

PII: S0040-4020(97)00175-0

# **Physical Properties of Atropisomeric 5-Deazaflavin Derivatives**

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**Abstract:** Optically active 5-deazaflavin derivatives with an axial chirality at the N(3) position have been synthesized. Kinetics for thermal enantiomerization and X-ray crystallographic analyses of these compounds have been carried out. In addition, all absolute configurations of their enantiomers have been determined on the basis of circular dichroism spectra. © 1997 Elsevier Science Ltd.

# **INTRODUCTION**

Flavoenzymes are enzymes which requires flavin adenine dinucleotide (FAD) or flavin mononucleotide (FMN) as a coenzyme and are responsible for oxidative metabolism of various biological substances. At the active site of flavoenzyme, the flavin coenzyme is located in a chiral environment binding to apoproteins through covalent or hydrogen bonding.<sup>2-4</sup>

On the basis of the results obtained from biological systems, several optically active compounds as models for a flavoenzyme have been synthesized and their stereochemical reactivities with various substrates,  $5^{-16}$  especially NAD(P)H analogs, have been studied widely. However, since most of absolute configurations of these model compounds have not been confirmed so far, absolute stereochemistry associated with the reaction of a model and a substrate has not been discussed in detail.

In the course of our studies on atropisomeric flavoenzyme models,  $^{17-21}$  thermal enantiomerization of 3-(2-substituted phenyl)-10-(4-*tert*-butylphenyl)pyrimido[4,5-*b*]quinoline-2,4(3*H*,10*H*)-dione (**1a** and **1f**) have been carried out.<sup>19</sup> The result indicates that the difference between a hydroxymethyl group and a methyl group has little influence on the energy barrier for rotation around the N(3)–C(aryl) bond because the benzylic carbon at the C(2') position is a primary one in both compounds. This assumption was supported by their X-ray crystal structures. Thus, we carried out thermal enantiomerizations and X-ray crystallographic analyses of some of these compounds to investigate the influence of substituent on the carbon at the benzylic position of the aryl group at the N(3) position.



Furthermore, we first determined the absolute configuration of 1h by X-ray crystallographic analysis<sup>18</sup> and those of 1a and 1f by chemical reactions.<sup>19</sup> Then, we determined absolute configurations of other atropisomeric 5-deazaflavin derivatives (1b-e and 1g) on the basis of the Cotton effect in CD spectra of 1a and 1f.

# **RESULTS AND DISCUSSION**

#### Syntheses and Optical Resolutions of 5-Deazaflavin Derivatives

The syntheses of 1b-d were carried out according to the procedure shown in Scheme 1. The syntheses of 1e and 1g were accomplished as reported in a previous paper.<sup>19</sup> The optical resolutions of  $1b-e^{23-26}$  and  $1g^{27}$  were carried out by HPLC on a chiral stationary phase (CHIRALCEL OD<sup>®</sup>). The specific rotations of the enantiomers of these compounds are listed in Table 1.

#### Thermal Enantiomerizations

It has been reported that the difference in activation parameter for thermal enantiomerization is small between 1a and 1f; the hydroxy group in 1f has little influence on the energy barrier for the rotation around the N(3)-C(aryl) bond.<sup>19</sup> The observation suggests that the hydroxy group in 1f is set away from the flavin skeleton so that the steric repulsion between the hydroxymethyl group and the flavin skeleton is minimized at least in a solution (*vide supra*).

Here, the thermal enantiomerization of 1b-e was studied kinetically in order to investigate the effect of substituent on the carbon at the benzylic position of the aryl group at the N(3) position. The enantiomerizations of 1b, 1c, and 1e were carried out in DMF, whereas that of 1d was conducted in N,N-dibutylformamide due to



Table 1. Specific Rotation of the Enantiomers of 1b-e and 1g

		$\left[\alpha\right]_{D}^{a}$		
compound <sup>b</sup>	<i>T</i> , °C	1st fraction	2nd fraction	
1b	26	+17.9°	~17.8°	
1 c	24	-24.9°	+23.7°	
1d	24	- 2.4°	+ 2.5°	
1e	25	-55.8°	+55.0°	
1 g	24	+ 7.8°	- 8.1°	

<sup>a</sup> c 0.500. <sup>b</sup> All enantiomers were obtained in >99% ee.

its high energy barrier. The rate constants are listed in Table 2. The Arrhenius and Eyring plots have excellent linear relationships (r > 0.9997). The activation parameters obtained from the plots are listed in Table 3.

