Cyclic Peptides. XV. Synthesis of [4-a-Hydroxyalanine]AM-toxin II

Kosaku Noda,* Junko Nakashima, Sannamu Lee,† and Nobuo Izumiya†

Laboratory of Biochemistry, Fukuoka Women's University, Kasumigaoka, Higashi-ku, Fukuoka 813

†Laboratory of Biochemistry, Faculty of Science, Kyushu University, 33, Hakozaki, Higashi-ku, Fukuoka 812

(Received August 12, 1982)

Synopsis. An AM-toxin II analog, cyclo(-L-Ala¹-L-Hmb²-L-App³-Hyala⁴-), with an α-hydroxyalanine (Hyala) residue in place of dehydroalanine in position 4 was synthesized from pyruvoyl-L-Ala-L-Hmb-L-App-amide (Hmb, 2-hydroxy-3-methylbutanoic acid; App, 2-amino-5-phenylpentanoic acid) by intramolecular condensation between the pyruvoyl and carbamoyl groups. The synthetic peptide did not show the toxic activity of AM-toxin II.

The cyclotetradepsipeptide AM-toxins are phytotoxic metabolites of Alternalia mali causing necrosis on apple leaves. The structure of AM-toxin II¹⁾ was shown to be $cyclo(-L-Ala^1-L-Hmb^2-L-App^3-\Delta Ala^4-).^{2}$ Previously, we attempted the synthesis of their analogs with L-Phe or L-Tyr residue replacing the uncommon aromatic amino acid in position 3 by the intramolecular condensation of pyruvoyl-L-Ala-L-Hmb-L-Phe-NH₂ (1b) or -L-Tyr-NH₂ (1c).³⁾ The product was, however, identified to be cyclotetrapeptide 2b or 2c containing a Hyala residue instead of Δ Ala expected (Fig. 1), and did not show the activity of AM-toxins. The inactivity of **2b** or **2c** is not attributed to the replacement of Δ Ala by Hyala, because [L-Phe³]AM-toxin⁴) and [L-Tyr-(Me)³]AM-toxin⁵⁾ were also almost inactive.

In order to examine the influence of substitution of Δ Ala by Hyala in position 4 on the activity, we have synthesized [Hyala⁴]AM-toxin II (2a). The Hyala residue was expected to contribute toward the toxic activity, because it might be a possible precursor of

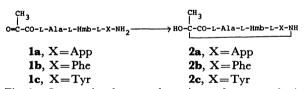


Fig. 1. Intramolecular condensation of pyruvoyl-tripeptide amides.

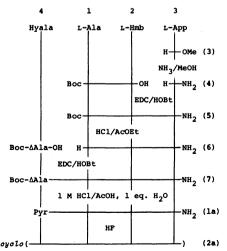


Fig. 2. Synthetic route for [Hyala⁴]AM-toxin II (1a).

ΔAla, and an AM-toxin I analog with L-Ser⁴, the structural isomer of Hyala, possessed a recognizable activity.⁶⁾

Figure 2 shows the reaction route for the synthesis. The tripeptide, Boc-L-Ala-L-Hmb-L-App-NH₂(5), was prepared by coupling of Boc-L-Ala-L-Hmb-OH and L-App amide (4) by the EDC method.⁷⁾ Removal of the Boc group of 5 followed by coupling with Boc-ΔAla-OH afforded tetrapeptide amide 7, which was converted to pyruvoyl-tripeptide amide 1a by the treatment with HCl in AcOH in the presence of an equivalent amount of water. Treatment of 1a with HF cyclized the peptide intramolecularly with the formation of Hyala residue to give the desired peptide 2a. Dimer and polymer formation was not observed in this reaction. After purification by chromatography with silicic acid, the homogeneity and identity of the product were established by thin-layer chromatography, mass spectroscopy and ¹H-NMR spectroscopy. The singlet peak assingned for the a-OH of Hyala residue in the NMR data suggests that the Hyala is of a single optical configuration of either L or D.

Bioassay showed that, even at high concentration, 2a did not show the toxic activity for the induction of necrosis on apple leaves. Thus, replacement of the Δ Ala residue by Hyala eliminated the biological activity. The possible conversion of serine residue to Δ Ala in the biological system was suggested from the previous results that [L-Ala⁴]AM-toxin I was inactive⁴⁾ while [L-Ser⁴]analog was active.⁶⁾ The present data indicates that there is no possibility of such a conversion of the Hyala residue to Δ Ala in the biological system. The chemical conversion of the Hyala residue to Δ Ala is under investigation.

