

Synthesis of glycosyl boranes and glycosyl borinates†

Andrea Vasella,* Wolfgang Wenger and Thennati Rajamannar

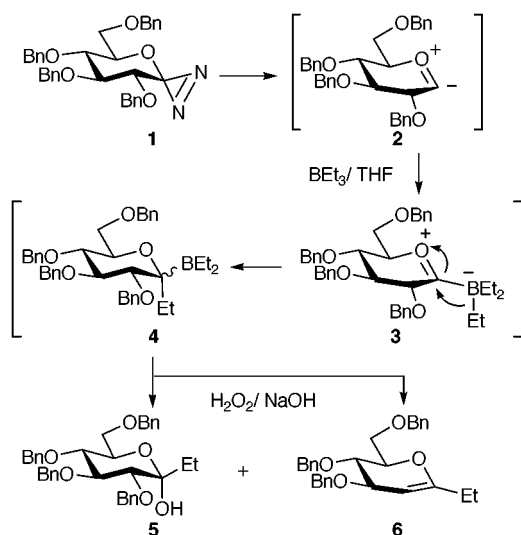
Laboratorium für Organische Chemie, ETH-Zentrum, Universitätstrasse 16, CH-8092 Zürich, Switzerland.

E-mail: vasella@sugar.org.chem.ethz.ch

Received (in Liverpool, UK) 5th August 1999, Accepted 17th September 1999

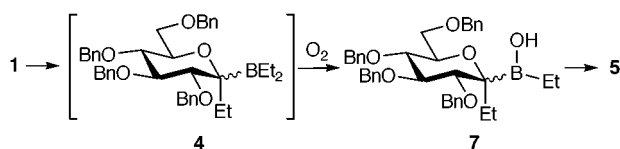
Insertion of glycosylidene carbenes into a B–C bond of BEt_3 leads to unstable glycosyl boranes, while insertion into a B–C bond of borinic esters yields stable anomeric glycosyl borinates.

Insertion of glycosylidene carbenes, generated by thermolysis or photolysis of glycosylidene diazirines, into HX bonds leads to *O*-, *C*- and *N*-glycosides,¹ glycosyl phosphines² and glycosyl stannanes.³ We speculated that reaction of a borane with the carbene **2** would lead to a zwitterion such as **3**; migration of a *B*-substituent—as proposed for the reaction of methoxycarbene with trialkylboranes⁴—should lead to the as yet unknown glycosyl boranes (Scheme 1).



Scheme 1

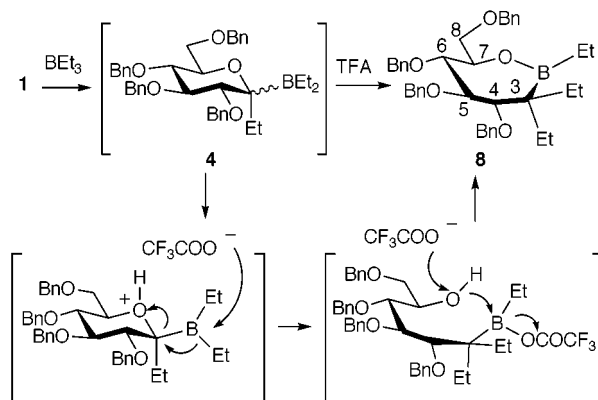
Thermolysis of the diazirine **1**⁵ in degassed THF in the presence of 1.5 equiv. of BEt_3 at 25 °C, followed by treatment with excess 30% alkaline H_2O_2 and aqueous work-up, yielded 55% of hemiacetal **5** and 13% of the *C*-ethylglucal **6**. These products suggest the intermediate formation of anomeric glycosyl boranes **4**. Treatment with alkaline H_2O_2 leads either (depending on the borane configuration?) to oxidation⁶ and (after anomerisation?) to the hemiacetal **5**, or to elimination of the C(1) boron substituent and the vicinal benzyloxy group.⁷ *trans*-Elimination is suggested by the observation (^{13}C NMR) that **6** is only formed after addition of alkaline H_2O_2 . Thermolysis of the diazirine **1** in the presence of 1.5 equiv. of BEt_3 in oxygen-containing THF yielded 25% of the borinic acid **7** after rapid chromatography (Scheme 2). The borinic acid **7**