The free energy of activation for thermal enantiomerization decreases in the order  $R = Bu' \approx CF_3 > Pr' > Et$ (> Me  $\approx CH_2OH$ )<sup>19</sup> due to steric effect. Although the free energy of activation of 1 c is similar to that of 1 e, the entropy of activation is much smaller for the former than the latter.<sup>28</sup> However, it is apparent from Table 3 that the entropy term does not contribute meaningfully to the energy barrier.

# X-ray Crystallographic Analyses

The X-ray crystallographic analyses of 1b and 1c have been accomplished successfully by the use of crystals which were obtained by recrystallization from ethanol. Unfortunately, the crystals of 1d, 1e, and 1g suitable for X-ray analyses were not obtained. The ORTEP drawings of 1b and 1c are illustrated in Figure 1 and the crystal data are summarized in Table 4.



		$k_{rot.}$ <b>1b</b> : R = CH <sub>2</sub> CH <sub>3</sub> <b>1c</b> : R = CH(CH <sub>3</sub> ) <sub>2</sub> <b>1d</b> : R = C(CH <sub>3</sub> ) <sub>3</sub> <b>1e</b> : R = CF <sub>3</sub>		N Bu'
	$k_{\rm rot} \times 10^6,  {\rm s}^{-1}$			
<i>T</i> , °C	1 b	1c	1d	1e
50 60 70 80 90 100 110 120	$\begin{array}{c} 2.62 \pm 0.02 \\ 10.0 \pm 0.1 \\ 32.1 \pm 0.2 \\ 102 \pm 2 \\ 275 \pm 4 \end{array}$	$5.34 \pm 0.04 \\ 16.3 \pm 0.1 \\ 46.5 \pm 0.5 \\ 117 \pm 2 \\ 316 \pm 6$		$\begin{array}{c} 2.03 \pm 0.03 \\ 6.05 \pm 0.09 \\ 19.5 \pm 0.4 \\ 58.2 \pm 1.4 \\ 155 \pm 2 \end{array}$
160 170 180 190 200			$\begin{array}{r} 4.77 \pm 0.06 \\ 10.8 \pm 0.1 \\ 23.9 \pm 0.4 \\ 51.6 \pm 0.7 \\ 116 \pm 1 \end{array}$	

Table 3. Activation Parameters for Enantiomerization of 1b-e

activation parameter	1b	1c	1d	1 e
$E_a$ , kcal/mol $\Delta G^{\ddagger}$ , kcal/mol $\Delta H^{\ddagger}$ , kcal/mol $\Delta S^{\ddagger}$ , kcal/mol•deg	$\begin{array}{r} 27.1 \pm 0.3 \\ 27.1 \pm 0.04 \\ 26.4 \pm 0.3 \\ -2.31 \pm 0.94 \end{array}$	$27.9 \pm 0.3 \\ 29.0 \pm 0.05 \\ 27.2 \pm 0.3 \\ -5.90 \pm 0.67$	$\begin{array}{r} 32.3 \pm 0.5 \\ 34.7 \pm 0.15 \\ 31.4 \pm 0.5 \\ -11.05 \pm 0.98 \end{array}$	$\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$

<sup>a</sup> At 25 °C.

A crystalline 1:1 inclusion complex composed of 1 c and an ethanol molecule through hydrogen bonding to the carbonyl oxygen at the C(2) position was obtained (Figure 1b). As expected, the benzylic protons of the aryl group at the N(3) position of 1b and 1c face toward the flavin skeleton in order to minimize the steric repulsion. The same is observed in the X-ray crystallographic structure of 1 f.<sup>19</sup> Since no particular effect can be predicted in a solution to change the conformation of these compounds observed in a solid state drastically, it will be safe to conclude that these structures are also the most stable ones in solutions exerting the observed order of activation parameters in Table 3.

In addition, the X-ray crystallographic structures elucidated herein offers a clear interpretation for the results from (net) hydride-transfer between 1-benzyl-1,4-dihydronicotinamide (BNAH) and the analogs of 1b, 1c, and 1d, which have a *p*-tolyl group at the N(10) position.<sup>13,14</sup> The reaction with the analogs of 1b and 1c takes place with a similar stereoselectivity (*syn/anti* = 24/76 for the analog of 1b and 20/80 for the analog of 1 c), whereas the reaction with the analog of 1 d takes place stereospecifically in its *anti* face (*syn/anti* = 1/99).<sup>14</sup> The results reveal that the stereoselectivity in (net) hydride-transfer reactions depends significantly on the effectiveness of steric blocking for the pyrimidine ring of a flavin molecule from the attack.