Experimental

Thin layer chromatography was carried out on silica gel G (Merck) with the following solvent systems: $R_{\rm f}^{\, 1}$, CHCl₃–MeOH (9:1); $R_{\rm f}^{\, 2}$, CHCl₃–MeOH–AcOH (85:10:5); $R_{\rm f}^{\, 3}$, n-BuOH–AcOH– pyridine–H₂O (4:1:1:2). Optical rotations were measured with a Union polarimeter PM-201, Mass spectra were taken on a Hitachi RMS-4 mass spectrometer and NMR spectra on a JEOL LNM PS-100 spectrometer.

H-L-App-OMe·HCl (3·HCl). This compound was obtained by the treatment of H-L-App-OH⁸⁾ with MeOH and SOCl₂ according to the procedure described in the literature; byield, 96%; mp 115 °C; $[a]_{D}^{25}$ +21.5° (c 2, EtOH); R_{t}^{1} 0.23; R_{t}^{2} 0.54.

Found: C, 59.08; H, 7.39; N, 5.83%. Calcd for C₁₂H₁₈-NO₂Cl: C, 59.13; H, 7.44; N, 5.75%.

H-L-App-N H_2 ·HCl (4·HCl). This compound was prepared from 3·HCl by the treatment with saturated N H_3 in MeOH according to the literature; 10) yield, 82%; mp 200 °C; [a] $_{\rm D}^{25}$ +17.0° (c 1, EtOH); $R_{\rm f}^2$ 0.05; $R_{\rm f}^3$ 0.67.

Found: C, 57.70; H, 7.55; N, 12.28%. Calcd for C₁₁H₁₇-N₂OCl: C, 57.76; H, 7.49; N, 12.25%.

Boc-L-Ala-L-Hmb-L-App-NH₂ (5). Boc-L-Ala-L-Hmb-OH⁶⁾ (1.45 g, 5 mmol), **4·HCl** (1.14 g, 5 mmol), HOBt (1.35 g, 10 mmol) and NEt₃ (0.70 ml, 5 mmol) was dissolved in DMF (10 ml). To the solution, EDC·HCl (1.15 g, 6 mmol) was added at 0 °C. The mixture was stirred for 1 h at 0 °C and overnight at room temperature, and evaporated *in vacuo*. The residue was treated with a mixture of water and AcOEt. The organic layer was washed with 4% NaHCO₃ and 10% citric acid, dried (Na₂SO₄), and evaporated. The product was recrystallized from AcOEt-ether-petroleum ether; yield, 1.67 g (72%); mp 94—95 °C; [a]_D²⁵ −40.0° (c 1, EtOH); $R_{\rm f}^{1}$ 0.40; $R_{\rm f}^{2}$ 0.63.

Found: C, 62.33; H, 8.16; N, 9.17%. Calcd for $C_{24}H_{37}$ - N_3O_6 : C, 62.18; H, 8.05; N, 9.07%.

 $Boc-\Delta Ala-L-Ala-L-Hmb-L-App-NH_2$ (7). (963 mg, 2.1 mmol) was treated with 2 M HCl in AcOEt (25 ml) for 1 h at room temperature, and the mixture was concentrated in vacuo, the concentration being repeated twice after addition of AcOEt. The solid residue was washed with ether by means of decantation, and dried in a desiccator. The hydrochloride of H-L-Ala-L-Hmb-L-App-NH₂ (6·HCl) was obtained as a powder. To the solution of $6 \cdot HCl$, Boc- ΔAla -OH3) (374 mg, 2 mmol), HOBt (338 mg, 2.5 mmol) and NEt₃ (0.28 ml, 2 mmol) in DMF (10 ml) was added EDC·HCl (383 mg, 2 mmol). The mixture was stirred for 1 h at 0 °C and overnight at room temperature, and evaporated. The residue was treated with a mixture of 20% NaCl and AcOEt. The organic layer was washed with 4% NaHCO3 and saturated NaCl, dried (Na₂SO₄), and evaporated. The product was recrystallized from MeOH-ether-petroleum ether; yield, 643 mg (60%); mp 95—97 °C; $[a]_D^{25}$ -21.0° (c 1, EtOH); R_f^{1} 0.41; $R_{\rm f}^2$ 0.64.