Scheme 2

was rapidly converted into the hemiacetal **5** (isolated in *ca.* 75%) by exposure to air, storage in oxygen-containing CDCl_3 , or treatment with alkaline H_2O_2 . The ^{11}B NMR of air-stable mixtures of **7** and 2 equiv. of PPh_3 shows a broad signal at δ 56.5, typical for dialkyl borinic acids.⁸ The ^1H NMR shows a broad singlet of an exchangeable H at δ 8.16, corresponding to *B*-OH, two doublets of quintets at δ 1.8 and 1.6 (2 H), a triplet at δ 0.9 (3 H) for the *C*-ethyl and a multiplet at δ 1.0–0.9 (5 H) of the *B*-Et group. The ^{13}C NMR signal for the boron substituted anomeric carbon is missing, as expected.⁹

The intermediate formation of a glycosyl borane was further evidenced by treating the product of thermolysis of **1** and BEt_3 in degassed THF with excess TFA, followed by aqueous work-up (Scheme 3). The resulting cyclic borinic ester **8**, isolated in 45%, showed a ^{11}B signal at δ 51, typical for *O*-alkyl borinates.¹⁰

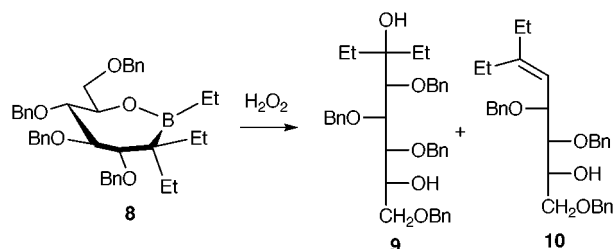


Scheme 3

The ^1H NMR spectrum of **8** shows signals for a $\text{BC}(\text{Et})_2$ moiety and a multiplet at δ 0.9–0.7 for a BEt group. *H*-C(7) is strongly shifted downfield due to the R_2BO substituent [*H*-C(5) in **6** at δ 4.02, *H*-C(7) in **8** at δ 4.77].

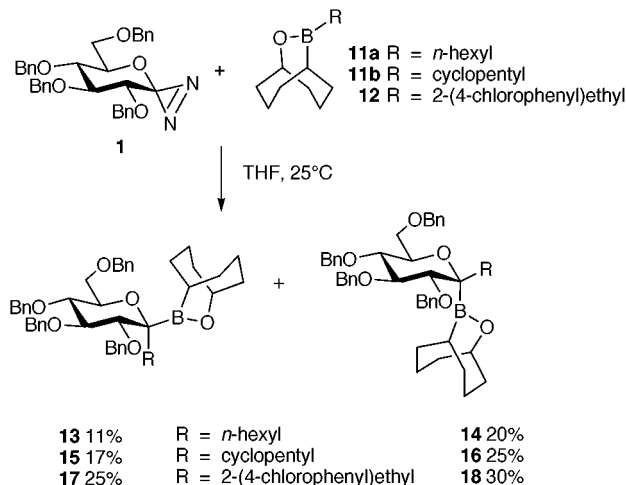
The formation of **8** is rationalised by protonation of the ring oxygen of **4**, nucleophilic attack of trifluoroacetate at boron, 1,2-migration of an ethyl substituent with concomitant ring-opening, and nucleophilic attack of *HO*-C(7) at boron. Treatment of **8** with alkaline H_2O_2 in THF leads to the diol **9** (37%) and the alkene **10** (39%) (Scheme 4).

To prepare glycosyl borinates, we exposed the diazirine **1** to the exceptionally stable borinates **11**–**12**¹¹ derived from 10-bora-9-oxabicyclo[3.3.2]decane (Scheme 5). This led to



Scheme 4

† Glycosylidene Carbenes Part 28. For Part 27, see ref. 1(b).



Scheme 5

diastereoisomeric mixtures **13/14** (31%; 35:65), **15/16** (42%; 40:60) and **17/18** (55%; 45:55) that were isolated by flash chromatography. The isomers **15/16** and **17/18** were separated by HPLC. The glycosyl borinates **13–18** were characterized by FAB-MS, ^{11}B NMR, ^1H NMR, ^{13}C NMR and IR spectroscopy. They were stable at -10°C for several weeks and not affected by air.

The configuration of **17** and **18** was deduced from NOE experiments (Fig. 1), with **17** showing NOEs between H-C(2) and H-C(5), H-C(2) and H-C(7), and H-C(1') and H-C(4), indicating an axial orientation of the anomeric alkyl group. In contradistinction, a small NOE between H-C(1') and H-C(5) and the lack of other NOEs > 1% for **18** indicate an equatorial orientation of the anomeric alkyl group.

