

Published on Web 06/21/2008

A Monomeric Mn^{III}-Peroxo Complex Derived Directly from Dioxygen

Ryan L. Shook,[†] William A. Gunderson,[‡] John Greaves,[†] Joseph W. Ziller,[†] Michael P. Hendrich,[‡] and A. S. Borovik*,†

Department of Chemistry, University of California—Irvine, 1102 Natural Sciences II, Irvine, California 92697-2025, and Department of Chemistry, Carnegie Mellon University, Pittsburgh, Pennsylvania 15213

Received April 15, 2008; E-mail: aborovik@uci.edu

The binding and activation of dioxygen is an essential process in synthetic and biological chemistry. 1 The activation processes are often proposed to involve formation of peroxometal complexes, as exemplified by bleomycin and the mono-oxygenases cytochromes P450.² It is generally agreed that the initial steps in the O₂ binding/ activation process in these enzymes involve a superoxoiron(III) intermediate that converts to a hydroperoxoiron(III) species through addition of an electron and proton. In this report, we demonstrate that a similar O₂ to peroxo conversion is operable in a synthetic manganese system.

The observation of synthetic monomeric peroxometal complexes is frequently difficult because of their inherent reactivity. This is especially true for peroxomanganese complexes, where the Mn^{IV}₂(*u*-1,2-peroxo) complex of Wieghardt is the only O2-derived system that has been structurally characterized.^{3,4} Others have found that treating Mn^{II} or Mn^{III} complexes with superoxides⁵ or peroxides⁶ produce systems with monomeric peroxomanganese centers—this approach has yielded a handful of complexes at low temperatures that were stable enough to be characterized. We have been investigating the interactions of dioxygen with manganese complexes containing intramolecular hydrogen bonding (H-bond) networks.⁷ Our systems utilize urea-based tripodal ligands that provide H-bond donors to coordinated O-atom species. The MnII complexes of these ligands bind and activate dioxygen producing monomeric oxomanganese complexes.^{7,8} We have developed a hybrid ligand (H₅bupa) that combines two urea arms with one carboxyamidopyridyl moiety9—the MnII complex of this ligand binds O₂ to produce a detectable peroxomanganese(III) species.

Preparation of the precursor 1 is outlined in Figure 1.¹⁰ Treating H₅bupa with 3 equiv of KH in dimethylacetamide (DMA) followed by 1 equiv of Mn(OAc)₂ afforded K[1] and 2 equiv of KOAc.

The molecular structure of 1 determined by X-ray diffraction shows a five-coordinate Mn^{II} complex, having a distorted trigonal bipyramidal geometry.10 The trigonal plane is defined by the deprotonated urea and pyridyl nitrogen atoms of [H₂bupa]³⁻; the apical N1 atom and carbonyl oxygen O1 from the deprotonated carboxamide occupy the axial positions. The remaining portions of the urea groups form the scaffolding of a cavity, in which NH groups are positioned inward toward atom O1. However, the N6(N7)···O1 distances are greater than 3.2 Å, distances that are too long for intramolecular H-bonds.

A new green species (2) is formed in approximately 50% yield¹¹ when [Mn^{II}H₂bupa]⁻ reacts with O₂ at room temperature. In DMA, the reaction is relatively slow (~30 min), yet the formation of 2 can be completed in approximately 10 min when 0.5 equiv of diphenylhydrazine (DPH) is added to the reaction mixture (Figure 1). The yield of 2 also increases to nearly 80% when using DPH,

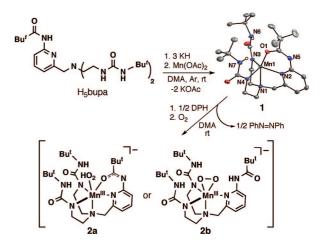


Figure 1. Preparative routes for 1 and 2, showing two possible tautomers for 2. The thermal ellipsoid plot of [Mn^{II}H₂bupa]⁻ is drawn at the 50% probability level, and non-urea hydrogen atoms are omitted for clarity. Selected distances (Å): Mn1-N1, 2.275(3); Mn1-N2, 2.214(3); Mn1-N3, 2.100(3); Mn1-N4, 2.125(3); Mn1-O1, 2.070(2).