# Determinations of the Absolute Configurations

We have determined the absolute configuration of 1h by X-ray crystallographic analysis<sup>18</sup> and those of 1a and 1f by chemical reactions.<sup>19</sup> and it will be reasonable to elucidate the absolute configurations of the other models. 1b-e, by measuring their circular dichroism (CD) spectra and comparing them with those of 1a and 1f.

The CD spectra of the enantiomers of 1a and 1f reveals that the (S)-enantiomer with a 2-substituted phenyl group at the N(3) position has a positive Cotton effect at around 400 nm (Figures 2a and 2f). A similar Cotton effect is also observed in the CD spectrum of (S)-1g derived from (S)-1f (Figure 2g).

The data of CD spectra of the enantiomers of 1b-e and 1g are shown in Table 5. The CD spectra shown in Figure 2 demonstrate that they have reliable Cotton effects to estimate the absolute configurations. The absolute configurations of the compounds that have been elucidated are listed in Table 6.

dFl	1b	<b>1 с•</b> C₂H₅OH
formula	$C_{29}H_{27}N_{3}O_{2}$	C <sub>30</sub> H <sub>29</sub> N <sub>3</sub> O <sub>2</sub> •C <sub>2</sub> H <sub>5</sub> OH
formula weight	449.55	509.65
crystal color, habit	yellow, prismatic	yellow, prismatic
crystal dimensions, mm	$0.50 \times 0.30 \times 0.20$	$0.50 \times 0.25 \times 0.20$
space group	$P2_1/n$ (#14)	$P2_1/n$ (#14)
a, Å	23.623(2)	15.291(2)
<i>b</i> , Å	12.429(3)	22.613(4)
<i>c</i> , Å	8.123(3)	8.257(3)
$\beta$ , deg	90.25(2)	101.23(2)
$V, Å^3$	2384(1)	2800(1)
Z value	4	4
$2\theta_{\rm max}$ , deg	120.1	120.1
no. total reflections	4032	4629
no. unique reflections	3738	4296
no. observations $(I > 3.00\sigma(I))$	2306	3015
no. variables	308	484
R	0.054	0.047
R <sub>w</sub>	0.076	0.063
S	1.93	1.98

Table 4. Summary of Crystallographic Data



Figure 1. ORTEP drawings of (a) 1b and (b) 1c-ethanol with displacement ellipsoids at 50% probability level. Only the (S)-enantiomer is illustrated in both drawings.





Table 5. Data of Circular Dichroism Spectra of 1a-g

compound	configuration		$\lambda_{\max}, \operatorname{nm}(\Delta \varepsilon_{\max})$	
	S	256 (+0.799)	272 (-1.243)	340 (+0.771)
	R	256 (-0.799)	271 (+1.159)	340 (-0.721)
1b	S	256 (+0.234)	273 (-2.210)	292 (+0.270)
		318 (-0.417)	415 (+0.551)	
	R	254 (-0.343)	273 (+2.210)	291 (-0.460)
		318 (+0.417)	400 (-0.552)	
1 c	S	253 (+0.551)	272 (~2.567)	293 (+0.226)
		318 (–0.315)	401 (+0.976)	
	R	255 (-0.399)	273 (+2.900)	292 (-0.200)
		319 (+0.563)	401 (–0.976)	
1d	S	266 (+0.254)	284 (-0.073)	297 (+0.056)
		324 (0.084)	399 (+0.042)	× ,
	R	265 (-0.239)	283 (+0.061)	298 (-0.052)
		324 (+0.084)	398 ( <del>~</del> 0.030)	
1e	S	260 (-1.845)	400 (+1.139)	
	R	261 (+1.880)	400 (–1.268)	
1f	S	272 (-0.640)	290 (+0.079)	317 (-0.519)
		407 (+0.679)		
	R	272 (+0.640)	290 (-0.039)	317 (+0.555)
		407 (–0.679)		
1 g	S	269 (–6.586)	424 (+1.400)	
8	R	269 (+6.671)	424 (–1.400)	





Figure 2. CD spectra of (a) 1a, (b) 1b, (c) 1c, (d) 1d, (e) 1e, (f) 1f, and (g) 1g in acetonitrile. The solid and broken lines represent the spectra of (S)- and (R)-enantiomers, respectively.

