4 . The acidified solution was refluxed for 4 h, alkalized with 10% NaOH, and then extracted with CHCl₃. The CHCl₃ layer was evaporated in vacuo to give a residue, which was recrystallized from iso-PrOH to afford 8.2 g (59%) of 3g. Data are summarized in Tables III and IV.

3,7-Dimethyl-1-[3-(4-morpholino)propyl]xanthine (3h)—A mixture of 6 (9.6 g, 3.2 mmol), morpholine (3.3 g, 3.8 mmol) and K_2CO_3 (5.28 g, 2.8 mmol) in acetone (80 ml) was refluxed for 4 h and evaporated *in vacuo* to give a residue, which was extracted with CHCl₃. The CHCl₃ layer was washed with H_2O and evaporated *in vacuo* to give a residue, which, after EtOH-HCl treatment, was recrystallized from EtOH to afford 7.5 g of $3h \cdot HCl$. Data are summarized in Tables III and IV.

TABLE VI. Spectral Data for Imidazole Derivatives (1)

Compd. No.	MS (M ⁺) . m/z	IR (cm ⁻¹)	1 H-NMR (δ)
1b · picric acid	238 (M ⁺ – picric acid)	3320, 3280 (NH), 1625 (CO)	0.87 (3H, t, $J=6$ Hz, CH ₃), 1.1—1.7 (8H, m, CH ₂ × 4), 2.84 (3H, s, NHC $\underline{\text{H}}_3$), 3.20 (2H, q, $J=6$ Hz, NHC $\underline{\text{H}}_2$), 3.87 (3H, s, N ₁ -CH ₃), 7.59 (1H, t, $J=6$ Hz, N $\underline{\text{H}}$ CH ₂), 8.59 (2H, s, Ph), 8.64 (1H, s, C ₂ -H) ^{a)}
1c · picric acid	238 (M ⁺ – picric acid)	3420, 3300 (NH), 1640 (CO)	0.85 (3H, t, J =7 Hz, CH ₃), 1.1—1.4 (6H, m, CH ₂ × 3), 1.5—1.9 (2H, m, CH ₂), 2.74 (3H, d, J =5 Hz, CONHCH ₃), 2.83 (3H, s, NHCH ₃), 4.28 (2H, t, J =7 Hz, N ₁ -CH ₂), 7.76 (1H, q, J =5 Hz, CONHCH ₃), 8.59 (2H, s, Ph), 8.74 (1H, s, C ₂ -H) ^a)
1d	244	3280 (NH), 1600 (CO)	2.84 (3H, s, NHC \underline{H}_3), 3.74 (1H, br s, N \underline{H} CH $_3$), 3.86 (3H, s, N $_1$ -CH $_3$), 4.58 (2H, d, J =6Hz, NHC \underline{H}_2), 7.18 (1H,s, C $_2$ -H), 7.32 (5H, s, Ph; 1H, br s, N \underline{H} CH $_2$)
1e	244	3310, 3260 (NH), 1620 (CO)	2.81 (3H, d, $J=5$ Hz, CONHC \underline{H}_3), 2.91 (3H, s, NHC \underline{H}_3), 4.05 (1H, br s, N \underline{H} CH $_3$), 5.44 (2H, s, N $_1$ -C \underline{H}_2), 6.68 (1H, br s, CON \underline{H} CH $_3$), 7.1—7.5 (5H, m, Ph; 1H, s, C $_2$ -H)
If pieric acid	268 (M ⁺ – picric acid)	3330, 3280 (NH), 1630 (CO), 1320 (NO ₂)	1.04 (3H, d, $J=6$ Hz, CHC \underline{H}_3), 1.1—1.6 (6H, m, CH $_2 \times 3$), 2.84 (3H, s, NHC \underline{H}_3), 3.19 (3H, s, OCH $_3$), 3.0—3.3 (2H, m, NHC \underline{H}_2 ; 1H, m, C \underline{H} CH $_3$), 3.87 (3H, s, N $_1$ -CH $_3$), 7.59 (1H, t, $J=6$ Hz, N \underline{H} CH $_2$), 8.58 (2H, s, Ph), 8.65 (1H, s, C $_2$ -H) a)