which is converted to azobenzene (>95% yield). Monitoring the reactions with optical spectroscopy shows that 2 has a visible absorbance band at $\lambda_{max} \approx 660$ nm and a shoulder at 490 nm (Figure S1).12 Similar spectra have been reported for MnIII complexes containing a coordinated peroxo ligand. 5d,6b

The oxygenation of 1 was followed by electron paramagnetic resonance (EPR) spectroscopy (Figure S2). Perpendicular-mode X-band EPR spectra of $\left[Mn^{II}H_{2}bupa\right]^{-}$ reveal the complex as a nearly axial S = 5/2 spin system with a large zero-field splitting constant of $D \sim 0.3 \text{ cm}^{-1}$. After exposure to O_2 , the S = 5/2 signaldecreases as a new parallel-mode EPR signal associated with 2 appears at a g value of 8.2 (Figure 2A). A quantitative simulation of the signal (Figure 2B) indicates an S = 2 ground state with a six-line (I = 5/2) hyperfine splitting of a = 57 G. Variable temperature studies determined that the signal is from the ground doublet with D = -2.0(5) cm⁻¹. The spin state, zero-field splitting, and hyperfine constant are in agreement with other known monomeric Mn^{III} species. 5d,13 Moreover, the negative sign for the axial zero-field splitting constant is consistent with tetragonally elongated octahedral coordination geometry. The simulations also indicate that 2 accounts for 80(10)% of the Mn in the sample. In perpendicular-mode, this sample also showed the signal of the initial Mn(II) complex (6%) and a mixed valent species at g = 2 (4%). The parallel-mode signal vanishes after prolonged incubation (6 h) at room temperature—the identity of the resultant species are under investigation.

Isotopic labeling studies support the presence of a peroxo ligand coordinated to the Mn^{III} center in 2. Solution FTIR spectra recorded at room temperature contained a peak at 885 cm⁻¹ for 2 prepared

[†] University of California—Irvine. † Carnegie Mellon University.

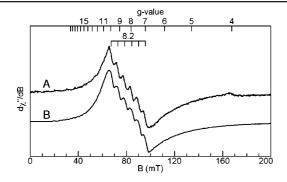


Figure 2. Parallel-mode EPR spectrum (A) and simulation (B) of 2 (10 mM in DMF) recorded at 11 K. Microwave frequency and power, 9.379 GHz, 0.2 mW; modulation, 10 G. Simulation parameters: S = 2, g = 2.0, D = -2 cm⁻¹, E/D = 0.13(3), A = 160 MHz.

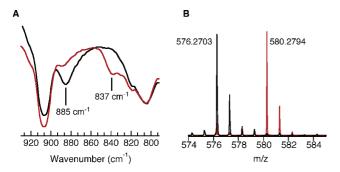


Figure 3. FTIR (A) and negative-mode ESI-MS (B) spectra of $\bf 2$ after exposure to $^{16}O_2$ (black) and $^{18}O_2$ (red) collected from DMA solutions at room temperature.

under a ¹⁶O₂ atmosphere (Figure 3A). The ¹⁸O-isotopomer can be prepared from ¹⁸O₂, causing a shift in the peak to 837 cm⁻¹. The observed vibrational change between the two isotopomers is as expected based on a harmonic O-O oscillator $(\nu(^{16}O_2)/\nu(^{18}O_2) =$ 1.06; calcd = 1.07). These vibrational values are in the range normally observed for other metal-based peroxo systems. For instance, the η^2 -peroxoMn^{III}(Tp)¹⁵ complexes of Kitajima, formed using H₂O₂, have FTIR-active peaks at 892 cm⁻¹ that were assigned to $\nu(O_2)$.^{6a} The electrospray ionization mass spectrum (ESI-MS) of 2 prepared with ¹⁶O₂ exhibits a strong ion with a mass-to-charge ratio (m/z) of 576.2703 (Figure 3B), a shift of 33 mass units from the peak associated with 1 (Figure S3). The mass and calculated isotopic distribution corresponds to the addition of a hydroperoxo ligand to 1 (calcd, 576.2706; Figure S4A). Furthermore, when 2 was prepared from $^{18}\mathrm{O}_2$, the molecular ion peak shifts by 4 mass units (Figure 3B) to a m/z of 580.2794 (calcd, 580.2792; Figure S4B).