compound	(+)-isomer	(–)-isomer	
<b>1a</b> <sup>19</sup>	R	S	
1 b	R	S	
1 c	R	S	
1d	R	S	
1 e	S	R	
<b>1 f</b> <sup>19</sup>	S	R	
1 g	S	R	

**Table 6.** Absolute Configurations of the Enantiomers of **1a**-g

### **EXPERIMENTAL SECTION**

# Instruments

Melting points (mp) were obtained using a Yanagimoto micro melting point apparatus and were uncorrected. Infrared (IR) spectra were recorded on a JASCO FT/IR-5300 spectrometer. <sup>1</sup>H NMR spectra were recorded on a Varian VXR 200 FT-NMR spectrometer with tetramethylsilane as an internal reference and all shifts are indicated in ppm. Elemental analyses were performed using a Yanaco MT-3 Elemental Analyzer. Optical rotations were measured on a JASCO DIP-181 digital polarimeter. Circular dichroism (CD) spectra were recorded on a JASCO J-720W spectropolarimeter.

## Materials

Organic reagents and solvents were purchased from Wako Pure Chemical Industries, Ltd., Nacalai Tesque, Inc., Tokyo Kasei Kogyo Co. Ltd. and Aldrich Chemical Co., Inc. Dry methanol was obtained by reflux and distillation on sodium. The syntheses of 1a and 1e-g were carried out as reported previously.<sup>19</sup>

# Preparation of 1-(2-Alkylphenyl)urea (2b-d)

2-Alkylaniline (200 mmol) was dissolved into acetic acid/water (2:3, 200 mL) and stirred at room temperature. To the solution, a suspension of sodium cyanate (400 mmol) in water (200 mL) was added slowly. A white precipitate of the product appeared quickly. The suspension was stirred for 1 h and poured into ice-water. The precipitate was collected by filtration, washed with water, and air dried. Recrystallization from ethyl acetate gave 2.

*1-(2-Ethylphenyl)urea* (**2b**): white needles; yield 82%; mp 189–190 °C; IR (KBr) 3443, 3312, 2967, 1649, 1605, 1547, 1354, 742 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.16 (t, J = 7.4 Hz, 3H), 2.57 (q, J = 7.4 Hz, 2H), 5.99 (brs, 2H), 6.92 (t, J = 7.4 Hz, 1H), 7.03–7.12 (m, 2H), 7.68 (brs, 1H), 7.75 (d, J = 7.8 Hz, 1H). Anal. Calcd for C<sub>9</sub>H<sub>12</sub>N<sub>2</sub>O: C, 65.83; H,7.37; N, 17.06%. Found: C, 65.67; H, 7.30; N, 17.05%.

*1-(2-Isopropylphenyl)urea* (**2**c): white plates; yield 60%; mp 166–167 °C; IR (KBr) 3434, 3326, 2963, 1657, 1611, 1522, 1449, 1354, 754 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.17 (d, J = 6.8 Hz, 6H), 3.13 (m, J = 6.8 Hz, 1H), 5.97 (brs, 2H), 6.96–7.12 (m, 2H), 7.21 (d, J = 7.4 Hz, 1H), 7.62 (d, J = 7.8 Hz, 1H), 7.73 (brs, 1H). Anal. Calcd for C<sub>10</sub>H<sub>14</sub>N<sub>2</sub>O: C, 67.39; H, 7.92; N, 15.72%. Found: C, 67.45; H, 7.98; N, 15.67%.

*I*-(2-tert-Butylphenyl)urea (2d): white needles; yield 89%; mp 177–178 °C; IR (KBr) 3515, 3424, 3312, 2965, 1653, 1622, 1591, 1534, 1445, 1364, 1339, 770, 758 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.35 (s, 9H), 5.94 (brs, 2H), 7.01–7.17 (m, 2H), 7.21–7.33 (m, 3H). Anal. Calcd for C<sub>11</sub>H<sub>16</sub>N<sub>2</sub>O: C, 68.72; H, 8.39; N, 14.57%. Found: C, 68.56; H, 8.48; N, 14.39%.

#### Preparation of 1-(2-Alkylphenyl)barbituric Acid (3b-d)

Sodium (90 mmol) was dissolved in dry methanol (75 mL) at 0 °C. To the solution, 2 (75 mmol) and

diethyl malonate (90 mmol) were added and the mixture was refluxed with stirring for 12 h under argon atmosphere. After being cooled, the solution was poured into ice-water and made strongly acidic with 2 M hydrochloric acid. The precipitate was collected by filtration, washed with water, and air dried. Recrystallization from methanol gave 3.