1g · picric acid	294 (M ⁺ – picric acid)	3320, 3290 (NH), 1710, 1625 (CO), 1315 (NO ₂)	1.1—1.7 (10H, m, CH ₂ ×5), 2.06 (3H, s, COCH ₃), 2.40 (2H, t, $J=7$ Hz, COCH ₂), 2.84 (3H, s, NHCH ₃), 3.19 (2H, q, $J=6$ Hz, NHCH ₂), 3.87 (3H, s, N ₁ -CH ₃), 7.58 (1H, t, $J=6$ Hz, NHCH ₂), 8.58 (2H, s, Ph), 8.64 (1H, s, C ₂ -H) ^a)
1h·2HCl	281 (M ⁺ -2HCl)		2.0—2.3 (2H, m, CH ₂), 2.94 (3H, s, NHC \underline{H}_3), 3.2—4.3 (12H, m, NCH ₂ ×4, OCH ₂ ×2), 3.93 (3H, s, N ₁ -CH ₃), 8.32 (1H, s, C ₂ -H) ^b)
li picric acid	302 (M ⁺ – picric acid)	3340, 3270 (NH), 1630 (CO), 1320 (NO ₂)	1.5—1.9 (4H, m, 'CH ₂ × 2), 2.85 (3H, s, NHC \underline{H}_3), 3.28 (2H, q, J =6 Hz, NHC \underline{H}_2), 3.88 (3H, s, N ₁ -CH ₃), 3.98 (2H, t, J =7 Hz, OCH ₂), 6.8—7.0 (3H, m, Ph-H ₃ ,H ₄ ,H ₅), 7.1—7.4 (2H, m, Ph-H ₂ ,H ₆), 7.62 (1H, t, J =6 Hz, N \underline{H} CH ₂), 8.58 (2H, s, Ph), 8.65 (1H, s, C ₂ -H) ^{a)}
1j	332	3360, 3290 (NH), 1630 (CO), 1235, 1030 (COC)	1.7—1.9 (4H, m, $CH_2 \times 2$), 2.86 (3H, s, $NHC\underline{H}_3$), 3.44 (2H, q, $J=6$ Hz, $NHC\underline{H}_2$), 3.75 (3H, s, OCH_3), 3.83 (3H, s, N_1 - CH_3), 3.95 (2H, t, $J=6$ Hz, OCH_2), 6.81 (4H, s, Ph), 7.16 (1H, s, C_2 -H; 1H, br s, $N\underline{H}CH_2$)
1k · picric acid	360 (M ⁺ – picric acid)		1.4—2.0 (8H, m, $CH_2 \times 4$), 2.99 (3H, s, $NHC\underline{H}_3$), 3.45 (2H, q, $J=6$ Hz, $NHC\underline{H}_2$), 3.75 (3H, s, OCH_3), 3.91 (2H, t, $J=6$ Hz, $OC\underline{H}_2$), 4.05 (3H, s, N_1 - CH_3), 6.72 (1H, t, $J=6$ Hz, $N\underline{H}CH_2$), 6.81 (4H, s, Ph), 7.82 (1H, br s, NH^+), 7.98 (1H, s, C_2 -H), 8.94 (2H, s, Ph)
11 picric acid	364 (M ⁺ – picric acid)		1.2—1.9 (8H, m, $CH_2 \times 4$), 2.85 (3H, s, $NHCH_3$), 3.23 (2H, q, $J=6$ Hz, $NHCH_2$), 3.88 (3H, s, N_1 - CH_3), 3.95)(2H, t, $J=6$ Hz, OCH_2), 6.92 (2H, td, $J=9$, 3 Hz, $Ph-H_2$, H_6), 7.30 (2H, td, $J=9$, 3 Hz, $Ph-H_3$, H_5), 7.61 (1H, t, $J=6$ Hz, $NHCH_2$), 8.59 (2H, s, Ph), 8.68 (1H, s, C_2 - $H)^{a}$)
1m·picric acid		3410, 3380 (NH), 1625 (CO), 1335, 1310 (NO ₂), 1265, 1225 (COC)	0.81 (3H, t, $J=7$ Hz, CH ₃), 1.6—1.9 (6H, m, CH ₂ × 3), 2.84 (3H, s, NHCH ₃), 3.26 (2H, t, $J=6$ Hz, NHCH ₂), 3.69 (3H, s, OCH ₃), 3.92 (2H, t, $J=6$ Hz, OCH ₂), 4.26 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 6.84 (4H, s, Ph), 7.82 (1H, t, J=6 Hz, NHCH ₂), 8.59 (2H, s, Ph), 8.73 (1H, s, C ₂ -H) ^a)