Preliminary reactivity studies indicate that 2 leads to the oxidative deformylation of aldehydes. For instance, treating 2 with cyclohexanecarboxaldehyde afforded cyclohexanone as the only GC-MS detectable product in an unoptimized yield of 40% (eq 1). Note that deformylation reactions are known for iron(III)¹⁶ and manganese(III)^{6b} peroxo complexes.

$$H \xrightarrow{2} H$$
 (1)

The spectroscopic, mass spectrometry, and reactivity results are consistent with 2 being a monomeric peroxomanganese(III) complex. A possible mechanism for its formation would involve a superoxomanganese(III) intermediate that reacts with solvent or external substrates, such as DPH, via a H-atom abstraction process to initially produce a η^1 -hydroperoxoMn(III) complex (Figure 1, 2a). The reduction and protonation of the superoxo ligand mirrors steps proposed during turnover in cytochrome P450. The tautomeric η^2 -peroxomanganese(III) species (Figure 1, **2b**) could be formed from 2a by intramolecular proton transfer from the hydroperoxo to the carboxamido component of the tripodal ligand. In this pathway, the pivaloylamide moiety is re-formed to provide an additional H-bond donor within the cavity. At present, we cannot distinguish between these two structural possibilities. Nevertheless, our findings establish that a mononuclear peroxoMn(III) can be produced from O₂ at room temperature.

Acknowledgment is made to the NIH (GM050781 to A.S.B.; GM77387 to M.P.H.) for financial support.

Supporting Information Available: Experimental details for all chemical reactions and figures for all spectra. This material is available free of charge via the Internet at http://pubs.acs.org.

References

- (1) Borovik, A. S.; Zart, M. K.; Zinn, P. J. In Activation of Small Molecules:
- Organometallic and Bioinorganic Perspectives, Tolman, W. B., Ed.; Wiley-VCH: Weinheim, Germany, 2006; pp 187–234, and references therein.