*1-(2-Ethylphenyl)barbituric Acid* (**3b**): white needles; yield 70%; mp 239–240 °C; IR (KBr) 3214, 3096, 2973, 2897, 1723, 1676, 1493, 1473, 799, 772, 721 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.07 (t, J = 7.4 Hz, 3H), 2.43 (q, J = 7.4 Hz, 2H), 3.79 (dd, J = 21.0, 49.6 Hz, 2H), 7.14–7.40 (m, 4H), 11.53 (brs, 1H). Anal. Calcd for C<sub>12</sub>H<sub>12</sub>N<sub>2</sub>O<sub>3</sub>: C, 62.06; H, 5.21; N, 12.06%. Found: C, 62,13; H, 5.13; N, 12.10%.

*1-(2-Isopropylphenyl)barbituric Acid* (**3**c): white needles; yield 83%; mp 213–214 °C; IR (KBr) 3231, 3100, 2976, 1676, 1493, 1335, 806, 768 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>)  $\delta$  1.08 (dd, *J* = 3.2, 6.4 Hz, 6H), 2.90 (m, *J* = 6.4 Hz, 1H), 3.79 (dd, *J* = 20.8, 46.0 Hz, 2H), 7.14 (d, *J* = 7.6 Hz, 1H), 7.23 (dt, *J* = 2.2, 7.0 Hz, 1H), 7.33–7.44 (m, 2H), 11.50 (brs, 1H). Anal. Calcd for C<sub>13</sub>H<sub>14</sub>N<sub>2</sub>O<sub>3</sub>: C, 63.40; H, 5.73; N, 11.38%. Found: C, 63.32; H, 5.62; N, 11.34%.

*I*-(2-tert-Butylphenyl)barbituric Acid (**3d**): white needles; yield 82%; mp 249–250 °C; IR (KBr) 3212, 2961, 1684, 1493, 1441. 1414, 1385, 1358, 775 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.23 (s, 9H), 3.81 (dd, J = 21.0, 72.2 Hz, 2H), 7.13 (d, J = 7.4 Hz, 1H), 7.25 (dt, J = 1.8, 7.4 Hz, 1H), 7.35 (dt, J = 1.8, 7.4 Hz, 1H), 7.54 (dd, J = 1.8, 7.8 Hz, 1H), 11.59 (brs, 1H). Anal. Calcd for C<sub>14</sub>H<sub>16</sub>N<sub>2</sub>O<sub>3</sub>: C, 64.60; H, 6.20; N, 10.76%. Found: C, 64.59; H, 6.08; N, 10.78%.

#### Preparation of 3-(2-Alkylphenyl)-6-chlorouracil (4b-d)

To a mixture of **3** (50 mmol) in phosphorus oxychloride (500 mmol), water (125 mmol) was added portionwise. The resulting mixture was heated with stirring at 60 °C for 24 h. Phosphorus oxychloride was removed under reduced pressure, and the residue was poured into ice-water. The mixture was neutralized with 2 M aqueous sodium hydroxide, and the precipitate was collected by filtration, washed with water, and air dried. Recrystallization from ethyl acetate gave **4**.

6-Chloro-3-(2-ethylphenyl)uracil (**4b**): white powder; yield 65%; mp 178–179 °C; IR (KBr) 3033, 2934, 2824, 1725, 1661, 1493, 1420, 1337, 810, 762 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.05 (t, J = 7.6 Hz, 3H), 2.34 (q, J = 7.6 Hz, 2H), 6.02 (s, 1H), 7.14 (d, J = 7.6 Hz, 1H), 7.20–7.44 (m, 3H), 12.56 (brs, 1H). Anal. Calcd for C<sub>12</sub>H<sub>11</sub>ClN<sub>2</sub>O<sub>2</sub>: C, 57.50; H, 4.42; N, 11.18%. Found: C, 57.64; H, 4.25; N, 10.97%.

6-Chloro-3-(2-isopropylphenyl)uracil (4c): white powder; yield 56%; mp 249–250 °C; IR (KBr) 3094, 2967, 2811, 1723, 1651, 1491, 1451, 1408, 860, 766 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.09 (dd, J = 1.8, 6.6 Hz, 6H), 2.67 (m, J = 6.6 Hz, 1H), 6.02 (s, 1H), 7.13 (d, J = 7.2 Hz, 1H), 7.25 (dt, J = 2.0, 7.4 Hz, 1H), 7.35–7.45 (m, 2H), 12.56 (brs, 1H). Anal. Calcd for C<sub>13</sub>H<sub>13</sub>ClN<sub>2</sub>O<sub>2</sub>: C, 58.99; H, 4.95; N, 10.58%. Found: C, 58.79; H, 4.92; N, 10.61%.