TABLE	VI. ((continued)
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Compd. No.	MS (M ⁺) m/z	IR (cm ⁻¹)	¹H-NMR (δ)
In · picric acid	388 (M ⁺ – picric acid)	3440, 3310 (NH), 1625 (CO), 1350, 1320 (NO ₂), 1225 (COC)	0.81 (3H, t, $J=7$ Hz, CH ₃), 1.2—1.9 (10H, m, CH ₂ ×5), 2.85 (3H, s, NHCH ₃), 3.23 (2H, q, $J=6$ Hz, NHCH ₂), 3.69 (3H, s, OCH ₃), 3.88 (2H, t, $J=6$ Hz, OCH ₂), 4.27 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 6.83 (4H, s, Ph), 7.77 (1H, t, $J=6$ Hz, NHCH ₂), 8.59 (2H, s, Ph), 8.75 (1H, s, C ₂ -H) ^a)
10	392	3350, 3210 (NH), 1640 (CO), 1235 (COC)	0.81 (3H, t, $J=7$ Hz, CH ₃), 1.3—2.0 (10H, m, CH ₂ × 5), 2.87 (3H, s, NHCH ₃), 3.38 (2H, q, $J=6$ Hz, NHCH ₂), 3.91 (2H, t, $J=6$ Hz, OCH ₂), 4.19 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 6.79 (2H, td, $J=9$, 3 Hz, Ph-H ₂ ,H ₆), 7.21 (2H, td, $J=9$, 3 Hz, Ph-H ₃ ,H ₅), 7.23 (1H, s, C ₂ -H), 7.32 (1H, br s, NHCH ₂)
1p	430	3420, 3290 (NH), 1605 (CO), 1230 (COC)	0.87 (3H, t, $J=7$ Hz, CH ₃), 1.1—1.9 (16H, m, CH ₂ × 8), 2.87 (3H, s, NHCH ₃), 3.38 (2H, q, $J=6$ Hz, NHCH ₂), 3.76 (3H, s, OCH ₃), 3.90 (2H, t, $J=6$ Hz, OCH ₂), 4.22 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 6.81 (4H, s, Ph), 7.22 (1H, s, C ₂ -H), 7.24 (1H, br s, NHCH ₂)
1q	434	3310, 3250 (NH), 1615 (CO), 1245 (COC)	0.87 (3H, t, $J=7$ Hz, CH ₃), 1.1—1.9 (16H, m, CH ₂ ×8), 2.87 (3H, s, NHCH ₃), 3.53 (2H, q, $J=6$ Hz, NHCH ₂), 3.92 (2H, t, $J=6$ Hz, OCH ₂), 4.22 (2H, t, $J=7$ Hz, N ₁ -CH ₂), 6.79 (2H, td, $J=9$, 3 Hz, Ph-H ₂ ,H ₆), 7.21 (2H, td, $J=9$, 3 Hz, Ph-H ₃ ,H ₅), 7.24 (1H, s, C ₂ -H; 1H, br s, NHCH ₂)

a) ¹H-NMR spectrum was measured in DMSO-d₆. b) ¹H-NMR spectrum was measured in D₂O.