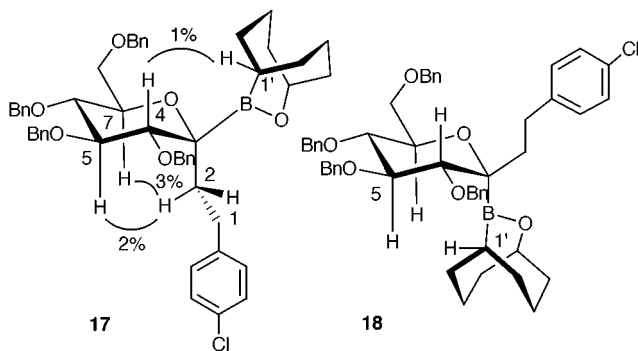


Fig. 1

We thank the Swiss National Science Foundation and F Hoffmann-La Roche, Basel, for generous financial support.

Notes and references

‡ *Synthesis of 17 and 18*: at 25°C , a solution of **12** (R,R' = H, 4-ClC₆H₄, 116 mg, 0.42 mmol) in abs. THF (3 ml) was treated portionwise with a cooled (dry ice, ca. -60°C) solution of **1** (77 mg, 0.14 mmol) in dry CH₂Cl₂ (0.8 ml) within 140 min, stirred for 2 h at 25°C until complete disappearance of **1**. Evaporation at 20°C and flash chromatography

(hexane–AcOEt–CH₂Cl₂ 18:1:1) gave **17/18** (61 mg, 55%, 45:55), which were separated by preparative HPLC (hexane–AcOEt 12:1; 9 ml min⁻¹). *Selected data for 17*: R_f (hexane–AcOEt–CH₂Cl₂ 4:1:1) 0.36; $[\alpha]_D^{25} +51.4$ (c 1.16, CH₂Cl₂); ν_{max} (CH₂Cl₂)/cm⁻¹ 3032w, 2927m, 2963m, 1604w, 1493m, 1453m, 1420m, 1364m, 1092s, 1027s; δ_{H} (300 MHz, CDCl₃) 7.62–7.11 (m, 20 arom. H), 4.87 (d, *J* 10.6, PhCH), 4.85 (d, *J* 10.9, PhCH), 4.83 (d, *J* 10.9, PhCH), 4.77 (d, *J* 10.9, PhCH), 4.71 (d, *J* 12.1, PhCH), 4.70–4.66 (m, BOCH), 4.68 (d, *J* 10.9, PhCH), 4.66 (d, *J* 10.9, PhCH), 4.64 (d, *J* 12.1, PhCH), 3.91 [t, *J* 8.7, H-C(5)], 3.74 [dd, *J* 11.2, 4.0, H-C(8)], 3.73 [dd, *J* 11.2, 1.9, H'-C(8)], 3.68 [d, *J* 9.0, irradi. at 3.91 → *d*, *J* ≈ 4, H-C(4)], 3.67–3.63 [m, H-C(7)], 3.58 [t, *J* 8.4, irradi. at 3.91 → *dd*, *J* ≈ 9, 3, H-C(6)], 2.65–2.56 [m, 2 H-C(1)], 2.37–2.27 [m, irradi. at 2.6 → *d*, *J* ≈ 10, H-C(2)], 2.22–2.18 (m, BCH), 2.01–1.91 [m, irradi. at 2.6 → *d*, *J* ≈ 10, H'-C(2)], 2.01–1.46 (m, 12 H); δ_{C} (75 MHz, CDCl₃, assignment based on $^1\text{H}/^{13}\text{C}$ COSY) 142.03 (s), 139.00 (2s), 138.69, 138.29, 131.12 (3s), 129.79–127.12 (several d), 84.42 [d, C(5)], 81.57 [d, C(4)], 79.53 [d, C(6)], 75.60, 75.13, 74.73 (3t, 3 PhCH₂), 74.19 (d, BOCH), 73.51 (t, PhCH₂), 72.24 [d, C(7)], 69.99 [t, C(8)], 32.41 (t), 30.63 (t), 30.19 [t, C(1)], 28.91 [t, C(2)], 27.54 (t), 25.63 (t), 22.89 (t), 21.91 (t), 21.30 (small br d, HCB), signal of C(3) hidden by noise; δ_{B} (160 MHz, CDCl₃) 52.02 (br s); *m/z* (FAB) 821 (< 1%, [*M* + Na]⁺), 799 (< 1, [*M* + H]⁺), 599 (38), 553 (43, [*M* – BnOBoc₈H₁₄ + H]⁺), 447 (44), 181 (100). For **18**: R_f (hexane–AcOEt–CH₂Cl₂ 10:1:1) 0.32; $[\alpha]_D^{25} +12.0$ (c 0.65, CH₂Cl₂); ν_{max} (CH₂Cl₂/cm⁻¹ 3032w, 2927m, 1492m, 1453m, 1418w, 1364m, 1093s, 1027m, 1015m; δ_{H} (300 MHz, CDCl₃) 7.38–6.99 (m, 20 arom. H), 4.98 (d, *J* 11.8, PhCH), 4.85 (d, *J* 10.9, PhCH), 4.84 (s, PhCH₂), 4.70 (d, *J* 11.8, PhCH), 4.71–4.67 (m, HCOB), 4.68 (d, *J* 12.1, PhCH), 4.62 (d, *J* 10.9, PhCH), 4.60 (d, *J* 12.1, PhCH), 3.96 [dt, *J* ≈ 9.6, 4.0, H-C(7)], 3.78–3.70 [m, H-C(5), 2 H-C(8)], 3.60 [t, *J* 9.6, irradi. at 3.96 → *d*, *J* ≈ 9, H-C(6)], 3.50 [d, *J* 9.4, H-C(4)], 2.74–2.67 [m, 2 H-C(1)], 2.25–2.21 (m, BCH), 2.05–1.95 [m, irradi. at 2.70 → *d*, *J* ≈ 12, H-C(2)], 1.99–1.26 (m, 13 H); δ_{C} (75 MHz, CDCl₃, assignment based on $^1\text{H}/^{13}\text{C}$ COSY) 141.75 (s), 139.26 (2s), 138.2, 137.8, 133.6 (3s), 129.9–127.14 (several d), 85.93 [d, C(5)], 84.90 [d, C(3)], 79.54 [d, C(6)], 76.00 [d, C(7)], 75.37, 75.09, 74.80 (3t, 3 PhCH₂); 74.06 (d, BOCH), 73.22 (t, PhCH₂), 69.93 [t, C(8)], 37.14 [t, C(2)], 31.82 (t), 30.91 (t), 29.33 [t, C(1)], 26.79 (t), 25.77 (t), 23.38 (small br d, BCH), 22.48 (t), 21.92 (t), signal of C(3) hidden by noise; δ_{B} (160 MHz, CDCl₃) 53.8 (br s); *m/z* (FAB) 821 (< 1%, [*M* + Na]⁺), 799 (< 1, [*M* + H]⁺), 553 (43, [*M* – BnOBoc₈H₁₄ + H]⁺), 461 (28), 401 (41), 325 (60), 281 (87), 181 (100).