3-(2-tert-Butylphenyl)-6-chlorouracil (4d): white powder; yield 67%; mp 199–200 °C; IR (KBr) 3077, 2913, 1726, 1659, 1493, 1416, 810, 766 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.22 (s, 9H), 6.03 (s, 1H), 7.05 (dd, J = 1.6, 7.6 Hz, 1H), 7.26 (dt, J = 1.6, 7.4 Hz, 1H), 7.35 (dt, J = 1.6, 8.0 Hz, 1H), 7.57 (dd, J = 1.6, 8.0 Hz, 1H), 12.56 (brs, 1H). Anal. Calcd for C<sub>14</sub>H<sub>15</sub>ClN<sub>2</sub>O<sub>2</sub>: C, 60.33; H, 5.42; N, 10.05%. Found: C, 60.28; H, 5.46; N, 10.11%.

# Preparation of 3-(2-Alkylphenyl)-6-(4-tert-butylanilino)uracil (5b-d)

A solution of 4 (25 mmol) and 4-*tert*-butylaniline (55 mmol) in butanol (50 mL) was refluxed with stirring for 6 h under argon atmosphere. After the mixture was cooled, the precipitate was collected by filtration and washed with methanol. Recrystallization from hot methanol gave 5.

6-(4-tert-Butylanilino)-3-(2-ethylphenyl)uracil (**5**b): white needles; yield 40%; mp 296–297 °C; IR (KBr) 3335, 2965, 1725, 1555, 1416, 1292, 799 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.08 (t, J = 7.6 Hz, 3H), 1.29 (s, 9H), 2.38 (q, J = 7.6 Hz, 2H), 4.85 (s, 1H), 7.07 (d, J = 7.2 Hz, 1H), 7.20 (d, J = 8.4 Hz, 2H), 7.25–7.34 (m, 3H), 7.43 (d, J = 8.4 Hz, 2H), 8.29 (brs, 1H), 10.63 (brs, 1H). Anal. Calcd for C<sub>22</sub>H<sub>25</sub>N<sub>3</sub>O<sub>2</sub>: C, 72.70; H, 6.93; N, 11.56%. Found: C, 72.64; H, 6.94; N, 11.51%.

6-(4-tert-Butylanilino)-3-(2-isopropylphenyl)uracil (5c): white needles; yield 65%; mp 296–297 °C; IR (KBr) 3333, 2959, 1728, 1647, 1595, 1557, 1420, 841, 801, 756 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ ) δ 1.10 (dd, J = 5.0, 6.8 Hz, 6H), 1.29 (s, 9H), 2.71 (m, J = 6.8 Hz, 1H), 4.84 (s, 1H), 7.05 (d, J = 7.8 Hz, 1H), 7.18–7.46 (m, 7H), 8.33 (brs, 1H), 10.62 (brs, 1H). Anal. Calcd for C<sub>23</sub>H<sub>27</sub>N<sub>3</sub>O<sub>2</sub>: C, 73.18; H, 7.21; N, 11.13%. Found: C, 73.17; H, 7.16; N, 11.05%.

6-(4-tert-Butylanilino)-3-(2-tert-butylphenyl)uracil (5d): white needles; yield 55%; mp >300 °C; IR (KBr) 3316, 2965, 1728, 1599, 1561, 1422, 1298, 758 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.25 (s, 9H), 1.29 (s, 9H), 4.86 (s, 1H), 6.92 (dd, J = 1.8, 7.4 Hz, 1H), 7.16–7.27 (m, 3H), 7.32 (dt, J = 1.8, 7.4 Hz, 1H), 7.43 (d, J = 8.6 Hz, 2H), 7.56 (dd, J = 1.6, 7.8 Hz, 1H), 8.32 (brs, 1H), 10.63 (brs, 1H). Anal. Calcd for C<sub>24</sub>H<sub>29</sub>N<sub>3</sub>O<sub>2</sub>: C, 73.63; H, 7.47; N, 10.73%. Found: C, 73.49; H, 7.50; N, 10.61%.