7-Alkyl-3-methyl-1-phenoxyalkylxanthines (3j—q)—Typical Procedure: Compound 8a (14.2 g, 0.055 mol) was added to a solution of 7a (9.0 g, 0.05 mol) and NaOH (2.1 g, 0.053 mol) in EtOH- H_2O (3:2 v/v, 200 ml), and the mixture was refluxed for 6 h. The reaction mixture was concentrated *in vacuo* to about one-fourth of the initial volume and alkalized with 10% NaOH. The alkaline mixture was extracted with CHCl₃, and the CHCl₃ layer was evaporated *in vacuo* to give a residue, which was purified by column chromatography using CHCl₃-EtOH (40:1, v/v) as the eluent to afford 3i.

Compounds 3k-q were prepared from the corresponding 8a-c and 7a-c in a similar manner and purified by recrystallization or column chromatography using the CHCl₃-EtOH mixture as the eluent. Data are summarized in Tables III and IV.

1-Substituted 4-Methylamino-5-substituted aminocarbonylimidazoles (1b—q)—Typical Procedure: A suspension of 3b (7.5 g, 0.028 mol) in 3 N NaOH (300 ml) was refluxed for 6 h and extracted with CHCl₃. The CHCl₃ layer was evaporated *in vacuo* to give a residue, which was purified by column chromatography using CHCl₃-EtOH (40:1, v/v) as the eluent to afford 2.6 g of 1b. Compound 1b was converted into the corresponding picrate using EtOH-picric acid.

Compounds 1c—q were prepared from the corresponding 3c—q in a similar manner and, after purification by column chromatography using the CHCl₃-EtOH mixture as the eluent, directly recrystallized or converted into the corresponding picrate or hydrochloride by EtOH-picric acid or EtOH-HCl treatment, respectively. Data are summarized in Tables V and VI.

1,3,6,7-Tetrahydro-1,3,6-trimethyl-7-oxoimidazo[4,5-c][1,2,6]thiadiazine 2-Oxide (2a)—Method a: $SOCl_2$ (3.1 g, 0.026 mol) was added dropwise to a solution of 1a (3.4 g, 0.02 mol) in dry pyridine (50 ml) with stirring at -10-0 °C and stirring was continued for a further 3 h at room temperature. The reaction mixture was evaporated *in vacuo* to give a residue, which was purified by column chromatography using $CHCl_3$ -EtOH (40:1, v/v) as the eluent to afford 3.7 g of 2a.

Method b: A solution of SOCl₂ (0.5 g, 4.2 mmol) in dry C_6H_6 (7 ml) was added dropwise to a solution of 1a (0.67 g, 4.0 mmol) and Et_3N (0.88 g, 8.7 mmol) in dry C_6H_6 (20 ml) with stirring at room temperature for 2 h, and subsequently the mixture was heated at 80 °C for 10 min, then cooled and filtered. The filtrate was evaporated in vacuo to give a residue, which was purified in the same manner as in method a to afford 0.34 g (40%) of 2a. Data are summarized in Tables I and II.

Hydrolysis of 2a—Method c: A solution of 2a (0.43 g, 2.0 mmol) in H₂O (25 ml) was stirred at room