- (a) A. Vasella, *Glycosylidene Carbenes*, in *Bioorganic Chemistry*, Vol. 3, Carbohydrates, ed. S. Hecht, OUP, New York, 1999, p. 56 and references therein; (b) M. Weber, A. Vasella, M. Textor and N. D. Spencer, *Helv. Chim. Acta*, 1998, **81**, 1359; (c) K. Briner and A. Vasella, *Helv. Chim. Acta*, 1992, **75**, 621; (d) P. Uhlmann and A. Vasella, *Helv. Chim. Acta*, 1992, **75**, 1979; (e) A. Vasella and C. A. A. Waldruff, *Helv. Chim. Acta*, 1991, **74**, 585; (f) A. Vasella, P. Dhar and C. Witzig, *Helv. Chim. Acta*, 1993, **76**, 1767; (g) T. Rajamannar and A. Vasella, unpublished results on the synthesis of *N*-glycosylsulfonamides.
- A. Vasella, G. Baudin and L. Panza, *J. Heteroatom Chem.*, 1991, **2**, 151.
- P. Uhlmann, D. Nanz, E. Bozo and A. Vasella, *Helv. Chim. Acta*, 1994, **77**, 1430.
- A. Suzuki, S. Nozawa, N. Miyaura and M. Itoh, *Tetrahedron Lett.*, 1969, 2955.
- K. Briner and A. Vasella, *Helv. Chim. Acta*, 1989, **72**, 1371.
- G. Zweifel and H. C. Brown, *Org. React.*, 1963, **13**, 1.
- D. J. Pasto and S. R. Snyder, *J. Org. Chem.*, 1966, **31**, 2777; D. S. Matteson and M. L. Peterson, *J. Org. Chem.*, 1987, **52**, 5116.
- H. Nöth and B. Wrackmeyer, *NMR: Basic Principles and Progress*, 1978, vol. 14, p. 140.
- B. Wrackmeyer, *Prog. Nucl. Magn. Reson. Spectrosc.*, 1979, **12**, 227.
- H. Nöth and B. Wrackmeyer, *NMR: Basic Principles and Progress*, 1978, vol. 14, p. 138.
- J. A. Soderquist and M. R. Najafi, *J. Org. Chem.*, 1986, **51**, 1330.

Communication 9/06400A