# Preparation of 3-(2-Alkylphenyl)-10-(4-tert-butylphenyl)pyrimido[4,5-b]quinoline-2,4(3H,10H)-dione (1b-d)

2-Fluorobenzaldehyde (12 mmol) and 5 (10 mmol) were dissolved in DMF (8 mL), and the solution was heated with stirring at 120 °C for 7 h. After the mixture was cooled, DMF was removed to dryness under reduced pressure. The residue was recrystallized from hexane/chloroform (1b and 1c) or chloroform/methanol (1d) to gave 1.

10-(4-tert-Butylphenyl)-3-(2-ethylphenyl)pyrimido[4,5-b]quinoline-2,4(3H,10H)-dione (1b): yellow powder; yield 58%; mp 279–280 °C; IR (KBr) 3449, 3049, 2965, 1709, 1659, 1620, 1566, 1535, 1491, 1453, 1412, 797, 745 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.17 (t, J = 7.6 Hz, 3H), 1.42 (s, 9H), 2.51 (q, J = 7.6 Hz, 2H), 6.89 (d, J = 8.6 Hz, 1H), 7.13 (d, J = 7.4 Hz, 1H), 7.22–7.48 (m, 6H), 7.61–7.68 (m, 3H), 7.94 (dd, J = 1.4, 7.6 Hz, 1H), 9.05 (s, 1H). Anal. Calcd for C<sub>29</sub>H<sub>27</sub>N<sub>3</sub>O<sub>2</sub>: C, 77.48; H, 6.05; N, 9.35%. Found: C, 77.24; H, 6.09; N, 9.30%.

*10-(4-tert-Butylphenyl)-3-(2-isopropylphenyl)pyrimido*[4,5-*b*]*quinoline-2,4(3H,10H)-dione* (**1**c): yellow needles; yield 46%; mp 281–282 °C; IR (KBr) 3480, 2963, 1711, 1663, 1618, 1566, 1535, 1491, 1453, 1414, 1370, 797, 758 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.19 (dd, J = 1.4, 7.0 Hz, 6H), 1.42 (s, 9H), 2.81 (m, J = 7.0 Hz, 1H), 6.92 (d, J = 8.8 Hz, 1H), 7.09 (dd, J = 1.4, 7.2 Hz, 1H), 7.22–7.33 (m, 3H), 7.39–7.47 (m, 3H), 7.60-7.68 (m, 3H), 7.93 (dd, J = 1.4, 8.0 Hz, 1H), 9.09 (s, 1H). Anal. Calcd for C<sub>30</sub>H<sub>29</sub>N<sub>3</sub>O<sub>2</sub>: C, 77.73; H, 6.31; N, 9.06%. Found: C, 77.64; H, 6.39; N, 9.02%.

3-(2-tert-Butylphenyl)-10-(4-tert-butylphenyl)pyrimido[4,5-b]quinoline-2,4(3H,10H)-dione (1d): yellow powder; yield 65%; mp >300 °C; IR (KBr) 3484, 2963, 1711, 1661, 1618, 1566, 1535, 1491, 1414, 1370, 797, 756 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.31 (s, 9H), 1.41 (s, 9H), 6.92 (d, J = 8.8 Hz, 1H), 6.99 (dd, J = 1.8, 7.4 Hz, 1H), 7.23–7.48 (m, 5H), 7.57–7.69 (m, 4H), 7.93 (dd, J = 1.4, 7.8 Hz, 1H), 9.08 (s, 1H). Anal. Calcd for C<sub>31</sub>H<sub>31</sub>N<sub>3</sub>O<sub>2</sub>•CH<sub>3</sub>OH: C, 75.24; H, 6.92; N, 8.24%. Found: C, 75.56; H, 6.69; N, 8.52%.

# Crystallographic Studies

The lattice parameters and intensity data were measured on a Rigaku AFC7R diffractometer and an 18 kW rotating anode generator with 8 kW Cu K $\alpha$  radiation. The structures were solved by the direct method, and non-hydrogen atoms were refined anisotropically. All calculations were performed using a Texsan crystallographic software package developed by Molecular Structure Corporation (1985 and 1992).

### Kinetic Measurements for Thermal Enantiomerization

Thermal enantiomerization was carried out by immersing a solution of a chiral compound in DMF or N,Ndibutylformamide (*ca.*  $1.0 \times 10^{-3}$  M) in a thermostated oil bath. At appropriate intervals, aliquots were withdrawn and subjected to HPLC analysis to measure ee. It has been confirmed by HPLC that no decomposition of the compound in the solution takes place throughout the reaction. The activation parameters were calculated according to the Arrhenius and Eyring equations.