Compd.	mp (°C) (Appearance)	Yield	Formula	Analysis (%) Calcd (Found)			MS (M ⁺)	IR (cm ⁻¹)	1 H-NMR (δ)
		(%)		C	Н	N	m/z		
15a	181—182 (Colorless needles)	21	C ₁₃ H ₁₂ N ₄ O			23.32 23.25)	240	1680 (CO)	3.47 (3H, s, N ₃ -CH ₃), 4.07 (3H, s, N ₇ -CH ₃), 7.4—7.7 (5H, m, Ph), 7.81 (1H, s, C ₈ -H)
15b HCl	154—156 (Colorless needles)	14	C ₁₈ H ₂₂ N ₄ O HCl			16.15 16.16)	310 (M ⁺ – HCl)		0.87 (3H, t, $J = 7$ Hz, CH ₃), 1.1—1.4 (4H, m CH ₂ × 2), 1.7—2.0 (2H m, CH ₂), 3.38 (3H, s, N ₃ -CH ₃), 4.2 (2H, t, J = 7 Hz, N ₇ -CH ₂), 7.5—7.7 (5H, m, Ph), 8.85 (1H, s, C ₈ -H), 9.2 (1H, br s, NH ⁺) ^{a)}
15c	180—181 (Colorless needles)	25	C ₁₉ H ₁₆ N ₄ O			17.71 17.71)	316	1685 (CO)	3.50 (3H, s, N ₃ -CH ₃), 5.64 (2H, s, N ₇ -CH ₂), 7.36 (5H, s, Ph), 7.50 (5H, s, C ₂ -Ph)
15d	247—249 (Yellow needles)	8	$C_{13}H_{11}N_5O_3$			24.55 24.32)	285	1680 (CO), 1520, 1340 (NO ₂)	3.50 (3H, s, N ₃ -CH ₃), 4.13 (3H, s, N ₇ -CH ₃), 7.77 (2H, td, J=9, 2 Hz Ph-H ₂ ,H ₆), 7.84 (1H, s C ₈ -H), 8.38 (2H, td, J= 9, 2 Hz, Ph-H ₂ ,H ₅)

TABLE VII. 3,7-Dihydro-6*H*-purin-6-one Derivatives (15)

temperature for 24 h; only 2a was detectable by TLC. Then, 0.1 N NaOH (20 ml) was added to the solution and the pH of the solution became neutral after 1 h. At this time, both 2a and 1a were detectable by TLC. Further 0.1 N NaOH (20 ml) was applied to the solution, and after 1 h 2a was no longer detected by TLC. The solution was extracted with CHCl₃ and the CHCl₃ layer was evaporated *in vacuo* to give a residue, which, after Et₂O-HCl treatment, was recrystallized from EtOH to afford 0.28 g (68%) of 1a HCl.

Method d: Compound 2a (0.32 g, 1.5 mmol) was added to 3% NaHCO₃ (20 ml) and the mixture was stirred at room temperature until 2a was no longer detectable by TLC (ca. 36 h). The solution was extracted with CHCl₃, and the CHCl₃ layer was evaporated *in vacuo* to give a residue, which was treated in the same manner as in method c to afford 0.23 g (75%) of 1a·HCl.

Method e: A solution (pH 1.5) of 2a (0.32 g, 1.5 mmol) in 0.1 N HCl (15 ml) was stirred at room temperature until 2a was hardly detected by TLC (ca. 3 d). The solution was evaporated in vacuo to give a residue, which was treated in the same manner as in method c to afford 0.24 g (78%) of $1a \cdot HCl$.

1,6-Disubstituted 1,3,6,7-Tetrahydro-3-methyl-7-oxoimidazo[4,5-c][1,2,6]thiadiazine 2-Oxides (2b-q)—Compounds 2b—q were prepared from SOCl₂ and the corresponding 1b—q in a similar manner to method a in the preparation of 2a and purified by column chromatography using the CHCl₃-EtOH mixture as the eluent. Data are summarized in Tables I and II.

5-Chloro-1,3,6,7-tetrahydro-1,3,6-trimethyl-7-oxoimidazo[4,5-c][1,2,6]thiadiazine 2-Oxide (10a)—Method f: A solution of 2a (0.86 g, 4.0 mmol) in SOCl₂ (35 ml) was gently refluxed for 24 h and evaporated *in vacuo* to give a residue, which was purified by column chromatography using CHCl₃-EtOH (25:2, v/v) as the eluent to afford 0.33 g (33%) of 10a.

Method g: A solution of SO_2Cl_2 (0.49 g, 3.6 mmol) in dry CCl_4 (5 ml) was added dropwise to a solution of **2a** (0.64 g, 3.0 mmol) in dry CCl_4 (15 ml) at room temperature, and the mixture was stirred for 2 h. The reaction mixture was poured into saturated NaHCO₃ and extracted with $CHCl_3$. The $CHCl_3$ layer was evaporated *in vacuo* to give a residue, which was treated in the same manner as in method f to afford 0.23 g (31%) of **10a**.