Found: C, 60.68; H, 7.62; N, 10.47%. Calcd for $C_{27}H_{40}$ - N_4O_7 : C, 60.88; H, 7.57; N, 10.52%.

Pyr-L-Ala-L-Hmb-L-App-NH₂ (1a). Compound 7 (533 mg, 1 mmol) was dissolved in 1 M HCl in AcOH (5 ml) containing an equivalent of water (18 μ l). After being kept for 10 min at 60 °C, the solution was evaporated. The residual solid was collected with the aid of ether; yield, 422 mg (96%). This compound was used in the cyclization reaction without further purification.

cyclo(-L-Ala-L-Hmb-L-App-Hyala-) (2a). A solution of **1a** (217 mg, 0.5 mmol) in anhydrous HF (5 ml) was stirred for 1 h at 0 °C. After HF was evaporated, the residual solid was dissolved in CHCl₃ and the solution chromatographed on

silicic acid (Mallinckrodt, 100 mesh) using a column (1.2 cm \times 10 cm) and a solvent system of CHCl₃-AcOEt (1:1). The main fractions (6—16 ml) were collected and evaporated. The residue was collected with the aid of ether; yield, 160 mg (74%); mp 124—126 °C; $[a]_{2}^{25}$ —44.0° (c 1, DMF); $R_{\rm f}^1$ 0.39; $R_{\rm f}^2$ 0.47. NMR (signals arising only from Hyala residue are given) (DMSO- $d_{\rm e}$) δ =2.38 (3H, s, HyalaCH₃), 7.04 (1H, s, HyalaOH), 7.35 (1H, s, HyalaNH).

Found: C, 60.77; H, 7.31; N, 9.63%; m/e 433. Calcd for $C_{22}H_{31}N_3O_6$: C, 60.95; H, 7.21; N, 9.69; M+, 433.

Biological Assay. Biological assays on apple leaves (susceptible cultivar, Indo) were carried out as described previously. Product **2a** did not show the toxicity at concentration up to 100 μg/ml, whereas synthetic or natural AMtoxin II showed at 0.02 μg/ml.

References

- 1) T. Ueno, T. Nakashima, Y. Hayashi, and H. Fukami, Agric. Biol. Chem., 39, 2081 (1975).
- 2) Abbreviations given by the IUPAC-IUB Commission, J. Biol. Chem., 247, 977 (1972), have been used throughout. Additional abbreviations: ΔAla, dehydroalanine; App, 2-amino-5-phenylpentanoic acid; EDC, 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide; Hmb, 2-hydroxy-3-methylbutanoic acid; HOBt, 1-hydroxybenzotriazole; Hyala, α-hydroxyalanine; Pyr, pyruvoyl; Tyr(Me), O-methyltyrosine.
- 3) K. Noda, Y. Shibata, E. Gross, Y. Shimohigashi, and N. Izumiya, Int. J. Pept. Protein Res., 18, 423 (1981).
- 4) Y. Shimohigashi, S. Lee, T. Kato, and N. Izumiya, Bull. Chem. Soc. Jpn., 51, 584 (1978).
- 5) Y. Shimohigashi, S. Lee, H. Aoyagi, T. Kato, and N. Izumiya, Int. J. Pept. Protein Res., 10, 197 (1977).
- 6) Y. Shimohigashi, S. Lee, H. Aoyagi, T. Kato, and N. Izumiya, Int. J. Pept. Protein Res., 10, 232 (1977).
- 7) J. C. Sheehan, J. Preston, and P. A. Cruickshank, J. Am. Chem. Soc., 87, 2492 (1965).
- 8) Y. Shimohigashi, S. Lee, and N. Izumiya, *Bull. Chem. Soc. Jpn.*, **49**, 3280 (1976).
- 9) M. Brenner and W. Huber, *Helv. Chim. Acta*, **36**, 1109 (1953).
- 10) J. P. Greenstein and M. Winitz, "Chemistry of the Amino Acids," John Wiley and Sons, Inc., New York (1961), Vol. 2, p. 1188.