## Determinations of the Absolute Configurations

The absolute configurations of the enantiomers of 1b-e, which were optically resolved by HPLC,<sup>21-24</sup> were determined by comparing their CD spectra with those of 1a and 1f, and that of 1g was determined by the

reaction of (S)-1f (>99% ee) with *tert*-butyldimethylsilyl chloride and imidazole in DMF according to the procedure mentioned in a previous paper.<sup>19</sup> The CD spectrum of this compound also confirmed the stereo-chemical assignment on the basis of the chemical reaction.

# ACKNOWLEDGEMENT

We are deeply indebted to Daicel Chemical Co., Ltd. for providing us a CHIRALCEL OD<sup>®</sup> column for HPLC. A part of this work was supported by a Grant-in-Aid for Scientific Research No. 07454194 from the Ministry of Education, Science, and Culture of Japan.

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- 22. Only one enantiomer is shown in the scheme for convenience.
- 23. Conditions for preparative chromatography of 1b: column, CHIRALCEL OD<sup>®</sup> (2 cm\$\u03c6 × 25 cm\$); eluent, ethanol; flow rate, 4.0 mL/min; detection, UV 254 nm; retention times, 48.2 and 69.6 min. Conditions for analytical chromatography of 1b: column, CHIRALCEL OD<sup>®</sup> (0.46 cm\$\u03c6 × 5 cm\$); eluent, ethanol; flow rate, 0.3 mL/min; detection, UV 254 nm; retention times, 9.0 and 13.2 min.
- 24. Conditions for preparative chromatography of 1 c: column, CHIRALCEL OD<sup>®</sup> (2 cm \u03c6 \u2225 cm); eluent, ethanol; flow rate, 3.0 mL/min; detection, UV 254 nm; retention times, 49.9 and 54.5 min. Conditions for analytical chromatography of 1 c: column, CHIRALCEL OD<sup>®</sup> (0.46 cm \u03c6 \u2225 5 cm + 0.46 cm \u03c6 \u2225 25 cm); eluent, 2-propanol; flow rate, 0.4 mL/min; detection, UV 254 nm; retention times, 33.8 and 42.1 min.
- 25. Conditions for preparative chromatography of 1d: column, CHIRALCEL OD<sup>®</sup> (2 cm \u03c6 x 25 cm); eluent, ethanol; flow rate, 1.5 mL/min; detection, UV 254 nm; retention times, 71.4 and 78.2 min. Conditions for analytical chromatography of 1d: column, CHIRALCEL OD<sup>®</sup> (0.46 cm \u03c6 x 5 cm + 0.46 cm \u03c6 x 25 cm); eluent, 2-propanol; flow rate, 0.4 mL/min; detection, UV 254 nm; retention times, 24.3 and 30.8 min.
- 26. Conditions for preparative chromatography of 1 e: column, CHIRALCEL OD<sup>®</sup> (2 cm \$\phi\$ × 25 cm); eluent, ethanol; flow rate, 1.5 mL/min; detection, UV 254 nm; retention times, 74.1 and 84.1 min. Conditions for analytical chromatography of 1 e: column, CHIRALCEL OD<sup>®</sup> (0.46 cm \$\phi\$ × 5 cm + 0.46 cm \$\phi\$ × 25 cm); eluent, 2-propanol; flow rate, 0.4 mL/min; detection, UV 254 nm; retention times, 27.8 and 40.6 min.
- 27. Conditions for preparative chromatography of 1 g: column, CHIRALCEL OD<sup>®</sup> (2 cm ∉ × 25 cm); eluent, ethanol; flow rate, 2.0 mL/min; detection, UV 254 nm; retention times, 47.0 and 85.6 min. Conditions for analytical chromatography of 1 g: column, CHIRALCEL OD<sup>®</sup> (0.46 cm ∉ × 5 cm); eluent, ethanol; flow rate, 0.2 mL/min; detection, UV 254 nm; retention times, 6.1 and 10.3 min.
- 28. The same is observed in the thermal enantiomerizations of 6-(2-substituted phenyl)-5-methyl-1,1-dimethylindans : Bott, G.; Field, L. D.; Sternhell, S. J. Am. Chem. Soc. 1980, 102, 5618-5626.

(Received in Japan 22 November 1996; accepted 18 February 1997)