1,6-Disubstituted 5-Chloro-1,3,6,7-tetrahydro-3-methyl-7-oxoimidazo[4,5-c][1,2,6]thiadiazine 2-Oxides (10d, i—q)—Compounds 10d, i—q were prepared from SO_2Cl_2 and the corresponding 2d, i—q in a similar manner to

a) 1 H-NMR spectrum was measured in DMSO- d_{6} .

method g in the preparation of 10a and purified by column chromatography using CHCl₃ or CHCl₃-EtOH mixture as the eluent. Data are summarized in Tables I and II.

4-(N-Acetyl-N-methylamino)-1-methyl-5-methylaminocarbonylimidazole (11)—A solution of 2a (0.54 g, 2.5 mmol) in dry AcOH (3 ml) was refluxed for 2h and evaporated in vacuo to give a residue, which was purified by column chromatography using CHCl₃-EtOH (20:1, v/v) as the eluent to afford 0.31 g (59%) of 11 as colorless needles (from Et₂O-EtOH). The ¹H-NMR data of 11 were identical with those of the compound reported by Hoskinson. ¹⁴)

1,2,3,7-Tetrahydro-1,3,7-trimethyl-2-(2-nitrophenyl)-6*H*-purin-6-one (13)——A mixture of 2a (0.32 g, 1.5 mmol) and 12 (0.31 g, 2.0 mmol) was heated at 145—150 °C for 2.5 h. The gas generated during the reaction was introduced into aqueous NaOH. The aqueous NaOH solution was positive to the qualitative reaction with sulfite. After cooling, the reaction mixture was purified by column chromatography using $CHCl_3$ -EtOH (20:1, v/v) as the eluent to give 0.11 g (24%) of 13. The IR spectrum of 13 was consistent with that of the compound previously synthesized by us.²⁾

2,7-Disubstituted 3,7-Dihydro-3-methyl-6H-purin-6-one (15)—Typical Procedure: A mixture of **2a** (1.00 g, 4.67 mmol) and **14a** (4.0 ml, excess) was heated at 150—160 °C for 40 min. The gas generated during the reaction was introduced into aqueous NaOH. The aqueous NaOH solution was positive to the qualitative reaction with sulfite. After cooling, the reaction mixture was poured into saturated NaHCO₃ and extracted with CHCl₃. The CHCl₃ layer was evaporated *in vacuo* to give a residue, which was purified by column chromatography using CHCl₃–EtOH (20:1, v/v) to afford **15a**. ¹³C-NMR (CDCl₃) δ : 33.6 (qd, J=142, 1 Hz, N₇-C), 33.9 (q, J=142 Hz, N₃-C), 114.0 (m, C₅), 128.3 (md, J=162 Hz, ph-C₂,C₆ or ph-C₃,C₅), 128.5 (md, J=162 Hz, ph-C₂,C₆ or ph-C₃,C₅), 128.8 (md, J=161 Hz, ph-C₄), 135.6 (m, ph-C₁), 144.3 (qd, J=20, 4 Hz, C₈), 155.7 (m, C₂ or C₄ or C₆), 156.0 (m, C₂ or C₄ or C₆), 156.2 (m, C₂ or C₄ or C₆).

Compounds 15b—d were prepared from the corresponding 2a, c, e and 14a, b in the same manner as used for 15a. Data are summarized in Table VII.

Vascular Relaxing Effect—The effect was evaluated by a similar method to the one described in the previous paper.³⁾ Helical strips isolated from 13-week old SHR were mounted for isometric recording of tension in a 20-ml water-jacketed tissue bath $(37\pm0.5\,^{\circ}\text{C})$ containing oxygenated Kreb's bicarbonate solution. The strips were contracted by exposure to KCl $(30\,\text{mM})$. After the contraction had reached a plateau, one of the test compounds was added and then papaverine was added at the end of the experiments to obtain the maximum relaxation. The relaxation induced by papaverine was taken as 100%. The effect of each compound was expressed as the ED₅₀ value, which was determined from the dose–response curve. Data are listed in Table I.

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