 (a) Cytochrome P450: Structure, Mechanism, and Biochemistry, 3rd ed.; Ortiz de Montellano, P. R., Ed.; Kluwer Academic/Plenum Publishers: New York, 2005. (b) Comprehensive Coordination Chemistry II; Que, L., Jr., Tolman, W. B., Eds.; Elsevier: Oxford, 2004; Vol. 8. (c) Decker, A.; Chow, M. S.; Kemsley, J. N.; Lehnert, N.; Solomon, E. I. J. Am. Chem. Soc. 2006, 128, 4719-4733. (d) Groves, J. T.; Han, Y.-Z. In Cytochrome P-450. Structure, Mechanism and Biochemistry; Ortiz de Montellano R. R., Ed.;
- Plenum Press: New York, 1995; pp 3–48.
 (3) Bossek, U.; Weyhermüller, T.; Wieghardt, K.; Nuber, B.; Weiss, J. *J. Am.* Chem. Soc. 1990, 112, 6387-6388.
- (4) Dioxygen adducts of manganese porphyrins have been observed at low temperatures: (a) Weschler, C. J.; Hoffman, B. M.; Basolo, F. J. Am. Chem. Soc. 1975, 97, 5278–5280. (b) Hoffman, B. M.; Weschler, C. J.; Basolo, F. J. Am. Chem. Soc. 1976, 98, 5473-5482.
- (a) Shirazi, A.; Goff, H. M. J. Am. Chem. Soc. 1982, 104, 6318-6322. (b) Groves, J. T.; Watanabe, Y.; McMurry, T. J. J. Am. Chem. Soc. 1983, 105, 4489–4490. (c) VanAtta, R. B.; Strouse, C. E.; Hanson, L. K.; Valentine, J. S. J. Am. Chem. Soc. 1987, 109, 1425–1434. (d) Groni, S.; Blain, G.; Guillot, R.; Policar, C.; Anxolabehere-Mallart, E. Inorg. Chem. **2007**, 46, 1951–1953.
- (6) (a) Kitajima, N.; Komatsuzaki, H.; Hikichi, S.; Osawa, M.; Moro-oka, Y. (a) Khajinia, I., Kolinasudaki, I., Tinkolii, S., Osawa, W., Morobak, T. J. Am. Chem. Soc. 1994, 116, 11596–11597. (b) Seo, M. S.; Kim, J.; Xim, Y.; Annaraj, J.; Kim, Y.; Lee, Y.-M.; Kim, S.-J.; Kim, J.; Nam, W. Angew. Chem., Int. Ed. 2007, 46, 377–380.
- (7) Borovik, A. S. Acc. Chem. Res. 2005, 38, 54-61, and references therein. (8) Parsell, T. H.; Behan, R. K.; Hendrich, M. P.; Green, M. T.; Borovik, A. S. J. Am. Chem. Soc. 2006, 128, 8728-8729.
- (a) Wada, A.; Harata, M.; Hasegawa, K.; Jitsukawa, K.; Masuda, H.; Mukai, M.; Kitagawa, T.; Einaga, H. Angew. Chem., Int. Ed. 1998, 37, 798-799. (b) Mareque Rivas, J. C.; Salvagni, E.; Parsons, S. *Dalton Trans.* **2004**, 4185–4192. (c) Rudzka, K.; Arif, A. M.; Berreau, L. M. *J. Am. Chem. Soc.* **2007**, *128*, 17018–17023.
- (10) Full experimental details are found in Supporting Information.
- (11) Yields are obtained from EPR simulations using SpinCount developed by one of the authors (M.P.H.).
- (12) The extinction coefficient for this peak is less than 300 M⁻¹cm⁻
- (12) The Catherland Control of the Control of th Chem. Soc. 2000, 122, 3754-3761. (c) Campbell, K. A.; Lashley, M. R.; Wyatt, J. K.; Nantz, M. H.; Britt, R. D. J. Am. Chem. Soc. 2001, 123, 5710-5719. (d) Krzystek, J.; Telser, J.; Hoffman, B. M.; Brunel, L.-C.;
- Licoccia, S. J. Am. Chem. Soc. 2001, 123, 7890–7897.
 The difference of 48 cm⁻¹ between the ν(O₂) of the two isotopomers is similar to those reported for Fe^{III} OOH and Fe^{III} OO complexes: Roelfes, G.; Vrajmasu, V.; Chen, K.; Ho, R. Y. N.; Rohed, J.-U.; Zondervan, C.; Crois, R. M.; Schudde, E. P.; Lutz, M.; Spek, A. L.; Hage, R.; Feringa, B. L.; Münck, E.; Que, L., Jr *Inorg. Chem.* **2003**, *42*, 2639–2653.
- (15) Tp, hydrotris(3,5-iPr-pyrazolyl)borate.
 (16) (a) Vaz, A. D. N.; Roberts, E. S.; Coon, M. J. J. Am. Chem. Soc. 1991, 113, 5887–5889. (b) Selke, M.; Sisemore, M. F.; Valentine, J. S. J. Am. Chem. Soc. 1996, 118, 2008-2012. (c) Wertz, D. L.; Sisemore, M. F.; Selke, M.; Driscoll, J.; Valentine, J. S. J. Am. Chem. Soc. 1998, 120, 5331-5332. (d) Goto, Y.; Wada, S.; Morishima, I.; Wantanabe, Y. J. Inorg. Biochem. **1998**, 69, 241–247.

JA802775E